

# Package ‘embryogrowth’

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**Type** Package

**Title** Tools to Analyze the Thermal Reaction Norm of Embryo Growth

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**Description** Tools to analyze the embryo growth and the sexualisation thermal reaction norms. See <[doi:10.7717/peerj.8451](https://doi.org/10.7717/peerj.8451)> for tsd functions; see <[doi:10.1016/j.jtherbio.2014.08.005](https://doi.org/10.1016/j.jtherbio.2014.08.005)> for thermal reaction norm of embryo growth.

**Depends** deSolve, optimx, numDeriv, parallel, HelpersMG (>= 4.0),R (>= 4.0)

**Suggests** entropy, shiny, coda, polynom, car, lmf, gam, pbapply

**License** GPL-2

**LazyData** yes

**LazyLoad** yes

**Encoding** UTF-8

**RoxygenNote** 7.1.2

**NeedsCompilation** no

**Repository** CRAN

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## R topics documented:

embryogrowth-package . . . . .	3
calibrate.datalogger . . . . .	7
ChangeSSM . . . . .	8
DatabaseNestingArea . . . . .	11
DatabaseTSD . . . . .	12
DatabaseTSD.version . . . . .	14
dydt.exponential . . . . .	14

dydt.Gompertz . . . . .	15
dydt.linear . . . . .	16
FormatNests . . . . .	17
GenerateAnchor . . . . .	19
GenerateConstInc . . . . .	20
Generate_hatchling_metric . . . . .	21
GRTRN_MHmcmc . . . . .	22
HatchingSuccess.fit . . . . .	25
HatchingSuccess.InL . . . . .	27
HatchingSuccess.MHmcmc . . . . .	29
HatchingSuccess.MHmcmc_p . . . . .	31
HatchingSuccess.model . . . . .	33
hist.Nests . . . . .	34
hist.NestsResult . . . . .	35
info.nests . . . . .	36
integral.exponential . . . . .	43
integral.Gompertz . . . . .	45
integral.linear . . . . .	46
likelihoodR . . . . .	47
logLik.HatchingSuccess . . . . .	48
logLik.NestsResult . . . . .	50
logLik.STRN . . . . .	51
logLik.tsd . . . . .	52
movement . . . . .	53
MovingIncubation . . . . .	54
nest . . . . .	57
nobs.HatchingSuccess . . . . .	58
nobs.NestsResult . . . . .	59
plot.HatchingSuccess . . . . .	60
plot.NestsResult . . . . .	63
plot.tsd . . . . .	67
plotR . . . . .	70
plot_transition . . . . .	73
predict.HatchingSuccess . . . . .	75
predict.tsd . . . . .	76
P_TRT . . . . .	78
resultNest_3p_Dallwitz . . . . .	80
resultNest_3p_Weibull . . . . .	81
resultNest_4p_normal . . . . .	82
resultNest_4p_SSM . . . . .	83
resultNest_4p_SSM_Linear . . . . .	84
resultNest_4p_transition . . . . .	85
resultNest_4p_trigo . . . . .	86
resultNest_4p_weight . . . . .	87
resultNest_5p_Dallwitz . . . . .	88
resultNest_6p_SSM . . . . .	89
resultNest_mcmc_4p_SSM . . . . .	90
resultNest_mcmc_4p_SSM_Linear . . . . .	91

resultNest_mcmc_6p_SSM . . . . .	92
resultNest_mcmc_newp . . . . .	93
resultNest_newp . . . . .	95
searchR . . . . .	96
stages . . . . .	103
STRN . . . . .	107
STRN_MHmcmc . . . . .	113
summary.Nests . . . . .	116
switch.transition . . . . .	117
tempConst . . . . .	118
test.parallel . . . . .	119
TRN_MHmcmc_p . . . . .	120
tsd . . . . .	121
tsd_MHmcmc . . . . .	127
tsd_MHmcmc_p . . . . .	130
TSP.list . . . . .	132
uncertainty.datalogger . . . . .	133
web.tsd . . . . .	135
weightmaxentropy . . . . .	136

**Index****138**


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embryogrowth-package    *The package embryogrowth*

---

**Description**

Tools to analyze the embryo growth and the sexualisation thermal reaction norms.

The latest version of this package can always been installed using:

```
install.packages("http://max2.es.e.u-psud.fr/epc/conservation/CRAN/HelpersMG.tar.gz", repos=NULL,
type="source")
```

```
install.packages("http://max2.es.e.u-psud.fr/epc/conservation/CRAN/embryogrowth.tar.gz", repos=NULL,
type="source")
```



## Details

Fit a parametric function that describes dependency of embryo growth to temperature

Package:	embryogrowth
Type:	Package
Version:	8.2 build 1473
Date:	2021-09-16
License:	GPL (>= 2)
LazyLoad:	yes

## Author(s)

Marc Girondot <marc.girondot@gmail.com>

## References

Girondot, M. & Kaska, Y. 2014. A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology* 45, 96-102.

Fuentes, M.M.P.B., Monsinjon, J., Lopez, M., Lara, P., Santos, A., dei Marcovaldi, M.A.G., Girondot, M., 2017. Sex ratio estimates for species with temperature-dependent sex determination differ according to the proxy used. *Ecological Modelling* 365, 55-67.

Girondot, M., Monsinjon, J., Guillon, J.-M., 2018. Delimitation of the embryonic thermosensitive period for sex determination using an embryo growth model reveals a potential bias for sex ratio prediction in turtles. *Journal of Thermal Biology* 73, 32-40.

Monsinjon, J., Jribi, I., Hamza, A., Ouerghi, A., Kaska, Y., Girondot, M., 2017. Embryonic growth rate thermal reaction norm of Mediterranean *Caretta caretta* embryos from two different thermal habitats, Turkey and Libya. *Chelonian Conservation and Biology* 16, 172-179.

## See Also

Delmas, V., Prevot-Julliard, A.-C., Pieau, C. & Girondot, M. 2008. A mechanistic model of temperature-dependent sex determination in a Chelonian, the European pond turtle. *Functional Ecology*, 22, 84-93.

Girondot, M., Ben Hassine, S., Sellos, C., Godfrey, M. & Guillon, J.-M. 2010. Modeling thermal influence on animal growth and sex determination in Reptiles: being closer of the target gives new views. *Sexual Development*, 4, 29-38.

Girondot, M. 1999. Statistical description of temperature-dependent sex determination using maximum likelihood. *Evolutionary Ecology Research*, 1, 479-486.

Girondot, M., & Kaska, Y. (2014). Nest temperatures in a loggerhead- nesting beach in Turkey is more determined by sea surface temperature than air temperature. *Journal of Thermal Biology*, 47, 13-18.

## Examples

```
## Not run:  
library("embryogrowth")
```

```

packageVersion("embryogrowth")
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(115.758929130522, 428.649022170996, 503.687251738993,
12.2621455821612, 306.308841227278, 116.35048615105), .Names = c("DHA",
"DHH", "DHL", "DT", "T12L", "Rho25"))
# or
x <- structure(c(118.431040984352, 498.205702157603, 306.056280989839,
118.189669472381), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)

#####
#
# The values of rK=2.093313 and M0=1.7 were used in
# Girondot, M. & Kaska, Y. 2014. A model to predict the thermal
# reaction norm for the embryo growth rate from field data. Journal of
# Thermal Biology. 45, 96-102.
#
# Based on recent analysis on table of development for both Emys orbicularis and
# Caretta caretta, best value for rK should be 1.209 and M0 should be 0.34.
# Girondot M, Monsinjon J, Guillon J-M (2018) Delimitation of the embryonic
# thermosensitive period for sex determination using an embryo growth model
# reveals a potential bias for sex ratio prediction in turtles. Journal of
# Thermal Biology 73: 32-40
#
# See the example in the stages datasets
#
#####

resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)
par(mar=c(4, 4, 1, 1))
plot(resultNest_4p_SSM$data[[1]][, 1]/60/24,resultNest_4p_SSM$data[[1]][, 2], bty="n", las=1,
      xlab="Days of incubation", ylab="Temperatures in °C",
      type="l", xlim=c(0,70),ylim=c(20, 35))
for (i in 2:resultNest_4p_SSM$data$IndiceT[["NbTS"]]) {
  par(new=TRUE)
  plot(resultNest_4p_SSM$data[[i]][, 1]/60/24,resultNest_4p_SSM$data[[i]][, 2],
      bty="n", las=1, xlab="", ylab="", type="l", xlim=c(0,70),ylim=c(20, 35), axes = FALSE)
}
par(mar=c(4, 4, 1, 1))
pMCMC <- TRN_MHmcmc_p(resultNest_4p_SSM, accept=TRUE)
# Take care, it can be very long, sometimes several days
resultNest_mcmc_4p_SSM <- GRTRN_MHmcmc(result=resultNest_4p_SSM,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1, n.adapt = 0,
thin=1, trace=TRUE)

```

```

data(resultNest_mcmc_4p_SSM)
out <- as.mcmc(resultNest_mcmc_4p_SSM)
# This out obtained after as.mcmc can be used with coda package
# plot() can use the direct output of GRTRN_MHmcmc() function.
plot(resultNest_mcmc_4p_SSM, parameters=1, xlim=c(0,550))
plot(resultNest_mcmc_4p_SSM, parameters=3, xlim=c(290,320))
# But rather than to use the SD for each parameter independantly, it is
# more logical to estimate the distribution of the curves
new_result <- ChangeSSM(resultmcmc = resultNest_mcmc_4p_SSM, result = resultNest_4p_SSM,
                        temperatures = seq(from = 20, to = 35, by = 0.1),
                        initial.parameters = NULL)
par(mar=c(4, 4, 1, 5)+0.4)

plotR(result = resultNest_4p_SSM, parameters = new_result$par,
      ylabH = "Temperatures\ndensity", ylimH=c(0, 0.3), atH=c(0, 0.1, 0.2),
      ylim=c(0, 3), show.hist=TRUE)

# Beautiful density plots

plotR(result = resultNest_4p_SSM,
      resultmcmc=resultNest_mcmc_4p_SSM,
      curve = "MCMC quantiles", show.density=TRUE)

plotR(resultNest_6p_SSM, resultmcmc=resultNest_mcmc_6p_SSM,
      ylim=c(0, 4), show.density=TRUE, show.hist=TRUE,
      curve = "MCMC quantiles",
      ylimH=c(0,0.5), atH=c(0, 0.1, 0.2))

## End(Not run)

```

---

calibrate.datalogger    *Calibrate data loggers and correct time series of temperatures*

---

## Description

Calibrate a time series of temperatures. Use or gam or glm. If no temperatures.series is given, it will use the read.temperatures.

## Usage

```

calibrate.datalogger(
  control.temperatures = stop("Control temperatures is missing"),
  read.temperatures = stop("Read temperatures must be indicated"),
  temperatures.series = NULL,
  gam = TRUE,
  se.fit = TRUE
)

```

**Arguments**

<code>control.temperatures</code>	The true temperatures during the calibration process
<code>read.temperatures</code>	The read temperatures during the calibration process
<code>temperatures.series</code>	The temperatures to be converted using calibration
<code>gam</code>	Does gam should be used (TRUE) or glm (FALSE).
<code>se.fit</code>	Do standard errors are to be returned

**Details**

`calibrate.datalogger` calibrates data loggers and correct time series of temperatures.

**Value**

The function will return a corrected time series of temperatures as a vector if `se.fit` is FALSE or a list if `se.fit` is TRUE.

**Author(s)**

Marc Girondot

**See Also**

Other Data loggers utilities: [movement\(\)](#), [uncertainty.datalogger\(\)](#)

**Examples**

```
## Not run:  
library(embryogrowth)  
calibrate.datalogger(control.temperatures=20:30,  
                    read.temperatures=(20:30)+rnorm(11))  
  
## End(Not run)
```

---

ChangeSSM

*Generate set of parameters for different forms of thermal norm of reaction*

---

**Description**

Generate a set of parameters for thermal reaction norm model.  
If `initial.parameters` is NULL and `resultmcmc` is not NULL, it will generate parameters and SE based on the average of the curves.



**Usage**

```
ChangeSSM(
  result = NULL,
  resultmcmc = NULL,
  temperatures = seq(from = 20, to = 35, by = 0.1),
  parameters = NULL,
  initial.parameters = NULL,
  fixed.parameters = NULL,
  outmcmc = "quantiles",
  progressbar = TRUE,
  ...
)
```

**Arguments**

result	A result obtained by searchR()
resultmcmc	A result obtained by GRTRN_MHmcmc()
temperatures	A vector with incubation temperatures in degrees Celsius
parameters	A vector of parameters for model to be converted. Not necessary if result is provided.
initial.parameters	NULL or a vector of parameters for initial model model to be fitted
fixed.parameters	NULL or a vector of parameters to be used but fixed
outmcmc	What statistic will be estimated if a mcmc is provided. Can be "mean-sd" or "quantiles".
progressbar	If TRUE, a progressbar is shown
...	A control list to be used with optim, see ?optim

**Details**

ChangeSSM convert different forms of thermal norm of reaction

**Value**

A vector with parameters or a result object formatted with new parameters is result is non null

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
data(resultNest_6p_SSM)
x1 <- resultNest_6p_SSM$par
data(resultNest_4p_SSM)
```

```

x2 <- resultNest_4p_SSM$par
temperaturesC <- (200:350)/10
s <- ChangeSSM(temperatures=temperaturesC, parameters=x1, initial.parameters=x2)
sY <- plotR(resultNest_6p_SSM, ylim=c(0,3), col="black", curve = "ML")
plotR(resultNest_4p_SSM, col="red", scaleY=sY, new=FALSE)
plotR(s$par, col="green", scaleY=sY, new=FALSE, curve = "ML")
legend("topleft", legend=c("r function to mimic", "Initial new r function",
"Fitted new r function"), lty=c(1, 1, 1), col=c("black", "red", "green"))
# Other example to fit anchored parameters
data(resultNest_4p_SSM)
x0 <- resultNest_4p_SSM$par
t <- hist(resultNest_4p_SSM, plot=FALSE)
x <- c(3.4, 3.6, 5.4, 5.6, 7.6, 7.5, 3.2)
names(x) <- seq(from=range(t$temperatures)[1], to=range(t$temperatures)[2],
length.out=7)
newx <- ChangeSSM(temperatures = (200:350)/10, parameters = x0,
initial.parameters = x,
control=list(maxit=5000))
# Example on how to generate a set of SSM parameters from anchored parameters
xanchor <- GenerateAnchor(nests=resultNest_4p_SSM)
x <- resultNest_4p_SSM$par
xanchor["294"] <- 0
xanchor["308"] <- 2.3291035
x <- ChangeSSM(parameters = xanchor,
initial.parameters = x, control=list(maxit=5000))
sY <- plotR(resultNest_4p_SSM$par, ylim = c(0,3), curve="ML")
plotR(xprime$par, col="red", scaleY=sY, new=FALSE, curve="ML")
legend("topleft", legend=c("Fitted parameters", "Constrained parameters"), lty=1,
col=c("black", "red"))
# Weibull model
x <- ChangeSSM(temperatures = (200:350)/10,
parameters = resultNest_4p_SSM$par,
initial.parameters = structure(c(73, 300, 26),
.Names = c("k", "lambda", "scale")),
control=list(maxit=1000))
# normal asymmetric model
x <- ChangeSSM(temperatures = (200:350)/10,
parameters = resultNest_4p_SSM$par,
initial.parameters = structure(c(3, 10, 8, 32),
.Names = c("Scale", "sdL", "sdH", "Peak")),
control=list(maxit=1000))
# trigonometric model
x <- ChangeSSM(temperatures = (200:350)/10,
parameters = resultNest_4p_SSM$par,
initial.parameters = structure(c(3, 20, 40, 32),
.Names = c("Max", "LengthB", "LengthE", "Peak")),
control=list(maxit=1000))

# example with a mcmc object, CI being 2.SD
# Note the symmetric CI
data(resultNest_mcmc_4p_SSM)
new_result <- ChangeSSM(resultmcmc = resultNest_mcmc_4p_SSM, result = resultNest_4p_SSM,
temperatures = seq(from = 20, to = 35, by = 0.1),

```

```

        outmcmc = "mean-sd",
        initial.parameters = NULL)

plotR(new_result, ylim=c(0, 3), curve="ML")
# example with a mcmc object, CI being defined by 2.5%-97.5% quantiles
# Note the asymmetric CI
data(resultNest_mcmc_4p_SSM)
new_result <- ChangeSSM(resultmcmc = resultNest_mcmc_4p_SSM, result = resultNest_4p_SSM,
                        temperatures = seq(from = 20, to = 35, by = 0.1),
                        outmcmc = "quantiles",
                        initial.parameters = NULL)

plotR(new_result, ylim=c(0, 3), curve="ML")
plotR(new_result, ylim=c(0, 3), curve="ML quantiles")

## End(Not run)

```

---

DatabaseNestingArea     *Database of RMU for marine turtles*

---

### Description

Database of RMU for marine turtles

### Usage

DatabaseNestingArea

### Format

A dataframe with raw data.

### Details

Database of RMU for marine turtles

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

Maria Sousa Martins <maria.esmartins@gmail.com>

### References

Wallace, B. P., DiMatteo, A. D., Hurley, B. J., Finkbeiner, E. M., Bolten, A. B., Chaloupka, M. Y., Hutchinson, B. J., Abreu-Grobois, F. A., Amorocho, D., Bjordal, K. A., Bourjea, J., Bowen, B. W., Duenas, R. B., Casale, P., Choudhury, B. C., Costa, A., Dutton, P. H., Fallabrino, A., Girard, A., Girondot, M., Godfrey, M. H., Hamann, M., Lopez-Mendilaharsu, M., Marcovaldi, M. A., Mortimer, J. A., Musick, J. A., Nel, R., Seminoff, J. A., Troeng, S., Witherington, B. & Mast, R. B. 2010. Regional management units for marine turtles: a novel framework for prioritizing conservation and research across multiple scales. *Plos One*, 5, e15465.

**Examples**

```
## Not run:
library(embryogrowth)
data(DatabaseNestingArea)

## End(Not run)
```

---

DatabaseTSD

*Database of TSD information for reptiles*


---

**Description**

Database of TSD information for reptiles  
The columns are:

- Species: Name of the species in binominal nomenclature
- Subspecies: Name of the subspecies
- Country: From which country the eggs come from
- Area: Name of the beach or region the eggs come from
- RMU: For marine turtles, name of the RMU for this population; see Wallace, B.P., DiMatteo, A.D., Hurley, B.J., Finkbeiner, E.M., Bolten, A.B., Chaloupka, M.Y., Hutchinson, B.J., Abreu-Grobois, F.A., Amorocho, D., Bjorndal, K.A., Bourjea, J., Bowen, B.W., Duenas, R.B., Casale, P., Choudhury, B.C., Costa, A., Dutton, P.H., Fallabrino, A., Girard, A., Girondot, M., Godfrey, M.H., Hamann, M., Lopez-Mendilaharsu, M., Marcovaldi, M.A., Mortimer, J.A., Musick, J.A., Nel, R., Seminoff, J.A., Troeng, S., Witherington, B., Mast, R.B., 2010. Regional management units for marine turtles: a novel framework for prioritizing conservation and research across multiple scales. Plos One 5, e15465.
- Incubation.temperature: Nominal incubation temperature
- Incubation.temperature.Constant: Does the incubation temperature was set as constant or CTE was reported
- Incubation.temperature.Accuracy: What is the accuracy of the measure of temperature
- Incubation.temperature.SD: Experimental SD of incubation temperatures
- Incubation.temperature.Amplitude: How much the temperature could fluctuate around nominal temperature
- Correction.factor: Difference between the incubator temperature and the eggs temperature
- IP.min: Shorter incubation period
- IP.max: Longer incubation period
- IP.mean: Mean incubation periods
- IP.SD: Standard deviation for incubation periods
- Total: Total number of eggs incubated
- Hatched: Number of hatchlings

- NotHatched: Number of embryos with development visible but dead during incubation
- Undeveloped: Number of embryos showing no development
- Intersexes: Number of individuals intersexes or ambiguous for sex phenotype
- Males: Number of individuals identified as males
- Females: Number of individuals identified as females
- Sexed: Number of sexed individuals
- Clutch: Identity or number of clutches
- Reference: Bibliographic reference
- Note: Diverse information for this incubation
- Version: Date of the last modification for each record

### Usage

DatabaseTSD

### Format

A dataframe with raw data.

### Details

Database of TSD information for marine turtles

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

### See Also

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

### Examples

```
## Not run:
library(embryogrowth)
data(DatabaseTSD)
DatabaseTSD.version()
totalIncubation_Lo <- subset(DatabaseTSD,
  Species=="Lepidochelys olivacea" & (!is.na(Sexed) & Sexed!=0),
  select=c("Males", "Females", "Incubation.temperature"))
tot_Lo <- with(totalIncubation_Lo, tsd(males=Males, females=Females,
  temperatures=Incubation.temperature), parameters.initial = c(P=30.5, S=-0.4))
predict(tot_Lo)

## End(Not run)
```

---

DatabaseTSD.version     *Version of database of TSD information for reptiles*

---

**Description**

Return the date of the most recent update of the database.

**Usage**

```
DatabaseTSD.version()
```

**Details**

Database of TSD information for marine turtles

**Value**

The date of the lastest updated version

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**See Also**

Other Functions for temperature-dependent sex determination: [DatabaseTSD](#), [P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

**Examples**

```
## Not run:  
library(embryogrowth)  
DatabaseTSD.version()  
  
## End(Not run)
```

---

dydt.exponential     *Return the derivative of the exponential function*

---

**Description**

Return the derivative of the exponential function  
dydt.exponential(t, size, parms)

**Usage**

```
dydt.exponential(t, size, parms)
```

**Arguments**

t	The time in any unit
size	The current size
parms	A vector with alpha and K values being c(alpha=x1, K=x2). K is not used.

**Details**

dydt.exponential returns the derivative of the exponential function.

**Value**

A list with the derivative

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(306.174998729436, 333.708348843241,
299.856306141849, 149.046870203155),
.Names = c("DHA", "DHH", "T12H", "Rho25"))
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_exponential <- searchR(parameters=x, fixed.parameters=NULL,
temperatures=formatted, derivate=dydt.exponential, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))

## End(Not run)
```

---

dydt.Gompertz

*Return the derivative of the Gompertz function*


---

**Description**

Return the derivative of the Gompertz function  
dydt.Gompertz(t, size, parms)

**Usage**

```
dydt.Gompertz(t, size, parms)
```

**Arguments**

t	The time in any unit
size	The current size
parms	A vector with alpha and K values being c(alpha=x1, K=x2)

**Details**

dydt.Gompertz returns the derivative of the Gompertz function.

**Value**

A list with the derivative

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(118.768297442004, 475.750095909406, 306.243694918151,
116.055824800264), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, derivate=dydt.Gompertz, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)

## End(Not run)
```

---

dydt.linear

*Return the derivative of the linear function*


---

**Description**

Return the derivative of the linear function  
dydt.linear(t, size, parms)



**Usage**

```
dydt.linear(t, size, parms)
```

**Arguments**

t	The time in any unit
size	The current size
parms	A vector with alpha being c(alpha=x1, K=x2). Only alpha is used.

**Details**

dydt.Linear returns the derivative of the linear function.

**Value**

A list with the derivative

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(306.174998729436, 333.708348843241, 299.856306141849,
149.046870203155), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_linear <- searchR(parameters=x, fixed.parameters=NULL,
temperatures=formatted, derivate=dydt.linear, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))

## End(Not run)
```

**Description**

Will create a dataset of class Nests to be used with searchR  
 FormatNests(nest, previous=x) with x being a previously formatted data.  
 The raw data must be organized being:  
 First column is the time in minutes since the beginning of incubation  
 Each column next is the trace of temperatures, one column for each nest.  
 For example, for two nests:  
 Time Nest1 Nest2  
 0 29.8 27.6  
 90 30.2 28.8  
 120 30.4 30.7  
 180 31.2 32.6  
 ...  
 65800 30.8 32.6  
 65890 30.2  
 65950 30.4

The Nest1 ends incubation at 65800 minutes whereas Nest2 ends incubation at 65950 (last row with temperature for each).

The parameter Weight is a vector: weight=c(Nest1=1, Nest2=1.2)

It can be used to format database already formatted with old format; in this case, just use data=xxx with xxx being the old format database.

**Usage**

```
FormatNests(
  data = stop("A dataset must be provided !"),
  previous = NULL,
  usemiddletime = FALSE,
  simplify = TRUE,
  weight = NULL
)
```

**Arguments**

data	Data to be newly formatted
previous	Data already formatted
usemiddletime	If TRUE, suppose that recorded temperatures are those at middle segment
simplify	If TRUE, simply the time series by removing identical time series of temperatures
weight	Named vector with weight for likelihood

**Details**

FormatNests creates a dataset of class "Nests" to be used with searchR

**Value**

A list with all the nests formatted to be used with searchR.

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest, previous=NULL)
formatted <- FormatNests(nest)

## End(Not run)
```

---

GenerateAnchor

*Generate a set of anchored parameters*

---

**Description**

Generate a set of anchored parameters.

It is important that the anchors (i.e. the temperatures used as anchors) encompass the highest and lowest temperatures that are present in nests.

The value for each anchor is  $R * 1E5$ . The 1E5 factor allows to value to be close to unity.

**Usage**

```
GenerateAnchor(
  temperatures = NULL,
  nests = NULL,
  parameters = NULL,
  number.anchors = 7
)
```

**Arguments**

temperatures	A vector with temperatures to serve as anchors
nests	Formatted nest data or result object obtained from searchR()
parameters	A set of parameters value
number.anchors	Number of anchors

**Details**

GenerateAnchor Generate a set of anchored parameters

**Value**

A vector with parameters

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
# Example to generate anchored parameters
newp <- GenerateAnchor()
newp <- GenerateAnchor(temperatures=seq(from=20,
  to=35, length.out=7))
newp <- GenerateAnchor(number.anchors=7)
data(nest)
formatted <- FormatNests(nest, previous=NULL)
newp <- GenerateAnchor(nests=formatted)
newp <- GenerateAnchor(nests=formatted, number.anchors=10)
data(resultNest_4p_SSM)
newp <- GenerateAnchor(nests=resultNest_4p_SSM, number.anchors=7)
newp <- GenerateAnchor(nests=resultNest_4p_SSM, temperatures=seq(from=20,
  to=35, length.out=10))
newp <- GenerateAnchor(nests=resultNest_4p_SSM, number.anchors=7)
newp <- c(newp, Scale=1)

## End(Not run)
```

---

GenerateConstInc

*Generate a data.frame with constant incubation temperature and incubation duration*

---

**Description**

Generate a data.frame from constant incubation temperature and incubation duration

**Usage**

```
GenerateConstInc(
  durations = stop("At least one incubation length must be provided"),
  temperatures = stop("At least one incubation temperature must be provided"),
  names = NULL
)
```

**Arguments**

`durations`      A vector with incubation durations  
`temperatures`    A vector with incubation temperatures  
`names`            A vector of column names

**Details**

GenerateConstInc generates a data.frame with constant incubation temperature and incubation duration

**Value**

A date.frame that can be used with FormatNests()

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
temp_cst <- GenerateConstInc(durations=c(150000, 100100, 100000),
  temperatures=c(28, 30.5, 30.6),
  names=c("T28", "T30.5", "T30.6"))

## End(Not run)
```

---

Generate\_hatchling\_metric

*Generate a data.frame that can be used as test value for searchR()*

---

**Description**

Generate a data.frame that can be used as test value for searchR()

**Usage**

```
Generate_hatchling_metric(
  series = stop("A result object or names of series must be provided"),
  hatchling.metric = NULL,
  previous = NULL
)
```

**Arguments**

series	Name of series or object from searchR()
hatchling.metric	Size or mass at hatching. Will be recycled if necessary
previous	Previous formatted test data

**Details**

Generate\_hatchling\_metric Generate a data.frame that can be used as test value for searchR()

**Value**

A data.frame with size or mass at hatching for each nest

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
testsize1 <- Generate_hatchling_metric(resultNest_4p_SSM)
testsize2 <- Generate_hatchling_metric(series=resultNest_4p_SSM,
hatchling.metric=c(Mean=39.3, SD=1.92))

## End(Not run)
```

---

GRTRN\_MHmcmc

*Metropolis-Hastings algorithm for Embryo Growth Rate Thermal Reaction Norm*

---

**Description**

Run the Metropolis-Hastings algorithm for data.

The number of iterations is  $n.iter+n.adapt+1$  because the initial likelihood is also displayed.

I recommend that  $thin=1$  because the method to estimate SE uses resampling.

If initial point is maximum likelihood,  $n.adapt = 0$  is a good solution.

To get the SE of the point estimates from `result_mcmc <- GRTRN_MHmcmc(result=try)`, use:

```
result_mcmc$SD
```

coda package is necessary for this function.

The parameters `intermediate` and `filename` are used to save intermediate results every 'intermediate' iterations (for example 1000). Results are saved in a file named `filename`.

The parameter `previous` is used to indicate the list that has been save using the parameters `intermediate` and `filename`. It permits to continue a mcmc search.

These options are used to prevent the consequences of computer crash or if the run is very very long and processes with user limited time.

**Usage**

```
GRTRN_MHmcmc(
  result = NULL,
  n.iter = 10000,
  parametersMCMC = NULL,
  n.chains = 1,
  n.adapt = 0,
  thin = 1,
```

```

trace = NULL,
traceML = FALSE,
parallel = TRUE,
adaptive = FALSE,
adaptive.lag = 500,
adaptive.fun = function(x) { ifelse(x > 0.234, 1.3, 0.7) },
intermediate = NULL,
filename = "intermediate.Rdata",
previous = NULL
)

```

### Arguments

result	An object obtained after a SearchR fit
n.iter	Number of iterations for each step
parametersMCMC	A set of parameters used as initial point for searching with information on priors
n.chains	Number of replicates
n.adapt	Number of iterations before to store outputs
thin	Number of iterations between each stored output
trace	TRUE or FALSE or period, shows progress
traceML	TRUE or FALSE to show ML
parallel	If true, try to use several cores using parallel computing
adaptive	Should an adaptive process for SDProp be used
adaptive.lag	Lag to analyze the SDProp value in an adaptive content
adaptive.fun	Function used to change the SDProp
intermediate	Period for saving intermediate result, NULL for no save
filename	If intermediate is not NULL, save intermediate result in this file
previous	Previous result to be continued. Can be the filename in which intermediate results are saved.

### Details

GRTRN\_MHmcmc runs the Metropolis-Hastings algorithm for data (Bayesian MCMC)

### Value

A list with resultMCMC being mcmc.list object, resultLnL being likelihoods and parametersMCMC being the parameters used

### Author(s)

Marc Girondot

## Examples

```

## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "T12H", "DHA", "DHH", "DHL", "Rho25"
#####
pfixed <- c(rK=1.208968)
M0 = 0.3470893
#####
# 4 parameters
#####
x c('DHA' = 109.31113503282113, 'DHH' = 617.80695919563857,
    'T12H' = 306.38890489505093, 'Rho25' = 229.37265815800225)

resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=M0,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), series=1,
embryo.stages="Caretta caretta.SCL")
#####
pMCMC <- TRN_MHmcmc_p(resultNest_4p_SSM, accept=TRUE)
# Take care, it can be very long; several days
resultNest_mcmc_4p_SSM <- GRTRN_MHmcmc(result=resultNest_4p_SSM,
    adaptive = TRUE,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
n.adapt = 0, thin=1, trace=TRUE)
# The SDProp in pMCMC at the beginning of the Markov chain can
# be considered as non-optimal. However, the values at the end
# of the Markov chain are better due to the use of the option
# adaptive = TRUE. Then a good strategy is to run again the MCMC
# with this final set:
pMCMC[, "SDProp"] <- resultNest_mcmc_4p_SSM$parametersMCMC$SDProp.end
# Also, take the set of parameters fitted as maximim likelihood
# as initial set of value
pMCMC[, "Init"] <- as.parameters(resultNest_mcmc_4p_SSM)
# Then I run again the MCMC; it will ensure to get the optimal distribution
resultNest_mcmc_4p_SSM <- GRTRN_MHmcmc(result=resultNest_4p_SSM,
    adaptive = TRUE,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
n.adapt = 0, thin=1, trace=TRUE)

data(resultNest_mcmc_4p_SSM)
out <- as.mcmc(resultNest_mcmc_4p_SSM)
# This out can be used with coda package
# Test for stationarity and length of chain
require(coda)
heidel.diag(out)

```



```

raftery.diag(out)
# plot() can use the direct output of GRTRN_MHmcmc() function.
plot(resultNest_mcmc_4p_SSM, parameters=1, xlim=c(0,550))
plot(resultNest_mcmc_4p_SSM, parameters=3, xlim=c(290,320))
# summary() permits to get rapidly the standard errors for parameters
# They are store in the result also.
se <- result_mcmc_4p_SSM$SD
# the confidence interval is better estimated by:
apply(out[[1]], 2, quantile, probs=c(0.025, 0.975))
# The use of the intermediate method is as followed;
# Here the total mcmc iteration is 10000, but every 1000, intermediate
# results are saved in file intermediate1000.Rdata:
resultNest_mcmc_4p_SSM <- GRTRN_MHmcmc(result=resultNest_4p_SSM,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
n.adapt = 0, thin=1, trace=TRUE,
intermediate=1000, filename="intermediate1000.Rdata")
# If run has been stopped for any reason, it can be resumed with:
resultNest_mcmc_4p_SSM <- GRTRN_MHmcmc(previous="intermediate1000.Rdata")
# Example to use of the epsilon parameter to get confidence level
resultNest_4p_epsilon <- resultNest_4p
resultNest_4p_epsilon$fixed.parameters <- c(resultNest_4p_epsilon$par,
resultNest_4p_epsilon$fixed.parameters)
resultNest_4p_epsilon$par <- c(epsilon = 0)
pMCMC <- TRN_MHmcmc_p(resultNest_4p_epsilon, accept = TRUE)
resultNest_mcmc_4p_epsilon <- GRTRN_MHmcmc(result = resultNest_4p_epsilon,
n.iter = 10000, parametersMCMC = pMCMC,
n.chains = 1, n.adapt = 0, thin = 1, trace = TRUE, parallel = TRUE)
data(resultNest_mcmc_4p_epsilon)
plot(resultNest_mcmc_4p_epsilon, parameters="epsilon", xlim=c(-11, 11), las=1)
plotR(resultNest_4p_epsilon, SE=c(epsilon = unname(resultNest_mcmc_4p_epsilon$SD)),
ylim=c(0, 3), las=1)

## End(Not run)

```

---

HatchingSuccess.fit     *Fit a hatching success model to data using maximum likelihood*

---

## Description

Set of functions to study the hatching success.

The first version of the model was published in:

Laloë, J.-O., Monsinjon, J., Gaspar, C., Touron, M., Genet, Q., Stubbs, J., Girondot, M. & Hays, G.C. (2020) Production of male hatchlings at a remote South Pacific green sea turtle rookery: conservation implications in a female-dominated world. *Marine Biology*, 167, 70.

The version available here is enhanced by using a double flexit model rather than a double logistic model. The flexit model is described here:

Abreu-Grobois, F.A., Morales-Mérida, B.A., Hart, C.E., Guillon, J.-M., Godfrey, M.H., Navarro, E. & Girondot, M. (2020) Recent advances on the estimation of the thermal reaction norm for sex ratios. *PeerJ*, 8, e8451.

**Usage**

```
HatchingSuccess.fit(  
  par = NULL,  
  data = stop("data must be provided"),  
  fixed.parameters = NULL,  
  column.Incubation.temperature = "Incubation.temperature",  
  column.Hatched = "Hatched",  
  column.NotHatched = "NotHatched",  
  hessian = TRUE  
)
```

**Arguments**

par	A set of parameters.
data	A dataset in a data.frame with a least three columns: Incubation.temperature, Hatched and NotHatched
fixed.parameters	A set of parameters that must not be fitted.
column.Incubation.temperature	Name of the column with incubation temperatures
column.Hatched	Name of the column with hatched number
column.NotHatched	Name of the column with not hatched number
hessian	Should Hessian matrix be estimated?

**Details**

HatchingSuccess.fit fits a hatching success model to data

**Value**

Return a object of class HatchingSuccess

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

**Examples**

```

## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

g.logistic <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g.logistic$par, data=totalIncubation_Cc)

plot(g.logistic)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10,
         K1.low=1, K2.low=-1, K1.high=1, K2.high=-1,
         MaxHS=0.8)

g.flexit <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g.flexit$par, data=totalIncubation_Cc)

compare_AICc(logistic=g.logistic, flexit=g.flexit)
plot(x=g.logistic, what = c("observations", "ML", "CI"), replicates=10000)

pMCMC <- HatchingSuccess.MHmcmc_p(result = g.logistic, accept = TRUE)
MCMC <- HatchingSuccess.MHmcmc(result = g.logistic, parametersMCMC = pMCMC,
                               n.iter = 100000,
                               adaptive = TRUE)

plot(MCMC, parameters = "S.low")
plot(MCMC, parameters = "S.high")
plot(MCMC, parameters = "P.low")
plot(MCMC, parameters = "deltaP")
plot(MCMC, parameters = "MaxHS")

plot(x=g.logistic, what = c("observations", "ML", "CI"),
     replicates=10000, resultmcmc=MCMC)

## End(Not run)

```

**Description**

Set of functions to study the hatching success.

**Usage**

```
HatchingSuccess.InL(  
  par,  
  data,  
  fixed.parameters = NULL,  
  column.Incubation.temperature = "Incubation.temperature",  
  column.Hatched = "Hatched",  
  column.NotHatched = "NotHatched"  
)
```

**Arguments**

<code>par</code>	A set of parameters.
<code>data</code>	A dataset in a data.frame with a least three columns: <code>Incubation.temperature</code> , <code>Hatched</code> and <code>NotHatched</code>
<code>fixed.parameters</code>	A set of parameters that must not be fitted.
<code>column.Incubation.temperature</code>	Name of the column with incubation temperatures
<code>column.Hatched</code>	Name of the column with hatched number
<code>column.NotHatched</code>	Name of the column with not hatched number

**Details**

`HatchingSuccess.InL` return `-log` likelihood of the data and the parameters

**Value**

Return `-log` likelihood of the data and the parameters

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
        P.low=25, deltaP=10, MaxHS=0.8)

HatchingSuccess.lnL(par=par, data=totalIncubation_Cc)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g$par, data=totalIncubation_Cc)

t <- seq(from=20, to=40, by=0.1)
CIq <- predict(g, temperature=t)

par(mar=c(4, 4, 1, 1), +0.4)
plot(g)

## End(Not run)
```

---

HatchingSuccess.MHmcmc

*Metropolis-Hastings algorithm for hatching success*

---

**Description**

Run the Metropolis-Hastings algorithm for hatching success.

The number of iterations is  $n.iter+n.adapt+1$  because the initial likelihood is also displayed.

I recommend that  $thin=1$  because the method to estimate SE uses resampling.

If initial point is maximum likelihood,  $n.adapt = 0$  is a good solution.

To get the SE from `result_mcmc <- HatchingSuccess.MHmcmc(result=try)`, use:

`result_mcmc$BatchSE` or `result_mcmc$TimeSeriesSE`

The batch standard error procedure is usually thought to be not as accurate as the time series methods.

Based on Jones, Haran, Caffo and Neath (2005), the batch size should be equal to  $\sqrt{n.iter}$ .

Jones, G.L., Haran, M., Caffo, B.S. and Neath, R. (2006) Fixed Width Output Analysis for Markov chain Monte Carlo, *Journal of the American Statistical Association*, 101:1537-1547.

coda package is necessary for this function.

The parameters `intermediate` and `filename` are used to save intermediate results every '`intermediate`' iterations (for example 1000). Results are saved in a file of name `filename`.

The parameter `previous` is used to indicate the list that has been save using the parameters `intermediate` and `filename`. It permits to continue a mcmc search.

These options are used to prevent the consequences of computer crash or if the run is very very long

and processes at time limited.

### Usage

```
HatchingSuccess.MHmcmc(
  result = stop("Give a result of HatchingSuccess.fit()"),
  n.iter = 10000,
  parametersMCMC = NULL,
  n.chains = 1,
  n.adapt = 0,
  thin = 1,
  trace = FALSE,
  traceML = FALSE,
  batchSize = sqrt(n.iter),
  adaptive = FALSE,
  adaptive.lag = 500,
  adaptive.fun = function(x) { ifelse(x > 0.234, 1.3, 0.7) },
  intermediate = NULL,
  filename = "intermediate.Rdata",
  previous = NULL
)
```

### Arguments

result	An object obtained after a SearchR fit
n.iter	Number of iterations for each step
parametersMCMC	A set of parameters used as initial point for searching with information on priors
n.chains	Number of replicates
n.adapt	Number of iterations before to store outputs
thin	Number of iterations between each stored output
trace	TRUE or FALSE or period, shows progress
traceML	TRUE or FALSE to show ML
batchSize	Number of observations to include in each batch fo SE estimation
adaptive	Should an adaptive process for SDProp be used
adaptive.lag	Lag to analyze the SDProp value in an adaptive content
adaptive.fun	Function used to change the SDProp
intermediate	Period for saving intermediate result, NULL for no save
filename	If intermediate is not NULL, save intermediate result in this file
previous	Previous result to be continued. Can be the filename in which intermediate results are saved.

### Details

HatchingSuccess.MHmcmc runs the Metropolis-Hastings algorithm for hatching success (Bayesian MCMC)

**Value**

A list with resultMCMC being mcmc.list object, resultLnL being likelihoods and parametersMCMC being the parameters used

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)
pMCMC <- HatchingSuccess.MHmcmc_p(g, accept=TRUE)
mcmc <- HatchingSuccess.MHmcmc(result=g, parameters = pMCMC,
                              adaptive=TRUE, n.iter=100000, trace=1000)

## End(Not run)
```

---

HatchingSuccess.MHmcmc\_p

*Generates set of parameters to be used with HatchingSuccess.MHmcmc()*

---

**Description**

Interactive script used to generate set of parameters to be used with HatchingSuccess.MHmcmc().

**Usage**

```
HatchingSuccess.MHmcmc_p(
  result = NULL,
  parameters = NULL,
```

```

    fixed.parameters = NULL,
    accept = FALSE
  )

```

### Arguments

result	An object obtained after a <code>HatchingSuccess.fit()</code> fit
parameters	A set of parameters. Replace the one from result
fixed.parameters	A set of fixed parameters. Replace the one from result
accept	If TRUE, the script does not wait user information

### Details

`HatchingSuccess.MHmcmc_p` generates set of parameters to be used with `HatchingSuccess.MHmcmc()`

### Value

A matrix with the parameters

### Author(s)

Marc Girondot

### See Also

Other Hatching success: [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

### Examples

```

## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)
pMCMC <- HatchingSuccess.MHmcmc_p(g, accept=TRUE)
mcmc <- HatchingSuccess.MHmcmc(result=g, parameters = pMCMC,
                              adaptive=TRUE, n.iter=100000, trace=1000)

## End(Not run)

```



---

HatchingSuccess.model *Return the hatching success according the set of parameters and temperatures*

---

## Description

Set of functions to study the hatching success.

## Usage

```
HatchingSuccess.model(par, temperature)
```

## Arguments

par	A set of parameters.
temperature	A vector of temperatures.

## Details

HatchingSuccess.model returns the hatching success according the set of parameters and temperatures

## Value

Return the hatching success according the set of parameters and temperatures

## Author(s)

Marc Girondot

## See Also

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

## Examples

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)
```

```
HatchingSuccess.lnL(par=par, data=totalIncubation_Cc)
g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)
HatchingSuccess.lnL(par=g$par, data=totalIncubation_Cc)
plot(g)
## End(Not run)
```

---

hist.Nests

*Show the histogram of temperatures with set of nests*

---

### Description

Show the histogram of temperatures with set of nests `hist(data)`

### Usage

```
## S3 method for class 'Nests'
hist(x, series = "all", ...)
```

### Arguments

<code>x</code>	Data formatted using <code>formatdata</code> .
<code>series</code>	Series to be used, logical (TRUE ou FALSE), numbers or names. If "all", all series are used.
<code>...</code>	Parameters used by <code>hist</code> function

### Details

`hist.Nests` shows the histogram of temperatures with set of nests

### Value

A list with an histogram object with information on histogram or NULL if no series was selected and the complete set of temperatures used.

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
h <- hist(formated, series="all")

## End(Not run)
```

---

hist.NestsResult      *Show the histogram of temperatures with set of nests*

---

**Description**

Show the histogram of temperatures with set of nests hist(data)

**Usage**

```
## S3 method for class 'NestsResult'
hist(x, series = "all", ...)
```

**Arguments**

x	Results obtained after searchR
series	Series to be used, logical (TRUE ou FALSE), numbers or names. If "all", all series are used.
...	Parameters used by hist function (example main="Title")

**Details**

hist.NestsResult shows the histogram of temperatures with set of nests

**Value**

A list with an histogram object with information on histogram or NULL if no series was selected and the complete set of temperatures used.

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**Examples**

```
## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
h <- hist(resultNest_4p_SSM, series=c(1:5))

## End(Not run)
```

**Description**

This function calculates many statistics about nests.

The `embryo.stages` is a named vector with relative size as compared to final size at the beginning of the stage. Names are the stages.

For example for SCL in *Caretta caretta*:

```
embryo.stages=structure(c(8.4, 9.4, 13.6, 13.8, 18.9, 23.5, 32.2, 35.2, 35.5, 38.5)/39.33),
.Names = c("21", "22", "23", "24", "25", "26", "27", "28", "29", "30", "31"))
```

indicates that the stages 21 begins at the relative size of 8.4/39.33 as compared to the final size.

Series can be indicated as the name of the series, their numbers or series or succession of TRUE or FALSE. "all" indicates that all series must be analyzed.

The likelihood object is just the total likelihood of the data in the model.

If one parameter is named "pipping\_emergence" it is used as the number of days between pipping and emergence to calculate the 1/3 and 2/3 of incubation.

The summary object is a data.frame composed of these elements with the suffix `.mean`, `.se` or `.quantile_x` with x from the parameter probs.

- `Temperature.max` Maximum temperature recorded during incubation
- `TimeWeighted.temperature` Average temperature during all incubation
- `GrowthWeighted.temperature` Average temperature weighted by the actual growth during all incubation
- `TimeWeighted.GrowthRateWeighted.temperature` Average temperature weighted by the growth rate during all incubation
- `TSP.TimeWeighted.temperature` Average temperature during the TSP
- `TSP.GrowthWeighted.temperature` Average temperature weighted by the actual growth during the TSP
- `TSP.TimeWeighted.GrowthRateWeighted.temperature` Average temperature weighted by the growth rate during the TSP
- `TSP.TimeWeighted.STRNWeighted.temperature` Average temperature weighted by the thermal reaction norm of sexualization during the TSP
- `TSP.GrowthWeighted.STRNWeighted.temperature` Average temperature weighted by actual growth and the thermal reaction norm of sexualization during the TSP
- `TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.temperature` Average temperature weighted by growth rate and the thermal reaction norm of sexualization during the TSP
- `TSP.length` TSP duration
- `TSP.begin` Beginning of the TSP
- `TSP.end` End of the TSP
- `TSP.PM.GrowthWeighted` Average of male probability for each temperature weighted by actual growth during the TSP

- `TSP.PM.TimeWeighted.GrowthRateWeighted` Average of male probability for each temperature weighted by growth rate during the TSP
- `TSP.PM.TimeWeighted` Average of male probability for each temperature during the TSP
- `Incubation.length` Incubation length duration
- `MiddleThird.length` Middle third incubation duration
- `MiddleThird.begin` Beginning of the middle third incubation duration
- `MiddleThird.end` End of the middle third incubation duration
- `MiddleThird.TimeWeighted.temperature` Average temperature during the middle third incubation
- `MiddleThird.GrowthWeighted.temperature` Average temperature weighted by the actual growth during the middle third incubation
- `MiddleThird.TimeWeighted.GrowthRateWeighted.temperature` Average temperature weighted by the growth rate during the middle third incubation
- `TSP.TimeWeighted.sexratio` Sex ratio based on average temperature during the TSP
- `TSP.GrowthWeighted.sexratio` Sex ratio based on average temperature weighted by the actual growth during the TSP
- `TSP.TimeWeighted.GrowthRateWeighted.sexratio` Sex ratio based on average temperature weighted by the growth rate during the TSP
- `TSP.TimeWeighted.STRNWeighted.sexratio` Sex ratio based on average temperature weighted by the thermal reaction norm of sexualization during the TSP
- `TSP.GrowthWeighted.STRNWeighted.sexratio` Sex ratio based on average temperature weighted by the actual growth and thermal reaction norm of sexualization during the TSP
- `TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio` Sex ratio based on average temperature weighted by the growth rate and the thermal reaction norm of sexualization during the TSP
- `MiddleThird.TimeWeighted.sexratio` Sex ratio based on average temperature during the middle third incubation
- `MiddleThird.GrowthWeighted.sexratio` Sex ratio based on average temperature weighted by actual growth during the middle third incubation
- `MiddleThird.TimeWeighted.GrowthRateWeighted.sexratio` Sex ratio based on average temperature weighted by growth rate during the middle third incubation
- `TimeWeighted.sexratio` Sex ratio based on average temperature during all incubation
- `GrowthWeighted.sexratio` Sex ratio based on average temperature weighted by actual growth during all incubation
- `TimeWeighted.GrowthRateWeighted.sexratio` Sex ratio based on average temperature weighted by growth rate during all incubation

If out is equal to summary, the return is a list with:

- `summary` is a `data.frame` with statistics for each nest.
- `dynamic.metric` object is a list composed of `data.frames` with the dynamics of growth for each nest. It showed only temperatures from original dataset.

- `summary.dynamic.metric` is a data.frame with the following columns with the suffix `.mean`, `.se` or `.quantile_x` with `x` from the parameter `probs`.

If `out` is equal to `metric`, the return is a list with:

- `dynamic.metric` object is a list composed of data.frames with the dynamics of growth for each nest
- `indices.dynamic.metric` is a data.frame with the following columns.

The object `summary.dynamic.metric` or `indices.dynamic.metric` is a data.frame with the following columns:

- `series` Name of the series
- `metric.begin.tsp` Metric at the beginning of TSP
- `metric.end.tsp` Metric at the end of TSP
- `hatchling.metric.mean` Average expected size of hatchlings
- `hatchling.metric.sd` standard deviation of expected size of hatchlings
- `time.begin.tsp` Time at the beginning of TSP
- `time.end.tsp` Time at the end of TSP
- `time.begin.middlethird` Time at the beginning of the middle third incubation
- `time.end.middlethird` Time at the end of the middle third incubation
- `stop.at.hatchling.metric` Take the value of NA if `stop.at.hatchling.metric` was FALSE. TRUE if at least one incubation series was longer than hatchling size and FALSE at contrary
- `stop.at.hatchling.metric` Take the value of NA if `stop.at.hatchling.metric` was FALSE. TRUE if at least one incubation series was longer than hatchling size and FALSE at contrary
- `stop.at.hatchling.metric` Take the value of NA if `stop.at.hatchling.metric` was FALSE. TRUE if at least one incubation series was longer than hatchling size and FALSE at contrary
- `stop.at.hatchling.metric` Take the value of NA if `stop.at.hatchling.metric` was FALSE. TRUE if at least one incubation series was longer than hatchling size and FALSE at contrary
- `stop.at.hatchling.metric` Take the value of NA if `stop.at.hatchling.metric` was FALSE. TRUE if at least one incubation series was longer than hatchling size and FALSE at contrary
- `stop.at.hatchling.metric` Take the value of NA if `stop.at.hatchling.metric` was FALSE. TRUE if at least one incubation series was longer than hatchling size and FALSE at contrary

If you indicate new set of temperatures, you must probably also indicate new `hatchling.metric` values.

Note: four species have predefined embryo stages. `embryo.stages` parameter can take the values:

- `Caretta caretta.SCL`
- `Chelonia mydas.SCL`
- `Emys orbicularis.SCL`
- `Emys orbicularis.mass`
- `Podocnemis expansa.SCL`

- `Generic.ProportionDevelopment`

But remember that mass is not the best proxy to describe the growth of an embryo because it can decrease if the substrate becomes dry.

The progress bar is based on both replicates and timeseries progress. It necessitates the `pbapply` package.

If `replicate.CI` is null or 0, only maximum likelihood is used and no confidence interval is calculated.

If `replicate.CI` is 1, one random value for the parameters is used but no confidence interval is calculated.

In other cases, `replicate.CI` random samples are used to estimate confidence interval.

## Usage

```
info.nests(
  x = NULL,
  parameters = NULL,
  NestsResult = NULL,
  resultmcmc = NULL,
  hessian = NULL,
  GTRN.CI = NULL,
  fixed.parameters = NULL,
  SE = NULL,
  temperatures = NULL,
  integral = NULL,
  derivate = NULL,
  hatchling.metric = NULL,
  stop.at.hatchling.metric = FALSE,
  M0 = NULL,
  series = "all",
  TSP.borders = NULL,
  embryo.stages = NULL,
  TSP.begin = 0,
  TSP.end = 0.5,
  replicate.CI = 0,
  weight = NULL,
  out = "likelihood",
  fill = NULL,
  probs = c(0.025, 0.5, 0.975),
  SexualisationTRN = NULL,
  SexualisationTRN.mcmc = NULL,
  SexualisationTRN.CI = NULL,
  metric.end.incubation = "observed",
  metabolic.heating = 0,
  temperature.heterogeneity = 0,
  progressbar = FALSE,
  warnings = TRUE,
  parallel = TRUE,
  tsd = NULL,
```

```

    tsd.CI = NULL,
    tsd.mcmc = NULL,
    zero = 1e-09
)

```

## Arguments

<code>x</code>	A set of parameters to model the embryo growth thermal reaction norm or a <code>NestsResult</code> object.
<code>parameters</code>	A set of parameters to model the embryo growth thermal reaction norm. It will replace the parameters included in <code>NestsResult</code> (same as <code>x</code> ).
<code>NestsResult</code>	A <code>NestsResult</code> object generated by <code>searchR</code> to model the embryo growth thermal reaction
<code>resultmcmc</code>	A mcmc result for embryo growth thermal reaction norm
<code>hessian</code>	An hessian matrix for embryo growth thermal reaction norm. It will replace the hessian matrix included in <code>NestResult</code> object.
<code>GTRN.CI</code>	How to estimate CI for embryo growth thermal reaction norm; can be <code>NULL</code> , <code>"SE"</code> , <code>"MCMC"</code> , or <code>"Hessian"</code> .
<code>fixed.parameters</code>	A set of fixed parameters to model the embryo growth thermal reaction norm. It will replace the fixed parameters included in <code>NestsResult</code> .
<code>SE</code>	Standard error for each parameter. It will replace the SE in <code>NestsResult</code> . Use <code>SE=NA</code> to remove SE from <code>NestResult</code>
<code>temperatures</code>	Timeseries of temperatures formatted using <code>formatNests()</code> . It will replace the one in <code>NestsResult</code> .
<code>integral</code>	Function used to fit embryo growth: <code>integral.Gompertz</code> , <code>integral.exponential</code> or <code>integral.linear</code> . It will replace the one in <code>NestsResult</code> .
<code>derivate</code>	Function used to fit embryo growth: <code>dydt.Gompertz</code> , <code>dydt.exponential</code> or <code>dydt.linear</code> . It will replace the one in <code>NestsResult</code> .
<code>hatchling.metric</code>	Mean and SD of size of hatchlings. It will replace the one in <code>NestsResult</code> .
<code>stop.at.hatchling.metric</code>	TRUE or FALSE. If TRUE, the model stops when proxy of size reached the mean <code>hatchling.metric</code> size.
<code>M0</code>	Measure of hatchling size proxy at laying date. It will replace the one in <code>NestsResult</code> .
<code>series</code>	The name or number of the series to be estimated.
<code>TSP.borders</code>	The limits of TSP in stages. See <code>embryo.stages</code> parameter.
<code>embryo.stages</code>	The embryo stages. At least <code>TSP.borders</code> stages must be provided to estimate TSP borders. See note.
<code>TSP.begin</code>	Where TSP begin during the stage of beginning? In relative proportion of the stage.
<code>TSP.end</code>	Where TSP begin during the stage of ending? In relative proportion of the stage.



replicate.CI	Number of replicates to estimate CI. See description
weight	Weights of the different nests to estimate likelihood. It will replace the ones in NestsResult.
out	Can take the values of "likelihood", "summary", "metric" or "dynamic".
fill	Number of minutes between two records. Create new one if they do not exist. NULL does not change the time of temperature recordings.
probs	Probabilities for metric quantiles.
SexualisationTRN	A set of parameters used to model sexualisation thermal reaction norm during TSP or a result of STRN()
SexualisationTRN.mcmc	A mcmc object obtained from STRN_MHmcmc() to generate variability for sexualisation thermal reaction norm during TSP
SexualisationTRN.CI	How to estimate CI of sexualisation thermal reaction norm. Can be NULL, "SE", "MCMC", or "Hessian".
metric.end.incubation	The metric at the end of incubation used to calibrate TSP size. Can be "hatching.metric", or "observed".
metabolic.heating	Degrees Celsius to be added at the end of incubation due to metabolic heating.
temperature.heterogeneity	SD of heterogeneity of temperatures. Can be 2 values, sd_low and sd_high and then HelpersMG::r2norm() is used.
progressbar	If FALSE, the progress bar is not shown (useful for using with sweave or knitr)
warnings	If FALSE, does not show warnings
parallel	If TRUE use parallel version for nests estimation
tsd	A object from tsd() that describe the thermal react norm of sex ratio at constant temperatures
tsd.CI	How to estimate CI for sex ratio thermal reaction norm; Can be NULL, "SE", "MCMC", or "Hessian".
tsd.mcmc	A object from tsd_MHmcmc() .
zero	Value to replace 0 or 1.

### Details

Calculate statistics about nests

### Value

Return or the total likelihood or a list with \$metric and \$summary depending on out parameter

### Author(s)

Marc Girondot

## Examples

```
## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
# Some basic calculations to show the advantage of parallel computing
system.time(summary.nests <- info.nests(x=resultNest_4p_SSM, out="summary",
  embryo.stages="Caretta caretta.SCL", replicate.CI=0, parallel=FALSE))
system.time(summary.nests <- info.nests(x=resultNest_4p_SSM, out="summary",
  embryo.stages="Caretta caretta.SCL", replicate.CI=0, parallel=TRUE))
system.time(summary.nests <- info.nests(x=resultNest_4p_SSM, out="summary",
  embryo.stages="Caretta caretta.SCL", replicate.CI=0, parallel=TRUE, progressBar=TRUE))
system.time(summary.nests <- info.nests(x=resultNest_4p_SSM, out="likelihood",
  embryo.stages="Caretta caretta.SCL", replicate.CI=0, parallel=TRUE, progressBar=FALSE))

# By default parallel computing is TRUE but progressBar is FALSE
# When out is "likelihood", it returns only the likelihood
# otherwise, it returns a list with 3 objects "summary",
#       "dynamic.metric", and "summary.dynamic.metric".

summary.nests <- info.nests(resultNest_4p_SSM, out="summary",
  embryo.stages="Caretta caretta.SCL",
  replicate.CI=100,
  resultmcmc=resultNest_mcmc_4p_SSM,
  GTRN.CI="MCMC",
  progressBar=TRUE)

summary.nests <- info.nests(resultNest_4p_SSM,
  embryo.stages="Caretta caretta.SCL",
  out="summary", replicate.CI=100,
  GTRN.CI="Hessian",
  progressBar=TRUE)

summary.nests <- info.nests(resultNest_4p_SSM,
  series = 1,
  embryo.stages="Caretta caretta.SCL",
  out="summary", replicate.CI=100,
  GTRN.CI="SE",
  progressBar=TRUE)

# Example of use of embryo.stages and TSP.borders:
summary.nests <- info.nests(resultNest_4p_SSM, out="summary",
  embryo.stages=c("10"=0.33, "11"=0.33, "12"=0.66, "13"=0.66),
  TSP.borders = c(10, 12),
  replicate.CI=100,
  progressBar=TRUE)

#####
# Sex ratio using Massey et al. method PM
#####

# Massey, M.D., Holt, S.M., Brooks, R.J., Rollinson, N., 2019. Measurement
# and modelling of primary sex ratios for species with temperature-dependent
```

```

# sex determination. J Exp Biol 222, 1-9.

CC_Mediterranean <- subset(DatabaseTSD, RMU=="Mediterranean" &
Species=="Caretta caretta" & (!is.na(Sexed) & Sexed!=0))
tsdL <- with (CC_Mediterranean, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature,
                                equation="logistic", replicate.CI=NULL))

PM <- info.nests(x=resultNest_4p_SSM,
                GTRN.CI="Hessian", tsd.CI="Hessian",
                embryo.stages="Caretta caretta.SCL", replicate.CI=100,
                out="summary", progressbar=TRUE, tsd=tsdL)

plot_errbar(x=PM$summary$TimeWeighted.temperature.mean,
            y=PM$summary$TSP.PM.GrowthWeighted.mean,
            y.minus=PM$summary$TSP.PM.GrowthWeighted.quantile_0.025,
            y.plus=PM$summary$TSP.PM.GrowthWeighted.quantile_0.975,
            xlab="CTE SCL growth",
            ylab="PM Massey et al. 2016", xlim=c(26, 32), ylim=c(0, 1), las=1)

# Relationship between growth and growth rate

infoall.df <- info.nests(x=resultNest_4p_SSM, out="summary",
                        embryo.stages="Caretta caretta.SCL",
                        replicate.CI=100,
                        resultmcmc=resultNest_mcmc_4p_SSM,
                        GTRN.CI="MCMC",
                        progressbar=TRUE)

layout(1)
plot(x=infoall.df$dynamic.metric[[1]][, "Time"],
     y=infoall.df$dynamic.metric[[1]][, "Metric_50%"],
     type="l", las=1, bty="n",
     xlab="Time in minute", ylab="Growth", ylim=c(0, 39), xlim=c(0, 100000))
lines(x=infoall.df$dynamic.metric[[1]][, "Time"],
      y=infoall.df$dynamic.metric[[1]][, "Metric_2.5%"], lty=2)
lines(x=infoall.df$dynamic.metric[[1]][, "Time"],
      y=infoall.df$dynamic.metric[[1]][, "Metric_97.5%"], lty=2)

## End(Not run)

```

---

integral.exponential *Return the derivative of the exponential function*

---

### Description

Return the derivative of the exponential function  
integral.exponential(t, size, parms)

**Usage**

```
integral.exponential(t, size, parms)
```

**Arguments**

t	The time in any unit
size	The current size
parms	A vector with alpha and K values being c(alpha=x1, K=x2). K is not used.

**Details**

integral.exponential returns the derivative of the exponential function.

**Value**

A list with the derivative

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(306.174998729436, 333.708348843241,
299.856306141849, 149.046870203155),
.Names = c("DHA", "DHH", "T12H", "Rho25"))
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_exponential <- searchR(parameters=x, fixed.parameters=NULL,
temperatures=formatd, derivate=dydt.exponential, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))

## End(Not run)
```

---

integral.Gompertz      *Return the result of the Gompertz function*

---

### Description

Return the result of the Gompertz function as a data.frame with two columns, time and metric  
 integral.Gompertz(t, size, parms)

### Usage

```
integral.Gompertz(t, size, parms)
```

### Arguments

t	The time in any unit
size	The current size
parms	A vector with alpha and K values being c(alpha=x1, K=x2)

### Details

integral.Gompertz returns the derivative of the Gompertz function.

### Value

A list with the derivative

### Author(s)

Marc Girondot

### Examples

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(118.768297442004, 475.750095909406, 306.243694918151,
116.055824800264), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)
```

```
## End(Not run)
```

---

```
integral.linear      Return the linear function
```

---

### Description

Return the derivative of the linear function  
 integral.linear(t, size, parms)

### Usage

```
integral.linear(t, size, parms)
```

### Arguments

t	The time in any unit
size	The current size
parms	A vector with alpha being c(alpha=x1)

### Details

integral.linear returns the linear function.

### Value

A list with the derivative

### Author(s)

Marc Girondot

### Examples

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(306.174998729436, 333.708348843241, 299.856306141849,
149.046870203155), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_linear <- searchR(parameters=x, fixed.parameters=NULL,
temperatures=formatted, derivate=dydt.linear, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
```

```
## End(Not run)
```

---

likelihoodR	<i>Estimate the likelihood of a set of parameters for nest incubation data</i>
-------------	--

---

## Description

Estimate the likelihood of a set of parameters for nest incubation data

## Usage

```
likelihoodR(
  result = NULL,
  parameters = NULL,
  fixed.parameters = NULL,
  temperatures = NULL,
  integral = NULL,
  derivate = NULL,
  hatchling.metric = NULL,
  M0 = NULL,
  hessian = FALSE,
  weight = NULL,
  parallel = TRUE,
  echo = TRUE
)
```

## Arguments

result	A object obtained after searchR or likelihoodR
parameters	A set of parameters
fixed.parameters	A set of parameters that will not be changed
temperatures	Timeseries of temperatures
integral	Function used to fit embryo growth: integral.Gompertz, integral.exponential or integral.linear
derivate	Function used to fit embryo growth: dydt.Gompertz, dydt.exponential or dydt.linear. It will replace the one in NestsResult.
hatchling.metric	Mean and SD of size of hatchlings
M0	Measure of hatchling size or mass proxi at laying date
hessian	If TRUE, the hessian matrix is estimated and the SE of parameters estimated.
weight	A named vector of the weight for each nest for likelihood estimation
parallel	If true, try to use several cores using parallel computing.
echo	If FALSE, does not display the result.

**Details**

likelihoodR estimates the likelihood of a set of parameters for nest incubation data

**Value**

A result object

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
# K for Gompertz must be set as fixed parameter or being a constant K
# or relative to the hatchling size rK
x <- structure(c(118.768297442004, 475.750095909406, 306.243694918151,
116.055824800264), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)
# K or rK are not used for integral.linear or integral.exponential
LresultNest_4p <- likelihoodR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)
LresultNest_4p <- likelihoodR(result=resultNest_4p_SSM)

## End(Not run)
```

---

logLik.HatchingSuccess

*Return -log L of a fit*

---

**Description**

Set of functions to study the hatching success.

**Usage**

```
## S3 method for class 'HatchingSuccess'
logLik(object, ...)
```



**Arguments**

object            The return of a fit done with fitHS.  
 ...              Not used

**Details**

logLik.HatchingSuccess returns  $-\log L$  of a fit

**Value**

Return  $-\log L$  of a fit

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

HatchingSuccess.lnL(par=par, data=totalIncubation_Cc)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g$par, data=totalIncubation_Cc)

t <- seq(from=20, to=40, by=0.1)
CIq <- predict(g, temperature=t)

par(mar=c(4, 4, 1, 1), +0.4)
plot(g)

## End(Not run)
```

---

logLik.NestsResult      *Return Log Likelihood of a fit generated by searchR*

---

### Description

Return Log Likelihood of a fit generated by searchR

### Usage

```
## S3 method for class 'NestsResult'  
logLik(object, ...)
```

### Arguments

object	A result file generated by searchR
...	Not used

### Details

logLik.NestsResult Return Log Likelihood of a fit

### Value

The Log Likelihood value of the fitted model and data

### Author(s)

Marc Girondot

### Examples

```
## Not run:  
library(embryogrowth)  
data(resultNest_4p_SSM)  
logLik(resultNest_4p_SSM)  
AIC(resultNest_4p_SSM)  
  
## End(Not run)
```

---

logLik.STRN	<i>Return Log Likelihood of a fit generated by STRN</i>
-------------	---

---

**Description**

Return Log Likelihood of a fit generated by STRN

**Usage**

```
## S3 method for class 'STRN'  
logLik(object, ...)
```

**Arguments**

object	A result file generated by STRN
...	Not used

**Details**

logLik.STRN Return Log Likelihood of a fit

**Value**

The Log Likelihood value of the fitted model and data

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:  
library(embryogrowth)  
data(resultNest_4p_SSM)  
logLik(resultNest_4p_SSM)  
AIC(resultNest_4p_SSM)  
  
## End(Not run)
```

---

`logLik.tsd`*Return Log Likelihood of a fit generated by tsd*

---

**Description**

Return Log Likelihood of a fit generated by tsd. The object has 3 attributes: nall, and nobs the number of observations, df, the number of fitted parameters.

**Usage**

```
## S3 method for class 'tsd'  
logLik(object, ...)
```

**Arguments**

<code>object</code>	A result file generated by tsd
<code>...</code>	Not used

**Details**

logLik.tsd Return Log Likelihood of a fit

**Value**

The Log Likelihood value of the fitted model and data

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:  
library(embryogrowth)  
m <- c(10, 14, 7, 4, 3, 0, 0)  
f <- c(0, 1, 2, 4, 15, 10, 13)  
t <- c(25, 26, 27, 28, 29, 30, 31)  
result <- tsd(males=m, females=f, temperatures=t)  
logLik(result)  
AIC(result)  
  
## End(Not run)
```

---

movement	<i>Analyze movement recorded within a nest with an accelerometer datalogger</i>
----------	---

---

**Description**

This function is used to evaluate significant movement within a nest.

**Usage**

```
movement(  
  x = stop("data.frame must be provided"),  
  col.time = "Time",  
  col.x = "x",  
  col.y = "y",  
  col.z = "z",  
  NumberRecordBeforeEmergence = 1900,  
  k = 4,  
  Windowsize = 15  
)
```

**Arguments**

x	A data.frame with 4 columns, one for time and three for x, y, and z position
col.time	Name of the column with time
col.x	Name of the column with x positions
col.y	Name of the column with y positions
col.z	Name of the column with z positions
NumberRecordBeforeEmergence	Number of records in quiet period
k	Factor to multiply SD to prevent false positive detection
Windowsize	Number of records used for moving average

**Details**

movement is a function that permits to analyze movement datalogger

**Value**

The function will return a list

**Author(s)**

Marc Girondot

**See Also**

Other Data loggers utilities: [calibrate.datalogger\(\)](#), [uncertainty.datalogger\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
mv <- movement(x=dataf,
               col.time="Time",
               col.x="x", col.y="y", col.z="z")

## End(Not run)
```

---

MovingIncubation

*Simulate incubation of a nest with the beginning of incubation varying*

---

**Description**

Simulate incubation of a nest with the beginning varying day by day  
 Temperatures must be in a data.frame with one column (Time) being the time and the second the temperatures (Temperature). A third column can indicate the temperature at the end of incubation (Temperature.end.incubation). Do not use FormatNests() for this dataframe.

**Usage**

```
MovingIncubation(
  NestsResult = NULL,
  resultmcmc = NULL,
  GTRN.CI = "Hessian",
  tsd = NULL,
  tsd.CI = NULL,
  tsd.mcmc = NULL,
  SexualisationTRN = NULL,
  SexualisationTRN.CI = "Hessian",
  SexualisationTRN.mcmc = NULL,
  temperatures.df = stop("A data.frame must be provided"),
  temperature.heterogeneity = 0,
  metabolic.heating = 0,
  average.incubation.duration = 60 * 1440,
  max.time = 100 * 24 * 60,
  skip = 1,
  parameters = NULL,
  fixed.parameters = NULL,
  SE = NULL,
  hessian = NULL,
  integral = NULL,
  derivate = NULL,
```

```

    hatchling.metric = NULL,
    M0 = NULL,
    embryo.stages = "Caretta caretta.SCL",
    TSP.borders = c(21, 26),
    TSP.begin = 0,
    TSP.end = 0.5,
    replicate.CI = 1,
    parallel = TRUE,
    progressbar = TRUE
  )

```

### Arguments

<code>NestsResult</code>	A result file generated by searchR
<code>resultmcmc</code>	A mcmc result. Will be used rather than SE if provided.
<code>GTRN.CI</code>	How to estimate CI for embryo growth thermal reaction norm; can be NULL, "SE", "MCMC", or "Hessian".
<code>tsd</code>	A object from <code>tsd()</code> that describe the thermal react norm of sex ratio at constant temperatures
<code>tsd.CI</code>	How to estimate CI for sex ratio thermal reaction norm; Can be NULL, "SE", "MCMC", or "Hessian".
<code>tsd.mcmc</code>	A object from <code>tsd_MHmcmc()</code>
<code>SexualisationTRN</code>	A model for sexualisation thermal reaction norm during TSP obtained using <code>STRN()</code>
<code>SexualisationTRN.CI</code>	How to estimate CI of sexualisation thermal reaction norm. Can be NULL, "SE", "MCMC", or "Hessian".
<code>SexualisationTRN.mcmc</code>	MCMC object for STRN.
<code>temperatures.df</code>	A data.frame with 2 or 3 columns: Times, Temperatures and Temperatures.end.incubation (facultative)
<code>temperature.heterogeneity</code>	SD of heterogeneity of temperatures. Can be 2 values, <code>sd_low</code> and <code>sd_high</code> and then <code>HelpersMG::r2norm()</code> is used.
<code>metabolic.heating</code>	Degrees Celsius to be added at the end of incubation due to metabolic heating
<code>average.incubation.duration</code>	The average time to complete incubation (not used if metabolic heating is setup)
<code>max.time</code>	Maximum time of incubation
<code>skip</code>	Number of data to skip between two runs
<code>parameters</code>	A set of parameters if result is not provided.
<code>fixed.parameters</code>	Another set of parameters if result is not provided.

SE	Standard error for each parameter if not present in result is not provided
hessian	A hessian matrix
integral	Function used to fit embryo growth: integral.Gompertz, integral.exponential or integral.linear
derivate	Function used to fit embryo growth: dydt.Gompertz, dydt.exponential or dydt.linear. It will replace the one in NestsResult.
hatchling.metric	Mean and SD of size of hatchlings as a vector ie hatchling.metric=c(Mean=xx, SD=yy)
M0	Measure of hatchling size proxi at laying date
embryo.stages	The embryo stages. At least TSP.borders stages must be provided to estimate TSP length
TSP.borders	The limits of TSP
TSP.begin	Where TSP begin during the stage of beginning? In relative proportion of the stage.
TSP.end	Where TSP begin during the stage of ending? In relative proportion of the stage.
replicate.CI	Number of randomizations to estimate CI
parallel	Should parallel computing be used. TRUE or FALSE
progressbar	Should a progress bar be shown ? TRUE or FALSE

### Details

MovingIncubation simulate incubation of a nest with the beginning varying day by day

### Value

A dataframe with informations about thermosensitive period length and incubation length day by day of incubation

### Author(s)

Marc Girondot

### Examples

```
## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
ti <- seq(from=0, to=(60*24*100), by=60)
temperatures <- rnorm(length(ti), 29, 5)
temperatures <- temperatures+ti/(60*24*100)/2
layout(mat=1:3)
parpre <- par(mar=c(4, 4, 1, 1)+0.4)
plot(ti/(60*24), temperatures, type="l", xlab="Days",
      ylab=expression("Nest temperature in ".*degree*."C"), bty="n", las=1)
# The sexualisation thermal reaction norm is calculated for South Pacific RMU
out <- MovingIncubation(NestsResult=resultNest_4p_SSM,
```



```

temperatures.df=data.frame(Time=ti, Temperature=temperatures),
metabolic.heating = 0,
SexualisationTRN = structure(c(71.922411148397, 613.773055147801,
318.059753164125, 120.327257089974),
.Names = c("DHA", "DHH", "T12H", "Rho25"))
with(out, plot(Time/(60*24), Incubation.length.mean/(60*24),
xlab="Days along the season",
ylab="Incubation duration",
type="l", bty="n", las=1, ylim=c(70, 80)))
with(out, plot(Time/(60*24), TSP.GrowthWeighted.STRNWeighted.temperature.mean,
xlab="Days along the season",
ylab=expression("CTE for sex ratio in "*degree*"C"),
type="l", bty="n", las=1, ylim=c(30, 31)))
par(mar=parpre)
layout(mat=c(1))

## End(Not run)

```

---

nest

*Timeseries of temperatures for nests*


---

## Description

Timeseries of temperatures for nests

## Usage

```
nest
```

## Format

A dataframe with raw data

## Details

Timeseries of temperatures for nests

## Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

## References

Girondot, M. & Kaska, Y. 2014. A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*. 45, 96-102.

**Examples**

```
## Not run:  
library(embryogrowth)  
data(nest)  
  
## End(Not run)
```

---

nobs.HatchingSuccess *Return number of observations of a fit*

---

**Description**

Set of functions to study the hatching success.

**Usage**

```
## S3 method for class 'HatchingSuccess'  
nobs(object, ...)
```

**Arguments**

object	The return of a fit done with fitHS.
...	Not used

**Details**

nobs.NestsResult Return number of observations of a fit

**Value**

Return number of observations of a fit

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

HatchingSuccess.lnL(par=par, data=totalIncubation_Cc)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g$par, data=totalIncubation_Cc)

plot(g)

nobs(g)

## End(Not run)
```

---

nobs.NestsResult	<i>Return number of observations of a fit</i>
------------------	---

---

**Description**

Return number of observations of a fit.  
 This function is used for `bbmle::ICtb()`.

**Usage**

```
## S3 method for class 'NestsResult'
nobs(object, ...)
```

**Arguments**

object	A result file generated by <code>searchR</code>
...	Not used

**Details**

`nobs.NestsResult` Return number of observations of a fit

**Value**

Return number of observations of a fit

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
logLik(resultNest_4p_SSM)
AIC(resultNest_4p_SSM)
nobs(resultNest_4p_SSM)

## End(Not run)
```

---

plot.HatchingSuccess *Plot results of HatchingSuccess.fit() that best describe hatching success*

---

**Description**

Plot the estimates that best describe hatching success.

If replicates is 0, it returns only the fitted model.

If replicates is null and resultmcmc is not null, it will use all the mcmc data.

if replicates is lower than the number of iterations in resultmcmc, it will use sequence of data regularly thined.

**Usage**

```
## S3 method for class 'HatchingSuccess'
plot(
  x,
  xlim = c(20, 40),
  ylim = c(0, 1),
  xlab = "Constant incubation temperatures",
  ylab = "Hatching success",
  bty = "n",
  las = 1,
  col.observations = "red",
  pch.observations = 19,
  cex.observations = 1,
  show.CI.observations = TRUE,
  col.ML = "black",
  lty.ML = 1,
  lwd.ML = 1,
  col.median = "black",
  lty.median = 2,
  lwd.median = 1,
```

```

col.CI = "black",
lty.CI = 3,
lwd.CI = 1,
replicates = NULL,
resultmcmc = NULL,
polygon = TRUE,
color.polygon = rgb(red = 0.8, green = 0, blue = 0, alpha = 0.1),
what = c("observations", "ML", "CI"),
...
)

```

### Arguments

x	A result file generated by HatchingSuccess.fit()
xlim	Range of temperatures
ylim	Hatching success range for y-axis
xlab	x label
ylab	y label
bty	bty graphical parameter
las	las graphical parameter
col.observations	Color of observations
pch.observations	Character used for observation (no observations if NULL)
cex.observations	Size of characters for observations
show.CI.observations	Should the confidence interval of the observations be shown ?
col.ML	Color of the maximum likelihood model
lty.ML	Line type of the maximum likelihood model (no line if NULL)
lwd.ML	Line width of the maximum likelihood model
col.median	Color of the median model
lty.median	Line type of the median model (no line if NULL)
lwd.median	Line width of the mean model
col.CI	Color of the 95% confidence interval lines
lty.CI	Line type of the 95% confidence interval lines (no line if NULL)
lwd.CI	Line width of the 95% confidence interval lines
replicates	Number of replicates to estimate confidence interval
resultmcmc	Results obtained using HatchingSuccess.MHmcmc()
polygon	If TRUE, confidence interval is shown as a polygon
color.polygon	The color used for polygon
what	Indicate what to plot: "observations", "ML", "CI"
...	Parameters for plot()

**Details**

`plot.HatchingSuccess` plot result of `HatchingSuccess.fit()` or `HatchingSuccess.MHmcmc()` that best describe hatching success

**Value**

Nothing

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [predict.HatchingSuccess\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

HatchingSuccess.lnL(par=par, data=totalIncubation_Cc)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g$par, data=totalIncubation_Cc)

plot(g, replicates=0)
plot(g, replicates=10000)

pMCMC <- HatchingSuccess.MHmcmc_p(g, accept=TRUE)
mcmc <- HatchingSuccess.MHmcmc(result=g, parameters = pMCMC,
                              adaptive=TRUE, n.iter=100000, trace=1000)

plot(g, resultmcmc=mcmc)
plot(g, resultmcmc=mcmc, pch.observations=NULL, lty.mean=NULL)

## End(Not run)
```

---

plot.NestsResult      *Plot the embryo growth*

---

### Description

Plot the embryo growth from one or several nests.

The embryo.stages is a named vector with relative size as compared to final size at the beginning of the stage. Names are the stages.

For example for SCL in Caretta caretta:

```
embryo.stages=structure(c(8.4, 9.4, 13.6, 13.8, 18.9, 23.5, 32.2, 35.2, 35.5, 38.5)/39.33),
.Names = c("21", "22", "23", "24", "25", "26", "27", "28", "29", "30", "31"))
```

indicates that the stages 21 begins at the relative size of 8.4/39.33.

Series can be indicated as the name of the series, its number or succession of TRUE or FALSE. "all" indicates that all series must be printed.

show.floritures parameter does not affect show.hatchling.metric option.

Note: Four species have predefined embryo stages. embryo.stages parameter can take the values:

- Caretta caretta.SCL
- Chelonia mydas.SCL
- Emys orbicularis.SCL
- Emys orbicularis.mass
- Podocnemis expansa.SCL
- Generic.ProportionDevelopment

### Usage

```
## S3 method for class 'NestsResult'
plot(
  x,
  ...,
  parameters = NULL,
  fixed.parameters = NULL,
  resultmcmc = NULL,
  hessian = NULL,
  SE = NULL,
  temperatures = NULL,
  integral = NULL,
  derivate = NULL,
  hatchling.metric = NULL,
  stop.at.hatchling.metric = FALSE,
  M0 = NULL,
  weight = NULL,
  series = "all",
  TSP.borders = NULL,
  embryo.stages = NULL,
```

```

STRN = NULL,
TSP.begin = 0,
TSP.end = 0.5,
replicate.CI = 100,
metric.end.incubation = "observed",
col.stages = "blue",
col.PT = "red",
col.TSP = "gray",
col.temperatures = "green",
col.S = "black",
lty.temperatures = 1,
lwd.temperatures = 2,
ylimT = NULL,
ylimS = NULL,
xlim = NULL,
show.stages = TRUE,
show.TSP = TRUE,
show.third = TRUE,
GTRN.CI = NULL,
show.metric = TRUE,
show.fioritures = TRUE,
show.temperatures = TRUE,
show.PT = TRUE,
PT = c(mean = NA, SE = NA),
show.hatchling.metric = TRUE,
add = FALSE,
lab.third = "2nd third of incubation",
at.lab.third = 10,
lab.PT = "PT",
lab.stages = "Stages",
at.lab.TSP = 8,
lab.TSP = "TSP",
mar = c(4, 5, 4, 5) + 0.3,
xlab = "Days of incubation",
ylabT = expression("Temperature in " * degree * "C"),
ylabS = "Embryo metric",
progress = TRUE,
parallel = TRUE
)

```

### Arguments

x	A result file generated by searchR
...	Parameters for plot()
parameters	A set of parameters if result is not provided.
fixed.parameters	Another set of parameters if result is not provided.
resultmcmc	A mcmc result. Will be used rather than SE if provided.



hessian	An Hessian matrix.
SE	Standard error for each parameter if result is not provided, or replace the one in NestsResult. Use SE=NA to remove SE from NestResult
temperatures	Timeseries of temperatures formatted using formatNests(). Will replace the one in result.
integral	Function used to fit embryo growth: integral.Gompertz, integral.exponential or integral.linear
derivate	Function used to fit embryo growth: dydt.Gompertz, dydt.exponential or dydt.linear. It will replace the one in NestsResult.
hatchling.metric	Mean and SD of size of hatchlings
stop.at.hatchling.metric	TRUE or FALSE. If TRUE, the model stops when proxy of size reached the mean hatchling.metric size.
M0	Measure of hatchling size proxi at laying date
weight	Weights of the different nests to estimate likelihood
series	The name or number of the series to be displayed. Only one series can be displayed at a time.
TSP.borders	The limits of TSP in stages. See embryo.stages parameter.
embryo.stages	The embryo stages. At least TSP.borders stages must be provided to estimate TSP borders. See note.
STRN	An object obtained from STRN()
TSP.begin	Where TSP begin during the stage of beginning? In relative proportion of the stage.
TSP.end	Where TSP begin during the stage of ending? In relative proportion of the stage.
replicate.CI	Number of replicates to estimate CI. If 1, no CI is estimated.
metric.end.incubation	The expected metric at the end of incubation. Used to calibrate TSP size. If NULL, take the maximum Mean of the hatchling.metric parameter. If NA, use the actual final size. Can be a vector and is recycled if necessary.
col.stages	The color of the stages
col.PT	The color of the pivotal temperature
col.TSP	The color of the TSP
col.temperatures	The color of the temperatures
col.S	The color of the size or mass. Can be a vector (useful when series="all" option).
lty.temperatures	Type of line for temperatures
lwd.temperatures	Width of line for temperatures
ylimT	Range of temperatures to be displayed
ylimS	Range of size to be displayed

xlim	Range of incubation days to be displayed
show.stages	TRUE or FALSE, does the embryo stages should be displayed?
show.TSP	TRUE or FALSE, does the TSP borders should be displayed?
show.third	TRUE or FALSE, does the first and second third borders should be displayed?
GTRN.CI	How to estimate CI; can be NULL, "SE", "MCMC", or "Hessian"
show.metric	TRUE or FALSE, does the plot of embryo metric is shown?
show.fioritures	If FALSE, set show.PT, show.temperatures, show.stages, show.TSP, show.third to FALSE, GTRN.CI to NULL
show.temperatures	TRUE or FALSE, does the temperatures should be displayed?
show.PT	TRUE or FALSE, does the pivotal temperature should be displayed?
PT	Value for pivotal temperature, mean and SE
show.hatchling.metric	TRUE or FALSE, does the hatchling size should be displayed
add	If TRUE, all the curves are shown on the same graph
lab.third	Label for 2nd third of incubation
at.lab.third	Position of Label for 2nd third of incubation [default=10]; y-lim is scaled by at.lab.third
lab.PT	Label for Pivotal Temperature
lab.stages	Label for Stages
at.lab.TSP	Position of Label for TSP [default=8]; y-lim is scaled by at.lab.third
lab.TSP	Label for the TSP
mar	Parameter mar used for plot
xlab	Label for axis
ylabT	Label for temperature axis
ylabS	Label for size axis
progress	If FALSE, the progress bar is not shown (useful for use with sweave or knitr)
parallel	Should parallel computing be used ? TRUE or FALSE
NestsResult	A NestsResult file generated by searchR

### Details

plot.NestsResult Plot the embryo growth

### Value

Nothing

### Author(s)

Marc Girondot

**Examples**

```

## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), series=1,
      SE=c(DHA=1.396525, DHH=4.101217, T12H=0.04330405, Rho25=1.00479),
      GTRN.CI = "SE", replicate.CI = 100,
      embryo.stages="Caretta caretta.SCL")
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), series=1,
      GTRN.CI = "Hessian", replicate.CI = 100,
      embryo.stages="Caretta caretta.SCL")
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), series=1,
      resultmcmc = resultNest_mcmc_4p_SSM,
      GTRN.CI = "MCMC", replicate.CI = 100,
      embryo.stages="Caretta caretta.SCL")
# to plot all the nest at the same time, use
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45),
      series="all", show.fioritures=FALSE, add=TRUE,
      embryo.stages="Caretta caretta.SCL")
# to use color different for series
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), add=TRUE,
      series="all", show.fioritures=FALSE, col.S=c(rep("black", 5), rep("red", 6)),
      embryo.stages="Caretta caretta.SCL")

# to plot all the temperature profiles

par(mar=c(4, 4, 1, 1))
plot(resultNest_4p_SSM$data[[1]][, 1]/60/24,
      resultNest_4p_SSM$data[[1]][, 2], bty="n",
      las=1, xlab="Days of incubation",
      ylab=expression("Temperatures in "*degree*"C"),
      type="l", xlim=c(0,70),ylim=c(20, 35))

for (i in 2:21) {
  par(new=TRUE)
  plot(resultNest_4p_SSM$data[[i]][, 1]/60/24,
        resultNest_4p_SSM$data[[i]][, 2], bty="n",
        las=1, xlab="", ylab="", type="l", xlim=c(0,70),
        ylim=c(20, 35), axes = FALSE)
}

## End(Not run)

```

plot.tsd

---

*Plot results of tsd() that best describe temperature-dependent sex determination*

---

**Description**

Plot the estimates that best describe temperature-dependent sex determination.

**Usage**

```
## S3 method for class 'tsd'
plot(
  x,
  ...,
  show.observations = TRUE,
  show.model = TRUE,
  males.freq = TRUE,
  show.PTRT = TRUE,
  las.x = 1,
  las.y = 1,
  lab.PT = paste("Pivotal ", x$type),
  resultmcmc = NULL,
  chain = 1,
  l = 0.05,
  replicate.CI = 10000,
  range.CI = 0.95,
  mar = c(4, 4, 4, 1) + 0.4,
  temperatures.plot = seq(from = 25, to = 35, by = 0.1),
  durations.plot = seq(from = 40, to = 70, by = 0.1),
  lab.TRT = paste0("Transitional range of ", x$type, "s l=", x$l * 100, "%"),
  col.TRT = "gray",
  col.TRT.CI = rgb(0.8, 0.8, 0.8, 0.5),
  col.PT.CI = rgb(0.8, 0.8, 0.8, 0.5),
  show.CI = TRUE,
  warn = TRUE
)
```

**Arguments**

<code>x</code>	A result file generated by <code>tsd()</code>
<code>...</code>	Parameters for <code>plot()</code>
<code>show.observations</code>	Should the observations be shown
<code>show.model</code>	Should the model be shown
<code>males.freq</code>	Should the graph uses males relative frequency [TRUE] or females [FALSE]
<code>show.PTRT</code>	Should the P and TRT information be shown
<code>las.x</code>	las parameter for x axis
<code>las.y</code>	las parameter for y axis
<code>lab.PT</code>	Label to describe pivotal temperature
<code>resultmcmc</code>	A result of <code>tsd_MHmcmc()</code>
<code>chain</code>	What chain to be used is <code>resultmcmc</code> is provided
<code>l</code>	Sex ratio limits to define TRT are <code>l</code> and <code>1-l</code>
<code>replicate.CI</code>	<code>replicate.CI</code> replicates from the hessian matrix to estimate CI

range.CI	The range of confidence interval for estimation, default=0.95
mar	The par("mar") parameter
temperatures.plot	Temperatures used for showing curves of sex ratio
durations.plot	Durations used for showing curves of sex ratio
lab.TRT	Label to describe transitional range of temperature
col.TRT	The color of TRT
col.TRT.CI	The color of CI of TRT based on range.CI
col.PT.CI	The color of CI of PT based on range.CI
show.CI	Do the CI for the curve should be shown
warn	Do the warnings must be shown ? TRUE or FALSE

### Details

plot.tsd plot result of tsd() that best describe temperature-dependent sex determination

### Value

Nothing

### Author(s)

Marc Girondot

### References

Girondot, M. 1999. Statistical description of temperature-dependent sex determination using maximum likelihood. *Evolutionary Ecology Research*, 1, 479-486.

Godfrey, M.H., Delmas, V., Girondot, M., 2003. Assessment of patterns of temperature-dependent sex determination using maximum likelihood model selection. *Ecoscience* 10, 265-272.

Hulin, V., Delmas, V., Girondot, M., Godfrey, M.H., Guillon, J.-M., 2009. Temperature-dependent sex determination and global change: are some species at greater risk? *Oecologia* 160, 493-506.

Girondot M., Submitted. On the concept of embryological thermosensitive period for sex determination in reptiles.

### See Also

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD](#), [P\\_TRT\(\)](#), [TSP.list](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

**Examples**

```
## Not run:
CC_AtlanticSW <- subset(DatabaseTSD, RMU=="Atlantic, SW" &
                        Species=="Caretta caretta" & (!is.na(Sexed) & Sexed!=0))
tsdL <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="logistic"))

plot(tsdL)

## End(Not run)
```

---

plotR

---

*Show the fitted growth rate dependent on temperature and its density*


---

**Description**

Show the fitted growth rate dependent on temperature and its density.

The curve "ML quantiles" is based on delta method.

The curve "ML" just shows the fitted model.

The curve "MCMC quantiles" uses the mcmc replicates to build the quantiles.

The curve "MCMC mean-SD" uses the mcmc replicates to build a symmetric credibility interval.

The parameter curve is case insensitive. If only parameters is given, curve must be ML.

**Usage**

```
plotR(
  result = NULL,
  resultmcmc = NULL,
  chain = 1,
  parameters = NULL,
  fixed.parameters = NULL,
  temperatures = NULL,
  curve = "ML quantiles",
  set.par = 1,
  ylim = c(0, 5),
  xlim = c(20, 35),
  hessian = NULL,
  replicate.CI = 1000,
  cex.lab = par("cex"),
  cex.axis = par("cex"),
  scaleY = "auto",
  lty = 1,
  ltyCI = 3,
  lwd = 1,
  lwdCI = 1,
  col = "black",
  col.polygon = "grey",
```

```

polygon = FALSE,
probs = 0.95,
colramp = colorRampPalette(c("white", rgb(red = 0.5, green = 0.5, blue = 0.5))),
bandwidth = c(0.1, 0.01),
pch = "",
main = "",
xlab = expression("Temperature in " * degree * "C"),
ylab = NULL,
bty = "n",
las = 1,
by.temperature = 0.1,
show.density = FALSE,
new = TRUE,
show.hist = FALSE,
ylimH = NULL,
atH = NULL,
ylabH = "Temperature density",
breaks = "Sturges",
log.hist = FALSE,
mar = NULL
)

```

### Arguments

result	A result object or a list of result objects
resultmcmc	A result object from GRTN_MHmcmc() function
chain	The chain to use in resultmcmc
parameters	A set of parameters - Has the priority over result
fixed.parameters	A set of fixed parameters
temperatures	A set of temperatures - Has the priority over result
curve	What curve to show: "MCMC quantiles" or "MCMC mean-SD" based on mcmc or "ML" or "ML quantiles" or "ML mean-SE" for maximum-likelihood. Or "none"
set.par	1 or 2 to designate with set of parameters to show
ylim	Range of values for y-axis
xlim	Range of values for x-axis
hessian	An hessian matrix
replicate.CI	Number of replicates to estimate confidence interval with Hessian if delta method failed
cex.lab	cex value for axis
cex.axis	cex value for axis
scaleY	Scaling factor for y axis or "auto"
lty	The type of lines

ltyCI	The type of lines
lwd	The type of lines
lwdCI	The type of lines
col	The color of the lines
col.polygon	The color of the polygon
polygon	If TRUE, confidence interval is shown as a polygon with color
probs	Confidence or credibility interval to show
colramp	Ramp function accepting an integer as an argument and returning n colors.
bandwidth	numeric vector (length 1 or 2) of smoothing bandwidth(s). If missing, a more or less useful default is used. bandwidth is subsequently passed to function bkde2D.
pch	Character for outliers
main	Title of the graph
xlab	Label for x axis
ylab	Label for y axis
bty	Box around the plot
las	Orientation for labels in y axis
by.temperature	Step to build the temperatures
show.density	TRUE or FALSE for use with Hessian or MCMC
new	Should the graphics be a new one (TRUE) or superimposed to a previous one (FALSE)
show.hist	TRUE or FALSE
ylimH	Scale of histogram using ylimH=c(min, max)
atH	Position of ticks for scale of histogram
ylabH	Label for histogram scale
breaks	See ?hist
log.hist	SHould the y scale for hist is log ?
mar	The value of par("mar"). If null, it will use default depending on show.dist. If NA, does not change par("mar").

### Details

plotR shows the fitted growth rate dependent on temperature and the density of the mcmc

### Value

The value of scaleY to be used with other plotR function

### Author(s)

Marc Girondot



**Examples**

```

## Not run:
library(embryogrowth)
# Note that the confidence interval is not the same for mcmc and ml quantiles
plotR(result = resultNest_4p_SSM,
      resultmcmc=resultNest_mcmc_4p_SSM,
      curve = "MCMC quantiles", ylim=c(0, 8))
plotR(resultNest_4p_SSM, curve="ML quantiles", ylim=c(0, 6))
#####
plotR(resultmcmc=resultNest_mcmc_4p_SSM, ylim=c(0, 10),
      curve = "MCMC quantiles", show.density=TRUE)
#####
plotR(resultmcmc=resultNest_mcmc_4p_SSM,
      curve = "MCMC quantiles", polygon=TRUE)
#####
plotR(resultmcmc=resultNest_mcmc_6p_SSM, ylim=c(0,8),
      curve = "MCMC quantiles", polygon=TRUE, col.polygon = rgb(0, 1, 0, 1))
plotR(resultmcmc=resultNest_mcmc_4p_SSM, ylim=c(0,8),
      curve = "MCMC quantiles", polygon=TRUE, col.polygon = rgb(1, 0, 0, 0.5),
      new=FALSE)
legend("topleft", legend=c("SSM 4 parameters", "SSM 6 parameters"),
      pch=c(15, 15), col=c(rgb(1, 0, 0, 0.5), rgb(0, 1, 0, 1)))
#####
sy <- plotR(resultmcmc=resultNest_mcmc_4p_SSM, ylim=c(0, 8),
      curve = "MCMC quantiles", show.density=FALSE)
plotR(resultmcmc=resultNest_mcmc_6p_SSM, col="red", ylim=c(0, 8),
      curve = "MCMC quantiles", show.density=FALSE,
      new=FALSE, scaleY=sy)
#####
sy <- plotR(result=resultNest_6p_SSM, curve="none",
      scaleY=1E5,
      ylim=c(0, 8),
      show.hist = TRUE, new = TRUE, mar=c(4, 4, 1, 4))
#####
plotR(result=resultNest_6p_SSM, curve="ML",
      ylim=c(0, 8),
      show.hist = TRUE, ylimH=c(0,1), atH=c(0, 0.1, 0.2))
#####
plotR(result = resultNest_4p_SSM, ylim=c(0, 8),
      resultmcmc=resultNest_mcmc_4p_SSM,
      show.density = TRUE,
      curve = "MCMC quantiles")
#####
plotR(resultmcmc=resultNest_mcmc_4p_SSM, ylim=c(0, 8),
      curve = "MCMC quantiles", show.density=TRUE, scaleY=1E5)

## End(Not run)

```

**Description**

Plot the transition function

**Usage**

```
plot_transition(result = NULL, parameters = NULL, sizes = c(0, 40), ...)
```

**Arguments**

result	A result object
parameters	Set of parameters. If both result and parameters are indicated, parameters have priority.
sizes	The range of possible sizes
...	Parameters for plot() such as main= or ylim=

**Details**

plot\_transition show fonction used for transition

**Value**

Nothing

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
data(resultNest_4p_SSM)
# Get a set of parameters without transition
x1 <- resultNest_4p_SSM$par
# Generate a set of parameters with transition
x2 <- switch.transition(x1)
x2 <- x2[names(x2)!="transition_P"]
x2["transition_S"] <- 4
pfixed <- c(rK=2.093313, transition_P=20)
resultNest_4p_transition <- searchR(parameters=x2, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_transition)
# show the model for smallest size
plotR(resultNest_4p_transition, ylim=c(0,0.3))
# show the model for larger sizes
plotR(resultNest_4p_transition, set.par=2, ylim=c(0,0.3))
# plot model for both together
```

```

plotR(resultNest_4p_transition, set.par=c(1,2), ylim=c(0,0.3),
      col=c("red", "black"), legend=list("Initial", "End"))
plot_transition(result=resultNest_4p_transition, las=1, sizes=c(0,40))
compare_AIC(one.model=list(resultNest_4p_SSM), two.models=list(resultNest_4p_transition))
# Note that the model with fitted transition_P is trivial. Embryos grow fast until
# they reach hatchling size and then growth rate becomes null!

## End(Not run)

```

---

predict.HatchingSuccess

*Return prediction based on a model fitted with HatchingSuccess.fit()*

---

### Description

Set of functions to study the hatching success.  
 If replicates is 0, it returns only the fitted model.  
 If replicates is null and resultmcmc is not null, it will use all the mcmc data.  
 if replicates is lower than the number of iterations in resultmcmc, it will use sequence of data regularly thined.

### Usage

```

## S3 method for class 'HatchingSuccess'
predict(
  object,
  ...,
  temperature = NULL,
  probs = c(0.025, 0.5, 0.975),
  replicates = NULL,
  resultmcmc = NULL,
  chain = 1
)

```

### Arguments

object	The return of a fit done with HatchingSuccess.fit().
...	Not used
temperature	A vector of temperatures.
probs	Quantiles.
replicates	Number of replicates to estimate the confidence interval.
resultmcmc	Results obtained using HatchingSuccess.MHmcmc()
chain	Chain to use in resultmcmc

### Details

predict.HatchingSuccess returns prediction based on a model fitted with HatchingSuccessfit()

**Value**

Return a matrix with prediction based on a model fitted with `HatchingSuccess.fit()`

**Author(s)**

Marc Girondot

**See Also**

Other Hatching success: [HatchingSuccess.MHmcmc\\_p\(\)](#), [HatchingSuccess.MHmcmc\(\)](#), [HatchingSuccess.fit\(\)](#), [HatchingSuccess.lnL\(\)](#), [HatchingSuccess.model\(\)](#), [logLik.HatchingSuccess\(\)](#), [nobs.HatchingSuccess\(\)](#), [plot.HatchingSuccess\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
totalIncubation_Cc <- subset(DatabaseTSD,
                             Species=="Caretta caretta" &
                             Note != "Sinusoidal pattern" &
                             !is.na(Total) & Total != 0)

par <- c(S.low=0.5, S.high=0.3,
         P.low=25, deltaP=10, MaxHS=0.8)

HatchingSuccess.lnL(x=par, data=totalIncubation_Cc)

g <- HatchingSuccess.fit(par=par, data=totalIncubation_Cc)

HatchingSuccess.lnL(par=g$par, data=totalIncubation_Cc)

plot(g)

## End(Not run)
```

---

predict.tsd

*Estimate sex ratio according to constant incubation temperature*

---

**Description**

Estimate sex ratio according to constant incubation temperature

The data.frame has the temperatures or durations in columns and the quantiles in rows.

Note that incubation duration is a very bad proxy for sex ratio. See Georges, A., Limpus, C. J. & Stoutjesdijk, R. 1994. Hatchling sex in the marine turtle *Caretta caretta* is determined by proportion of development at a temperature, not daily duration of exposure. *J. Exp. Zool.*, 270, 432-444.

If replicate.CI is 0 or NULL, point estimate for maximum likelihood is returned.

**Usage**

```
## S3 method for class 'tsd'
predict(
  object,
  temperatures = NULL,
  durations = NULL,
  SD.temperatures = NULL,
  SD.durations = NULL,
  resultmcmc = NULL,
  chain = 1,
  replicate.CI = 10000,
  probs = c(0.025, 0.5, 0.975),
  ...
)
```

**Arguments**

object	A result file generated by tsd
temperatures	A vector of temperatures
durations	A vector of durations
SD.temperatures	SD of temperatures
SD.durations	SD of durations
resultmcmc	A result of tsd_MHmcmc()
chain	What chain to be used is resultmcmc is provided
replicate.CI	Number of replicates to estimate CI
probs	The quantiles to be returned, default=c(0.025, 0.5, 0.975)
...	Not used

**Details**

predict.tsd Estimate sex ratio according to constant incubation temperature

**Value**

A data.frame with informations about sex-ratio

**Author(s)**

Marc Girondot

**See Also**

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD](#), [P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
m <- c(10, 14, 7, 4, 3, 0, 0)
f <- c(0, 1, 2, 4, 15, 10, 13)
t <- c(25, 26, 27, 28, 29, 30, 31)
result <- tsd(males=m, females=f, temperatures=t)
plot(result)
predict(result, temperatures=c(25, 31), replicate.CI = 10000)
predict(result, temperatures=c(25, 31), SD.temperatures = c(1, 2), replicate.CI = 10000)
d <- c(72, 70, 65, 63, 62, 60, 59)
result <- tsd(males=m, females=f, durations=d)
predict(result, durations=c(67, 68), replicate.CI = 10000)

## End(Not run)
```

---

P\_TRT

---

*Estimate the transitional range of temperatures based on a set of parameters*


---

**Description**

Estimate the parameters that best describe the thermal reaction norm for sex ratio when temperature-dependent sex determination occurs.

It can be used also to evaluate the relationship between incubation duration and sex ratio.

The parameter  $l$  was defined in Girondot (1999). The TRT is defined from the difference between the two boundary temperatures giving sex ratios of  $l$  and  $1 - l$ .

In Girondot (1999),  $l$  was 0.05 and then the TRT was defined as being the range of temperatures producing from 5 If  $l$  is null, TRT is not estimated and only sex ratio is estimated.

if replicate.CI is null or 0, no replicate is used and only point estimate is done.

Standard error is calculated using resampling based on the Hessian matrix with Cholesky decomposition or using MCMC chain if resultmcmc is provided. In the former case, a replicate.CI random sample of the MCMC results will be used.

**Usage**

```
P_TRT(
  x = NULL,
  resultmcmc = NULL,
  fixed.parameters = NULL,
  chain = 1,
  equation = NULL,
  l = 0.05,
  replicate.CI = NULL,
  temperatures = NULL,
  durations = NULL,
  SD.temperatures = NULL,
```

```

SD.durations = NULL,
probs = c(0.025, 0.5, 0.975),
warn = TRUE
)

```

### Arguments

x	Set of parameters or a result of tsd()
resultmcmc	A result of tsd_MHmcmc()
fixed.parameters	Set of fixed parameters
chain	What chain to be used is resultmcmc is provided
equation	What equation should be used. Must be provided if x is not a result of tsd()
l	Sex ratio limits to define TRT are l and 1-l. If NULL, TRT is not estimated.
replicate.CI	If a result of tsd() is provided, use replicate.CI replicates from the hessian matrix to estimate CI
temperatures	If provided returns the sex ratio and its quantiles for each temperature
durations	If provided returns the sex ratio and its quantiles for each duration
SD.temperatures	SD of temperatures
SD.durations	SD of durations
probs	Probabilities used to estimate quantiles
warn	Do the warnings must be shown ? TRUE or FALSE

### Details

P\_TRT estimates the transitional range of temperatures based on a set of parameters

### Value

A list with a P\_TRT object containing a matrix with lower and higher bounds for TRT, TRT and P and a P\_TRT\_quantiles object with quantiles for each and a sexratio\_quantiles object

### Author(s)

Marc Girondot

### References

- Girondot, M. 1999. Statistical description of temperature-dependent sex determination using maximum likelihood. *Evolutionary Ecology Research*, 1, 479-486.
- Godfrey, M.H., Delmas, V., Girondot, M., 2003. Assessment of patterns of temperature-dependent sex determination using maximum likelihood model selection. *Ecoscience* 10, 265-272.
- Hulin, V., Delmas, V., Girondot, M., Godfrey, M.H., Guillon, J.-M., 2009. Temperature-dependent sex determination and global change: are some species at greater risk? *Oecologia* 160, 493-506.

**See Also**

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

**Examples**

```
## Not run:
library("embryogrowth")
CC_AtlanticSW <- subset(DatabaseTSD, RMU=="Atlantic, SW" &
                        Species=="Caretta caretta" & Sexed!=0)
tsdL <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="logistic"))

P_TRT(tsdL)
P_TRT(tsdL, replicate.CI=1000)
P_TRT(tsdL, replicate.CI=1000, temperatures=20:35)
P_TRT_out <- P_TRT(tsdL, replicate.CI=1000, temperatures=c(T1=20, T2=35))
attributes(P_TRT_out$sexratio_quantiles)$temperatures
P_TRT(tsdL$par, equation="logistic")
pMCMC <- tsd_MHmcmc_p(tsdL, accept=TRUE)
# Take care, it can be very long
result_mcmc_tsd <- tsd_MHmcmc(result=tsdL,
                              parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
                              n.adapt = 0, thin=1, trace=FALSE, adaptive=TRUE)
P_TRT(result_mcmc_tsd, equation="logistic")

## End(Not run)
```

---

resultNest\_3p\_Dallwitz

*Fit using the nest database*

---

**Description**

Fit using the nest database

**Usage**

```
resultNest_3p_Dallwitz
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database



**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot M, Monsinjon J, Guillon J-M (2018) Delimitation of the embryonic thermosensitive period for sex determination using an embryo growth model reveals a potential bias for sex ratio prediction in turtles. *Journal of Thermal Biology* 73: 32-40

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
x <- structure(c(4.88677476830268, 20.4051904475743, 31.5173105860335),
.Names = c("Dallwitz_b1", "Dallwitz_b2", "Dallwitz_b3"))
pfixed <- c(rK=1.208968)
resultNest_3p_Dallwitz <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(result=resultNest_3p_Dallwitz, show.hist = TRUE,
ylim=c(0, 8), curve="ML quantiles")

## End(Not run)
```

---

resultNest\_3p\_Weibull *Result of the fit using the nest database using Weibull function*

---

**Description**

Fit using the nest database using Weibull function. The model is:  
rT <- dweibull(T, shape=abs(parms["k"]),  
scale=abs(parms["lambda"])\*parms["scale"]\*1E-5

**Usage**

```
resultNest_3p_Weibull
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database using Weibull function

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# Weibull model
x <- ChangeSSM(temperatures = (200:350)/10,
               parameters = resultNest_4p_SSM$par,
               initial.parameters = structure(c(73.4009010417375, 304.142079511996,
                                               27.4671689276281),
                                             .Names = c("k", "lambda", "scale")),
               control=list(maxit=1000))
pfixed <- c(rK=2.093313)
resultNest_3p_Weibull <- searchR(parameters=x$par, fixed.parameters=pfixed,
                                temperatures=formatted, integral=integral.Gompertz, M0=1.7,
                                hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_3p_Weibull, ylim=c(0, 3))
plotR(resultNest_3p_Weibull, ylim=c(0, 3), ylimH = c(0, 0.9), show.hist=TRUE)
compare_AIC(SSM=resultNest_4p_SSM, Weibull=resultNest_3p_Weibull)

## End(Not run)
```

---

resultNest\_4p\_normal    *Result of the fit using the nest database using asymmetric normal function*

---

**Description**

Fit using the nest database using asymmetric normal function

**Usage**

```
resultNest_4p_normal
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database using asymmetric normal function

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
x <- ChangeSSM(temperatures = (200:350)/10,
               parameters = resultNest_4p_SSM$par,
               initial.parameters = structure(c(3, 7, 11, 32),
                                             .Names = c("Scale", "sdL", "sdH", "Peak")),
               control=list(maxit=1000))
pfixed <- c(rK=2.093313)
resultNest_4p_normal <- searchR(parameters=x$par, fixed.parameters=pfixed,
                               temperatures=formatted, integral=integral.Gompertz, M0=1.7,
                               hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_4p_normal, ylim=c(0, 3))
plotR(resultNest_4p_normal, ylim=c(0, 3), ylimH = c(0, 0.9), show.hist=TRUE)
compare_AIC(SSM=resultNest_4p_SSM, Asymmetric.normal=resultNest_4p_normal)

## End(Not run)
```

---

resultNest\_4p\_SSM      *Fit using the nest database*

---

**Description**

Fit using the nest database

**Usage**

```
resultNest_4p_SSM
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot M, Monsinjon J, Guillon J-M (2018) Delimitation of the embryonic thermosensitive period for sex determination using an embryo growth model reveals a potential bias for sex ratio prediction in turtles. *Journal of Thermal Biology* 73: 32-40

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
x <- structure(c(109.683413821537, 614.969219372661, 306.386903812694,
  229.003478775323), .Names = c("DHA", "DHH", "T12H", "Rho25"))
pfixed <- c(rK=1.208968)
resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
  temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
  hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(result=resultNest_4p_SSM, show.hist = TRUE,
  ylim=c(0, 8), curve="ML quantiles")

## End(Not run)
```

---

resultNest\_4p\_SSM\_Linear

*Fit using the nest database*

---

**Description**

Fit using the nest database

**Usage**

```
resultNest_4p_SSM_Linear
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database with linear progression

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
pfixed <- NULL
M0 = 0
#####
# 4 parameters SSM
#####
x <- c('DHA' = 64.868697530424186, 'DHH' = 673.18292743646771,
      'T12H' = 400.90952554047749, 'Rho25' = 82.217237723502123)
resultNest_4p_SSM_Linear <- searchR(parameters=x, fixed.parameters=pfixed,
  temperatures=formatted, integral=integral.linear, M0=M0,
  hatchling.metric=c(Mean=39.33, SD=1.92)/39.33)

## End(Not run)
```

---

```
resultNest_4p_transition
```

*Result of the fit using the nest database using transition*

---

**Description**

Fit using the nest database using transition

**Usage**

```
resultNest_4p_transition
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database using transition

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
data(resultNest_4p_transition)

## End(Not run)
```

---

resultNest\_4p\_trigo     *Result of the fit using the nest database using trigonometric function*

---

**Description**

Fit using the nest database using trigonometric function

**Usage**

```
resultNest_4p_trigo
```

**Format**

A list with fitted information about data(nest)

**Details**

Result of the fit using the nest database using trigonometric function

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
x <- ChangeSSM(temperatures = (200:350)/10,
               parameters = resultNest_4p_SSM$par,
               initial.parameters = structure(c(3, 20, 40, 32),
                                             .Names = c("Max", "LengthB", "LengthE", "Peak")),
               control=list(maxit=1000))
pfixed <- c(rK=2.093313)
```

```

resultNest_4p_trigo <- searchR(parameters=x$par, fixed.parameters=pfixed,
                             temperatures=formatted, integral=integral.Gompertz, M0=1.7,
                             hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_4p_trigo, ylim=c(0, 3))
plotR(resultNest_4p_trigo, ylim=c(0, 3), ylimH = c(0, 0.9), show.hist=TRUE)
compare_AIC(SSM=resultNest_4p_SSM, trigonometric=resultNest_4p_trigo)

## End(Not run)

```

---

resultNest\_4p\_weight *Fit using the nest database with weight*

---

### Description

Fit using the nest database with weight

### Usage

```
resultNest_4p_weight
```

### Format

A list with fitted information about data(nest)

### Details

Result of the fit using the nest database with weight

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

### References

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

### Examples

```

## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
w <- weightmaxentropy(formatted, control_plot=list(xlim=c(20,36)))
x <- structure(c(118.768297442004, 475.750095909406, 306.243694918151,
116.055824800264), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)
# K or rK are not used for integral.linear or integral.exponential
resultNest_4p_weight <- searchR(parameters=x,

```

```

fixed.parameters=pfixed, temperatures=formatted,
integral=integral.Gompertz, M0=1.7, hatchling.metric=c(Mean=39.33, SD=1.92),
method = "BFGS", weight=w)
data(resultNest_4p_weight)
plotR(resultNest_4p_weight, ylim=c(0,0.50), xlim=c(15, 35))

## End(Not run)

```

---

resultNest\_5p\_Dallwitz

*Fit using the nest database*

---

### Description

Fit using the nest database

### Usage

```
resultNest_5p_Dallwitz
```

### Format

A list with fitted information about data(nest)

### Details

Result of the fit using the nest database

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

### References

Girondot M, Monsinjon J, Guillon J-M (2018) Delimitation of the embryonic thermosensitive period for sex determination using an embryo growth model reveals a potential bias for sex ratio prediction in turtles. *Journal of Thermal Biology* 73: 32-40

### Examples

```

## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
x <- structure(c(4.91191231405918, 12.7453211281394, 31.2670410811077,
  5.7449376569153, -0.825689964543813), .Names = c("Dallwitz_b1",
  "Dallwitz_b2", "Dallwitz_b3", "Dallwitz_b4", "Dallwitz_b5"))
pfixed <- c(rK=1.208968)
resultNest_5p_Dallwitz <- searchR(parameters=x, fixed.parameters=pfixed,

```



```

temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(result=resultNest_5p_Dallwitz, show.hist = TRUE,
      ylim=c(0, 8), curves="ML quantiles")

## End(Not run)

```

---

resultNest\_6p\_SSM      *Fit using the nest database*

---

### Description

Fit using the nest database

### Usage

```
resultNest_6p_SSM
```

### Format

A list with fitted information about data(nest)

### Details

Result of the fit using the nest database

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

### References

Girondot M, Monsinjon J, Guillon J-M (2018) Delimitation of the embryonic thermosensitive period for sex determination using an embryo growth model reveals a potential bias for sex ratio prediction in turtles. *Journal of Thermal Biology* 73: 32-40

### Examples

```

## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
x <- structure(c(104.954347370542, 3447.10062406071, 661.269363920423,
96.3871849546537, 306.456389026151, 232.105840347154), .Names = c("DHA",
"DHH", "DHL", "DT", "T12L", "Rho25"))
pfixed <- c(rK=1.208968)
resultNest_6p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
hatchling.metric=c(Mean=39.33, SD=1.92))

```

```
plotR(result=resultNest_6p_SSM, show.hist = TRUE,
      ylim=c(0, 8), curve="ML")

## End(Not run)
```

---

```
resultNest_mcmc_4p_SSM
```

*Result of the mcmc using the nest database*

---

## Description

Fit using the nest database

## Usage

```
resultNest_mcmc_4p_SSM
```

## Format

A list of class mcmcComposite with mcmc result for data(nest) with 4 parameters and Gompertz model of growth

## Details

Result of the mcmc using the nest database

## Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

## References

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

## Examples

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(118.431040984352, 498.205702157603, 306.056280989839,
118.189669472381), .Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)
```

```

resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
  temperatures=formatted, integral=integral.Gompertz, M0=1.7,
  test=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)
pMCMC <- TRN_MHmcmc_p(resultNest_4p_SSM, accept=TRUE)
# Take care, it can be very long, sometimes several days
resultNest_mcmc_4p_SSM <- GRTRN_MHmcmc(result=resultNest_4p_SSM,
  adaptive = TRUE,
  parametersMCMC=pMCMC, n.iter=10000, n.chains = 1, n.adapt = 0,
  thin=1, trace=TRUE)
data(resultNest_mcmc_4p_SSM)
1-rejectionRate(as.mcmc(resultNest_mcmc_4p_SSM))
as.parameters(resultNest_mcmc_4p_SSM)
layout(mat=matrix(1:4, nrow = 2))
plot(resultNest_mcmc_4p_SSM, parameters = "all", scale.prior = TRUE, las = 1)
layout(mat=1)
plotR(resultNest_4p_SSM, resultmcmc=resultNest_mcmc_4p_SSM, ylim=c(0,4),
  main="Schoolfield, Sharpe & Magnuson 4-parameters", show.density=TRUE)

## End(Not run)

```

---

```
resultNest_mcmc_4p_SSM_Linear
```

*Result of the mcmc using the nest database*

---

## Description

Fit using the nest database

## Usage

```
resultNest_mcmc_4p_SSM_Linear
```

## Format

A list of class mcmcComposite with mcmc result for data(nest) with 4 parameters and linear model of growth

## Details

Result of the mcmc using the nest database

## Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**Examples**

```

## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
pfixed <- NULL
M0 = 0
#####
# 4 parameters SSM
#####
x <- c('DHA' = 64.868697530424186, 'DHH' = 673.18292743646771,
      'T12H' = 400.90952554047749, 'Rho25' = 82.217237723502123)
resultNest_4p_SSM_Linear <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.linear, M0=M0,
hatchling.metric=c(Mean=39.33, SD=1.92)/39.33)
data(resultNest_4p_SSM_Linear)
pMCMC <- TRN_MHmcmc_p(resultNest_4p_SSM_Linear, accept=TRUE)
# Take care, it can be very long, sometimes several days
resultNest_mcmc_4p_SSM_Linear <- GRTRN_MHmcmc(result=resultNest_4p_SSM_Linear,
adaptive = TRUE,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1, n.adapt = 0,
thin=1, trace=TRUE)
data(resultNest_mcmc_4p_SSM_Linear)
1-rejectionRate(as.mcmc(resultNest_mcmc_4p_SSM_Linear))
as.parameters(resultNest_mcmc_4p_SSM_Linear)
layout(mat=matrix(1:4, nrow = 2))
plot(resultNest_mcmc_4p_SSM_Linear, parameters = "all", scale.prior = TRUE, las = 1)
layout(mat=1)
plotR(resultNest_4p_SSM_Linear, resultmcmc=resultNest_mcmc_4p_SSM_Linear, ylim=c(0,4),
main="Schoolfield, Sharpe & Magnuson 4-parameters", show.density=TRUE)

## End(Not run)

```

---

```
resultNest_mcmc_6p_SSM
```

*Result of the mcmc using the nest database*

---

**Description**

Fit using the nest database

**Usage**

```
resultNest_mcmc_6p_SSM
```

**Format**

A list of class `mcmcComposite` with mcmc result for `data(nest)` with 6 parameters and Gompertz model of growth

**Details**

Result of the mcmc using the nest database

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
x <- structure(c(115.758929130522, 428.649022170996, 503.687251738993,
12.2621455821612, 306.308841227278, 116.35048615105), .Names = c("DHA",
"DHH", "DHL", "DT", "T12L", "Rho25"))
pfixed <- c(rK=2.093313)
resultNest_6p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=1.7,
hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_6p)
pMCMC <- TRN_MHmcmc_p(resultNest_6p_SSM, accept=TRUE)
# Take care, it can be very long, sometimes several days
resultNest_mcmc_6p_SSM <- GRTRN_MHmcmc(result=resultNest_6p_SSM,
adaptive = TRUE,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1, n.adapt = 0,
thin=1, trace=TRUE)
data(resultNest_mcmc_6p_SSM)
plot(resultNest_mcmc_6p_SSM, parameters="T12L", main="", xlim=c(290, 320), bty="n")

## End(Not run)
```

---

resultNest\_mcmc\_newp    *Result of the mcmc using the nest database with anchored parameters*

---

**Description**

Fit using the nest database with anchored parameters

**Usage**

```
resultNest_mcmc_newp
```

**Format**

A list of class `mcmcComposite` with mcmc result for `data(nest)` with anchored parameters

**Details**

Result of the mcmc using the nest database with anchored parameters

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
newp <- GenerateAnchor(nests=formatted, number.anchors=7)
pfixed <- c(rK=2.093313)
resultNest_newp <- searchR(parameters=newp, fixed.parameters=pfixed,
  temperatures=formatted, integral=integral.Gompertz, M0=1.7,
  hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_newp)
pMCMC <- TRN_MHmcmc_p(resultNest_newp, accept=TRUE)
# Take care, it can be very long, sometimes several days
resultNest_mcmc_newp <- GRTRN_MHmcmc(result=resultNest_newp,
  parametersMCMC=pMCMC, n.iter=10000, n.chains = 1, n.adapt = 0,
  thin=1, trace=TRUE)
data(resultNest_mcmc_newp)
data(resultNest_4p_SSM)
newp <- GenerateAnchor(nests=resultNest_4p_SSM, number.anchors=7)
# Here the confidence interval is built based on anchored parameters
plotR(resultNest_4p_SSM, parameters=newp, SE=resultNest_mcmc_newp$SD,
  ylim=c(0,4), ylimH=c(0,0.4), show.hist=TRUE, curve="ML quantiles")
# Here the confidence interval is built based on parametric SSM equation
data(resultNest_4p_SSM)
plotR(resultNest_4p_SSM, SE=resultNest_mcmc_4p_SSM$SD,
  ylim=c(0,4), ylimH=c(0,0.4), show.hist=TRUE, curve="ML quantiles")
plot(resultNest_mcmc_newp, las=1, xlim=c(0,30), parameters="294",
  breaks=c(0, 1.00095, 2.0009, 3.00085, 4.0008, 5.00075, 6.0007, 7.00065, 8.0006, 9.00055,
  10.0005, 11.00045, 12.0004, 13.00035, 14.0003, 15.00025, 16.0002, 17.00015, 18.0001,
  19.00005, 20))
```

```
plot(resultNest_mcmc_newp, las=1, xlim=c(0,30), parameters="296.333333333333")
plot(resultNest_mcmc_newp, las=1, xlim=c(0,30), parameters=3)

## End(Not run)
```

---

resultNest\_newp      *Fit using the nest database with anchored parameters*

---

### Description

Fit using the nest database with anchored parameters

### Usage

```
resultNest_newp
```

### Format

A list with fitted information from data(nest) with anchored parameters

### Details

Result of the fit using the nest database with anchored parameters

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

### References

Girondot, M., & Kaska, Y. (2014). A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology*, 45, 96-102. doi: 10.1016/j.jtherbio.2014.08.005

### Examples

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
newp <- GenerateAnchor(nests=formatted, number.anchors=7)
pfixed <- c(rK=2.093313)
resultNest_newp <- searchR(parameters=newp, fixed.parameters=pfixed,
  temperatures=formatted, integral=integral.Gompertz, M0=1.7,
  hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_newp)
plotR(resultNest_newp)

## End(Not run)
```

---

 searchR

---

*Fit the parameters that best represent nest incubation data.*


---

## Description

Fit the parameters that best represent data.

hatchling.metric can be a data.frame with two columns Mean and SD and rownames with the nest name.

If SD is na, then least square criteria is used for fitting.

Function to fit thermal reaction norm can be expressed as :

- a 4-parameters Schoolfield, Sharpe, and Magnuson model (1981) with DHH, DHA, T12H, and Rho25;
- a 6-parameters Schoolfield, Sharpe, and Magnuson model (1981) with T12L, DT, DHH, DHL, DHA, and Rho25;
- Each of these two first models can be combined as low and high sets of parameters by adding the \_L suffix to one set. Then you must add also transition\_S and transition\_P parameters and then the growth rate is  $1/(1+\exp((1/\text{transition\_S})*(P-\text{transition\_P})))$  with P being the proportion of development;
- The Rho25\_b control the effect of hygrometry (or Rho25\_b\_L) (It is not fully functional still);
- a Weibull function with k (shape), lambda (scale) and theta parameters;
- a normal function with Peak, Scale, and sd parameters;
- an asymmetric normal function with Peak, Scale, sdH and sdL parameters;
- a symmetric trigonometric function with Length, Peak, and Max;
- an asymmetric trigonometric function with LengthB, LengthE, Peak, and Max.
- Dallwitz-Higgins model (1992) can be used using Dallwitz\_b1, Dallwitz\_b2, Dallwitz\_b3, Dallwitz\_b4 and Dallwitz\_b5 parameters.
- If Dallwitz\_b4 is not included, Dallwitz\_b4 = 6 will be used.
- If Dallwitz\_b5 is not included, Dallwitz\_b5 = 0.4 will be used.
- It is possible also to add the parameter epsilon and then the model becomes  $X + \text{epsilon}$  with X being any of the above model;
- It is possible also to add the parameter epsilon\_L and then the model becomes  $X\_L + \text{epsilon\_L}$  with X\_L being any of the above model with suffix \_L;
- If the name of the parameter is a number, then the model is a polynom anchored with the rate being the parameter value at this temperature (the name). see ChangeSSM() function.

## Usage

```
searchR(
  parameters = stop("Initial set of parameters must be provided"),
  fixed.parameters = c(rK = 1.208968),
  temperatures = stop("Formatted temperature must be provided !"),
```



```

integral = integral.Gompertz,
derivate = NULL,
hatchling.metric = c(Mean = 39.33, SD = 1.92),
M0 = 0.3470893,
saveAtMaxiter = FALSE,
fileName = "intermediate",
weight = NULL,
control = list(trace = 1, REPORT = 100, maxit = 500)
)

```

### Arguments

parameters	A set of parameters used as initial point for searching
fixed.parameters	A set of parameters that will not be changed
temperatures	Timeseries of temperatures after formatted using FormatNests()
integral	Function used to fit embryo growth: integral.Gompertz, integral.exponential or integral.linear
derivate	Function used to fit embryo growth: derivate.Gompertz, derivate.exponential or derivate.linear
hatchling.metric	A vector with Mean and SD of size of hatchlings, ex. hatchling.metric=c(Mean=39, SD=3). Can be a data.frame also. See description
M0	Measure of hatchling size or mass proxi at laying date
saveAtMaxiter	If True, each time number of iteration reach maxiter, current data are saved in file with filename name
fileName	The intermediate results are saved in file with fileName.Rdata name
weight	A named vector of the weight for each nest for likelihood estimation
control	List for control parameters for optimx

### Details

searchR fits the parameters that best represent nest incubation data.

### Value

A result object

### Author(s)

Marc Girondot

## References

- Angilletta, M.J., 2006. Estimating and comparing thermal performance curves. *Journal of Thermal Biology* 31, 541-545.
- Dallwitz, M.J., Higgins, J.P., 1992. User's guide to DEVAR. A computer program for estimating development rate as a function of temperature. *CSIRO Aust Div Entomol Rep* 2, 1-23.
- Georges, A., Beggs, K., Young, J.E., Doody, J.S., 2005. Modelling development of reptile embryos under fluctuating temperature regimes. *Physiological and Biochemical Zoology* 78, 18-30.
- Girondot, M., Kaska, Y., 2014. A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology* 45, 96-102.
- Schoolfield, R.M., Sharpe, P.J., Magnuson, C.E., 1981. Non-linear regression of biological temperature-dependent rate models based on absolute reaction-rate theory. *Journal of Theoretical Biology* 88, 719-731.

## Examples

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "DT", "DHA", "DHH", "DHL", "Rho25"
# K for Gompertz must be set as fixed parameter or being a constant K
# or relative to the hatchling size rK
#####
# From Girondot M, Monsinjon J, Guillon J-M (2018) Delimitation of the
# embryonic thermosensitive period for sex determination using an embryo
# growth model reveals a potential bias for sex ratio prediction in turtles.
# Journal of Thermal Biology 73: 32-40
# rK = 1.208968
# M0 = 0.3470893
#####
pfixed <- c(rK=1.208968)
M0 = 0.3470893
#####
# 4 parameters SSM
#####
x <- c('DHA' = 109.31113503282113, 'DHH' = 617.80695919563857,
      'T12H' = 306.38890489505093, 'Rho25' = 229.37265815800225)

resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz, M0=M0,
hatchling.metric=c(Mean=39.33, SD=1.92))

data(resultNest_4p_SSM)
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), series=1,
embryo.stages="Caretta caretta.SCL")

#####
```

```

# 6 parameters SSM
#####
x <- structure(c(106.567809092008, 527.359011254683, 614.208632495199,
2720.94506457237, 306.268259715624, 120.336791245212), .Names = c("DHA",
"DHH", "DHL", "DT", "T12L", "Rho25"))

#####
# example of data.frame for hatchling.metric
#####
thatchling.metric <- data.frame(Mean=rep(39.33, formatted$IndiceT["NbTS"]),
SD=rep(1.92, formatted$IndiceT["NbTS"]),
row.names=names(formatted)[1:formatted$IndiceT["NbTS"]])
# It is sometimes difficult to find a good starting point for
# SSM 6 parameters model. This function helps to find it based on a previousl
# fitted model.

x <- ChangeSSM(temperatures = (200:350)/10,
parameters = resultNest_4p_SSM$par,
initial.parameters = structure(c(106.567809092008,
527.359011254683, 614.208632495199,
4.94506457237, 306.268259715624, 120.336791245212),
.Names = c("DHA", "DHH", "DHL", "DT", "T12L", "Rho25")),
control=list(maxit=1000))

resultNest_6p_SSM <- searchR(parameters=x$par, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.Gompertz,
M0=M0,
hatchling.metric=thatchling.metric)

plotR(resultNest_6p_SSM, ylim=c(0, 1))

data(resultNest_6p_SSM)
pMCMC <- TRN_MHmcmc_p(resultNest_6p_SSM, accept=TRUE)
# Take care, it can be very long, sometimes several days
resultNest_mcmc_6p_SSM <- GRTRN_MHmcmc(result=resultNest_6p_SSM,
parametersMCMC=pMCMC,
n.iter=10000,
n.chains = 1,
n.adapt = 0,
thin=1,
trace=TRUE)

#####
# compare_AIC() is a function from the package "HelpersMG"
compare_AIC(test1=resultNest_4p_SSM, test2=resultNest_6p_SSM)
#####

#####
##### Example as linear progression of development
##### The development progress goes from 0 to 1
#####

pfixed <- NULL

```

```

M0 = 0

#####
# 4 parameters SSM
#####
x <- c('DHA' = 64.868697530424186, 'DHH' = 673.18292743646771,
      'T12H' = 400.90952554047749, 'Rho25' = 82.217237723502123)
resultNest_4p_SSM_Linear <- searchR(parameters=x, fixed.parameters=pfixed,
temperatures=formatted, integral=integral.linear, M0=M0,
hatchling.metric=c(Mean=39.33, SD=1.92)/39.33)
plotR(resultNest_4p_SSM_Linear, ylim=c(0, 2), scaleY= 100000)
plot(resultNest_4p_SSM_Linear, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,1.1),
series=1, embryo.stages="Generic.ProportionDevelopment")

tc <- GenerateConstInc(duration=300*24*60, temperatures = 28)
tc_f <- FormatNests(tc)
plot(resultNest_4p_SSM_Linear, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,1.1),
series=1, embryo.stages="Generic.ProportionDevelopment", temperatures=tc_f)

#####
##### with new parametrization based on anchor
##### This is a non-parametric version
#####

data(resultNest_4p_SSM)
x0 <- resultNest_4p_SSM$par
t <- range(hist(resultNest_4p_SSM, plot=FALSE)$temperatures)

x <- getFromNamespace(".SSM", ns="embryogrowth")(T=seq(from=t[1],
                                                    to=t[2],
                                                    length.out=7),
                                                    parms=x0)[[1]]*1E5

names(x) <- as.character(seq(from=t[1],
                              to=t[2],
                              length.out=7))

M0 <- 0.3470893
pfixed <- c(rK=1.208968)
resultNest_newp <- searchR(parameters=x, fixed.parameters=pfixed,
                           temperatures=formatted,
                           integral=integral.Gompertz, M0=M0,
                           hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_newp, ylim=c(0, 2),
      xlim=c(23, 34), ylimH=c(0, 3), show.hist=TRUE)
compare_AIC(test4p=resultNest_4p_SSM,
            test6p=resultNest_6p_SSM,
            testAnchor=resultNest_newp)

#####
# example with thermal reaction norm fitted from Weibull function
#####

x <- ChangeSSM(temperatures = (200:350)/10,

```

```

parameters = resultNest_4p_SSM$par,
initial.parameters = structure(c(73.4009010417375, 304.142079511996,
                                27.4671689276281),
                                .Names = c("k", "lambda", "scale")),
control=list(maxit=1000))
M0 <- 0.3470893
pfixed <- c(rK=1.208968)
resultNest_3p_Weibull <- searchR(parameters=x$par, fixed.parameters=pfixed,
                                temperatures=formatted, integral=integral.Gompertz, M0=M0,
                                hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_3p_Weibull, ylim=c(0,6), col="Black")
compare_AIC(SSM=resultNest_4p_SSM, Weibull=resultNest_3p_Weibull)

#####
# example with thermal reaction norm fitted from asymmetric normal function
#####

x <- ChangeSSM(temperatures = (200:350)/10,
               parameters = resultNest_4p_SSM$par,
               initial.parameters = structure(c(3, 7, 11, 32),
                                               .Names = c("Scale", "sdL", "sdH", "Peak")),
               control=list(maxit=1000))
M0 <- 0.3470893
pfixed <- c(rK=1.208968)
resultNest_4p_normal <- searchR(parameters=x$par, fixed.parameters=pfixed,
                                temperatures=formatted, integral=integral.Gompertz, M0=M0,
                                hatchling.metric=c(Mean=39.33, SD=1.92))

#####
# example with thermal reaction norm fitted from trigonometric model
#####

x <- ChangeSSM(temperatures = (200:350)/10,
               parameters = resultNest_4p_SSM$par,
               initial.parameters = structure(c(3, 20, 40, 32),
                                               .Names = c("Max", "LengthB", "LengthE", "Peak")),
               control=list(maxit=1000))
M0 <- 0.3470893
pfixed <- c(rK=1.208968)
resultNest_4p_trigo <- searchR(parameters=x$par, fixed.parameters=pfixed,
                                temperatures=formatted, integral=integral.Gompertz, M0=M0,
                                hatchling.metric=c(Mean=39.33, SD=1.92))

#####
# example with thermal reaction norm fitted from Dallwitz model
#####
# See: Dallwitz, M.J., Higgins, J.P., 1992. User's guide to DEVAR. A computer
# program for estimating development rate as a function of temperature. CSIRO Aust
# Div Entomol Rep 2, 1-23.

# Note that Dallwitz model has many problems and I recommend to not use it:
# - The 3-parameters is too highly constraint
# - The 5 parameters produced infinite outputs for some sets of parameters

```

```

x <- c('Dallwitz_b1' = 4.8854060791241816,
      'Dallwitz_b2' = 20.398366565842029,
      'Dallwitz_b3' = 31.510995256647092)
M0 <- 0.3470893
pfixed <- c(rK=1.208968)
resultNest_3p_Dallwitz <- searchR(parameters=x, fixed.parameters=pfixed,
                                temperatures=formatted, integral=integral.Gompertz, M0=M0,
                                hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_3p_Dallwitz, ylim=c(0,6))

x <- c('Dallwitz_b1' = 4.9104386262684656,
      'Dallwitz_b2' = 7.515425231891359,
      'Dallwitz_b3' = 31.221784599026638,
      'Dallwitz_b4' = 7.0354467023505682,
      'Dallwitz_b5' = -1.5955717975708577)
pfixed <- c(rK=1.208968)
resultNest_5p_Dallwitz <- searchR(parameters=x, fixed.parameters=pfixed,
                                temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
                                hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_5p_Dallwitz, ylim=c(0,3), scaleY=10000)

xp <- resultNest_6p_SSM$par
xp["Rho25"] <- 233
pfixed <- c(rK=1.208968)
resultNest_6p_SSM <- searchR(parameters=xp, fixed.parameters=pfixed,
                              temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
                              hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_6p_SSM, ylim=c(0,8))

xp <- ChangeSSM(parameters = resultNest_3p_Dallwitz$par,
                initial.parameters = resultNest_4p_SSM$par)
pfixed <- c(rK=1.208968)
resultNest_4p_SSM <- searchR(parameters=xp$par, fixed.parameters=pfixed,
                              temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
                              hatchling.metric=c(Mean=39.33, SD=1.92))
plotR(resultNest_4p_SSM, ylim=c(0,6))

compare_AIC(Dallwitz3p=resultNest_3p_Dallwitz, Dallwitz5p=resultNest_5p_Dallwitz,
            SSM=resultNest_4p_SSM, SSM=resultNest_6p_SSM)

#####
# Example with thermal reaction norm of proportion of development
# fitted from Dallwitz model
# see Woolgar, L., Trocini, S., Mitchell, N., 2013. Key parameters describing
# temperature-dependent sex determination in the southernmost population of loggerhead
# sea turtles. Journal of Experimental Marine Biology and Ecology 449, 77-84.
#####

x <- structure(c(1.48207559695689, 20.1100310234046, 31.5665036287242),
              .Names = c("Dallwitz_b1", "Dallwitz_b2", "Dallwitz_b3"))
resultNest_PropDev_3p_Dallwitz <- searchR(parameters=x, fixed.parameters=NULL,
                                          temperatures=formatted, integral=integral.linear, M0=0,

```

```

      hatchling.metric=c(Mean=1, SD=NA))
plotR(resultNest_PropDev_3p_Dallwitz, ylim=c(0, 1.5), curve="ML")
plot(x=resultNest_PropDev_3p_Dallwitz, ylimS=c(0,1), xlim=c(0,60), series=2,
     embryo.stages="Generic.ProportionDevelopment")

x <- structure(c(1.48904182113431, 10.4170365155993, 31.2591665490154,
6.32355497589913, -1.07425378667104), .Names = c("Dallwitz_b1",
"Dallwitz_b2", "Dallwitz_b3", "Dallwitz_b4", "Dallwitz_b5"))
resultNest_PropDev_5p_Dallwitz <- searchR(parameters=x, fixed.parameters=NULL,
    temperatures=formatted, integral=integral.linear, M0=0,
    hatchling.metric=c(Mean=1, SD=NA))
plotR(resultNest_PropDev_5p_Dallwitz, ylim=c(0, 1.5))
plot(x=resultNest_PropDev_5p_Dallwitz, ylimS=c(0,1), xlim=c(0,60), series=2,
     embryo.stages="Generic.ProportionDevelopment")

plotR(resultNest_PropDev_3p_Dallwitz, ylim=c(0, 1.5), curve="ML")
plotR(resultNest_PropDev_5p_Dallwitz, ylim=c(0, 1.5), curve="ML", new=FALSE, col="red")
compare_AICc(Dallwitz3p=resultNest_PropDev_3p_Dallwitz,
             Dallwitz5p=resultNest_PropDev_5p_Dallwitz)

## End(Not run)

```

---

stages	<i>Database of of embryonic development and thermosensitive period of development for sex determination</i>
--------	---

---

### Description

Database of embryonic development and thermosensitive period of development for sex determination.

### Usage

stages

### Format

A list with dataframes including attributes

### Details

Database of embryonic development and thermosensitive period of development for sex determination

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

## References

- Pieau, C., Dorizzi, M., 1981. Determination of temperature sensitive stages for sexual differentiation of the gonads in embryos of the turtle, *Emys orbicularis*. *Journal of Morphology* 170, 373-382.
- Yntema, C.L., Mrosovsky, N., 1982. Critical periods and pivotal temperatures for sexual differentiation in loggerhead sea turtles. *Canadian Journal of Zoology-Revue Canadienne de Zoologie* 60, 1012-1016.
- Kaska, Y., Downie, R., 1999. Embryological development of sea turtles (*Chelonia mydas*, *Caretta caretta*) in the Mediterranean. *Zoology in the Middle East* 19, 55-69.
- Greenbaum, E., 2002. A standardized series of embryonic stages for the emydid turtle *Trachemys scripta*. *Canadian Journal of Zoology-Revue Canadienne de Zoologie* 80, 1350-1370.
- Magalhães, M.S., Vogt, R.C., Sebben, A., Dias, L.C., de Oliveira, M.F., de Moura, C.E.B., 2017. Embryonic development of the Giant South American River Turtle, *Podocnemis expansa* (Testudines: Podocnemididae). *Zoomorphology*.

## See Also

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD.P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

## Examples

```
## Not run:
library(embryogrowth)
data(stages)
names(stages)
levels(as.factor(stages$Species))
# Version of database
stages$Version[1]
kaska99.SCL <- subset(stages, subset=(Species == "Caretta caretta"),
  select=c("Stage", "SCL_Mean_mm", "SCL_SD_mm", "Days_Begin", "Days_End"))

kaska99.SCL[kaska99.SCL$Stage==31, "Days_Begin"] <- 51
kaska99.SCL[kaska99.SCL$Stage==31, "Days_End"] <- 62
kaska99.SCL <- na.omit(kaska99.SCL)
kaska99.SCL[which(kaska99.SCL$Stage==31), "Stage"] <- c("31a", "31b", "31c")
kaska99.SCL <- cbind(kaska99.SCL,
  Days_Mean=(kaska99.SCL[, "Days_Begin"]+kaska99.SCL[, "Days_End"])/2)
kaska99.SCL <- cbind(kaska99.SCL,
  Days_SD=(kaska99.SCL[, "Days_End"]-kaska99.SCL[, "Days_Begin"])/4)
Gompertz <- function(x, par) {
  K <- par["K"]
  rT <- par["rT"]
  X0 <- par["X0"]
  y <- abs(K)*exp(log(abs(X0)/abs(K))*exp(-rT*x))
  return(y)
}

ML.Gompertz <- function(x, par) {
  par <- abs(par)
```



```

y <- Gompertz(x, par)
return(sum(-dnorm(y, mean=kaska99.SCL[, "SCL_Mean_mm"],
                 sd=kaska99.SCL[, "SCL_SD_mm"], log=TRUE)))
}

parIni <- structure(c(48.66977358, 0.06178453, 0.38640902),
                  .Names = c("K", "rT", "X0"))

fitsize.SCL <- optim(parIni, ML.Gompertz, x=kaska99.SCL[, "Days_Mean"], hessian = TRUE)

# Estimation of standard error of parameters using Hessian matrix
sqrt(diag(solve(fitsize.SCL$hessian)))

# Estimation of standard error of parameters using Bayesian concept and MCMC
pMCMC <- structure(list(Density = c("dunif", "dunif", "dunif"),
                      Prior1 = c(0, 0, 0), Prior2 = c(90, 1, 2),
                      SDProp = c(1, 1, 1),
                      Min = c(0, 0, 0), Max = c(90, 1, 2),
                      Init = fitsize.SCL$par),
                  .Names = c("Density", "Prior1", "Prior2", "SDProp", "Min", "Max", "Init"),
                  row.names = c("K", "rT", "X0"), class = "data.frame")

Bayes.Gompertz <- function(data, x) {
  x <- abs(x)
  y <- Gompertz(data, x)
  return(sum(-dnorm(y, mean=kaska99.SCL[, "SCL_Mean_mm"],
                  sd=kaska99.SCL[, "SCL_SD_mm"], log=TRUE)))
}

mcmc_run <- MHalgoGen(n.iter=50000, parameters=pMCMC, data=kaska99.SCL[, "Days_Mean"],
                    likelihood=Bayes.Gompertz, n.chains=1, n.adapt=100, thin=1, trace=1,
                    adaptive = TRUE)

plot(mcmc_run, xlim=c(0, 90), parameters="K")
plot(mcmc_run, xlim=c(0, 1), parameters="rT")
plot(mcmc_run, xlim=c(0, 2), parameters="X0")

1-rejectionRate(as.mcmc(mcmc_run))

par <- mcmc_run$resultMCMC[[1]]

outsp <- t(apply(par, MARGIN = 1, FUN=function(x) Gompertz(0:70, par=x)))

rangqtiles <- apply(outsp, MARGIN=2, function(x) {quantile(x, probs=c(0.025, 0.5, 0.975))})

par(mar=c(4, 4, 2, 1))
plot_errbar(x=kaska99.SCL[, "Days_Mean"], y=kaska99.SCL[, "SCL_Mean_mm"],
            errbar.y = 2*kaska99.SCL[, "SCL_SD_mm"], bty="n", las=1,
            ylim=c(0, 50), xlab="Days", ylab="SCL mm",
            xlim=c(0, 70), x.plus = kaska99.SCL[, "Days_End"],
            x.minus = kaska99.SCL[, "Days_Begin"])

lines(0:70, rangqtiles["2.5%", ], lty=2)

```

```

lines(0:70, rangqtiles["97.5%", ], lty=2)
lines(0:70, rangqtiles["50%", ], lty=3)

text(x=50, y=10, pos=4, labels=paste("K=", format(x = fitsize.SCL$par["K"], digits = 4)))
text(x=50, y=12.5, pos=4,
     labels=paste("rK=", format(x = fitsize.SCL$par["K"]/39.33, digits = 4)))
text(x=50, y=15, pos=4, labels=paste("X0=", format(x = fitsize.SCL$par["X0"], digits = 4)))
title("Univariate normal distribution")

# Using a multivariate normal distribution

library(mvtnorm)

ML.Gompertz.2D <- function(x, par) {
  par <- abs(par)
  y <- Gompertz(x, par)
  L <- 0
  for (i in seq_along(y)) {
    sigma <- matrix(c(kaska99.SCL$SCL_SD_mm[i]^2, 0, 0, kaska99.SCL$Days_SD[i]^2),
                    nrow=2, byrow=TRUE,
                    dimnames=list(c("SCL_SD_mm", "Days_SD"), c("SCL_SD_mm", "Days_SD")))
    L <- L -dmvnorm(x=c(SCL_SD_mm=kaska99.SCL$SCL_Mean_mm[i],
                      Days_SD=kaska99.SCL$Days_Mean[i]),
                  mean= c(SCL_SD_mm=y[i], Days_SD=kaska99.SCL$Days_Mean[i]),
                  sigma=sigma, log=TRUE)
  }
  return(L)
}

parIni <- structure(c(48.66977358, 0.06178453, 0.38640902),
                  .Names = c("K", "rT", "X0"))

fitsize.SCL.2D <- optim(parIni, ML.Gompertz.2D, x=kaska99.SCL[, "Days_Mean"], hessian = TRUE)

# Estimation of standard error of parameters using Hessian matrix
sqrt(diag(solve(fitsize.SCL.2D$hessian)))

# Estimation of standard error of parameters using Bayesian concept and MCMC
Bayes.Gompertz.2D <- function(data, x) {
  x <- abs(x)
  y <- Gompertz(data, x)
  L <- 0
  for (i in seq_along(y)) {
    sigma <- matrix(c(kaska99.SCL$SCL_SD_mm[i]^2, 0, 0, kaska99.SCL$Days_SD[i]^2),
                    nrow=2, byrow=TRUE,
                    dimnames=list(c("SCL_SD_mm", "Days_SD"), c("SCL_SD_mm", "Days_SD")))
    L <- L - dmvnorm(x=c(SCL_SD_mm=kaska99.SCL$SCL_Mean_mm[i],
                      Days_SD=kaska99.SCL$Days_Mean[i]),
                  mean= c(SCL_SD_mm=y[i], Days_SD=kaska99.SCL$Days_Mean[i]),
                  sigma=sigma, log=TRUE)
  }
  return(L)
}

```

```

pMCMC <- structure(list(Density = c("dunif", "dunif", "dunif"),
  Prior1 = c(0, 0, 0), Prior2 = c(90, 1, 2),
  SDProp = c(1, 1, 1),
  Min = c(0, 0, 0), Max = c(90, 1, 2),
  Init = fitsize.SCL.2D$par),
  .Names = c("Density", "Prior1", "Prior2", "SDProp", "Min", "Max", "Init"),
  row.names = c("K", "rT", "X0"), class = "data.frame")
mcmc_run.2D <- MHalgoGen(n.iter=50000, parameters=pMCMC, data=kaska99.SCL[, "Days_Mean"],
  likelihood=Bayes.Gompertz.2D, n.chains=1, n.adapt=100, thin=1, trace=1,
  adaptive = TRUE)

plot(mcmc_run.2D, xlim=c(0, 90), parameters="K")
plot(mcmc_run.2D, xlim=c(0, 1), parameters="rT")
plot(mcmc_run.2D, xlim=c(0, 2), parameters="X0")

1-rejectionRate(as.mcmc(mcmc_run.2D))

par <- mcmc_run.2D$resultMCMC[[1]]

outsp <- t(apply(par, MARGIN = 1, FUN=function(x) Gompertz(0:70, par=x)))

rangqtiles <- apply(outsp, MARGIN=2, function(x) {quantile(x, probs=c(0.025, 0.5, 0.975))})

par(mar=c(4, 4, 2, 1))
plot_errbar(x=kaska99.SCL[, "Days_Mean"], y=kaska99.SCL[, "SCL_Mean_mm"],
  errbar.y = 2*kaska99.SCL[, "SCL_SD_mm"], bty="n", las=1,
  ylim=c(0, 50), xlab="Days", ylab="SCL mm",
  xlim=c(0, 70), x.plus = kaska99.SCL[, "Days_End"],
  x.minus = kaska99.SCL[, "Days_Begin"])

lines(0:70, rangqtiles["2.5%", ], lty=2)
lines(0:70, rangqtiles["97.5%", ], lty=2)
lines(0:70, rangqtiles["50%", ], lty=3)

text(x=50, y=10, pos=4,
  labels=paste("K=", format(x = fitsize.SCL.2D$par["K"], digits = 4)))
text(x=50, y=12.5, pos=4,
  labels=paste("rK=", format(x = fitsize.SCL.2D$par["K"]/39.33, digits = 4)))
text(x=50, y=15, pos=4,
  labels=paste("X0=", format(x = fitsize.SCL.2D$par["X0"], digits = 4)))
title("Multivariate normal distribution")

## End(Not run)

```

### Description

Estimate the parameters that best describe the sexualisation thermal reaction norm within the TSP. The `sexratio` parameter is a character string which can be:

- `TSP.TimeWeighted.sexratio.mean` Sex ratio based on average temperature during the TSP
- `TSP.GrowthWeighted.sexratio.mean` Sex ratio based on average temperature weighted by the actual growth during the TSP
- `TSP.TimeWeighted.GrowthRateWeighted.sexratio.mean` Sex ratio based on average temperature weighted by the growth rate during the TSP
- `TSP.TimeWeighted.STRNWeighted.sexratio.mean` Sex ratio based on average temperature weighted by the thermal reaction norm of sexualization during the TSP
- `TSP.GrowthWeighted.STRNWeighted.sexratio.mean` Sex ratio based on average temperature weighted by the actual growth and thermal reaction norm of sexualization during the TSP
- `TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean` Sex ratio based on average temperature weighted by the growth rate and the thermal reaction norm of sexualization during the TSP
- `MiddleThird.TimeWeighted.sexratio.mean` Sex ratio based on average temperature during the middle third incubation
- `MiddleThird.GrowthWeighted.sexratio.mean` Sex ratio based on average temperature weighted by actual growth during the middle third incubation
- `MiddleThird.TimeWeighted.GrowthRateWeighted.sexratio.mean` Sex ratio based on average temperature weighted by growth rate during the middle third incubation
- `TimeWeighted.sexratio.mean` Sex ratio based on average temperature during all incubation
- `GrowthWeighted.sexratio.mean` Sex ratio based on average temperature weighted by actual growth during all incubation
- `TimeWeighted.GrowthRateWeighted.sexratio.mean` Sex ratio based on average temperature weighted by growth rate during all incubation
- `TSP.PM.TimeWeighted.mean` Average sex ratio based on temperature during the TSP
- `TSP.PM.GrowthWeighted.mean` Average sex ratio based on temperature weighted by the actual growth during the TSP
- `TSP.PM.TimeWeighted.GrowthRateWeighted.mean` Average sex ratio based on temperature weighted by the growth rate during the TSP

If information for sex is not known for some timeseries, set NA for `Sexed`.

`Sexed`, `Males` and `Females` must be vectors with names. The names must be the same as the names of timeseries of temperatures in `EmbryoGrowthTRN`.

Only two of these 3 parameters are required: `Males`, `Females` and `Sexed`

Note: four species have predefined embryo stages. `embryo.stages` parameter can take the values:

- `Caretta caretta.SCL`
- `Chelonia mydas.SCL`

- `Emys orbicularis.SCL`
- `Emys orbicularis.mass`
- `Podocnemis expansa.SCL`
- `Generic.ProportionDevelopment`

A fifth name fitted must be used when limits of TSP are fitted using `BeginTSP` and `EndTSP` parameters.

The parameters that can be used in STRN are:

`BeginTSP`, `EndTSP` are the logit of the proportion of development;

To ensure that `BeginTSP < EndTSP`, it is better to use:

`BeginTSP`, `LengthTSP` and then `EndTSP` is estimated using `BeginTSP + abs(LengthTSP)`

`DHA`, `DHH`, `T12H` are the SSM parameters of sexualisation thermal reaction norm;

`dbeta_mu`, `dbeta_v` are the beta mean and variance of the impact of sexualisation according to TSP progress.

Or any parameter that can be used in a TSD model.

### Usage

```
STRN(
  Initial_STRN = NULL,
  fixed.parameters = NULL,
  EmbryoGrowthTRN = stop("Embryo Growth Thermal Reaction Norm must be provided"),
  TSP.borders = NULL,
  embryo.stages = NULL,
  TSP.begin = 0,
  TSP.end = 0.5,
  tsd = NULL,
  equation = "logistic",
  Sexed = NULL,
  Males = NULL,
  Females = NULL,
  sexratio = "TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean",
  fill = 60,
  parallel = TRUE,
  itnmax = 1000,
  method = c("Nelder-Mead", "BFGS"),
  control = list(trace = 1, REPORT = 10),
  zero = 1e-09,
  verbose = FALSE,
  hessian = TRUE
)
```

### Arguments

`Initial_STRN` Values for initial model of Sexualisation Thermal Reaction Norm or tsd model

`fixed.parameters`

Value for Sexualisation Thermal Reaction Norm or tsd model that will not be changed

EmbryoGrowthTRN	The Embryo Growth Thermal Reaction Norm obtained with searchR()
TSP.borders	The limits of TSP in stages. See embryo.stages parameter.
embryo.stages	The embryo stages. At least TSP.borders stages must be provided to estimate TSP borders. See note.
TSP.begin	Where TSP begin during the stage of beginning? In relative proportion of the stage.
TSP.end	Where TSP begin during the stage of ending? In relative proportion of the stage.
tsd	The model used to predict sex ratio, obtained from tsd()
equation	If tsd parameter is not provided, equation and parameters in Initial_STRN for tsd model must be provided.
Sexed	The number of sexed embryos with names identifying timeseries
Males	The number of males embryos with names identifying timeseries
Females	The number of females embryos with names identifying timeseries
sexratio	The sex ratio to be used
fill	See info.nests()
parallel	Should parallel computing for info.nests() be used
itnmax	Maximum number of iterations for each method; if 0, just return the likelihood
method	Methods to be used with optimx
control	List for control parameters for optimx
zero	The value to replace a null sex ratio
verbose	If TRUE, will show all intermediate parameters during fit
hessian	If TRUE, the Hessian approximation is estimated at the end of the fit.

### Details

STRN estimates the parameters that best describe the sexualisation thermal reaction norm within the TSP

### Value

The list with object return by optim()

### Author(s)

Marc Girondot

### Examples

```
## Not run:
library(embryogrowth)
MedIncubation_Cc <- subset(DatabaseTSD, Species=="Caretta caretta" &
RMU=="Mediterranean" & Sexed!=0)
Med_Cc <- tsd(males=MedIncubation_Cc$Males,
```

```

        females=MedIncubation_Cc$Females,
        temperatures=MedIncubation_Cc$Incubation.temperature,
        par=c(P=29.5, S=-0.1))
plot(Med_Cc, xlim=c(25, 35))
males <- c(7, 0, 0, 0, 0, 5, 6, 3, 5, 3, 2, 3, 0, 0, 0, 0, 0, 0, 0, 0)
names(males) <- rev(rev(names(resultNest_4p_SSM$data))[-(1:2)])
sexed <- rep(10, length(males))
names(sexed) <- rev(rev(names(resultNest_4p_SSM$data))[-(1:2)])

Initial_STRN <- c('DHA' = 1174.6461503413307,
                 'DHH' = 2001.0619192107047,
                 'T12H' = 3731.353104743393)

fp <- c(Rho25=100)
fitSTRN <- STRN(Initial_STRN=Initial_STRN,
               EmbryoGrowthTRN=resultNest_4p_SSM,
               tsd=Med_Cc,
               embryo.stages="Caretta caretta.SCL",
               Sexed=sexed, Males=males,
               fixed.parameters=fp,
               sexratio="TSP.GrowthWeighted.STRNWeighted.sexratio.mean")
plotR(fitSTRN, curve = "ML", ylim=c(0,2))
plotR(fitSTRN)
out <- info.nests(NestsResult=resultNest_4p_SSM,
                 SexualisationTRN=fitSTRN,
                 SexualisationTRN.CI="Hessian",
                 embryo.stages="Caretta caretta.SCL",
                 GTRN.CI="Hessian",
                 tsd=Med_Cc,
                 tsd.CI="Hessian",
                 replicate.CI=100,
                 progressBar=TRUE,
                 warnings=TRUE,
                 out="summary")$summary
# CTE with growth-weighted temperature average
plot(Med_Cc, xlim=c(25, 35))
points(x=out[, "TSP.GrowthWeighted.STRNWeighted.temperature.mean"], y=males/sexed,
       col="red", pch=19)
legend("topright", legend=c("CTE with growth-weighted and Sexualization TRN"),
       pch=19, col=c("red"))

# Fit the beginning and end of TSP

Initial_STRN <- c('BeginTSP' = invlogit(0.33),
                 'EndTSP' = invlogit(0.66))

fp <- NULL
fitSTRN <- STRN(Initial_STRN=Initial_STRN,
               EmbryoGrowthTRN=resultNest_4p_SSM,
               tsd=Med_Cc,
               embryo.stages="fitted",
               Sexed=sexed, Males=males,
               fixed.parameters=fp,
               sexratio="TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean")
invlogit(fitSTRN$par)

```

```

invlogit(fitSTRN$par-2*fitSTRN$SE)
invlogit(fitSTRN$par+2*fitSTRN$SE)

Initial_STRN <- c('dbeta_mu' = logit(0.5),
                 'dbeta_v' = 1/12)

fp <- NULL
fitSTRN <- STRN(Initial_STRN=Initial_STRN,
               EmbryoGrowthTRN=resultNest_4p_SSM,
               tsd=Med_Cc,
               embryo.stages="Caretta caretta.SCL",
               Sexed=sexed, Males=males,
               fixed.parameters=fp,
               sexratio="TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean")
mu <- invlogit(fitSTRN$par["dbeta_mu"]),
v <- abs(fitSTRN$par["dbeta_v"])
shape1 <- mu * (((mu * (1 - mu))/v) - 1)
shape2 <- shape1 * (1 - mu)/mu

plot(seq(from=0, to=1, length.out=100),
     dbeta(seq(from=0, to=1, length.out=100),
           shape1=shape1, shape2=shape2),
     type="l", xlab="Progress of TSP",
     ylab="Force of sexualisation", bty="n", ylim=c(0, 6), las=1)

Initial_STRN <- c('dbeta_mu' = 7.2194053298953236,
                 'dbeta_v' = 0.00050390986089928467)

fp <- NULL
fitSTRN <- STRN(Initial_STRN=Initial_STRN,
               EmbryoGrowthTRN=resultNest_4p_SSM,
               tsd=Med_Cc,
               embryo.stages="Caretta caretta.SCL",
               Sexed=sexed,
               Males=males,
               fixed.parameters=fp,
               sexratio="TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean")

mu <- invlogit(fitSTRN$par["dbeta_mu"]),
v <- abs(fitSTRN$par["dbeta_v"])
shape1 <- mu * (((mu * (1 - mu))/v) - 1)
shape2 <- shape1 * (1 - mu)/mu

plot(seq(from=0, to=1, length.out=100),
     dbeta(seq(from=0, to=1, length.out=100),
           shape1=shape1, shape2=shape2),
     type="l", xlab="Progress of TSP",
     ylab="Force of sexualisation", bty="n", ylim=c(0, 0.04), las=1)

Initial_STRN <- c('dbeta_mu' = logit(0.5),
                 'dbeta_v' = 1/12)

L <- STRN(Initial_STRN=NULL,
         fixed.parameters=Initial_STRN,
         EmbryoGrowthTRN=resultNest_4p_SSM,
         tsd=Med_Cc,
         embryo.stages="Caretta caretta.SCL",

```



```

Sexed=sexed
Males=males
sexratio="TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean")
Initial_STRN <- c('dbeta_mu' = logit(0.6),
                 'dbeta_v' = 1/12)
L <- STRN(Initial_STRN=NULL
         fixed.parameters=Initial_STRN
         EmbryoGrowthTRN=resultNest_4p_SSM
         tsd=Med_Cc
         embryo.stages="Caretta caretta.SCL"
         Sexed=sexed
         Males=males
         sexratio="TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean")
Initial_STRN <- c('dbeta_mu' = 7.2192972077000004,
                 'dbeta_v' = 0.00050396969999999997)
L <- STRN(Initial_STRN=NULL
         fixed.parameters=Initial_STRN
         EmbryoGrowthTRN=resultNest_4p_SSM
         tsd=Med_Cc
         embryo.stages="Caretta caretta.SCL"
         Sexed=sexed
         Males=males
         sexratio="TSP.TimeWeighted.GrowthRateWeighted.STRNWeighted.sexratio.mean")
      mu <- invlogit(fitSTRN$par["dbeta_mu"]),
      v <- abs(fitSTRN$par["dbeta_v"])
      shape1 <- mu * (((mu * (1 - mu))/v) - 1)
      shape2 <- shape1 * (1 - mu)/mu

tsp_progress <- seq(from=0, to=1, length.out=100)
plot(tsp_progress,
     dbeta(tsp_progress,
           shape1=shape1, shape2=shape2),
     type="l", xlab="Progress of TSP",
     ylab="Force of sexualisation", bty="n", ylim=c(0, 0.04), las=1)
segments(x0=0, x1=1, y0=0, y1=0, lty=2)

## End(Not run)

```

STRN\_MHmcmc

*Metropolis-Hastings algorithm for Sexualisation Thermal Reaction Norm*

### Description

Run the Metropolis-Hastings algorithm for Sexualisation Thermal Reaction Norm.

The number of iterations is  $n.iter+n.adapt+1$  because the initial likelihood is also displayed.

I recommend that  $thin=1$  because the method to estimate SE uses resampling.

If initial point is maximum likelihood,  $n.adapt = 0$  is a good solution.

To get the SE of the point estimates from `result_mcmc <- STRN_MHmcmc(result=try)`, use:

`result_mcmc$SD`

coda package is necessary for this function.

The parameters `intermediate` and `filename` are used to save intermediate results every 'intermediate' iterations (for example 1000). Results are saved in a file of name `filename`.

The parameter `previous` is used to indicate the list that has been save using the parameters `intermediate` and `filename`. It permits to continue a mcmc search.

These options are used to prevent the consequences of computer crash or if the run is very very long and processes at time limited.

If `fill` is NA, it will use the stored fill value in result.

## Usage

```
STRN_MHmcmc(
  result = NULL,
  n.iter = 10000,
  parametersMCMC = NULL,
  n.chains = 1,
  n.adapt = 0,
  thin = 1,
  trace = NULL,
  traceML = FALSE,
  batchSize = sqrt(n.iter),
  adaptive = FALSE,
  adaptive.lag = 500,
  adaptive.fun = function(x) { ifelse(x > 0.234, 1.3, 0.7) },
  parallel = TRUE,
  intermediate = NULL,
  filename = "intermediate.Rdata",
  previous = NULL,
  fill = NA
)
```

## Arguments

<code>result</code>	An object obtained after a STRN fit
<code>n.iter</code>	Number of iterations for each step
<code>parametersMCMC</code>	A set of parameters used as initial point for searching with information on priors
<code>n.chains</code>	Number of replicates
<code>n.adapt</code>	Number of iterations before to store outputs
<code>thin</code>	Number of iterations between each stored output
<code>trace</code>	TRUE or FALSE or period, shows progress
<code>traceML</code>	TRUE or FALSE to show ML
<code>batchSize</code>	Number of observations to include in each batch fo SE estimation
<code>adaptive</code>	Should an adaptive process for SDProp be used
<code>adaptive.lag</code>	Lag to analyze the SDProp value in an adaptive content
<code>adaptive.fun</code>	Function used to change the SDProp

parallel	Should parallel computing for info.nests() be used
intermediate	Period for saving intermediate result, NULL for no save
filename	If intermediate is not NULL, save intermediate result in this file
previous	Previous result to be continued. Can be the filename in which intermediate results are saved.
fill	Parameters to be sent to STRN().

### Details

STRN\_MHmcmc runs the Metropolis-Hastings algorithm for STRN (Bayesian MCMC)

### Value

A list with resultMCMC being mcmc.list object, resultLnL being likelihoods and parametersMCMC being the parameters used

### Author(s)

Marc Girondot

### Examples

```
## Not run:
library(embryogrowth)
MedIncubation_Cc <- subset(DatabaseTSD, Species=="Caretta caretta" &
RMU=="Mediterranean" & Sexed!=0)
Med_Cc <- tsd(males=MedIncubation_Cc$Males,
             females=MedIncubation_Cc$Females,
             temperatures=MedIncubation_Cc$Incubation.temperature,
             par=c(P=29.5, S=-0.1))
plot(Med_Cc, xlim=c(25, 35))
males <- c(7, 0, 0, 0, 0, 5, 6, 3, 5, 3, 2, 3, 0, 0, 0, 0, 0, 0, 0, 0)
names(males) <- rev(rev(names(resultNest_4p_SSM$data))[-(1:2)])
sexed <- rep(10, length(males))
names(sexed) <- rev(rev(names(resultNest_4p_SSM$data))[-(1:2)])
Initial_STRN <- resultNest_4p_SSM$par[c("DHA", "DHH", "T12H")]
Initial_STRN <- structure(c(582.567096666926, 2194.0806711639, 3475.28414940385),
                        .Names = c("DHA", "DHH", "T12H"))
fp <- c(Rho25=100)
fitSTRN <- STRN(Initial_STRN=Initial_STRN,
              EmbryoGrowthTRN=resultNest_4p_SSM,
              tsd=Med_Cc,
              embryo.stages="Caretta caretta.SCL",
              Sexed=sexed, Males=males,
              fixed.parameters=fp,
              sexratio="TSP.GrowthWeighted.STRNWeighted.sexratio")

pMCMC <- TRN_MHmcmc_p(fitSTRN, accept=TRUE)
pMCMC[, "Density"] <- "dunif"
pMCMC[, "Prior2"] <- pMCMC[, "Max"]<- 10000
pMCMC[, "Prior1"] <- pMCMC[, "Min"] <- 1
```

```

outMCMC <- STRN_MHmcmc(result = fitSTRN, n.iter = 10000, parametersMCMC = pMCMC,
                      n.chains = 1, n.adapt = 0, thin = 1, trace = TRUE,
                      adaptive = TRUE, adaptive.lag = 500,
                      intermediate = 1000,
                      filename = "intermediate_mcmcSTRN.Rdata")
plot(outMCMC, parameters=1)
plot(outMCMC, parameters=2)
plot(outMCMC, parameters=3)
1-rejectionRate(as.mcmc(x = outMCMC))

## End(Not run)

```

---

summary.Nests

*Summarize the information from a Nests object.*


---

### Description

Summarize the information from a Nests object:  
- Name of the nests, total incubation length and average temperature

### Usage

```

## S3 method for class 'Nests'
summary(object, ...)

```

### Arguments

object	A object obtained after FormatNests()
...	Not used

### Details

summary.Nests Summarize the information from a Nests object

### Value

None

### Author(s)

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest, previous=NULL)
summary(formatted)

## End(Not run)
```

---

switch.transition	<i>Add a transition parameter on a set of parameters or remove it</i>
-------------------	---

---

**Description**

Add a transition parameter on a set of parameters or remove it

**Usage**

```
switch.transition(parameters = stop("A set of parameters must be supplied"))
```

**Arguments**

parameters      A vector with parameters

**Details**

switch.transition Add a transition parameter on a set of parameters or remove it

**Value**

A vector with parameters

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
data(resultNest_6p_SSM)
# Get a set of parameters without transition
x1 <- resultNest_6p_SSM$par
# Generate a set of parameters with transition
x2 <- switch.transition(x1)
# Generate a set of parameters without transition
x3 <- switch.transition(x3)

## End(Not run)
```

tempConst

*Timeseries of constant temperatures for nests***Description**

Timeseries of temperatures for nests

**Usage**

```
tempConst
```

**Format**

A dataframe with raw data.

**Details**

Timeseries of constant temperatures for nests

**Author(s)**

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

**References**

Girondot, M., Kaska, Y., 2014. A model to predict the thermal reaction norm for the embryo growth rate from field data. *Journal of Thermal Biology* 45, 96-102.

**Examples**

```
## Not run:
library(embryogrowth)
# Same as:
# GenerateConstInc(durations = rep(104*60*24, 11),
# temperatures = 25:35,
# names = paste0("T",25:35))
data(tempConst)
tempConst_f <- FormatNests(tempConst)

data(nest)
formatted <- FormatNests(nest)
x <- structure(c(109.683413821537, 614.969219372661, 306.386903812694,
  229.003478775323), .Names = c("DHA", "DHH", "T12H", "Rho25"))

# See the stages dataset examples for justification of M0 and rK

pfixed <- c(rK=1.208968)
resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
  temperatures=formatted, integral=integral.Gompertz, M0=0.3470893,
```

```

hatchling.metric=c(Mean=39.33, SD=1.92))

plotR(result=resultNest_4p_SSM, show.hist = TRUE,
      ylim=c(0, 8), curve="ML quantiles")

# Now use the fitted parameters from resultNest_4p_SSM with
# the constant incubation temperatures:

plot(resultNest_4p_SSM, temperatures=tempConst_f,
     stop.at.hatchling.metric=TRUE, series="T30", xlim=c(0,50),
     ylimT=c(22, 32), hatchling.metric=c(Mean=39.33, SD=1.92),
     embryo.stages="Caretta caretta.SCL")

plot(resultNest_4p_SSM, temperatures=tempConst_f,
     stop.at.hatchling.metric=TRUE, series="T25", xlim=c(0,120),
     ylimT=c(22, 32), hatchling.metric=c(Mean=39.33, SD=1.92),
     embryo.stages="Caretta caretta.SCL")

## End(Not run)

```

---

test.parallel	<i>Estimate the likelihood of a set of parameters for nest incubation data with or without parallel computing option</i>
---------------	--

---

### Description

Estimate the likelihood of a set of parameters for nest incubation data with or without parallel computing option. It uses the user time from the print result of system.time() function.

### Usage

```
test.parallel(result = stop("A ResultNest object must be provided"))
```

### Arguments

result            A object obtained after searchR or likelihoodR

### Details

test.parallel estimates the likelihood of a set of parameters for nest incubation data with or without parallel computing option

### Value

The gain or loss of computing time using parallel version

### Author(s)

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(resultNest_4p_SSM)
test.parallel(resultNest_4p_SSM)

## End(Not run)
```

---

TRN_MHmcmc_p	<i>Generates set of parameters to be used with GRTRN_MHmcmc() or STRN_MHmcmc()</i>
--------------	--

---

**Description**

Interactive script used to generate set of parameters to be used with GRTRN\_MHmcmc() or STRN\_MHmcmc().

**Usage**

```
TRN_MHmcmc_p(
  result = NULL,
  parameters = NULL,
  fixed.parameters = NULL,
  accept = FALSE
)
```

**Arguments**

result	An object obtained after a SearchR fit
parameters	A set of parameters. Replace the one from result
fixed.parameters	A set of fixed parameters. Replace the one from result
accept	If TRUE, the script does not wait user information

**Details**

TRN\_MHmcmc\_p generates set of parameters to be used with GRTRN\_MHmcmc() or STRN\_MHmcmc()

**Value**

A matrix with the parameters

**Author(s)**

Marc Girondot



## Examples

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
# The initial parameters value can be:
# "T12H", "DHA", "DHH", "Rho25"
# Or
# "T12L", "T12H", "DHA", "DHH", "DHL", "Rho25"
#####
pfixed <- c(rK=1.208968)
M0 = 0.3470893
#####
# 4 parameters
#####
x <- structure(c(105.966881676793, 613.944134764125, 306.449533440186,
                118.193882815108), .Names = c("DHA", "DHH", "T12H", "Rho25"))
resultNest_4p_SSM <- searchR(parameters=x, fixed.parameters=pfixed,
                             temperatures=formatted, integral=integral.Gompertz, M0=M0,
                             hatchling.metric=c(Mean=39.33, SD=1.92))
data(resultNest_4p_SSM)
plot(resultNest_4p_SSM, xlim=c(0,70), ylimT=c(22, 32), ylimS=c(0,45), series=1,
      embryo.stages="Caretta caretta.SCL")
#####
pMCMC <- TRN_MHmcmc_p(resultNest_4p_SSM, accept=TRUE)

## End(Not run)
```

tsd

*Estimate the parameters that best describe temperature-dependent sex determination*

## Description

Estimate the parameters that best describe the thermal reaction norm for sex ratio when temperature-dependent sex determination occurs.

It can be used also to evaluate the relationship between incubation duration and sex ratio.

The parameter  $l$  was defined in Girondot (1999). The TRT is defined from the difference between the two boundary temperatures giving sex ratios of  $l$  and  $1 - l$ , respectively:

For logistic model (Girondot, 1999), it follows

$$TRT_l = abs(S K_l)$$

where  $K_l$  is a constant equal to  $2 \log\left(\frac{l}{1-l}\right)$ .

In Girondot (1999),  $l$  was 0.05 and then the TRT was defined as being the range of temperatures producing from 5% to 95% of each sex.

For other models, TRT is calculated numerically.

The basic model is logistic one. This model has the particularity to have a symmetric shape around  $P$ .

The other models have been built to alleviate this constraint. Hill and A-logistic models can be asymmetric, but it is impossible to control independently the low and high transitions. Hulin model is asymmetric but the control of asymmetry is difficult to manage. If asymmetric model is selected, it is always better to use flexit model.

$$\text{ifdose} < P \text{ then } (1 + (2^{K1} - 1) * \exp(4 * S1 * (P - x)))^{(-1/K1)}$$

$$\text{ifdose} > P \text{ then } 1 - ((1 + (2^{K2} - 1) * \exp(4 * S2 * (x - P)))^{(-1/K2)})$$

with:

$$S1 = S / ((4/K1) * (2^{(-K1)})^{(1/K1 + 1)} * (2^{K1} - 1))$$

$$S2 = S / ((4/K2) * (2^{(-K2)})^{(1/K2 + 1)} * (2^{K2} - 1))$$

## Usage

```
tsd(
  df = NULL,
  males = NULL,
  females = NULL,
  N = NULL,
  temperatures = NULL,
  durations = NULL,
  l = 0.05,
  parameters.initial = c(P = NA, S = -2, K = 0, K1 = 1, K2 = 0),
  males.freq = TRUE,
  fixed.parameters = NULL,
  equation = "logistic",
  replicate.CI = 10000,
  range.CI = 0.95,
  SE = TRUE,
  replicate.NullDeviance = 1000,
  control = list(maxit = 1000),
  print = TRUE
)
```

## Arguments

df	A dataframe with at least two columns named males, females or N and temperatures, Incubation.temperature or durations column
males	A vector with male numbers
females	A vector with female numbers
N	A vector with total numbers
temperatures	The constant incubation temperatures used to fit sex ratio
durations	The duration of incubation or TSP used to fit sex ratio
l	Sex ratio limits to define TRT are l and 1-l (see Girondot, 1999)
parameters.initial	Initial values for P, S or K search as a vector, ex. c(P=29, S=-0.3)

males.freq	If TRUE data are shown as males frequency
fixed.parameters	Parameters that will not be changed
equation	Can be "logistic", "Hill", "A-logistic", "Hulin", "Double-A-logistic", "flexit", "GSD", "logit", "probit"
replicate.CI	Number of replicates to estimate confidence intervals
range.CI	The range of confidence interval for estimation, default=0.95
SE	If FALSE, does not estimate SE of parameters. Can be use when something wrong happens.
replicate.NullDeviance	Number of replicates to estimate null distribution of deviance
control	List of parameters used in optim.
print	Should the results be printed at screen? TRUE (default) or FALSE

## Details

tsd estimates the parameters that best describe temperature-dependent sex determination

## Value

A list the pivotal temperature, transitional range of temperatures and their SE

## Author(s)

Marc Girondot

## References

- Girondot, M. 1999. Statistical description of temperature-dependent sex determination using maximum likelihood. *Evolutionary Ecology Research*, 1, 479-486.
- Godfrey, M.H., Delmas, V., Girondot, M., 2003. Assessment of patterns of temperature-dependent sex determination using maximum likelihood model selection. *Ecoscience* 10, 265-272.
- Hulin, V., Delmas, V., Girondot, M., Godfrey, M.H., Guillon, J.-M., 2009. Temperature-dependent sex determination and global change: are some species at greater risk? *Oecologia* 160, 493-506.
- Abreu-Grobois, F.A., Morales-Mérida, B.A., Hart, C.E., Guillon, J.-M., Godfrey, M.H., Navarro, E., Girondot, M., 2020. Recent advances on the estimation of the thermal reaction norm for sex ratios. *PeerJ* 8, e8451.

## See Also

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD.P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#)

## Examples

```
## Not run:
library(embryogrowth)
CC_AtlanticSW <- subset(DatabaseTSD, RMU=="Atlantic, SW" &
                        Species=="Caretta caretta" & (!is.na(Sexed) & Sexed!=0) &
                        !is.na(Correction.factor))
tsdL <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="logistic", replicate.CI=NULL))
tsdH <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="Hill", replicate.CI=NULL))
tsdR <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="A-logistic", replicate.CI=NULL))
tsdF <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="Flexit", replicate.CI=NULL))
tsdDR <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="Double-A-logistic", replicate.CI=NULL))
gsd <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                temperatures=Incubation.temperature-Correction.factor,
                                equation="GSD", replicate.CI=NULL))
compare_AIC(Logistic_Model=tsdL, Hill_model=tsdH, Alogistic_model=tsdR,
            flexit=tsdF,
            DoubleAlogistic_model=tsdDR, GSD_model=gsd)
compare_AICc(Logistic_Model=tsdL, Hill_model=tsdH, Alogistic_model=tsdR,
            DoubleAlogistic_model=tsdDR, GSD_model=gsd, factor.value = -1)
compare_BIC(Logistic_Model=tsdL, Hill_model=tsdH, Alogistic_model=tsdR,
            DoubleAlogistic_model=tsdDR, GSD_model=gsd, factor.value = -1)
#####
eo <- subset(DatabaseTSD, Species=="Emys orbicularis", c("Males", "Females",
              "Incubation.temperature"))

eo_Hill <- with(eo, tsd(males=Males, females=Females,
                      temperatures=Incubation.temperature,
                      equation="Hill"))
eo_Hill <- tsd(df=eo, equation="Hill", replicate.CI=NULL)
eo_logistic <- tsd(eo, replicate.CI=NULL)
eo_Alogistic <- with(eo, tsd(males=Males, females=Females,
                            temperatures=Incubation.temperature,
                            equation="a-logistic", replicate.CI=NULL))
### The Hulin model is a modification of A-logistic (See Hulin et al. 2009)

##### Caution
### It should not be used anymore as it can produce unexpected results
par <- eo_Alogistic$par
names(par)[which(names(par)=="K")] <- "K2"
par <- c(par, K1=0)
eo_Hulin <- with(eo, tsd(males=Males, females=Females,
                        parameters.initial=par,
```

```

                                temperatures=Incubation.temperature,
                                equation="Hulin", replicate.CI=NULL))

### The Double-A-logistic model is a A-logistic model with K1 and K2 using respectively
### below and above P

##### Caution
### The curve is not smooth at pivotal temperature

par <- eo_Alogistic$par
names(par)[which(names(par)=="K")] <- "K2"
par <- c(par, K1=as.numeric(par["K2"])*0.8)
par["K2"] <- par["K2"]*0.8
eo_Double_Alogistic <- with(eo, tsd(males=Males, females=Females,
                                parameters.initial=par,
                                temperatures=Incubation.temperature,
                                equation="Double-a-logistic", replicate.CI=NULL))

### The flexit model is modeled with K1 and K2 using respectively
### below and above P and smooth transition at P; S is the slope at P

par <- c(eo_logistic$par["P"], 1/4*eo_logistic$par["S"], K1=1, K2=1)
eo_flexit <- with(eo, tsd(males=Males, females=Females,
                        parameters.initial=par,
                        temperatures=Incubation.temperature,
                        equation="flexit", replicate.CI=NULL))

compare_AIC(Logistic=eo_logistic, Hill=eo_Hill, Alogistic=eo_Alogistic,
            Hulin=eo_Hulin, Double_Alogistic=eo_Double_Alogistic,
            flexit=eo_flexit)
## Note that SE for lower limit of TRT is wrong
plot(eo_flexit)
## To get correct confidence interval, check \code{tsd_MHmcmc()}.

### Note the asymmetry of the Double-A-logistic and flexit models
predict(eo_Double_Alogistic,
        temperatures=c(eo_Double_Alogistic$par["P"]-0.2, eo_Double_Alogistic$par["P"]+0.2))
predict(eo_Double_Alogistic)

(p <- predict(eo_flexit,
              temperatures=c(eo_flexit$par["P"]-0.3, eo_flexit$par["P"]+0.3)))
p["50%", 1]-0.5; 0.5-p["50%", 2]
predict(eo_flexit)

### It can be used also for incubation duration
CC_AtlanticSW <- subset(DatabaseTSD, RMU=="Atlantic, SW" &
                        Species=="Caretta caretta" & Sexed!=0)
tsdL_IP <- with (CC_AtlanticSW, tsd(males=Males, females=Females,
                                durations=IP.mean,
                                equation="logistic", replicate.CI=NULL))
plot(tsdL_IP, xlab="Incubation durations in days")
# Example with Chelonia mydas
cm <- subset(DatabaseTSD, Species=="Chelonia mydas" & !is.na(Sexed), c("Males", "Females",

```

```

                                "Incubation.temperature", "RMU"))
tsd(subset(cm, subset=RMU=="Pacific, SW"))
tsd(subset(cm, subset=RMU=="Pacific, Northwest"))
tsd(subset(cm, subset=RMU=="Atlantic, S Caribbean"))

### Eretmochelys imbricata
Ei_PacificSW <- subset(DatabaseTSD, RMU=="Pacific, SW" &
                      Species=="Eretmochelys imbricata")
Ei_AtlanticW <- subset(DatabaseTSD, RMU=="Atlantic, W (Caribbean and E USA)" &
                      Species=="Eretmochelys imbricata")
Ei_AtlanticSW <- subset(DatabaseTSD, RMU=="Atlantic, SW" &
                       Species=="Eretmochelys imbricata")
Ei_PacSW <- tsd(Ei_PacificSW)
Ei_AtlW <- tsd(Ei_AtlanticW)
Ei_AtlSW <- tsd(Ei_AtlanticSW)

plot(Ei_PacSW, xlim=c(27, 33), show.PTRT = FALSE, main=expression(italic("Eretmochelys imbricata")))
par(new=TRUE)
plot(Ei_AtlW, xlim=c(27, 33), col="red", xlab="", ylab="",
     axes=FALSE, xaxt="n", show.PTRT = FALSE, errbar.col="red")
par(new=TRUE)
plot(Ei_AtlSW, xlim=c(27, 33), col="blue", xlab="", ylab="", axes=FALSE,
     xaxt="n", show.PTRT = FALSE, errbar.col="blue")
legend("topright", legend=c("Pacific, SW", "Atlantic, W", "Atlantic, SW"), lty=1,
      col=c("black", "red", "blue"))

### Chelonia mydas
Cm_PacificSW <- subset(DatabaseTSD, RMU=="Pacific, SW" & !is.na(Sexed) &
                      Species=="Chelonia mydas")
Cm_PacificNW <- subset(DatabaseTSD, RMU=="Pacific, NW" & !is.na(Sexed) &
                      Species=="Chelonia mydas")
Cm_AtlanticSC <- subset(DatabaseTSD, RMU=="Atlantic, S Caribbean" & !is.na(Sexed) &
                       Species=="Chelonia mydas")
Cm_IndianSE <- subset(DatabaseTSD, RMU=="Indian, SE" & !is.na(Sexed) &
                      Species=="Chelonia mydas")
Cm_PacSW <- tsd(Cm_PacificSW)
Cm_PacNW <- tsd(Cm_PacificNW)
Cm_IndSE <- tsd(Cm_IndianSE)
Cm_AtlSC <- tsd(Cm_AtlanticSC)

plot(Cm_PacSW, xlim=c(24, 34), show.PTRT = FALSE, main=expression(italic("Chelonia mydas")))
par(new=TRUE)
plot(Cm_PacNW, xlim=c(24, 34), col="red", xlab="", ylab="",
     axes=FALSE, xaxt="n", show.PTRT = FALSE, errbar.col="red")
par(new=TRUE)
plot(Cm_IndSE, xlim=c(24, 34), col="blue", xlab="", ylab="",
     axes=FALSE, xaxt="n", show.PTRT = FALSE, errbar.col="blue")
par(new=TRUE)
plot(Cm_AtlSC, xlim=c(24, 34), col="green", xlab="", ylab="",
     axes=FALSE, xaxt="n", show.PTRT = FALSE, errbar.col="green")

# To fit a TSDII or FMF TSD pattern, you must indicate P_low, S_low, P_high, and S_high
# for logistic model and P_low, S_low, K1_low, K2_low, P_high, S_high, K1_high, and K2_high for

```

```

# flexit model
# The model must be 0-1 for low and 1-0 for high with P_low < P_high

Chelydra_serpentina <- subset(DatabaseTSD, !is.na(Sexed) & (Sexed != 0) &
                             Species=="Chelydra serpentina")

model_TSDII <- tsd(Chelydra_serpentina, males.freq=FALSE,
                  parameters.initial=c(P_low=21, S_low=0.3, P_high=28, S_high=-0.4),
                  equation="logistic")
plot(model_TSDII, lab.TRT = "TRT 1 = 5 %")
priors <- tsd_MHmcmc_p(result=model_TSDII, accept=TRUE)
out_mcmc <- tsd_MHmcmc(result=model_TSDII, n.iter=10000, parametersMCMC=priors)
plot(model_TSDII, resultmcmc=out_mcmc, lab.TRT = "TRT 1 = 5 %")
predict(model_TSDII, temperatures=25:35)

# Podocnemis expansa
Podocnemis_expansa <- subset(DatabaseTSD, !is.na(Sexed) & (Sexed != 0) &
                             Species=="Podocnemis expansa")
Podocnemis_expansa_Valenzuela_2001 <- subset(Podocnemis_expansa,
                                             Reference=="Valenzuela, 2001")
PeL2001 <- tsd(df=Podocnemis_expansa_Valenzuela_2001)
# The pivotal temperature is 32.133 °C (CI 95% 31.495;32.766)
# In Valenzuela, 2001: "Using data from the present study alone,
# the critical temperature was 32.2 °C by both methods and the 95%
# confidence limits were 31.4 °C and 32.9 °C."
# Data are close but not identical to what was published.

# The pivotal temperature calculated by maximum likelihood and by inverse
# prediction from logistic regression, was 32.6°C using raw data from
# 1991 (N. Valenzuela, unpublished data) and from this study. The lower
# and upper 95% confidence limits of the pivotal temperature were 32.2°C
# and 33.2°C,

Podocnemis_expansa_Valenzuela_1997 <- subset(Podocnemis_expansa,
                                             subset=((Reference=="Lance et al., 1992; Valenzuela et al., 1997") |
                                                    (Reference=="Valenzuela, 2001")) &
                                             (!is.na(Sexed)) & (Sexed != 0))

PeL1997 <- tsd(df=Podocnemis_expansa_Valenzuela_1997)

## End(Not run)

```

---

tsd\_MHmcmc

*Metropolis-Hastings algorithm for Sex ratio*


---

## Description

Run the Metropolis-Hastings algorithm for tsd.  
 Deeply modified from a MCMC script by Olivier Martin (INRA, Paris-Grignon).

The number of iterations is  $n.iter+n.adapt+1$  because the initial likelihood is also displayed. I recommend that  $thin=1$  because the method to estimate SE uses resampling. If initial point is maximum likelihood,  $n.adapt = 0$  is a good solution. To get the SE from `result_mcmc <- tsd_MHmcmc(result=try)`, use: `result_mcmc$BatchSE` or `result_mcmc$TimeSeriesSE`. The batch standard error procedure is usually thought to be not as accurate as the time series methods. Based on Jones, Haran, Caffo and Neath (2005), the batch size should be equal to  $\sqrt{n.iter}$ . Jones, G.L., Haran, M., Caffo, B.S. and Neath, R. (2006) Fixed Width Output Analysis for Markov chain Monte Carlo, *Journal of the American Statistical Association*, 101:1537-1547. coda package is necessary for this function. The parameters `intermediate` and `filename` are used to save intermediate results every 'intermediate' iterations (for example 1000). Results are saved in a file of name `filename`. The parameter `previous` is used to indicate the list that has been save using the parameters `intermediate` and `filename`. It permits to continue a mcmc search. These options are used to prevent the consequences of computer crash or if the run is very very long and processes at time limited.

## Usage

```
tsd_MHmcmc(
  result = stop("A result of tsd() fit must be provided"),
  n.iter = 10000,
  parametersMCMC = NULL,
  n.chains = 1,
  n.adapt = 0,
  thin = 1,
  trace = FALSE,
  traceML = FALSE,
  batchSize = sqrt(n.iter),
  adaptive = FALSE,
  adaptive.lag = 500,
  adaptive.fun = function(x) { ifelse(x > 0.234, 1.3, 0.7) },
  intermediate = NULL,
  filename = "intermediate.Rdata",
  previous = NULL
)
```

## Arguments

<code>result</code>	An object obtained after a SearchR fit
<code>n.iter</code>	Number of iterations for each step
<code>parametersMCMC</code>	A set of parameters used as initial point for searching with information on priors
<code>n.chains</code>	Number of replicates
<code>n.adapt</code>	Number of iterations before to store outputs
<code>thin</code>	Number of iterations between each stored output



trace	TRUE or FALSE or period, shows progress
traceML	TRUE or FALSE to show ML
batchSize	Number of observations to include in each batch fo SE estimation
adaptive	Should an adaptive process for SDProp be used
adaptive.lag	Lag to analyze the SDProp value in an adaptive content
adaptive.fun	Function used to change the SDProp
intermediate	Period for saving intermediate result, NULL for no save
filename	If intermediate is not NULL, save intermediate result in this file
previous	Previous result to be continued. Can be the filename in which intermediate results are saved.

### Details

tsd\_MHmcmc runs the Metropolis-Hastings algorithm for tsd (Bayesian MCMC)

### Value

A list with resultMCMC being mcmc.list object, resultLnL being likelihoods and parametersMCMC being the parameters used

### Author(s)

Marc Girondot

### See Also

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD.P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\(\)](#)

### Examples

```
## Not run:
library(embryogrowth)
eo <- subset(DatabaseTSD, Species=="Emys orbicularis", c("Males", "Females",
"Incubation.temperature"))

eo_logistic <- tsd(eo)
pMCMC <- tsd_MHmcmc_p(eo_logistic, accept=TRUE)
# Take care, it can be very long
result_mcmc_tsd <- tsd_MHmcmc(result=eo_logistic,
parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
n.adapt = 0, thin=1, trace=FALSE, adaptive=TRUE)
# summary() permits to get rapidly the standard errors for parameters
summary(result_mcmc_tsd)
plot(result_mcmc_tsd, parameters="S", scale.prior=TRUE, xlim=c(-3, 3), las=1)
plot(result_mcmc_tsd, parameters="P", scale.prior=TRUE, xlim=c(25, 35), las=1)

plot(eo_logistic, resultmcmc = result_mcmc_tsd)

1-rejectionRate(as.mcmc(result_mcmc_tsd))
```

```

raftery.diag(as.mcmc(result_mcmc_tsd))
heidel.diag(as.mcmc(result_mcmc_tsd))
library(car)
o <- P_TRT(x=eo_logistic, resultmcmc=result_mcmc_tsd)
outEo <- dataEllipse(x=o$P_TRT[, "PT"],
                    y=o$P_TRT[, "TRT"],
                    levels=c(0.95),
                    draw=FALSE)
plot(x = o$P_TRT[, "PT"],
     y=o$P_TRT[, "TRT"],
     pch=".", las=1, bty="n",
     xlab="Pivotal temperature",
     ylab=paste0("TRT ", as.character(100*eo_logistic$1), "%"),
     xlim=c(28.4, 28.6),
     ylim=c(0.8, 1.8))
lines(outEo[, 1], outEo[, 2], col="green", lwd=2)
legend("topleft", legend = c("Emys orbicularis", "95% confidence ellipse"),
      pch=c(19, NA), col=c("black", "green"), lty=c(0, 1), lwd=c(0, 2))

logistic <- function(x, P, S) {
  return(1/(1+exp((1/S)*(P-x))))
}

q <- as.quantile(result_mcmc_tsd, fun=logistic,
                xlim=seq(from=25, to=35, by=0.1), nameparxlim="x")
plot(x=seq(from=25, to=35, by=0.1), y=q[1, ], type="l", las=1,
     xlab="Temperatures", ylab="Male proportion", bty="n")
lines(x=seq(from=25, to=35, by=0.1), y=q[2, ])

## End(Not run)

```

---

tsd\_MHmcmc\_p

*Generates set of parameters to be used with tsd\_MHmcmc()*


---

## Description

Interactive script used to generate set of parameters to be used with `tsd_MHmcmc()`.

## Usage

```

tsd_MHmcmc_p(
  result = stop("An output from tsd() must be provided"),
  accept = FALSE
)

```

## Arguments

<code>result</code>	An object obtained after a <code>tsd</code> fit
<code>accept</code>	If TRUE, the script does not wait user information

**Details**

tsd\_MHmcmc\_p generates set of parameters to be used with tsd\_MHmcmc()

**Value**

A matrix with the parameters

**Author(s)**

Marc Girondot

**See Also**

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD](#), [P\\_TRT\(\)](#), [TSP.list](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
eo <- subset(DatabaseTSD, Species=="Emys orbicularis", c("Males", "Females",
  "Incubation.temperature"))
eo_logistic <- with(eo, tsd(males=Males, females=Females,
  temperatures=Incubation.temperature))
pMCMC <- tsd_MHmcmc_p(eo_logistic, accept=TRUE)
# Take care, it can be very long
result_mcmc_tsd <- tsd_MHmcmc(result=eo_logistic,
  parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
  n.adapt = 0, thin=1, trace=FALSE, adaptive=TRUE)
# summary() permits to get rapidly the standard errors for parameters
summary(result_mcmc_tsd)
plot(result_mcmc_tsd, parameters="S", scale.prior=TRUE, xlim=c(-3, 3), las=1)
plot(result_mcmc_tsd, parameters="P", scale.prior=TRUE, xlim=c(25, 35), las=1)

eo_flexit <- with(eo, tsd(males=Males, females=Females,
  parameters.initial=c(eo_logistic$par["P"],
    1/(4*eo_logistic$par["S"])),
  K1=1, K2=1),
  temperatures=Incubation.temperature,
  equation="flexit", replicate.CI=NULL))
pMCMC <- tsd_MHmcmc_p(eo_flexit, accept=TRUE)
result_mcmc_tsd <- tsd_MHmcmc(result=eo_flexit,
  parametersMCMC=pMCMC, n.iter=10000, n.chains = 1,
  n.adapt = 0, thin=1, trace=FALSE, adaptive=TRUE)
# summary() permits to get rapidly the standard errors for parameters
summary(result_mcmc_tsd)
plot(result_mcmc_tsd, parameters="S", scale.prior=TRUE, xlim=c(-3, 3), las=1)
plot(result_mcmc_tsd, parameters="P", scale.prior=TRUE, xlim=c(25, 35), las=1)
plot(result_mcmc_tsd, parameters="K1", scale.prior=TRUE, xlim=c(-10, 10), las=1)
plot(result_mcmc_tsd, parameters="K2", scale.prior=TRUE, xlim=c(-10, 10), las=1)

plot(eo_flexit, resultmcmc = result_mcmc_tsd)
```

```
## End(Not run)
```

---

TSP.list	<i>Database of thermosensitive period of development for sex determination</i>
----------	--

---

### Description

Database of thermosensitive period of development for sex determination.

This database can be used with the functions `plot()` or `info.nests()`.

The attributes `TSP.begin.stages` and `TSP.end.stages` for each dataframe give respectively the first and the last stages for TSP. Then the metrics for the limits of TSP are the average sizes before and after the TSP (see example, below).

If the metric for the stages before the TSP or after the TSP is not known, it will use the available information.

### Usage

```
TSP.list
```

### Format

A list with dataframes including attributes

### Details

Database of thermosensitive period of development for sex determination

### Author(s)

Marc Girondot <marc.girondot@universite-paris-saclay.fr>

### References

Mrosovsky, N., Pieau, C., 1991. Transitional range of temperature, pivotal temperatures and thermosensitive stages for sex determination in reptiles. *Amphibia-Reptilia* 12, 169-179.

### See Also

Other Functions for temperature-dependent sex determination: [DatabaseTSD.version\(\)](#), [DatabaseTSD](#), [P\\_TRT\(\)](#), [plot.tsd\(\)](#), [predict.tsd\(\)](#), [stages](#), [tsd\\_MHmcmc\\_p\(\)](#), [tsd\\_MHmcmc\(\)](#), [tsd\(\)](#)

**Examples**

```
## Not run:
library(embryogrowth)
data(TSP.list)
names(TSP.list)
reference <- "Emys_orbicularis.mass"
metric <- TSP.list[[reference]]
TSP.begin <- attributes(TSP.list[[reference]])$TSP.begin.stages
TSP.end <- attributes(TSP.list[[reference]])$TSP.end.stages
# Metric at the beginning of the TSP
del <- ifelse(all(metric$stages == TSP.begin - 1)==FALSE, 0, 1)
(metric$metric[metric$stages == TSP.begin - del] +
  metric$metric[metric$stages == TSP.begin]) / 2
# Metric at the end of the TSP
del <- ifelse(all(metric$stages == TSP.begin + 1)==FALSE, 0, 1)
(metric$metric[metric$stages == TSP.end] +
  metric$metric[metric$stages == del + TSP.end]) / 2

## End(Not run)
```

---

uncertainty.datalogger

*Uncertainty of average temperatures obtained using temperature data logger*

---

**Description**

Calculate the uncertainty of average temperature dependent on the characteristics of a data logger and sampling rate.

The temperature is supposed to be uniformly distributed with min and max being -accuracy and +accuracy.

**Usage**

```
uncertainty.datalogger(
  max.time = 0,
  sample.rate = 0,
  accuracy = 0.5,
  resolution = 1,
  replicates = 10000,
  method = function(x) { 2 * qnorm(0.975) * sd(x) }
)
```

**Arguments**

`max.time` being the maximum time to record in minutes  
`sample.rate` The sample rates in minutes

accuracy	The accuracy of the data logger in °C
resolution	The resolution of the data logger in °C
replicates	The number of replicates to estimate uncertainty.
method	The fonction that will be used to return the uncertainty.

### Details

uncertainty.datalogger Calculate the uncertainty of the average temperature calculated using data gathered by a data logger.

### Value

The function will return the uncertainty of the average temperature for the considered period as being the 95

### Author(s)

Marc Girondot

### See Also

Other Data loggers utilities: [calibrate.datalogger\(\)](#), [movement\(\)](#)

### Examples

```
## Not run:
library(embryogrowth)
# Exemple using the hypothesis of Gaussian distribution
uncertainty.datalogger(sample.rate=30, accuracy=1, resolution=0.5,
  method=function(x) {2*qnorm(0.975)*sd(x)})
# Example without hypothesis about distribution, using quantiles
uncertainty.datalogger(sample.rate=30, accuracy=1, resolution=0.5,
  method=function(x) {quantile(x, probs=c(0.975))-
    quantile(x, probs=c(0.025))})

par(mar=c(4, 4, 1, 1))
plot(x=10:120, uncertainty.datalogger(sample.rate=10:120,
  accuracy=0.5,
  resolution=1),
  las=1, bty="n", type="l",
  xlab="Sample rate in minutes",
  ylab=expression("Uncertainty in "*degree*"C"),
  ylim=c(0, 0.15), xlim=c(0, 120))
lines(x=10:120, uncertainty.datalogger(sample.rate=10:120,
  accuracy=1,
  resolution=0.5), col="red")
lines(x=10:120, uncertainty.datalogger(sample.rate=10:120,
  accuracy=1,
  resolution=1), col="blue")
lines(x=10:120, uncertainty.datalogger(sample.rate=10:120,
  accuracy=0.5,
  resolution=0.5), col="yellow")
```

```
legend("topleft", legend=c("Accuracy=0.5, resolution=0.5",
                           "Accuracy=0.5, resolution=1",
                           "Accuracy=1, resolution=0.5",
                           "Accuracy=1, resolution=1"), lty=1,
      col=c("yellow", "black", "red", "blue"),
      cex=0.6)

## End(Not run)
```

---

web.tsd

*Run a shiny application for basic functions of tsd function*

---

## Description

Run a shiny application for basic functions of tsd function.

## Usage

```
web.tsd()
```

## Details

web.tsd runs a shiny application for basic functions of tsd function

## Value

Nothing

## Author(s)

Marc Girondot

## Examples

```
## Not run:
library(embryogrowth)
web.tsd()

## End(Not run)
```

---

weightmaxentropy	<i>Search for the weights of the nests which maximize the entropy of nest temperatures distribution</i>
------------------	---

---

### Description

Search for the weights of the nests which maximize the entropy of nest temperatures distribution. Entropy is measured by Shanon index.  
 Entropy method must be entropy.empirical because it is the only method insensitive to scaling.  
 If no weight is given, the initial weight is uniformly distributed.  
 Use control\_optim=list(trace=0) for not show progress of search report.

### Usage

```
weightmaxentropy(
  temperatures = stop("Temperature data must be provided !"),
  weight = NULL,
  entropy.method = entropy::entropy.empirical,
  plot = TRUE,
  control_optim = list(trace = 0, maxit = 500),
  control_plot = NULL,
  control_entropy = NULL,
  col = c("black", "red")
)
```

### Arguments

temperatures	Timeseries of temperatures formated using FormatNests()
weight	A named vector of the initial weight search for each nest for likelihood estimation
entropy.method	Entropy function, for example entropy::entropy.empirical. See package entropy for description
plot	Do the plot of temperatures before and after weight must be shown ? TRUE or FALSE
control_optim	A list with control paramaters for optim function
control_plot	A list with control paramaters for plot function
control_entropy	A list with control paramaters for entropy function
col	Colors for unweighted and weighted distributions

### Details

Search for the weights of the nests which maximize the entropy of nest temperatures distribution



**Value**

A named vector of weights

**Author(s)**

Marc Girondot

**Examples**

```
## Not run:
library(embryogrowth)
data(nest)
formatted <- FormatNests(nest)
w <- weightmaxentropy(formatted, control_plot=list(xlim=c(20,36)))
x <- structure(c(120.940334922916, 467.467455887442,
306.176613681557, 117.857995419495),
.Names = c("DHA", "DHH", "T12H", "Rho25"))
# pfixed <- c(K=82.33) or rK=82.33/39.33
pfixed <- c(rK=2.093313)
# K or rK are not used for dydt.linear or dydt.exponential
resultNest_4p_weight <- searchR(parameters=x,
fixed.parameters=pfixed, temperatures=formatted,
integral=integral.Gompertz, M0=1.7, hatchling.metric=c(Mean=39.33, SD=1.92),
method = "BFGS", weight=w)
data(resultNest_4p_weight)
plotR(resultNest_4p_weight, ylim=c(0,0.50), xlim=c(15, 35))
# Standard error of parameters can use the GRTRN_MHmcmc() function

## End(Not run)
```

# Index

- \* **Data loggers utilities**
  - calibrate.datalogger, 7
  - movement, 53
  - uncertainty.datalogger, 133
- \* **Ecology**
  - embryogrowth-package, 3
- \* **Embryo**
  - embryogrowth-package, 3
- \* **Functions for temperature-dependent sex determination**
  - DatabaseTSD, 12
  - DatabaseTSD.version, 14
  - P\_TRT, 78
  - plot.tsd, 67
  - predict.tsd, 76
  - stages, 103
  - tsd, 121
  - tsd\_MHmcmc, 127
  - tsd\_MHmcmc\_p, 130
  - TSP.list, 132
- \* **Gompertz**
  - embryogrowth-package, 3
- \* **Growth**
  - embryogrowth-package, 3
- \* **Hatching success**
  - HatchingSuccess.fit, 25
  - HatchingSuccess.lnL, 27
  - HatchingSuccess.MHmcmc, 29
  - HatchingSuccess.MHmcmc\_p, 31
  - HatchingSuccess.model, 33
  - logLik.HatchingSuccess, 48
  - nobs.HatchingSuccess, 58
  - plot.HatchingSuccess, 60
  - predict.HatchingSuccess, 75
- \* **Sex-determination**
  - embryogrowth-package, 3
- \* **Temperature**
  - embryogrowth-package, 3
- \* **datasets**
  - DatabaseNestingArea, 11
  - DatabaseTSD, 12
  - nest, 57
  - resultNest\_3p\_Dallwitz, 80
  - resultNest\_3p\_Weibull, 81
  - resultNest\_4p\_normal, 82
  - resultNest\_4p\_SSM, 83
  - resultNest\_4p\_SSM\_Linear, 84
  - resultNest\_4p\_transition, 85
  - resultNest\_4p\_trigo, 86
  - resultNest\_4p\_weight, 87
  - resultNest\_5p\_Dallwitz, 88
  - resultNest\_6p\_SSM, 89
  - resultNest\_mcmc\_4p\_SSM, 90
  - resultNest\_mcmc\_4p\_SSM\_Linear, 91
  - resultNest\_mcmc\_6p\_SSM, 92
  - resultNest\_mcmc\_newp, 93
  - resultNest\_newp, 95
  - stages, 103
  - tempConst, 118
  - TSP.list, 132
- calibrate.datalogger, 7, 54, 134
- ChangeSSM, 8
- DatabaseNestingArea, 11
- DatabaseTSD, 12, 14, 69, 77, 80, 104, 123, 129, 131, 132
- DatabaseTSD.version, 13, 14, 69, 77, 80, 104, 123, 129, 131, 132
- dydt.exponential, 14
- dydt.Gompertz, 15
- dydt.linear, 16
- embryogrowth-package, 3
- FormatNests, 17
- Generate\_hatchling\_metric, 21
- GenerateAnchor, 19
- GenerateConstInc, 20

- GRTRN\_MHmcmc, 22
- HatchingSuccess.fit, 25, 28, 31–33, 49, 58, 62, 76
- HatchingSuccess.lnL, 26, 27, 31–33, 49, 58, 62, 76
- HatchingSuccess.MHmcmc, 26, 28, 29, 32, 33, 49, 58, 62, 76
- HatchingSuccess.MHmcmc\_p, 26, 28, 31, 31, 33, 49, 58, 62, 76
- HatchingSuccess.model, 26, 28, 31, 32, 33, 49, 58, 62, 76
- hist.Nests, 34
- hist.NestsResult, 35
  
- info.nests, 36
- integral.exponential, 43
- integral.Gompertz, 45
- integral.linear, 46
  
- likelihoodR, 47
- logLik.HatchingSuccess, 26, 28, 31–33, 48, 58, 62, 76
- logLik.NestsResult, 50
- logLik.STRN, 51
- logLik.tsd, 52
  
- movement, 8, 53, 134
- MovingIncubation, 54
  
- nest, 57
- nobs.HatchingSuccess, 26, 28, 31–33, 49, 58, 62, 76
- nobs.NestsResult, 59
  
- P\_TRT, 13, 14, 69, 77, 78, 104, 123, 129, 131, 132
- plot.HatchingSuccess, 26, 28, 31–33, 49, 58, 60, 76
- plot.NestsResult, 63
- plot.tsd, 13, 14, 67, 77, 80, 104, 123, 129, 131, 132
- plot\_transition, 73
- plotR, 70
- predict.HatchingSuccess, 26, 28, 31–33, 49, 58, 62, 75
- predict.tsd, 13, 14, 69, 76, 80, 104, 123, 129, 131, 132
  
- resultNest\_3p\_Dallwitz, 80
- resultNest\_3p\_Weibull, 81
- resultNest\_4p\_normal, 82
- resultNest\_4p\_SSM, 83
- resultNest\_4p\_SSM\_Linear, 84
- resultNest\_4p\_transition, 85
- resultNest\_4p\_trigo, 86
- resultNest\_4p\_weight, 87
- resultNest\_5p\_Dallwitz, 88
- resultNest\_6p\_SSM, 89
- resultNest\_mcmc\_4p\_SSM, 90
- resultNest\_mcmc\_4p\_SSM\_Linear, 91
- resultNest\_mcmc\_6p\_SSM, 92
- resultNest\_mcmc\_newp, 93
- resultNest\_newp, 95
  
- searchR, 96
- stages, 13, 14, 69, 77, 80, 103, 123, 129, 131, 132
- STRN, 107
- STRN\_MHmcmc, 113
- summary.Nests, 116
- switch.transition, 117
  
- tempConst, 118
- test.parallel, 119
- TRN\_MHmcmc\_p, 120
- tsd, 13, 14, 69, 77, 80, 104, 121, 129, 131, 132
- tsd\_MHmcmc, 13, 14, 69, 77, 80, 104, 123, 127, 131, 132
- tsd\_MHmcmc\_p, 13, 14, 69, 77, 80, 104, 123, 129, 130, 132
- TSP.list, 13, 14, 69, 77, 80, 104, 123, 129, 131, 132
  
- uncertainty.datalogger, 8, 54, 133
  
- web.tsd, 135
- weightmaxentropy, 136