

Package ‘clinfun’

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Suggests survival

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calogrank	<i>Survival curves analysis of covariance</i>
-----------	---

Description

Logrank test to compare survival curves adjusting for covariates

Usage

```
calogrank(ftime, fstatus, grp, cvt, strat=NULL)
```

Arguments

ftime	failure times
fstatus	status indicator
grp	group indicator
cvt	continuous covariates used for adjusted analysis
strat	stratification variable

Details

calogrank is the covariate adjusted version of k-sample survdiff. The function in its current form only does basic error checking.

References

Heller G. and Venkatraman E.S. (2004) A nonparametric test to compare survival distributions with covariate adjustment. *JRSS-B* 66, 719-733.

Examples

```
## Not run:  library(survival)
data(pbc)
pbc1 <- pbc
pbc1$trt[pbc1$trt == -9] <- NA
pbc1$copper[pbc1$copper == -9] <- NA
calogrank(pbc1$time, pbc1$status, pbc1$trt, pbc1[,c("copper")])
calogrank(pbc1$time, pbc1$status, pbc1$trt, pbc1[,c("protime", "copper")])
## End(Not run)
```

`coxphCPE`*Gonen \& Heller Concordance Probability Estimate*

Description

Calculates the Concordance Probability Estimate for a Cox model.

Usage

```
coxphCPE(phfit)
```

Arguments

`phfit` output from a proportional hazards fit.

Value

`coxphCPE` returns a vector with `CPE`, `smooth.CPE` & `se.CPE` which are the estimate, the smoothed estimate and its standard error respectively.

References

Gonen M and Heller G. (2005) Concordance probability and discriminatory power in proportional hazards regression. *Biometrika* 92, 965-970.

Examples

```
## Not run: library(survival)
data(pbc)
pbcfit <- coxph(Surv(time, status==2) ~ trt + log(copper), pbc,
  subset=(trt>0 & copper>0))
coxphCPE(pbcfit)

## End(Not run)
```

`coxphERR`*Heller Explained Relative Risk*

Description

Calculates the contribution of a subset of covariates to the explained relative risk derived from the full Cox proportional hazards model.

Usage

```
coxphERR(phfit, ngamma=NULL)
```

Arguments

phfit	The output from a proportional hazards fit.
ngamma	A vector of indices corresponding to covariates of interest. If missing (default), the explained relative risk is computed for the full model.

Details

The object phfit should be the result of a call to coxph with the option x=TRUE.

Value

The function coxphERR returns the vector (ERR, se.ERR). The first component ERR represents the contribution of a subset of covariates to the explained relative risk estimate of the full model. If a set of covariates is not provided, then it computes the estimate of the full model. The second component se.ERR is the standard error of the estimate.

References

Heller G. (2012) A measure of explained risk in the proportional hazards model. *Biostatistics*

Examples

```
## Not run:
library(survival)
ovarianfit <- coxph(Surv(futime, fustat)~age+resid.ds+rx+ecog.ps,data=ovarian,x=T)
# Compute the explained relative risk (ERR) and its standard error (se.ERR) for the full model.
coxphERR(ovarianfit)
# Compute the contribution of age and ECOG performance status to the explained relative risk. # Age and ECOG perform
coxphERR(ovarianfit, c(1,4))

## End(Not run)
```

 coxphQuantile

Survival time quantile as a function of covariate

Description

Draws a quantile curve of survival distribution as a function of covariate.

Usage

```
coxphQuantile(phfit, xrange, p=0.5, whichx=1, otherx=NULL, ...)
```

Arguments

phfit	output from a proportional hazards fit.
xrange	the range of covariate values for which the quantiles of survival times are computed.
p	the probability level for the quantile (default is median).
whichx	if there are more than one covariates in the Cox model, the one chosen for the quantile plot.
otherx	the values for other covariates in the Cox model. If missing uses their average values.
...	additional parameters to be passed on to the lines command.

Details

This function is used to draw quantile curves. It requires a plot of the data (time & covariate of interest) to be present. See example.

It invisibly returns the observed failure times and the covariate values at which the estimated survival probability is (exactly) p.

References

Heller G. and Simonoff J.S. (1992) Prediction in censored survival data: A comparison of the proportional hazards and linear regression models. *Biometrics* 48, 101-115.

Examples

```
## Not run:  library(survival)
data(pbc)
pbcfit <- coxph(Surv(time, status==2) ~ trt + log(copper), pbc,
  subset=(trt>0 & copper>0))
plot(log(pbc$copper[pbc$trt>0 & pbc$copper>0]), pbc$time[pbc$trt>0 &
  pbc$copper>0], pch=c("o","x")[1+pbc$status[pbc$trt>0 & pbc$copper>0]],
  xlab="log Copper", ylab="Survival time")
coxphQuantile(pbcfit, c(2.5,6), whichx=2, otherx=1)
coxphQuantile(pbcfit, c(2.5,6), p=0.75, whichx=2, otherx=2, col=2)
## End(Not run)
```

Description

Calculates sample size, effect size and power based on Fisher's exact test

Usage

```

fe.ssize(p1,p2,alpha=0.05,power=0.8,r=1,npm=5,mmax=1000)
CPS.ssize(p1,p2,alpha=0.05,power=0.8,r=1)
fe.mdor(ncase,ncontrol,pcontrol,alpha=0.05,power=0.8)
fe.power(d, n1, n2, p1, alpha = 0.05)
or2pcase(pcontrol, OR)

```

Arguments

p1	response rate of standard treatment
p2	response rate of experimental treatment
d	difference = p2-p1
pcontrol	control group probability
n1	sample size for the standard treatment group
n2	sample size for the standard treatment group
ncontrol	control group sample size
ncase	case group sample size
alpha	size of the test (default 5%)
power	power of the test (default 80%)
r	treatments are randomized in 1:r ratio (default r=1)
npm	the sample size program searches for sample sizes in a range (+/- npm) to get the exact power
mmax	the maximum group size for which exact power is calculated
OR	odds-ratio

Details

CPS.ssize returns Casagrande, Pike, Smith sample size which is a very close to the exact. Use this for small differences p2-p1 (hence large sample sizes) to get the result instantaneously.

fe.ssize return a 2x3 matrix with CPS and Fisher's exact sample sizes with power.

fe.mdor return a 3x2 matrix with Schlesselman, CPS and Fisher's exact minimum detectable odds ratios and the corresponding power.

fe.power returns a Kx2 matrix with probabilities (p2) and exact power.

or2pcase give the probability of disease among the cases for a given probability of disease in controls (pcontrol) and odds-ratio (OR).

References

Casagrande, JT., Pike, MC. and Smith PG. (1978). An improved approximate formula for calculating sample sizes for comparing two binomial distributions. *Biometrics* 34, 483-486.

Fleiss, J. (1981) *Statistical Methods for Rates and Proportions*.

Schlesselman, J. (1987) Re: Smallest Detectable Relative Risk With Multiple Controls Per Case. *Am. J. Epi.*

Description

Functions to calculate sample size for group sequential designs

Usage

```
gsdesign.binomial(ifrac, pC, pE, sig.level = 0.05, power = 0.8,
  delta.eb=0.5, delta.fb = NULL, alternative = c("two.sided", "one.sided"),
  tol=0.00001, ...)
gsdesign.normal(ifrac, delta, sd = 1, sig.level = 0.05, power = 0.8,
  delta.eb = 0.5, delta.fb = NULL, alternative = c("two.sided", "one.sided"),
  tol=0.00001, ...)
gsdesign.survival(ifrac, haz.ratio, sig.level = 0.05, power = 0.8,
  delta.eb = 0.5, delta.fb = NULL, alternative = c("two.sided", "one.sided"),
  tol=0.00001, ...)
```

Arguments

ifrac	information fraction or the ratio of current sample size or number of events to the total sample size or number of events. This should be an increasing vector of numbers from 0 to 1 with the last one being 1. If just 1 is given a fixed sample design is derived.
pC	prob of success of the standard therapy (for binomial data)
pE	prob of success of the experimental therapy (for binomial data)
delta	true difference in means (for normal data)
sd	standard deviation (for normal data)
haz.ratio	hazard ratio (for survival comparison)
sig.level	significance level (type I error probability)
power	power of test (1 minus type II error probability)
delta.eb	power for efficacy boundary in the Pocock (power=0) to O'Brien-Fleming (power=0.5) family (default is 0.5)
delta.fb	power for futility boundary in the Pocock (power=0) to O'Brien-Fleming (power=0.5) family (default is NULL i.e. no futility boundary is requested.)
alternative	one- or two-sided test.
tol	tolerance level for multivariate normal probability computation.
...	additional options passed on the pmvnorm function.

Details

The futility boundary is not returned when delta.fb is not specified i.e. stopping for futility is not requested. The futility boundary is non-binding. That is the significance level is not adjusted to account for early stopping for utility. This makes the test a bit conservative in that the true size is less than the nominal level.

Value

a list with ifrac, sig.level, power, alternative, delta.eb, delta.fb and:

efbdry	the critical value to use at the different looks.
futbdry	the critical value to use at the different looks.
sample.size	the sample size per arm for binomial/normal data.
num.events	the total number of failures which should be converted to number of subjects using censoring proportion.

ktau	<i>Kendall's tau-b estimate</i>
------	---------------------------------

Description

Calculates the Kendall's tau-b.

Usage

```
ktau(x, y)
```

Arguments

x	first variable
y	second variable

Details

ktau computes the same quantity as `cor(x, y, method="kendall")`. It uses a faster algorithm than pairwise comparisons used by `cor`.

Value

ktau returns Kendall's tau-b.

Examples

```
set.seed(1234)
x <- rnorm(20000); y <- x+rnorm(20000)
cor(x, y, method="k")
clinfun:::ktau(x,y)
```

oc.twostage.bdry *Two-stage boundary operating characteristics*

Description

Calculates the operating characteristics of a two-stage boundary.

Usage

```
oc.twostage.bdry(pu, pa, r1, n1, r, n)
```

Arguments

pu	unacceptable response rate
pa	response rate that is desirable
r1	first stage threshold to declare treatment undesirable
n1	first stage sample size
r	overall threshold to declare treatment undesirable
n	total sample size

Value

oc.twostage.bdry returns the type I and II error rates as well as the probability of early termination and expected sample size under pu for a specific boundary.

permtests *Permutation versions of some common tests*

Description

Small sample tests using the permutation reference distributions.

Usage

```
permlogrank(formula, data, subset, na.action, rho=0, nperm=5000)
jonckheere.test(x, g, alternative = c("two.sided", "increasing",
  "decreasing"))
```

Arguments

nperm	number of permutations for the reference distribution
formula, data, subset, na.action, rho	see survdiff for details
x, g	data and group vector
alternative	means are monotonic (two.sided), increasing, or decreasing

Details

permlogrank is the permutation version of k-sample survdiff

jonckheere.test is the exact (permutation) version of the Jonckheere-Terpstra test. It uses the statistic

$$\sum_{k < l} \sum_{ij} I(X_{ik} < X_{jl}) + 0.5I(X_{ik} = X_{jl}),$$

where i, j are observations in groups k and l respectively. The asymptotic version is equivalent to `cor.test(x, g, method="k")`.

ph2simon

Simon's 2-stage Phase II design

Description

Calculates Optimal and Minimax 2-stage Phase II designs given by Richard Simon

Usage

```
ph2simon(pu, pa, ep1, ep2, nmax=100)
## S3 method for class 'ph2simon'
print(x, ...)
## S3 method for class 'ph2simon'
plot(x, ...)
```

Arguments

pu	unacceptable response rate
pa	response rate that is desirable
ep1	threshold for the probability of declaring drug desirable under p0
ep2	threshold for the probability of rejecting the drug under p1
nmax	maximum total sample size (default 100; can be at most 500)
x	object returned by ph2simon
...	arguments to be passed onto plot and print commands called within

Value

ph2simon returns a list with pu, pa, alpha, beta and nmax as above and:

out	matrix of best 2 stage designs for each value of total sample size n. the 6 columns are: r1, n1, r, n, EN(p0), PET(p0)
-----	--

The "print" method formats and returns the minimax and optimal designs. The "plot" plots the expected sample size against the maximum sample size as in Jung et al., 2001

References

- Simon R. (1989). Optimal Two-Stage Designs for Phase II Clinical Trials. *Controlled Clinical Trials* 10, 1-10.
- Jung SH, Carey M and Kim KM. (2001). Graphical Search for Two-Stage Designs for Phase II Clinical Trials. *Controlled Clinical Trials* 22, 367-372.

See Also

[twostage.inference](#), [oc.twostage.bdry](#)

Examples

```
ph2simon(0.2, 0.4, 0.1, 0.1)
ph2simon(0.2, 0.35, 0.05, 0.05)
ph2simon(0.2, 0.35, 0.05, 0.05, nmax=150)
```

ph2single

Exact single stage Phase II design

Description

Calculates the exact one stage Phase II design

Usage

```
ph2single(pu,pa,ep1,ep2,nsoln=5)
```

Arguments

pu	unacceptable response rate
pa	response rate that is desirable
ep1	threshold for the probability of declaring drug desirable under p0
ep2	threshold for the probability of rejecting the drug under p1
nsoln	number of designs with given alpha and beta

Value

ph2single returns a data frame with variables: n, r, and the Type I and Type II errors.

 power.ladesign

Power of k-sample rank test under Lehmann alternative

Description

Functions to calculate the power of rank tests for animal studies.

Usage

```
power.ladesign(gsize, odds.ratio, sig.level = 0.05, statistic =
  c("Kruskal-Wallis", "Jonckheere-Terpstra"), alternative =
  c("two.sided", "one.sided"), nrep=1e+6)
## S3 method for class 'ladesign'
print(x,...)
```

Arguments

gsize	sample size of the k (= length of vector) groups.
odds.ratio	odds ratio parameters for the k-1 groups. The first group is considered the control.
sig.level	the significance level of the test (default = 0.05)
statistic	the test statistic for the k-group comparison. Is one of Kruskal-Wallis (default) or Jonckheere-Terpstra.
alternative	one- or two-sided test. Valid only for the Jonckheere-Terpstra test.
nrep	number of reps (default 1 million) for Monte Carlo.
x	object of class ladesign returned by power.ladesign
...	arguments to be passed on left for S3 method consistency.

Details

Although the power for Jonckheere-Terpstra test is calculated for any set of odds ratio, the test is meant for monotone alternative. Thus it is preferable to specify odds ratios that are monotonically increasing with all values larger than 1 or decreasing with all values smaller than 1.

Value

returns a list with objects group.size, odds.ratio, statistic, sig.level and power. The "print" method formats the output.

References

Heller G. (2006). Power calculations for preclinical studies using a K-sample rank test and the Lehmann alternative hypothesis. *Statistics in Medicine* 25, 2543-2553.

Examples

```
power.ladesign(c(9,7), 4, statistic="K")
power.ladesign(c(9,7,9), c(2,4), statistic="J")
power.ladesign(c(9,7,9), c(2,4), statistic="J", alt="o")
```

pselect

Probability of selection under pick the winner rule

Description

Calculates the probability of selecting the treatment with the higher response rate under the pick the winner rule.

Usage

```
pselect(n, p, min.diff=NULL, min.resp=NULL)
```

Arguments

n	sample size for each treatment arm. This is either a single integer or a vector of two integers for the special case of comparing two treatments with unequal sample sizes
p	vector of response rates for the treatments.
min.diff	this is the number of responses or the rate by which the best treatment should be superior to the others to be chosen. This must be a positive integer or a rate between 0 and 1. If missing it defaults to 1 for the equal sample size case but quits with a warning for the unequal sample size case.
min.resp	the minimum number of responses in each treatment arm for it to be considered further. If missing defaults to 0.

Value

the function returns a list with:

prob.none.worthy

is the probability that no treatment has the minimum number of responses specified in min.resp. this element is present only if min.resp is greater than 0 for at least one arm.

prob.inconclusive

this is the probability that the best treatment has the requisite min.resp responses but exceeds the second best by less than min.diff responses (rate) provided the second best also has at least min.resp responses.

prob.selection

this is a matrix which for each treatment gives the response probability and the probability of selecting it i.e. the number of responses in the chosen arm is at least min.resp and either none of the remaining arms exceed the min.resp threshold or the chosen (best) arm is better than the second best by at least min.diff responses (rate).

References

Simon R, Wittes RE, Ellenberg SS. (1985). Randomized phase II clinical trials. *Cancer Treat Rep* 69, 1375-1381.

Examples

```
pselect(18, c(0.2, 0.2, 0.2)) # selection when no difference i.e. type I error
pselect(18, c(0.2, 0.2, 0.4)) # selection probability
pselect(26, c(0.2, 0.2, 0.4), min.diff=2, min.resp=3)
pselect(c(27,54), c(0.5, 0.65), min.diff=0.05) # unequal sample size case
pselect(c(27,54), c(0.5, 0.65), min.diff=0.05, min.resp=c(14,27)) # unequal sample size case
```

roc.area.test

Nonparametric area under the ROC curve

Description

Computes the nonparametric area under the ROC curve and its variance based on U-statistic theory (DDCP).

Usage

```
roc.area.test(markers, status)
## S3 method for class 'roc.area.test'
print(x, ...)
```

Arguments

markers	The marker values for each subject. If there are more than one markers then this should be a matrix.
status	binary disease status indicator
x	object of class roc.area.test output from this function.
...	optional arguments to the print function.

Details

It calculates the area and its variance. For more than one marker it calculates the statistic to test for the equality of all AUCs. This statistic has a standard normal reference distribution for two variables and chi-square with number of variables minus 1.

Value

a list with the following elements

area	estimated area.
var	estimated variance (matrix).

stat test statistic for equality of AUCs. Is not returned when only one diagnostic marker is present.

p.value the p-value for the test of equality (2-sided).

df the degrees of freedom of the chi-square.

The "print" method formats and returns the output.

References

DeLong, E. R., D. M. DeLong, and D. L. Clarke-Pearson. 1988. Comparing the areas under two or more correlated receiver operating characteristic curves: A nonparametric approach. *Biometrics* 44:837-845.

Examples

```
g <- rep(0:1, 50)
x <- rnorm(100) + g
y <- rnorm(100) + g
z <- rnorm(100) + g
roc.area.test(cbind(x,y), g)
roc.area.test(cbind(x,y,z), g)
y1 <- y + 0.75*g
roc.area.test(cbind(x,y1), g)
```

roc.curve	<i>Empirical ROC curve</i>
-----------	----------------------------

Description

Computes the empirical ROC curve for a diagnostic tool.

Usage

```
roc.curve(marker, status, method=c("empirical"))
## S3 method for class 'roc.curve'
print(x, ...)
## S3 method for class 'roc.curve'
plot(x, ...)
## S3 method for class 'roc.curve'
lines(x, ...)
```

Arguments

marker the marker values for each subject.

status binary disease status indicator

method the method for estimating the ROC curve. Currently only the empirical curve is implemented.

x object of class roc.area.test output from this function.

... optional arguments to the print, plot and lines functions.

Details

The computation is based on assuming that larger values of the marker is indicative of the disease. So for a given threshold x_0 , TPR is $P(\text{marker} \geq x_0 | \text{status} = 1)$ and FPR is $P(\text{marker} \geq x_0 | \text{status} = 0)$. This function computes the empirical estimates of TPR and FPR.

Value

a list with the following elements

tpr	true positive rates for all thresholds.
fpr	true positive rates for all thresholds.
marker	the diagnostic marker being studied.
status	binary disease

The "print" method returns the nonparametric AUC and its s.e.

The "plot" and "lines" methods can be used to draw a new plot and add to an existing plot of ROC curve.

Examples

```
g <- rep(0:1, 50)
x <- rnorm(100) + g
y <- rnorm(100) + 1.5*g
o <- roc.curve(x, g)
plot(o)
lines(roc.curve(y, g), col=2)
```

roc.perm.test

Permutation test to compare ROC curve

Description

Computes the test statistic and permutation reference distribution for comparing paired or unpaired ROC curves.

Usage

```
roc.perm.test(marker, status, marker2=NULL, group=NULL, nperm=2500, mp=NULL)
## S3 method for class 'roc.perm.test'
print(x, ...)
## S3 method for class 'roc.perm.test'
plot(x, ...)
```

Arguments

marker	marker values for each subject.
status	binary disease status indicator.
marker2	second diagnostic marker for the same subjects (paired).
group	indicator of which diagnostic test was used (unpaired).
nperm	number of permutations for the reference distribution.
mp	mixing proportion for the unpaired case when proportion of diseased subjects can differ.
x	object of class roc.perm.test output from this function.
...	optional arguments to print and plot functions.

Details

This function implements the permutation method described in the Venkatraman and Begg (1996) paper for the paired case and the Venkatraman (2000) paper for the unpaired case.

The function detects whether the data are paired or unpaired by testing which of the options marker2 and group is specified. If both are missing it will stop with an error message. At present exactly one should be missing.

Value

an object of class roc.perm.test with the following elements

ostat	test statistic from the observed data.
pstat	test statistic from permuted data.
p.value	the p-value for the test of equality (2-sided).

The "print" method formats and returns the statistic and p-value. The "plot" method plots the density from the permutation reference distribution and marks the location of the observed statistic.

References

Venkatraman, E.S. and Begg, C.B. (1996). A distribution-free procedure for comparing receiver operating characteristic curves from a paired experiment. *Biometrika* 83, 835-848.

Venkatraman, E.S. (2000) A permutation test to compare receiver operating characteristic curves. *Biometrics* 56(4):1134-8.

Examples

```
d <- rep(0:1, 50)
x <- rnorm(100) + 1.2*d
y <- rnorm(100) + 1.2*d
oo <- roc.perm.test(x, d, marker2=y)
plot(oo)
oo <- roc.perm.test(c(x,y), c(d,d), group=rep(1:2,each=100))
plot(oo)
```

 ROCAnalysis

Functions to plot and compare ROC curves.

Description

These functions can be used for nonparametric analysis of ROC curves.

Details

The relevant functions are `roc.curve`, `roc.area.test` and `roc.perm.test`. See the individual functions for usage details.

 toxbdry

Stopping rule for toxicity monitoring

Description

Computes a stopping rule and its operating characteristics for toxicity monitoring based repeated significance testing.

Usage

```

toxbdry(pLo, pHi, n, cP0=0.1, cP1=0.9, ngrid=6, niter=5,
        priority=c("null", "alt"))
bdrycross.prob(n, r, ptox)
## S3 method for class 'toxbdry'
print(x, ...)

```

Arguments

pLo	the toxicity rate that is acceptable.
pHi	the toxicity rate that is too high and hence unacceptable.
n	vector of times (sample size) when toxicity is monitored.
r	vector of maximum acceptable toxicities corresponding to n.
ptox	the toxicity rates for which the operating characteristics are calculated.
cP0	boundary crossing probability under pLo i.e. type I error or the probability of declaring a treatment with toxicity rate pLo unacceptable.
cP1	boundary crossing probability under pHi i.e. power or the probability of declaring a treatment with toxicity rate pHi unacceptable.
ngrid	the number of toxicity rates from pLo to pHi for which the operating characteristics are computed.
niter	the number of iterations run to obtain the boundary.

priority	the error threshold to prioritize when the max sample size is too small to have both error thresholds satisfied. Default is the null i.e. error under pLo.
x	object returned by the function toxbdry.
...	additional arguments to print.

Value

the function returns a list with:

looks	when toxicity is monitored - same as input n.
lo.bdry	lower boundary is a vector of maximum acceptable number of toxicities corresponding the number of subjects in n. The boundary crossing probability for this is slightly above cP0.
hi.bdry	upper boundary is a vector of maximum acceptable number of toxicities corresponding the number of subjects in n. The boundary crossing probability for this is slightly below cP0.
bdry.oc	the operating characteristics i.e the toxicity rate, the probability of crossing, stopping (i.e. cross before the last observation) and the expected sample size for both the low (lo) and high (hi) boundaries.
bdry.alpha	the alpha levels for testing at each look for the two boundaries.

stopping for toxicity is done when the number of toxicities exceeda the boundary i.e. the boundary gives the maximum acceptable number.

Examples

```

toxbdry(0.2, 0.35, c(20,40,60,75))
toxbdry(0.2, 0.3, c(20,40,60,75), cP0=0.15, cP1=0.8)
toxbdry(0.1, 0.3, 2:30) # continuous monitoring
toxbdry(0.1, 0.3, 2:25, priority="alt") # prioritize cP1 error threshold

```

twostage.inference *Inference following a two-stage design for binary response*

Description

Calculates the p-value, UMVUE and CI for the data from a study using a two stage design for response.

Usage

```
twostage.inference(x, r1, n1, n, pu, alpha=0.05)
```

Arguments

x	number of responses observed at the end of the study
r1	first stage threshold to declare treatment undesirable
n1	first stage sample size
n	total sample size
pu	unacceptable response rate (null hypothesis)
alpha	the confidence level. For consistency with the design use the same value from the design. (default is 0.05)

Value

twostage.inference returns the UMVUE (Jung & Kim, 2004), p-value and CI (Koyama & Chen, 2008). The CI has confidence level $1-2*\alpha$ and the one-sided $(1-\alpha)$ interval consistent with the design is obtained by changing the upper confidence limit (UCL) to 1.

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