

Package ‘cate’

October 5, 2015

Type Package

Title High Dimensional Factor Analysis and Confounder Adjusted Testing and Estimation

Version 1.0.4

Date 2015-10-02

Description Provides several methods for factor analysis in high dimension (both $n, p \gg 1$) and methods to adjust for possible confounders in multiple hypothesis testing.

Imports MASS, esaBcv, ruv, sva, corpcor, leapp

Suggests knitr, ggplot2, gridExtra

License GPL-2

LazyData TRUE

VignetteBuilder knitr

NeedsCompilation no

Author Jingshu Wang [aut],
Qingyuan Zhao [aut, cre]

Maintainer Qingyuan Zhao <qyzhao@stanford.edu>

Repository CRAN

Date/Publication 2015-10-05 12:37:43

R topics documented:

cate-package	2
adjust.latent	2
cate	3
est.confounder.num	5
fa.em	7
fa.pc	7
factor.analysis	8
gen.sim.data	9
gender.sm	11
wrapper	11

Index	14
--------------	-----------

cate-package	<i>High dimensional factor analysis and confounder adjusted testing and estimation (CATE)</i>
--------------	---

Description

Provides several methods for factor analysis in high dimension (both $n, p \gg 1$) and methods to adjust for possible confounders in multiple hypothesis testing.

See Also

[factor.analysis](#), [cate](#)

adjust.latent	<i>Adjust for latent factors, after rotationn</i>
---------------	---

Description

Adjust for latent factors, after rotationn

Usage

```
adjust.latent(corr.margin, n, X.cov, Gamma, Sigma, method = c("rr", "nc",
  "lqs"), psi = psi.huber, nc = NULL, nc.var.correction = TRUE)
```

Arguments

corr.margin	marginal correlations, $p \times d$ matrix
n	sample size
X.cov	estimated second moment of X, $d \times d$ matrix
Gamma	estimated confounding effects, $p \times r$ matrix
Sigma	diagonal of the estimated noise covariance, $p \times 1$ vector
method	adjustment method
psi	derivative of the loss function in robust regression, choices are <code>psi.huber</code> , <code>psi.bisquare</code> and <code>psi.hampel</code>
nc	position of the negative controls
nc.var.correction	correct asymptotic variance based on our formula

Details

The function essentially runs a regression of `corr.margin ~ Gamma`. The sample size `n` is needed to have the right scale.

This function should only be called if you know what you are doing. Most of the time you want to use the main function [cate](#) to adjust for confounders.

Value

a list of objects

alpha estimated alpha, $r \times d1$ matrix

beta estimated beta, $p \times d1$ matrix

beta.cov.row estimated row covariance of beta, a length p vector

beta.cov.col estimated column covariance of beta, a $d1 \times d1$ matrix

See Also

[cate](#)

cate

The main function for confounder adjusted testing

Description

The main function for confounder adjusted testing

Usage

```
cate(formula, X.data = NULL, Y, r, fa.method = c("ml", "pc", "esa"),
     adj.method = c("rr", "nc", "lqs", "naive"), psi = psi.huber, nc = NULL,
     nc.var.correction = TRUE, calibrate = TRUE)
```

```
cate.fit(X.primary, X.nuis = NULL, Y, r, fa.method = c("ml", "pc", "esa"),
        adj.method = c("rr", "nc", "lqs", "naive"), psi = psi.huber, nc = NULL,
        nc.var.correction = TRUE, calibrate = TRUE)
```

Arguments

formula	a formula indicating the known covariates including both primary variables and nuisance variables, which are separated by . The variables before are primary variables and the variables after are nuisance variables. It's OK if there is no nuisance variables, then is not needed and formula becomes a typical formula with all the covariates considered primary. An intercept term will still be automatically added as a nuisance variable for the latter case.
X.data	the data frame used for formula
Y	outcome, $n \times p$ matrix
r	number of latent factors, can be estimated using the function <code>est.confounder.num</code>
fa.method	factor analysis method
adj.method	adjustment method
psi	derivative of the loss function in robust regression

<code>nc</code>	position of the negative controls, if $d_0 > 1$, this should be a matrix with 2 columns
<code>nc.var.correction</code>	correct asymptotic variance based on our formula
<code>calibrate</code>	if TRUE, use the Median and the Mean Absolute Deviation(MAD) to calibrate the test statistics
<code>X.primary</code>	primary variables, $n \times d_0$ matrix or data frame
<code>X.nuis</code>	nuisance covarites, $n \times d_1$ matrix

Details

Ideally `nc` can either be a vector of numbers between 1 and p , if $d_0 = 1$ or the negative controls are the same for every treatment variable, or a 2-column matrix specifying which positions of β are known to be zero. But this is yet implemented.

Value

a list of objects

alpha estimated alpha, $r \times d_1$ matrix

alpha.p.value asymptotic p-value for the global chi squared test of alpha, a vector of length d_1

beta estimated beta, $p \times d_1$ matrix

beta.cov.row estimated row covariance of beta, a length p vector

beta.cov.col estimated column covariance of beta, a $d_1 \times d_1$ matrix

beta.t asymptotic z statistics for beta

beta.p.value asymptotic p-values for beta, based on `beta.t`

Y.tilde the transformed outcome matrix, an $n \times p$ matrix

Gamma estimated factor loadings, $p \times r$ matrix

Z estimated latent factors

Sigma estimated noise variance matrix, a length p vector

Functions

- `cate.fit`: Basic computing function called by `cate`

References

J. Wang, Q. Zhao, T. Hastie, and A. B. Owen (2015). Confounder adjustment in multiple testing. *arXiv:1508.04178*.

See Also

[wrapper](#) for wrapper functions of some existing methods.

Examples

```
## simulate a dataset with 100 observations, 1000 variables and 5 confounders
data <- gen.sim.data(n = 100, p = 1000, r = 5)
X.data <- data.frame(X1 = data$X1)

## linear regression without any adjustment
output.naive <- cate(~ X1, X.data, Y = data$Y, r = 0, adj.method = "naive")
## confounder adjusted linear regression
output <- cate(~ X1, X.data, Y = data$Y, r = 5)
## plot the histograms of unadjusted and adjusted regression statistics
par(mfrow = c(1, 2))
hist(output.naive$beta.t)
hist(output$beta.t)
## simulate a dataset with 100 observations, 1000 variables and 5 confounders
data <- gen.sim.data(n = 100, p = 1000, r = 5)
## linear regression without any adjustment
output.naive <- cate.fit(X.primary = data$X1, X.nuis = NULL, Y = data$Y,
                        r = 0, adj.method = "naive")
## confounder adjusted linear regression
output <- cate.fit(X.primary = data$X1, X.nuis = NULL, Y = data$Y, r = 5)
## plot the histograms of unadjusted and adjusted regression statistics
par(mfrow = c(1, 2))
hist(output.naive$beta.t)
hist(output$beta.t)
```

est.confounder.num *Estimate the number of confounders*

Description

Estimate the number of confounders

Usage

```
est.confounder.num(formula, X.data = NULL, Y, method = c("bcv", "ed"),
                  rmax = 20, nRepeat = 20, bcv.plot = TRUE, log = "")
```

```
est.factor.num(Y, method = c("bcv", "ed"), rmax = 20, nRepeat = 12,
              bcv.plot = TRUE, log = "")
```

Arguments

formula a formula indicating the known covariates including both primary variables and nuisance variables, which are separated by |. The variables before | are primary variables and the variables after | are nuisance variables. It's OK if there is no nuisance variables, then | is not needed and formula becomes a typical formula with all the covariates considered primary. An intercept term will still be automatically added as a nuisance variable for the latter case.

X.data	the data frame used for formula
Y	outcome, n*p matrix
method	method to estimate the number of factors. There are currently two choices, "ed" is the eigenvalue difference method proposed by Onatski (2010) and "bcv" is the bi-cross-validation method proposed by Owen and Wang (2015). "bcv" tends to estimate more weak factors and takes longer time
rmax	the maximum number of factors to consider. If the estimated number of factors is rmax, then users are encouraged to increase rmax and run again. Default is 20.
nRepeat	the number of repeats of bi-cross-validation. A larger nRepeat will result in a more accurate estimate of the bcv error, but will need longer time to run.
bcv.plot	whether to plot the relative bcv error versus the number of estimated ranks. The relative bcv error is the entrywise mean square error divided by the average of the estimated noise variance.
log	if log = "y", then the y-axis of the bcv plot is in log scale.

Value

if method is "ed", then return the estimated number of confounders/factors. If method is "bcv", then return the a list of objects

r estimated number of confounders/factors

errors the relative bcv errors of length 1 + rmax

Functions

- `est.factor.num`: Estimate the number of factors

References

A. B. Owen and J. Wang (2015), Bi-cross-validation for factor analysis. *arXiv:1503.03515*.

A. Onatski (2010), Determining the number of factors from empirical distribution of eigenvalues. *The Review of Economics and Statistics* 92(4).

Examples

```
## example for est.confounder.num
data <- gen.sim.data(n = 50, p = 100, r = 5)
X.data <- data.frame(X1 = data$X1)
est.confounder.num(~ X1, X.data, data$Y, method = "ed")
est.confounder.num(~ X1, X.data, data$Y, method = "bcv")
## example for est.factor.num
n <- 50
p <- 100
r <- 5
Z <- matrix(rnorm(n * r), n, r)
Gamma <- matrix(rnorm(p * r), p, r)
Y <- Z %*% t(Gamma) + rnorm(n * p)
```

```
est.factor.num(Y, method = "ed")  
est.factor.num(Y, method = "bcv")
```

fa.em

Factor analysis via EM algorithm to maximize likelihood

Description

Factor analysis via EM algorithm to maximize likelihood

Usage

```
fa.em(Y, r, tol = 1e-06, maxiter = 1000)
```

Arguments

Y	data matrix, a n*p matrix
r	number of factors
tol	a tolerance scale of change of log-likelihood for convergence in the EM iterations
maxiter	maximum iterations

References

Bai, J. and Li, K. (2012). Statistical analysis of factor models of high dimension. *The Annals of Statistics* 40, 436-465. See <http://www.mathworks.com/matlabcentral/fileexchange/28906-factor-analysis/content/fa.m> for a MATLAB implementation of the EM algorithm.

See Also

[factor.analysis](#) for the main function.

fa.pc

Factor analysis via principal components

Description

Factor analysis via principal components

Usage

```
fa.pc(Y, r)
```

Arguments

Y	data matrix, a n*p matrix
r	number of factors

See Also

[factor.analysis](#) for the main function.

factor.analysis	<i>Factor analysis</i>
-----------------	------------------------

Description

The main function for factor analysis with potentially high dimensional variables. Here we implement some recent algorithms that is optimized for the high dimensional problem where the number of samples n is less than the number of variables p .

Usage

```
factor.analysis(Y, r, method = c("ml", "pc", "esa"))
```

Arguments

Y	data matrix, a n*p matrix
r	number of factors
method	algorithm to be used

Details

The three methods are quasi-maximum likelihood (ml), principal component analysis (pc), and factor analysis using an early stopping criterion (esa).

The ml is iteratively solved the Expectation-Maximization algorithm using the PCA solution as the initial value. See Bai and Li (2012) and for more details. For the esa method, see Owen and Wang (2015) for more details.

Value

a list of objects

Gamma estimated factor loadings

Z estimated latent factors

Sigma estimated noise variance matrix

References

Bai, J. and Li, K. (2012). Statistical analysis of factor models of high dimension. *The Annals of Statistics* 40, 436-465. Owen, A. B. and Wang, J. (2015). Bi-cross-validation for factor analysis. *arXiv:1503.03515*.

See Also

[fa.pc](#), [fa.em](#), [ESA](#)

Examples

```
## a factor model
n <- 100
p <- 1000
r <- 5
Z <- matrix(rnorm(n * r), n, r)
Gamma <- matrix(rnorm(p * r), p, r)
Y <- Z %*% t(Gamma) + rnorm(n * p)

## to check the results, verify the true factors are in the linear span of the estimated factors.
pc.results <- factor.analysis(Y, r = 5, "pc")
sapply(summary(lm(Z ~ pc.results$Z)), function(x) x$r.squared)

ml.results <- factor.analysis(Y, r = 5, "ml")
sapply(summary(lm(Z ~ ml.results$Z)), function(x) x$r.squared)

esa.results <- factor.analysis(Y, r = 5, "esa")
sapply(summary(lm(Z ~ esa.results$Z)), function(x) x$r.squared)
```

gen.sim.data

Generate simulation data set

Description

gen.sim.data generates data from the following model $Y = X_0 \text{Beta}_0^T + X_1 \text{Beta}_1^T + Z \text{Gamma}^T + E \text{Sigma}^{1/2}$, $Z|X_0, X_1 = X_0 \text{Alpha}_0^T + X_1 \text{Alpha}_1^T + D$, $\text{cov}(X_0, X_1) \sim \text{Sigma}_X$

Usage

```
gen.sim.data(n, p, r, d0 = 0, d1 = 1, X.dist = c("binary", "normal"),
  alpha = matrix(0.5, r, d0 + d1), beta = NULL, beta.strength = 1,
  beta.nonzero.frac = 0.05, Gamma = NULL, Gamma.strength = sqrt(p),
  Gamma.beta.cor = 0, Sigma = 1, seed = NULL)
```

Arguments

n	number of observations
p	number of observed variables
r	number of confounders
d0	number of nuisance regression covariates
d1	number of primary regression covariates
X.dist	the distribution of X, either "binary" or "normal"
alpha	association of X and Z, a $r \times d$ vector ($d = d0 + d1$)
beta	treatment effects, a $p \times d$ vector
beta.strength	strength of beta
beta.nonzero.frac	if beta is not specified, fraction of nonzeros in beta
Gamma	confounding effects, a $p \times r$ matrix
Gamma.strength	strength of Gamma, more precisely the mean of square entries of $\text{Gamma} \times \text{alpha}$
Gamma.beta.cor	the "correlation" (proportion of variance explained) of beta and Gamma
Sigma	noise variance, a $p \times p$ matrix or $p \times 1$ vector or a single real number
seed	random seed

Value

a list of objects

X0 matrix of nuisance covariates

X1 matrix of primary covariates

Y matrix Y

Z matrix of confounders

alpha regression coefficients between X and Z

beta regression coefficients between X and Y

Gamma coefficients between Z and Y

Sigma noise variance

beta.nonzero.pos the nonzero positions in beta

r number of confounders

gender.sm	<i>Gender study dataset</i>
-----------	-----------------------------

Description

This genetics dataset is used to demonstrate the usage of `cate` in the vignette. It was originally extracted by Gagnon-Bartsch and Speed (2012) as an example of confounded multiple testing. The data included in this package contains only 500 genes that are sampled from the original 12600 genes, besides keeping all the spike-in controls.

References

<http://www.stat.berkeley.edu/~johann/ruv/index.html> Vawter, M. P., S. Evans, P. Choudary, H. Tomita, J. Meador-Woodruff, M. Molnar, J. Li, J. F. Lopez, R. Myers, D. Cox, et al. (2004). Gender-specific gene expression in post-mortem human brain: localization to sex chromosomes. *Neuropsychopharmacology* 29(2), 373-384. Gagnon-Bartsch, J. A. and T. P. Speed (2012). Using control genes to correct for unwanted variation in microarray data. *Biostatistics* 13(3), 539-552.

wrapper	<i>Wrapper functions for some previous methods</i>
---------	--

Description

These functions provide an uniform interface to three existing methods: SVA, RUV, LEAPP. The wrapper functions transform the data into desired forms and call the corresponding functions in the package [sva](#), [ruv](#), [leapp](#).

Usage

```
sva.wrapper(formula, X.data = NULL, Y, r, sva.method = c("irw", "two-step"),
  B = 5)
```

```
ruv.wrapper(formula, X.data = NULL, Y, r, nc, lambda = 1,
  ruv.method = c("RUV2", "RUV4", "RUVinv"))
```

```
leapp.wrapper(formula, X.data = NULL, Y, r, search.tuning = F,
  ipod.method = c("hard", "soft"))
```

Arguments

<code>formula</code>	a formula indicating the known covariates including both primary variables and nuisance variables, which are separated by <code> </code> . The variables before <code> </code> are primary variables and the variables after <code> </code> are nuisance variables. It's OK if there is no nuisance variables, then <code> </code> is not needed and <code>formula</code> becomes a typical formula with all the covariates considered primary. An intercept term will still be automatically added as a nuisance variable for the latter case.
----------------------	---

X.data	the data frame used for formula
Y	outcome, n*p matrix
r	number of latent factors, can be estimated using the function <code>est.confounder.num</code>
sva.method	parameter for <code>sva</code> . whether to use an iterative reweighted algorithm (<code>irw</code>) or a two-step algorithm (<code>two-step</code>).
B	parameter for <code>sva</code> . the number of iterations of the <code>irwsva</code> algorithm
nc	parameter for <code>ruv</code> functions: position of the negative controls
lambda	parameter for <code>RUVinv</code>
ruv.method	either using <code>RUV2</code> , <code>RUV4</code> or <code>RUVinv</code> functions
search.tuning	logical parameter for <code>leapp</code> , whether using BIC to search for tuning parameter of IPOD.
ipod.method	parameter for <code>leapp</code> . "hard": hard thresholding in the IPOD algorithm; "soft": soft thresholding in the IPOD algorithm

Details

The `beta.p.values` returned is a length `p` vector, each for the overall effects of all the primary variables.

Only 1 variable of interest is allowed for `leapp.wrapper`. The method can be slow.

Value

All functions return `beta.p.value` which are the p-values after adjustment. For the other returned objects, refer to `cate` for their meaning.

Examples

```
## this is the simulation example in Wang et al. (2015).
n <- 100
p <- 1000
r <- 2
set.seed(1)
data <- gen.sim.data(n = n, p = p, r = r,
  alpha = rep(1 / sqrt(r), r),
  beta.strength = 3 * sqrt(1 + 1) / sqrt(n),
  Gamma.strength = c(seq(3, 1, length = r)) * sqrt(p),
  Sigma = 1 / rgamma(p, 3, rate = 2),
  beta.nonzero.frac = 0.05)
X.data <- data.frame(X1 = data$X1)
sva.results <- sva.wrapper(~ X1, X.data, data$Y,
  r = r, sva.method = "irw")
ruv.results <- ruv.wrapper(~ X1, X.data, data$Y, r = r,
  nc = sample(data$beta.zero.pos, 30), ruv.method = "RUV4")
leapp.results <- leapp.wrapper(~ X1, X.data, data$Y, r = r)
cate.results <- cate(~ X1, X.data, data$Y, r = r)

## p-values after adjustment
par(mfrow = c(2, 2))
```

```
hist(sva.results$beta.p.value)
hist(ruv.results$beta.p.value)
hist(leapp.results$beta.p.value)
hist(cate.results$beta.p.value)

## type I error
mean(sva.results$beta.p.value[data$beta.zero.pos] < 0.05)

## power
mean(sva.results$beta.p.value[data$beta.nonzero.pos] < 0.05)

## false discovery proportion for sva
discoveries.sva <- which(p.adjust(sva.results$beta.p.value, "BH") < 0.2)
fdp.sva <- length(setdiff(discoveries.sva, data$beta.nonzero.pos)) / max(length(discoveries.sva), 1)
fdp.sva
```

Index

*Topic **data**

gender.sm, 11

adjust.latent, 2

cate, 2, 3, 3, 12

cate-package, 2

ESA, 9

est.confounder.num, 5

est.factor.num (est.confounder.num), 5

fa.em, 7, 9

fa.pc, 7, 9

factor.analysis, 2, 7, 8, 8

gen.sim.data, 9

gender.sm, 11

leapp, 11, 12

leapp.wrapper (wrapper), 11

ruv, 11, 12

ruv.wrapper (wrapper), 11

RUV2, 12

RUV4, 12

RUVinv, 12

sva, 11, 12

sva.wrapper (wrapper), 11

wrapper, 4, 11