Package ‘PAMpal’

July 12, 2024

Type Package

Title Load and Process Passive Acoustic Data

Version 1.2.1

Maintainer Taiki Sakai <taiki.sakai@noaa.gov>


License GNU General Public License

Encoding UTF-8

LazyData true

Depends R (>= 3.5.0), dplyr (>= 1.1.1)

Imports PamBinaries (>= 1.3.0), PAMmisc (>= 1.12.0), tuneR, seewave, gam, data.table, RSQLite, purrr, methods, signal, tidyr, ggplot2, knitr, xml2, rlang, reticulate, lubridate, geosphere, shiny

RoxygenNote 7.3.1

Suggests testthat

NeedsCompilation no

Author Taiki Sakai [aut, cre],
Jay Barlow [ctb],
Emily Griffiths [ctb],
Michael Oswald [ctb],
Simone Baumann-Pickering [ctb],
Julie Oswald [ctb]

Repository CRAN

Date/Publication 2024-07-11 22:50:02 UTC
# Contents

- AcousticEvent-class .................................................. 3
- AcousticStudy-class .................................................... 4
- addAnnotation ............................................................. 4
- addBinaries ............................................................... 6
- addCalibration ............................................................ 7
- addDatabase .............................................................. 8
- addFPOD .................................................................. 9
- addFunction .............................................................. 10
- addGps ................................................................ 11
- addHydrophoneDepth ................................................... 12
- addMeasures .............................................................. 13
- addNote ................................................................... 14
- addRecordings ............................................................. 15
- addSettings ............................................................... 16
- addWaveHeight ........................................................... 17
- bindStudies ............................................................... 18
- calculateAverageSpectra .............................................. 19
- calculateEchoDepth .................................................... 21
- calculateICI ............................................................... 23
- calculateModuleData .................................................... 25
- checkStudy .............................................................. 26
- export_banter ............................................................ 27
- exStudy ................................................................. 28
- filter.AcousticStudy .................................................... 29
- filterEchoDepths ........................................................ 30
- getBinaryData .......................................................... 31
- getClipData ............................................................... 32
- getDetectorData ......................................................... 33
- getWarnings ............................................................. 35
- is.AcousticEvent ......................................................... 36
- is.AcousticStudy ......................................................... 36
- is.PAMpalSettings ....................................................... 36
- loadPamguardXML ....................................................... 37
- markAnnotated .......................................................... 37
- matchEnvData.AcousticEvent-method ............................. 39
- matchTimeData .......................................................... 41
- PAMpal.accessors ......................................................... 42
- PAMpalSettings ......................................................... 47
- PAMpalSettings-class .................................................. 48
- plotDataExplorer ........................................................ 49
- plotGram ................................................................. 49
- plotWaveform ............................................................ 51
- processPgDetections ..................................................... 53
- removeBinaries ........................................................... 55
- removeCalibration ....................................................... 56
- removeDatabase .......................................................... 57
AcousticEvent-class

Description
An S4 class storing acoustic detections from an Acoustic Event as well as other related metadata

Slots
- id: unique id or name for this event
- detectors: a list of data frames that have acoustic detections and any measurements calculated on those detections. Each data frame is named by the detector that made the detection
- localizations: a named list storing localizations, named by method
- settings: a list of recorder settings
- species: a list of species classifications for this event, named by classification method (ie. BANTER model, visual ID)
- files: a list of files used to create this object, named by the type of file (ie. binaries, database)
- ancillary: a list of miscellaneous extra stuff. Store whatever you want here

Author(s)
Taiki Sakai <taiki.sakai@noaa.gov>
**AcousticStudy-class  AcousticStudy Class**

**Description**

An S4 class storing acoustic data from an entire AcousticStudy

**Slots**

- `id` a unique id for the study
- `events` a list of AcousticEvent objects with detections from the AcousticStudy
- `files` a list of folders and files containing the AcousticStudy data
- `gps` a data frame of gps coordinates for the entire AcousticStudy
- `pps` the PAMPalSettings object used to create this object
- `settings` a named list of various settings for detectors, localizers, etc.
- `effort` something about effort lol
- `models` a place to store any models run on your data
- `ancillary` miscellaneous extra data

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>

---

**addAnnotation  Add Annotation Data to an AcousticStudy Object**

**Description**

Adds annotation data to an AcousticStudy object, usually in preparation for exporting to an external source. Most pieces of the annotation form will be filled in by the user when the function is called, but `lat`, `lon`, `time_start`, `time_end`, `freq_low`, `freq_high`, `source_id`, and `annotation_id` will be filled in automatically based on data in each AcousticEvent. Annotations are stored for each event in the ancillary slot.

**Usage**

```r
addAnnotation(x, anno, verbose = TRUE)
```

```r
prepAnnotation(
  x,
  specMap = NULL,
  mode = c("event", "detection"),
  interactive = FALSE,
)```
addAnnotation

getAnnotation(x)
checkAnnotation(x)
export_annomate(x, file = NULL)
matchRecordingUrl(anno, rec)

Arguments

x an AcousticStudy or AcousticEvent object
anno an annotation dataframe
verbose logical flag to print messages
specMap data.frame to map species ids in x to names to be used for the annotation (e.g. from 'ZC' to 'Ziphius cavirostris'). Dataframe must have columns old and new
mode one of 'event' or 'detection' to create annotation for events or detections
interactive logical flag to fill annotation data interactively (not recommended)
... additional named arguments to fill in annotation data. If names match a column in the annotation, that value will be used for all events or detections in the annotation
file file to write a CSV of the annotations to, if NULL (default) then no file will be written
rec dataframe of recording url information. Must have column recording_url. If clips were created using writeEventClips, then must have column filename containing the wav file names. Other column names will be automatically parsed from there. If wav files are from another source, must contain columns matchId

Value

The same object as x with an $annotation item added the the ancillary slot of each event in x

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
addBinaries

Add Binaries to a PAMpalSettings Object

Description

Adds more binary files to the "binaries" slot of a PAMpalSettings object. Interactively asks user to supply folder location if not provided.

Usage

addBinaries(pps, folder = NULL, verbose = TRUE)

Arguments

pps a PAMpalSettings object to add binary files to
folder a folder of binaries to add, all subfolders will also be added
verbose logical flag to show messages

Value

the same PAMpalSettings object as pps, with the binary files contained in folder added to the "binaries" slot. Only binary files for Click Detector and WhistlesMoans modules will be added, since these are the only types PAMpal currently knows how to process

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

# not recommended to create PPS like this, for example only
pps <- new('PAMpalSettings')
binFolder <- system.file('extdata', 'Binaries', package='PAMpal')
pps <- addBinaries(pps, binFolder)
pps
addCalibration

Add a Calibration File to a PAMpalSettings Object

Description

Adds a new calibration function to the "calibration" slot of a PAMpalSettings object. Interactively asks user to supply file and other parameters if not supplied.

Usage

addCalibration(
  pps,
  calFile = NULL,
  module = "ClickDetector",
  calName = NULL,
  all = FALSE,
  units = NULL
)

applyCalibration(pps, module = "ClickDetector", all = FALSE)

Arguments

pps  a PAMpalSettings object to add a database to

calFile  a calibration file name. Must be csv format with two columns. The first column must be the frequency (in Hz), and the second column must be the sensitivity (in dB), and the columns should be labeled Frequency and Sensitivity. Can also be supplied as a dataframe in which case the calName argument should also be set

module  the Pamguard module type this calibration should be applied to, for now this is only for ClickDetector modules. This is left as an option for future-proofing purposes but should not be needed.

calName  the name to assign to the calibration function, defaults to the file name and only needs to be set if supplying a dataframe instead of a csv file

all  logical flag whether or not to apply calibration to all functions without asking individually, recommended to stay as FALSE

units  a number from 1 to 3 specifying the units of the calibration file, number corresponds to dB re V/uPa, uPa/Counts, or uPa/FullScale respectively. A NULL (default) or other value will prompt user to select units.

Details

When adding a calibration, you will be asked what units your calibration value is in. The wave clips stored by Pamguard are values from -1 to 1, so if your calibration is expecting different units then this needs to be accounted for in order to get an accurate SPL value. For V / uPa you must know the voltage range of your recording equipment, and for calibrations expecting Count data you must
know the bit rate of your recordings. If your calibration is already relative to full-scale, then nothing needs to be adjusted. If you don’t know the units of your calibration and you are only interested in relative dB levels, then you can select the Full-Scale options.

The calibration function created takes frequency (in Hz) as input and outputs the associated dB value that needs to be added to correct the power spectrum of a signal. If the input is a matrix or dataframe, the first column is assumed to be frequency.

Value

the same PAMpalSettings object as pps, with the calibration function added to the calibration slot.

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

```r
pps <- new('PAMpalSettings')
calFile <- system.file('extdata', 'calibration.csv', package='PAMpal')
pps <- addCalibration(pps, calFile, all = TRUE, units=3)
calClick <- function(data, calibration=NULL) {
  standardClickCalcs(data, calibration=calibration, filterfrom_khz = 0)
}
pps <- addFunction(pps, calClick, module = 'ClickDetector')
pps <- applyCalibration(pps, all=TRUE)
pps
```

---

**addDatabase**

Add a Database to a PAMpalSettings Object

**Description**

Adds a new function to the “function” slot in a PAMpalSettings object. Interactively asks for database files if none are supplied as input.

**Usage**

```r
addDatabase(pps, db = NULL, verbose = TRUE)
```

**Arguments**

- **pps** a PAMpalSettings object to add a database to
- **db** database(s) to add, or single directory containing databases
- **verbose** logical flag to show messages
addFPOD

Value

the same PAMpalSettings object as pps, with the database db added to the "db" slot

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

# not recommended to create a pps like this, for example only
pps <- new('PAMpalSettings')
db <- system.file('extdata', 'Example.sqlite3', package='PAMpal')
pps <- addDatabase(pps, db)
pps

addFPOD

Add FPOD Detector to an AcousticStudy

Description

Adds data from FPOD detector CSV files to an AcousticStudy object as new detectors of type "fpod"

Usage

addFPOD(x, fpod, detectorName = "FPOD")

Arguments

x an AcousticStudy object
fpod path(s) to CSV files containing FPOD detector output
detectorName name for the detector, the default 'FPOD' should be fine unless you want to differentiate between multiple FPOD detectors

Details

FPOD detections are added to events based on their times. All detections between the start and end times events. NOTE: most PAMpal functions were designed with only PAMGuard data in mind, there is a chance that adding FPOD detections will cause other advanced functionality to not work.

Behavior is slightly different depending on how the original AcousticStudy was created. For those processed with mode='db', the start and end times for each event are just determined by the times of detections within the event.

For those processed with mode='recording' or mode='time', the start and end times for each event are determined by the start/end times of the recording files or the grouping file provided initially. This means that it is possible that there are events which initially had zero PAMGuard detections that now have FPOD detections. In these cases a new AcousticEvent will be created that only has FPOD detections, these events may not work with a variety of other PAMpal functions.
addFunction

**Value**
the same object as x with FPOD detector data added

**Author(s)**
Taiki Sakai <taiki.sakai@noaa.gov>

---

**addFunction**  *Add a Function to a PAMpalSettings Object*

**Description**
Adds a new function to the "function" slot in a PAMpalSettings object. Must be run interactively, user will be prompted to assign values for any parameters in the function to be added

**Usage**
addFunction(pps, fun, module = NULL, verbose = TRUE, ...)

**Arguments**
- **pps**
a PAMpalSettings object to add a function to
- **fun**
function to add OR another PAMpalSettings object. In this case all functions from the second object will be added to pps
- **module**
Pamguard module output this function should act on, one of ClickDetector, WhistlesMoans, Cepstrum, or GPLDetector. If NULL (default), user will be prompted to select which module it applies to
- **verbose**
logical flag to show messages
- **...**
named arguments to pass to function being added

**Value**
the same PAMpalSettings object as pps, with the function fun added to the "functions" slot

**Author(s)**
Taiki Sakai <taiki.sakai@noaa.gov>

**Examples**

# not recommended to create a pps like this, for example only
pps <- new('PAMpalSettings')
if(interactive()) pps <- addFunction(pps, standardClickCalcs)
pps <- addFunction(pps, roccaWhistleCalcs, module='WhistlesMoans')
addGps

Add GPS Locations to an AcousticStudy

Description

Add GPS Lat / Long to an AcousticStudy or AcousticEvent. If GPS data is not present in any of the databases, user will interactively be asked to provide GPS data to add.

Usage

addGps(x, gps = NULL, thresh = 3600, ...)

### S4 method for signature 'data.frame'
addGps(x, gps, thresh = 3600, keepDiff = FALSE, ...)

### S4 method for signature 'AcousticEvent'
addGps(x, gps = NULL, thresh = 3600, ...)

### S4 method for signature 'list'
addGps(x, gps = NULL, thresh = 3600, ...)

### S4 method for signature 'AcousticStudy'
addGps(x, gps = NULL, thresh = 3600, ...)

### S4 method for signature 'ANY'
addGps(x, gps = NULL, thresh = 3600, ...)

Arguments

- **x**: data to add GPS coordinates to. Must have a column UTC, and can also have an optional column Channel.
- **gps**: a data frame of GPS coordinates to match to data from x. Must have columns UTC, Latitude, Longitude, and optionally Channel. If not provided and x is an AcousticEvent or AcousticStudy object, then the gps data will be read from the databases contained in the files slot of x.
- **thresh**: maximum time apart in seconds for matching GPS coordinates to data, if the closest coordinate is more than thresh apart then the Latitude and Longitude values will be set to NA.
- **...**: additional arguments for other methods
- **keepDiff**: logical flag to keep time difference column (between GPS time and data time).

Details

Latitude and Longitude coordinates will be matched to the data by interpolating between points in the provided GPS data. After the interpolating is done, the time difference between the matched rows is checked and any that are greater than the set threshold are set to NA. This is done to prevent
accidentally matching weird things if an incomplete set of GPS data is provided. An approximate
distance between the interpolated points and the closest known GPS point is provided as a "gp-
sUncertainty" column (distance in meters).

If x is an AcousticEvent or AcousticStudy, then gps can be omitted and will be read from the
databases contained in the files slot of x. If x is an AcousticStudy, then the gps data will also be
saved to the gps slot of the object, and an additional argument bounds can be provided. This is a
length two vector of POSIXct class times that will bound the times of gps data to store, gps data
outside this range will not be stored (to reduce the potentially very large amount of data stored in
the gps slot)

Value

the same data as x, with Lat/Long data added. AcousticStudy objects will have all GPS data used
added to the "gps" slot, and all AcousticEvents will have Latitude and Longitude added to all de-
tector dataframes

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
  # need to update database file to local directory
db <- system.file('extdata', 'Example.sqlite3', package='PAMpal')
exStudy <- updateFiles(exStudy, db=db, bin=NA, verbose=FALSE)
exStudy <- addGps(exStudy)
head(gps(exStudy))

addHydrophoneDepth

Add Hydrophone Depth Data to an AcousticStudy

Description

Add hydrophone depth to an AcousticStudy or AcousticEvent

Usage

addHydrophoneDepth(x, depth = NULL, depthCol = NULL, thresh = 60, ...)

Arguments

x an AcousticStudy to add depth data to

depth a CSV or data frame of depth values to match to data from x. Must have column
UTC, and a column containing depth data to be specified by depthCol. If not
provided and x is an AcousticEvent or AcousticStudy object, then the depth
data will be read from the databases contained in the files slot of x
addMeasures

Description

Adds "measures" to an AcousticStudy or AcousticEvent. A "measure" is an event-level variable that will be exported alongside data from that event.

Usage

addMeasures(x, measures, replace = TRUE)

getMeasures(x)
addNote

Arguments

x 
an AcousticStudy or AcousticEvent object

measures the measures to add. Can either be a named list, where names match event
names of x or a dataframe with column eventId matching the event names of
x. If a list, every item within the list must also be named by the variable name.
All other data within measures will be added as new measures

replace logical flag whether or not to replace

Value

object of same class as x with measures added

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
measList <- list('Example.OE1' = list(a=1, b=2),
                 'Example.OE2' = list(a=2, b=3)
                 )
exMeasure <- addMeasures(exStudy, measList)
print(getMeasures(exMeasure))
measDf <- data.frame(eventId = c('Example.OE1', 'Example.OE2'),
                     a=4:5,
                     b=6:7)
exMeasure <- addMeasures(exMeasure, measDf, replace=TRUE)
getMeasures(exMeasure)

Description

Adds a note to an AcousticEvent or AcousticStudy. Notes can either be accessed with the "get-
Notes" function, or up to 6 notes will be printed when the object is printed

Usage

addNote(x, to = c("study", "event"), evNum = 1, label = NULL, note)

getNotes(x)
Add Recordings to an AcousticStudy Object

Description

Adds recording files to an AcousticStudy object, runs interactively to allow users to select files if they are not provided. No actual recordings are stored, a dataframe containing information on the start and end times of the recording files is added to the object.

Usage

addRecordings(x, folder = NULL, log = FALSE, progress = TRUE)
**Arguments**

- **x**: a `AcousticStudy` object to add recordings to.
- **folder**: a folder of recordings to add. If `NULL`, user will be prompted to select a folder of recordings for each database present in `x`. If a single folder, this will be applied to all databases. If multiple folders, length must be equal to the number of databases and they will be applied to each database in the provided order.
- **log**: (optional) log files for SoundTrap recordings. These are used to adjust apparent lengths of recordings for missing data. If `NULL`, user will be prompted to provide a folder (selecting no folder is a valid option here). If `FALSE` this step will be skipped. If a single folder or multiple folders will be applied similar to `folder`.
- **progress**: logical flag to show progress bars.

**Value**

The same object as `x` with recording information added to the `files` slots. The information added is a dataframe containing the start and end times of recording.

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>

**Examples**

```r
data(exStudy)
recs <- system.file('extdata', 'Recordings', package='PAMpal')
exStudy <- addRecordings(exStudy, folder=recs, log=FALSE, progress=FALSE)
files(exStudy)$recordings
```

---

**addSettings**

*Add Settings to a PAMpalSettings Object*

**Description**

Adds settings to a PAMpalSettings object, usually from an XML file created by Pamguard’s "Export XML Configuration".

**Usage**

```r
addSettings(pps, settings = NULL, type = c("xml", "list"), verbose = TRUE)
```

**Arguments**

- **pps**: a `PAMpalSettings` object to add settings to.
- **settings**: settings to add, either an XML file or a
- **type**: one of `xml` or `list` indicating type of settings to add
- **verbose**: logical flag to show messages
addWaveHeight

Value
the same PAMpalSettings object as pps, with a new list of settings replacing what was previously in the "settings" slot

Author(s)
Taiki Sakai <taiki.sakai@noaa.gov>

Examples
# not recommended to create PPS like this, for example only
pps <- new('PAMpalSettings')
xmSettings <- system.file('extdata', 'Example.xml', package='PAMpal')
pps <- addSettings(pps, xmSettings, type='xml')

Description
Add Wave Height Data to an AcousticStudy

Usage
addWaveHeight(x, height, thresh = 3600)

Arguments
x an AcousticStudy to add height data to
height either a single numeric value, or a dataframe with column UTC and either column waveHeight specifying height (m) at that time, or beaufort specifying the beaufort sea state at that time
thresh maximum time apart in seconds for matching height to data, if the closest value is more than thresh apart then the height value will be set to NA

Details
height values will be matched to the data by using data.table's rolling join with roll='nearest'. After the join is done, the time difference between the matched rows is checked and any that are greater than the set threshold are set to NA. This is done to prevent accidentally matching weird things if an incomplete set of height data is provided.

Value
the same as x, with wave height data added. All AcousticEvents will have height data added to all detector dataframes as column waveHeight
Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
# need to update database file to local directory
exStudy <- addWaveHeight(exStudy, height=.5)
getClickData(exStudy[1])

bindStudies

Combine AcousticStudy Objects

Description

Combines multiple AcousticStudy objects (or lists of these) into a single object

Usage

bindStudies(…)

Arguments

… AcousticStudy objects, or a list of AcousticStudy objects

Details

All events will be combined into one large list of events. Files, settings, effort, models, GPS, and ancillary fields will be combined using the squishList function from the PAMmisc package (dataframes are combined, vectors are appended). The id is changed by pasting all IDs together along with a note that they have been combined. Note that the PAMPalSettings object in the pps slot is just left as the pps in the first AcousticStudy to be combined, and thus is not representative of the new combined AcousticStudy

Value

A single AcousticStudy object

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
calculateAverageSpectra

Calculate Average Spectra of Clicks

Description

Calculates the average spectra of all the clicks present in an event

Usage

```r
calculateAverageSpectra(
  x,
  evNum = 1,
  calibration = NULL,
  wl = 512,
  channel = 1:2,
  filterfrom_khz = 0,
  filterto_khz = NULL,
  sr = NULL,
  snr = 0,
  norm = TRUE,
  plot = TRUE,
  noise = FALSE,
  decimate = 1,
  sort = FALSE,
  mode = "spec",
  title = NULL,
  ylim = NULL,
  flim = NULL,
  cmap = hcl.colors(30, "YlOrRd", rev = TRUE),
  brightness = 0,
  contrast = 0,
  q = 0.01,
  showBreaks = TRUE,
  ...
)
```

Arguments

- **x**: an `AcousticEvent` or `AcousticStudy` object
- **evNum**: if `x` is a study, the event index number to calculate the average spectra for. Note that this is the index in the order that they appear in the `AcousticStudy` object, not the actual event number. Alternatively full event names can be used
- **calibration**: a calibration function to apply, if desired
- **wl**: the size of the click clips to use for calculating the spectrum. If greater than the clip present in the binary, clip will be zero padded
calculateAverageSpectra

channel channel(s) to include in calculations. Currently does not correspond to actual channel in instrument, just the order present in the binary file

filterfrom_khz frequency in khz of highpass filter to apply, or the lower bound of a bandpass filter if filterto_khz is not NULL

filterto_khz if a bandpass filter is desired, set this as the upper bound. If only a highpass filter is desired, leave as the default NULL value. Currently only highpass and bandpass filters are supported, so if filterfrom_khz is left as zero then this parameter will have no effect

sr a sample rate to use if the sample rate present in the database needs to be overridden (typically not needed)

snr minimum signal-to-noise ratio to be included in the average, in dB. SNR is calculated as difference between the signal and noise spectra at the peak frequency of the signal. This can be inaccurate if noise is inaccurate (see noise for issues with noise calculations)

norm logical flag to normalize dB magnitude to maximum of 0

plot logical flag whether or not to plot the result. This will create two plots, the first is a concatenated spectrogram where the y-axis is frequency and the x-axis is click number. The second plot is the average spectrogram of all clicks, the y-axis is dB, x-axis is frequency. Can be a vector of length two to create only one of the two plots

noise logical flag to plot an average noise spectrum. This is estimated by taking a window of length wl immediately before click. Since there are only a limited number of samples saved in the Pamguard binary files, this can be very inaccurate when wl is a large proportion of the total samples saved. In these cases the noise floor will appear nearly identical to the signal, reducing wl can help get a more accurate noise floor.

decimate integer factor to reduce sample rate by

sort logical flag to sort concatenated spectrogram by peak frequency

mode one of 'spec' or 'ceps' to plot the spectrum or cepstrum

title replacement titles for plots, can be length vector of length two to provide separate titles

ylim optional y limits for mean spectra plot

flim optional frequency limits for both plots

cmap colors to use for concatenated click spectrogram, either a palette function or vector of colors

brightness value from -255 to 255, positive values increase brightness, negative values decrease brightness of concatenated spectrogram image

contrast value from -255 to 255, positive values increase contrast, negative values decrease contrast of concatenated spectrogram image

q lower and upper quantiles to remove for scaling concatenated spectrogram. Or if a single value, then quantiles q and 1-q will be used. Ex. if q=.01, then the bottom 1 plotting the image. This is done purely for cosmetic reasons, no output data is affected

showBreaks logical flag to show lines separating events when plotting multiple events

... optional args
Value

invisibly returns a list with six items: freq - the frequency, UID - the UID of each click, avgSpec - the average spectra of the event, allSpec - the individual spectrum of each click in the event as a matrix with each spectrum in a separate column, avgNoise - the average noise spectra, allNoise - the individual noise spectrum for each click.

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
# need to update binary file locations to users PAMPal installation
binUpd <- system.file('extdata', 'Binaries', package='PAMPal')
dbUpd <- system.file('extdata', package='PAMPal')
exStudy <- updateFiles(exStudy, bin = binUpd, db=dbUpd)
avSpec <- calculateAverageSpectra(exStudy)
str(avSpec$avgSpec)
range(avSpec$freq)
str(avSpec$allSpec)

Description

Calculate the estimated depth of echolocation clicks using surface reflected echoes. This uses the time delay between the received signal and its surface echo to estimate the depth of a calling animal. Requires that a set of waveform clips has been created using writeEventClips, and that events have been localized.

Usage

calculateEchoDepth(
  x,
  wav,
  clipLen = 0.03,
  spParams = NULL,
  soundSpeed = 1500,
  hpDepthError = 1,
  locType = "PGTargetMotion",
  plot = TRUE,
  nPlot = 400,
  nCol = 5,
  plotDir = NULL,
  progress = TRUE,
Arguments

- **x**: `AcousticStudy` object
- **wav**: either folder containing wave clips or list of wave clip files
- **clipLen**: length (seconds) of clip to analyze
- **spParams**: list of species-specific parameters, see details
- **soundSpeed**: sound speed (meters/second) to use for calculations
- **hpDepthError**: maximum error (meters) in hydrophone depth measurement
- **locType**: name of localization, note that this function is not computing any localization, only using previously calculated
- **plot**: logical flag to create summary plots
- **nPlot**: number of waveform plots to create for summary
- **nCol**: number of columns for waveform summary plot
- **plotDir**: directory to store plot outputs, default `NULL` will result in no plots being created
- **progress**: logical flag to show progress bar
- **verbose**: logical flag to show messages

Details

spParams allows for species-specific filtering and acceptable echo time delays to be specified. These are provided as a list with elements `freqLow` and `freqHigh` specifying the lower and upper ends of a bandpass filter to apply to the signals (in Hz), which can aid in properly detecting the echoes. Parameters `minTime` and `maxTime` can also be supplied to define ranges on allowed time delay values. Alternatively if `maxTime` is `NULL` or not present it will be calculated from the hydrophone geometry, and `minTime` can be calculated from geometry by providing `minDepth` and `maxRange` as the minimum detectable depth and maximum detectable range (in meters).

If the same values for these parameters should be used for all detections in `x`, then `spParams` can be provided as a list with each parameter named, e.g.

```
list(freqLow=10e3, freqHigh=50e3, minTime=.001, maxTime=NULL)
```

If different values should be used for different species, then `spParams` must be a named list where the names match the species in `x`, providing a separate list of values for each species. e.g.

```
list(Zc=list(freqLow=10e3, freqHigh=50e3, minTime=.001, maxTime=NULL),
     Pm=list(freqLow=2e3, freqHigh=16e3, minTime=.001, maxTime=NULL))
```
calculateICI

Calculate Inter-Click Interval

Value

the AcousticStudy object x with estimated dive depth outputs added for each detection that had a matching wav clip file in wav. Detections that either did not have matching wav files or did not have localizations will have NA for all dive depth outputs. The depth outputs are

- **maxTime**: Delay time with maximum correlation value
- **pair2Time**: Delay time with second highest correlation value
- **pair3Time**: Delay time with third highest correlation value
- **maxMag**: Correlation magnitude for "maxTime"
- **pair2Mag**: Correlation magnitude for "pair2Time"
- **pair3Mag**: Correlation magnitude for "pair3Time"
- **maxDepth**: Calculated depth for "maxTime"
- **pair2Depth**: Calculated depth for "pair2Time"
- **pair3Depth**: Calculated depth for "pair3Time"

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

```r
# example not run because it requires access to large files not present
# in the package testing material
## Not run:
study <- addRecordings(study, folder='path/to/recordings')
wavPath <- 'path/to/wavFiles'
writeEventClips(study, outDir=wavPath, mode='detection')
study <- calculateEchoDepth(study, wav=wavPath)
## End(Not run)
```

Description

Calculate inter-click interval for click data
Usage

calculateICI(
  x,
  time = c("UTC", "peakTime"),
  callType = c("click", "whistle", "cepstrum", "gpl"),
  verbose = TRUE,
  ...
)

## S4 method for signature 'AcousticStudy'
calculateICI(
  x,
  time = c("UTC", "peakTime"),
  callType = c("click", "whistle", "cepstrum", "gpl"),
  verbose = TRUE,
  ...
)

## S4 method for signature 'AcousticEvent'
calculateICI(
  x,
  time = c("UTC", "peakTime"),
  callType = c("click", "whistle", "cepstrum", "gpl"),
  verbose = TRUE,
  ...
)

getICI(x, type = c("value", "data"))

Arguments

x  a AcousticStudy object, a list of AcousticEvent objects, or a single AcousticEvent object
time the time measurement to use. start will use the UTC value, peak will use the peakTime value if present (currently present in standardClickCalcs, this is the time of the peak of the waveform)
callType the call type to calculate ICI for, usually this is click but also allows users to specify whistle or cepstrum to calculate this using other detector data
verbose logical flag to print messages
... not currently used
type the type of data to return, one of 'value' or 'data'. 'value' returns the single ICI value for each detector, 'data' returns all the individual ICI values used to calculate the number returned by 'value'

Details

Calculates the ICI for each individual detector and across all detectors. ICI calculation is done by ordering all individual detections by time, then taking the difference between consecutive detections
and approximating the mode value.

**Value**

the same object as x, with ICI data added to the "ancillary" slot of each AcousticEvent. Two items will be added. $ici contains all of the individual inter-click intervals used to calculate the ICI, as well as an "All" ICI using all the combined data. $measures will also have a ICI measurement added for each detector, this will be the single modal value. Data in the $measures spot can be exported easily to modeling algorithms. getICI will just return either the values stored in $measures for type = 'value' or a dataframe of the individual ICI values used to calculate these (with columns indicating separate Channels, eventIds, and detectorNames) for type = 'data'

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>

**Examples**

```r
# setting up example data
data(exStudy)
exStudy <- calculateICI(exStudy)
# each event has its ICI data stored separately, these are 0
# because there is only a single click in this event
ancillary(exStudy[[1]])$ici
# also saves it in measures that will get exported for modeling
ancillary(exStudy[[1]])$measures
```

---

**calculateModuleData**  
*Run Custom Calculations on Pamguard Module Data*

**Description**

Run a list of custom calculations on a Pamguard binary file.

**Usage**

```r
calculateModuleData(
  binData,
  binFuns = list(ClickDetector = list(standardClickCalcs)),
  settings = NULL
)
```

**Arguments**

- `binData`: Pamguard binary data as read in by `loadPamguardBinaryFile`
- `binFuns`: A named list of functions to run on each Pamguard module. Currently supported modules are 'ClickDetector' and 'WhistlesMoans', a sample input for binFuns would be `list('ClickDetector'=list(clickFun1, clickFun2), 'WhistlesMoans'=list(wmFun1))`
- `settings`: a list of settings from a Pamguard XML file
checkStudy

Value

A data frame with one row for each channel of each detection. Each row will have the UID, channel number, and name of the detector. Clicks of different classifications are treated as different detectors for this purpose, with the classification label number appended to the detector name. The number of columns will depend on the results of the calculations from the supplied binFuns.

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

checkStudy  Check an AcousticStudy Object for Issues

Description

Checks for any possible issues in an AcousticStudy object, issuing warnings and saving the messages

Usage

checkStudy(x, maxLength = Inf, maxSep = 60 * 60 * 2)

Arguments

x  an AcousticStudy object
maxLength  events with length greater than this value in seconds will trigger a warning
maxSep  events containing consecutive detections greater than maxSep seconds apart will trigger a warning. This is used to check for situations where detections were possibly added to the incorrect event.

Details

This function is called at the end of processPgDetections with default parameters, but can also be called later to investigate issues specific to each user’s data. For example, if you are expecting to process data where all recordings were duty cycled to record 2 out of every 10 minutes, then setting maxLength = 60*2 will alert you to any events that are longer than the 2 minute duty cycle. For continuously recorded data, the maxSep argument can be used to identify situations where there are large gaps between detections in a single event, since this could mean that detections were accidentally added to the incorrect event number during processing.

Value

returns a list of warning messages

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

data(exStudy)

# checks if any peak frequencies are 0, so we'll force this
exStudy[[1]][[1]]$peak <- 0
checkStudy(exStudy)
checkStudy(exStudy, maxLength = 1, maxSep = 1)

export_banter

Export Data for a BANTER Model

Description

Exports data from an AcousticStudy into the format required to run a BANTER model from the "banter" package

Usage

export_banter(
  x, 
  dropVars = NULL, 
  dropSpecies = NULL, 
  training = TRUE, 
  verbose = TRUE 
)

Arguments

x a AcousticStudy object or a list of AcousticEvent objects
dropVars a vector of the names of any variables to remove
dropSpecies a vector of the names of any species to exclude
training logical flag whether or not this will be used as a training data set, or a value between 0 and 1 specifying what percent of the data should be used for training (with the rest set aside for testing). If TRUE or greater than 0, must contain species ID. NOTE: if value is not 0, 1, TRUE, or FALSE, output will be further split into training and test items within the list output
verbose logical flag to show summary and informational messages

Value

a list with three items, events, detectors, and na. If value of training is not 0, 1, TRUE, or FALSE, output will be split into training and test lists that contain events and detectors. events is a dataframe with two columns. event.id is a unique identifier for each event, taken from the names of the event list. species is the species classification, taken from the species slot labelled id. detectors is a list of data frames containing all the detections and measurements. There is one list
for each unique detector type found in the detectors slots of \( x \). The data frames will only have columns with class numeric, integer, factor, or logical, and will also have columns named UID, Id, parentID, sampleRate, Channel, angle, and angleError, removed so that these are not treated as parameters for the banter random forest model. The dataframes will also have columns event.id and call.id added. na contains the UIDs and Binary File names for any detections that had NA values. These cannot be used in the random forest model and are removed from the exported dataset.

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

```r
# setting up example data
data(exStudy)
exStudy <- setSpecies(exStudy, method='pamguard')
banterData <- export_banter(exStudy)
# drop some variables
banterLess <- export_banter(exStudy, dropVars = c('peak', 'duration'))
```

---

**exStudy**

*Example AcousticStudy Object*

**Description**

An example AcousticStudy object created using the example PAMpalSettings object provided with the package. Processed with mode='db'

**Usage**

```r
data(exStudy)
```

**Format**

a AcousticStudy object containing two AcousticEvent objects
**filter.AcousticStudy**  
*Filter an AcousticStudy or AcousticEvent Object*

**Description**

Apply dplyr-like filtering to the detections of an AcousticStudy or AcousticEvent object, with a special case for filtering by species for an AcousticStudy.

**Usage**

```r
## S3 method for class 'AcousticStudy'
filter(.data, ..., .preserve = FALSE)
```

**Arguments**

- `.data`: AcousticStudy or AcousticEvent to filter
- `...`: Logical expressions, syntax is identical to `filter`. There are special cases to filter by environmental variables, species ID, database, or detector name. See details.
- `.preserve`: not used

**Details**

Most expression provided will be used to filter out detections based on calculated parameters.

If the name of an environmental variable added using `matchEnvData.AcousticStudy-method` is provided, will filter to only events with environmental variables matching those conditions.

If a provided logical expression uses "species" or "Species", then events will be filtered using the species present in the $id of the species slot of each event.

If a provided logical expression uses "database" or "Database", then only events with databases matching the expression in `files(.data)$db` will remain.

If a provided logical expression uses "detector" or "Detector", then only detections from detectors with names matching the expression will remain in events. Any events left with no detections will be removed.

**Value**

The original `.data` object, filtered by the given logical expressions

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

```r
# create example data
data(exStudy)
exStudy <- setSpecies(exStudy, method='manual', value=letters[1:2])
filterData <- filter(exStudy, peak < 20)
getDetectorData(filterData)$click

filterData <- filter(exStudy, species == 'a')
species(filterData[[1]])
```

---

filterEchoDepths  
*Filter Candidate Echo Depths*

Description

Filter out possible echo depths from `calculateEchoDepth` based on maximum depth, autocorrelation magnitude, and maximum swim speed criteria. Requires that `calculateEchoDepth` has been run first. This function adds a `keepClick` column to the data to track which detections should be used for further depth analysis by marking them as `FALSE` to be excluded or `TRUE` to be used.

Usage

```r
filterEchoDepths(
  x, 
  time = 30, 
  depth = NULL, 
  speed = NULL, 
  maxDepth = 4000, 
  minCorr = 0.01
)
```

Arguments

- `x`: an `AcousticStudy` object that has been processed with `calculateEchoDepth`
- `time`: maximum time apart (seconds) for detections. Detections with no no other detection within `time` seconds will be marked as `FALSE`
- `depth`: maximum depth difference (meters) between consecutive clicks, this value should be determined by maximum swim speed
- `speed`: as an alternative to providing `depth`, the swim speed (meters / second) can be provided and then `depth` will be calculated as `time * speed`
- `maxDepth`: calculated depth values greater than this will be marked `FALSE`
- `minCorr`: detections with autocorrelation magnitude less than this will be marked as `FALSE`
getBinaryData

Value

the AcousticStudy x with detections marked with column keepClick as TRUE or FALSE depending if they pass the filter parameters

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

# example not run because \link{calculateEchoDepth} must be run first, # and it requires a large amount of data not stored in the package
## Not run:
study <- calculateEchoDepth(study, wav='path/to/wavFiles')
study <- filterEchoDepths(study, time=30, speed=50/30, maxDepth=4000)

## End(Not run)

getBinaryData  Get Raw Binary Data for Detections

Description

Fetches matching binary data from a single or multiple detections in an AcousticEvent object

Usage

getBinaryData(
  x,
  UID,
  type = c("click", "whistle", "cepstrum", "gpl"),
  quiet = FALSE,
  ...
)

Arguments

x a AcousticStudy object, a list of AcousticEvent objects, or a single AcousticEvent object
UID the UID(s) of the individual detections to fetch the binary data for
type detection type
quiet logical flag to quiet some warnings, used internally and should generally not be changed from default FALSE
... additional arguments to pass to loadPamguardBinaryFile
getClipData

Description
Reads audio clips containing sounds from events or detections

Usage

getClipData(
  x,
  buffer = c(0, 0.1),
  mode = c("event", "detection"),
  channel = 1,
  useSample = FALSE,
  fixLength = FALSE,
  fillZeroes = TRUE,
  progress = TRUE,
  verbose = TRUE,
  FUN = NULL,
  ...
)

Arguments

x       AcousticStudy object containing data to read wav clips for
buffer  amount before and after each event to also include in the clip, in seconds. Can either be a vector of length two specifying how much to buffer before and after (first number should be negative), or a single value if the buffer amount should be identical.
mode    either 'event' or 'detection' specifying whether to create wav clips of entire events or individual detections

Value
a list of PamBinary objects for each UID

Author(s)
Taiki Sakai <taiki.sakai@noaa.gov>

Examples
data(exStudy)
binData <- getBinaryData(exStudy, UID = 8000003)
# works with multiple UIDs, if UIDs arent present they will be ignored
binData <- getBinaryData(exStudy, UID = c(8000003, 529000024, 1))
getDetectorData

channelchannel(s) of clips to write

useSamplelogical flag to use startSample information in binaries instead of UTC time for start of detections. This can be slightly more accurate (~1ms) but will take longer

fixLengthlogical flag to fix the output clip length to a constant value. If TRUE, then output clip length is entirely determined by the buffer value, as if the detection or event had zero length. E.g. buffer=c(-2,1) will produce clips 3 seconds long, starting 2 seconds before the detection/event start time.

fillZeroeslogical flag to fill gaps in non-consecutive clips with zeroes. If FALSE, will give warnings when attempting to retrieve clips spanning non-consecutive files and return no clip data

progresslogical flag to show progress bar

verboselogical flag to show summary messages

FUNoptional function to apply to wav clips. This function takes default inputs wav, a Wave class object, name the name of the detection or event, time the start and end time of the clip, channel as above, mode as above, and additional args...

...optional arguments to pass to FUN

Value

A named list of wav clips

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
recs <- system.file('extdata', 'Recordings', package='PAMpal')
exStudy <- addRecordings(exStudy, folder=recs, log=FALSE, progress=FALSE)
## Not run:
# not running so that no wav clips are written to disk
wavs <- getClipData(exStudy, mode='event')

## End(Not run)
getDetectorData

Usage

getDetectorData(x, measures = TRUE)
getClickData(x, measures = TRUE)
getWhistleData(x, measures = TRUE)
getCepstrumData(x, measures = TRUE)
getGPLData(x, measures = TRUE)
getFPODData(x, measures = TRUE)
nDetections(x, distinct = FALSE)
nClicks(x, distinct = FALSE)
nWhistles(x)
nCepstrum(x)
nGPL(x)

Arguments

x      data to extract detector data from, either an AcousticStudy, AcousticEvent or list of AcousticEvent object
measures  logical flag whether or not to append measures to detector dataframes
distinct  logical flag to only return number of distinct click detections

Details

The purpose of this function is to extract your data out of PAMpal’s S4 classes and put them into an easier format to work with. The output will be a list of up to three data frames, one for each call type found in your data. Each different call type will have had different processing applied to it by processPgDetections. Additionally, each detector will have its associated event id, the name of the detector, and the species id attached to it (species will be NA if not set). All detections from each call type will be combined into a single large data frame

Value

A list of data frames containing all detection data from x, named by call type (‘click’, ‘whistle’, or ‘cepstrum’).

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
getWarnings

Examples

```r
data(exStudy)
dets <- getDetectorData(exStudy)
names(dets)
str(dets$click)
# works on single events as well
oneDets <- getDetectorData(exStudy[[1]])
str(oneDets$click)
```

getWarnings  

*Get Warning Messages*

Description

Accessor to easily get all warning messages for `x`

Usage

```r
getWarnings(x)
```

Arguments

- `x` an `AcousticStudy` or `AcousticEvent` object

Value

a list of warning messages, named by the function call that created the warning

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

```r
## Not run:
data(exStudy)
# This will trigger a warning, then we can access it
exStudy <- filter(exStudy, species == "test")
getWarnings(exStudy)

## End(Not run)
```
is.PAMpalSettings

---

is.AcousticEvent  *Check if an Object is an AcousticEvent*

**Description**

Function to check if an object is an AcousticEvent

**Usage**

`is.AcousticEvent(x)`

**Arguments**

- `x`  
  object to check

---

is.AcousticStudy  *Check if an Object is an AcousticStudy*

**Description**

Function to check if an object is an AcousticStudy

**Usage**

`is.AcousticStudy(x)`

**Arguments**

- `x`  
  object to check

---

is.PAMpalSettings  *Check if an Object is a PAMpalSettings*

**Description**

Function to check if an object is a PAMpalSettings

**Usage**

`is.PAMpalSettings(x)`

**Arguments**

- `x`  
  object to check
loadPamguardXML

Load Pamguard XML Settings

Description

Loads in relevant settings and formats for use in PAMpal

Usage

loadPamguardXML(x)

Arguments

x an XML file created by Pamguard’s "Export XML Configuration"

Value

A list with settings for audio sources (sound acquisition, decimators, FFT, and cepstrum) and
detectors (click detector and whistle and moan detector). Also stores the entire XML file as raw and the file name as file

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

xmlFile <- system.file('extdata', 'Example.xml', package='PAMpal')
xmlList <- loadPamguardXML(xmlFile)
str(xmlList)

markAnnotated

Mark Detections as Annotated

Description

Marks detections within an AcousticStudy as being within the bounds of an annotation box. Annotations can either be read in from the “Spectrogram Annotation” module of PAMguard, or supplied as a separate dataframe. Detections must be entirely contained within the annotation bounds.
Usage

markAnnotated(
  x,
  anno = NULL,
  tBuffer = 0,
  fBuffer = 0,
  table = "Spectrogram_Annotation"
)

Arguments

x an AcousticStudy object
anno annotations to read from. If NULL, will be read in from the PAMguard database. If a data.frame, must have columns start and end in UTC, and column id. Can additionally have columns fmin and fmax to apply frequency bounds (values in Hz).
tBuffer additional buffer value to add on to annotation time bounds in seconds. If a single number, the number of seconds to extend the bounds by on the start and end of each annotation. Can also be a vector of two to extend different values on the start and end. This can be useful if original bounding boxes were drawn very close to the desired detections since any small portion of a signal outside the box will cause it to be excluded.
fBuffer additional buffer value to add to annotation frequency bounds in Hz. If a single number, the number of Hz to extend bounds by on lower and upper end of boxes. Can also be a vector of two to extend different values on lower and upper bounds. This can be useful if original bounding boxes were drawn very close to the desired detections since any small portion of a signal outside the box will cause it to be excluded.
table if anno is NULL, the name of the "Spectrogram Annotation" module table within the database.

Details

This adds new columns inAnno and annoId to all detector dataframes within the AcousticStudy. inAnno is a logical flag whether or not a given detection was fully contained in any annotation bounding box, and annoId lists the IDs of the boxes it matched. A detection is considered within an annotation only if it is entirely within the time and frequency bounds of the annotation. For GPL and whistle detections, the min and max frequency values are used. For click detections, only the peak frequency is used. For cesptrum detections, frequency bounds are ignored.

Value

the same object as x, but detectors have additional columns added

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

```r
data(exStudy)
annotation <- data.frame(start = min(getWhistleData(exStudy)$UTC),
                         fmin = c(16000, 17000),
                         fmax = c(17000, 18000))
annotation$end <- annotation$start + 1
exStudy <- markAnnotated(exStudy, annotation)
getWhistleData(exStudy)[c('UTC', 'duration', 'freqMin', 'freqMax', 'inAnno', 'annoId')]
```

Description

Extracts all variables from a netcdf file matching Longitude, Latitude, and UTC coordinates of the start of each AcousticEvent object. Matched values are stored in the "ancillary" slot of each event.

Usage

```r
# S4 method for signature 'AcousticEvent'
matchEnvData(
  data,
  nc = NULL,
  var = NULL,
  buffer = c(0, 0, 0),
  FUN = c(mean),
  fileName = NULL,
  progress = TRUE,
  depth = 0,
  ...
)

# S4 method for signature 'AcousticStudy'
matchEnvData(
  data,
  nc = NULL,
  var = NULL,
  buffer = c(0, 0, 0),
  FUN = c(mean),
  fileName = NULL,
  progress = TRUE,
  depth = 0,
  ...
)
```
Arguments

data an AcousticStudy or AcousticEvent object that must have GPS data added to it using the addGps functions

nc name of a netcdf file, ERDDAP dataset id, or an edinfo object

var (optional) vector of variable names

buffer vector of Longitude, Latitude, and Time (seconds) to buffer around each data-point. All values within the buffer will be used to report the mean, median, and standard deviation

FUN a vector or list of functions to apply to the data. Default is to apply mean, median, and standard deviation calculations

fileName (optional) file name to save downloaded nc file to. If not provided, then no nc files will be stored, instead small temporary files will be downloaded and then deleted. This can be much faster, but means that the data will need to be downloaded again in the future. If fileName is provided, then the function will attempt to download a single nc file covering the entire range of your data. If your data spans a large amount of time and space this can be problematic.

progress logical flag to show progress bar

depth depth values (meters) to use for matching, overrides any Depth column in the data or can be used to specify desired depth range when not present in data. Variables will be summarised over the range of these depth values. NULL uses all available depth values

... other parameters to pass to ncToData

Value

original data object with environmental data added to the ancillary slot of each event. Complete data will be stored in ancillary(data)$environmental, and the mean of each downloaded variable will be stored in ancillary(data)$measures so that it can be exported for modeling. For each event the coordinates associated with the earliest UTC value in that event are used to match

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
nec <- system.file('extdata', 'sst.nc', package='PAMmisc')
# suppressing warnings because nc coordinates dont align with this data,
# function warns of possible coordinate mismatch
exStudy <- suppressWarnings(matchEnvData(exStudy, nc=nec, progress=FALSE))
str(ancillary(exStudy[[1]])$environmental)
ancillary(exStudy[[1]])$measures
matchTimeData

**Description**

Match any time-based data (dataframe with a UTC column) to an AcousticStudy or AcousticEvent object.

**Usage**

```r
matchTimeData(
  x,
  data,
  mode = c("event", "detection"),
  thresh = Inf,
  interpolate = TRUE,
  replace = FALSE,
  keepDiff = FALSE
)
```

timeJoin(x, y, thresh = Inf, interpolate = TRUE, replace = FALSE)

**Arguments**

- **x**: AcousticStudy or AcousticEvent object to match data to, or a dataframe for timeJoin.
- **data**: a data frame to match to data to x. Must have column UTC, and optionally a column db if subsets of data should be matched only to parts of x with that database. All other columns will be considered variables to add to x.
- **mode**: one of "event" or "detection". "event" will match one set of variables per event, stored in the "measures" for that event. "detection" will match variables to every detection.
- **thresh**: maximum time apart in seconds for matching to data, if the closest value is more than thresh apart then the variable values will be set to NA.
- **interpolate**: logical flag whether or not to interpolate between points in data or just matched to nearest time.
- **replace**: one of TRUE, FALSE, or NA. If TRUE, all existing values with the same name as columns in data will be replaced. If FALSE no replacement occurs. If NA only values which are NA will be replaced with new values.
- **keepDiff**: logical flag to keep time difference data.
- **y**: dataframe to join to x.
Details

This function lets you match any arbitrary data to a PAMpal object as long as it has a time associated with it. Data will be attached to detector dataframes for mode=`detection` or to the event "measures" location for mode=`event` (this is where calculateICI and matchEnvData.AcousticStudy-method store their event data). These can be accessed with the getMeasures function and are also exported in the various "getXXX" functions (getClickData etc.) if measures=TRUE (default).

All columns in the provided data object will be treated as variables to add, with a few exceptions. There are a few reserved column names used by PAMpal that cannot be overridden (e.g. UID, eventId, species). Also any columns already existing in the PAMpal data will not be overridden unless replace is not FALSE. The column names in data will be used as the names for the added variables, so care should be taken to ensure these are informative enough for future use.

Value

the same data as x, with data added from data

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
addData <- data.frame(UTC = as.POSIXct('2018-03-20 15:25:10', tz='UTC'),
  newVariable = 26)
data <- matchTimeData(exStudy, addData, mode='detection')
getClickData(data)
data <- matchTimeData(exStudy, addData, mode='event')
getMeasures(data)

Description

Accessors for slots in AcousticEvent and AcousticStudy objects

Usage

settings(x, ...)

## S4 method for signature 'AcousticEvent'
settings(x, ...)

settings(x) <- value
## S4 replacement method for signature 'AcousticEvent'

settings(x) <- value

localizations(x, ...)

## S4 method for signature 'AcousticEvent'

localizations(x, ...)

localizations(x) <- value

## S4 replacement method for signature 'AcousticEvent'

localizations(x) <- value

id(x, ...)

## S4 method for signature 'AcousticEvent'

id(x, ...)

id(x) <- value

## S4 replacement method for signature 'AcousticEvent'

id(x) <- value

detectors(x, ...)

## S4 method for signature 'AcousticEvent'

detectors(x, ...)

detectors(x) <- value

## S4 replacement method for signature 'AcousticEvent'

detectors(x) <- value

species(x, ...)

## S4 method for signature 'AcousticEvent'

species(x, ...)

## S4 method for signature 'AcousticStudy'

species(x, type = "id", ...)

species(x) <- value

## S4 replacement method for signature 'AcousticEvent'

species(x) <- value

files(x, ...)
## S4 method for signature 'AcousticEvent'

files(x, ...)
iles(x) <- value

## S4 replacement method for signature 'AcousticEvent'
files(x) <- value

ancillary(x, ...)

## S4 method for signature 'AcousticEvent'

ancillary(x, ...)

ancillary(x) <- value

## S4 replacement method for signature 'AcousticEvent'
ancillary(x) <- value

## S4 method for signature 'AcousticEvent,ANY,ANY,ANY'

x[i]

## S4 replacement method for signature 'AcousticEvent,ANY,ANY,ANY'

x[i] <- value

## S4 method for signature 'AcousticEvent'

x$name

## S4 replacement method for signature 'AcousticEvent'

x$name <- value

## S4 method for signature 'AcousticEvent,ANY,ANY'

x[[i]]

## S4 replacement method for signature 'AcousticEvent,ANY,ANY,ANY'

x[[i]] <- value

## S4 method for signature 'AcousticStudy'

id(x, ...)

## S4 replacement method for signature 'AcousticStudy'

id(x) <- value

## S4 method for signature 'AcousticStudy'

files(x, ...)

## S4 replacement method for signature 'AcousticStudy'

files(x) <- value
gps(x, ...)
## S4 method for signature 'AcousticStudy'
gps(x, ...)

gps(x) <- value
## S4 replacement method for signature 'AcousticStudy'

## S4 method for signature 'AcousticStudy'
detectors(x, ...)
events(x, ...)
## S4 method for signature 'AcousticStudy'
events(x, ...)
events(x) <- value
## S4 replacement method for signature 'AcousticStudy'
events(x) <- value

## S4 method for signature 'AcousticStudy'
settings(x, ...)
## S4 replacement method for signature 'AcousticStudy'
settings(x) <- value

effort(x, ...)
## S4 method for signature 'AcousticStudy'
effort(x, ...)
effort(x) <- value
## S4 replacement method for signature 'AcousticStudy'
effort(x) <- value

## S4 method for signature 'AcousticStudy'
pps(x, ...)
## S4 method for signature 'AcousticStudy'
pps(x, ...)

pps(x) <- value
## S4 replacement method for signature 'AcousticStudy'
pps(x) <- value
## S4 method for signature 'AcousticStudy'
ancillary(x, ...)

## S4 replacement method for signature 'AcousticStudy'
ancillary(x) <- value

models(x, ...)

## S4 method for signature 'AcousticStudy'
models(x, ...)

models(x) <- value

## S4 replacement method for signature 'AcousticStudy'
models(x) <- value

## S4 method for signature 'AcousticStudy,ANY,ANY,ANY'
x[i]

## S4 replacement method for signature 'AcousticStudy,ANY,ANY,ANY'
x[i] <- value

## S4 method for signature 'AcousticStudy'
x$name

## S4 replacement method for signature 'AcousticStudy'
x$name <- value

## S4 method for signature 'AcousticStudy,ANY,ANY'
x[[i]]

## S4 replacement method for signature 'AcousticStudy,ANY,ANY,ANY'
x[[i]] <- value

### Arguments

- **x**: a `AcousticEvent` or `AcousticStudy` object
- **...**: other arguments to pass to methods
- **value**: value to assign with accessor
- **type**: species type to select
- **i**: index of the object to access
- **name**: name of the object to access

### Value

- **id**: a unique id or name for this object
settings a named list of settings for each detector and localization or recorder
detectors a list of detector data frames
localizations list of localizations
species list of species classifications
files list of files used to create this object
events a list of `AcousticEvent` objects
gps a dataframe containing gps data
pps the PAMpalSettings object used to create this
effort something about effort?
ancillary miscellaneous extra data

Author(s)
Taiki Sakai <taiki.sakai@noaa.gov>

---

**PAMpalSettings**  
*Constructor for PAMpalSettings Object*

**Description**
Create a PAMpalSettings object. Any values that are not supplied will be asked for interactively. Three processing functions will also be added by default: `standardClickCalcs`, `roccaWhistleCalcs`, and `standardCepstrumCalcs`

**Usage**
```
PAMpalSettings(
  db = NULL,
  binaries = NULL,
  settings = NULL,
  functions = NULL,
  verbose = TRUE,
  ...
)
```

**Arguments**
- `db` the full path to a Pamguard database file or folder of databases
- `binaries` a folder containing Pamguard binary files, all subfolders will also be added
- `settings` an XML settings file from Pamguard
- `functions` a named list of additional functions to add
- `verbose` logical flag to show messages
- `...` values to pass on to default `standardClickCalcs` function
PAMpalSettings-class

Value

A PAMpalSettings object

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

```r
# can be run with no arguments with popup menu selections
if(interactive()) pps <- PAMpalSettings()
db <- system.file('extdata', 'Example.sqlite3', package='PAMpal')
bin <- system.file('extdata', 'Binaries', package='PAMpal')
# or data folders can be supplied ahead of time
if(interactive()) pps <- PAMpalSettings(db=db, binaries=bin)
```

PAMpalSettings-class  PAMpalSettings Class

Description

An S4 class that stores settings related to all processing and analysis steps done in PAMpal. A PAMpalSettings object will be the main input to any major function in the PAMpal package.

Slots

db  the full path to a PamGuard database file

binaries a list with items "folder" containing the directory of the PamGuard binary files, and "list" containing the full path to each individual binary file.

functions a named list of functions to apply to data read in by PAMpal. Should be named by the PamGuard module the function should be applied to. Currently supports "ClickDetector", "WhistlesMoans", and "Cepstrum".

calibration a named list of calibration functions to apply while applying functions from the "functions" slot. Should named by the PamGuard module, same as the "functions"

settings a named list of settings, usually imported from Pamguard's "Export XML Configuration"

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
plotDataExplorer  Explore Data in an Interactive Shiny Plot

Description

Runs an interactive Shiny plot of detector data. Allows user to choose which numeric data to plot, and will allow user to both color and facet the plot by event number, detector name, or species.

Usage

plotDataExplorer(x)

Arguments

x  data to plot, can be an AcousticStudy, AcousticEvent, data.frame or a list of AcousticEvent objects

Value

nothing, just plots

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)

if(interactive()) plotDataExplorer(exStudy)

plotGram  Plot Spectrogram or Cepstrogram

Description

Plots either a spectrogram or cepstrogram and also overlays whistle or cepstral contours from the binary files.
Usage

plotGram(
  x,
  evNum = 1,
  start = NULL,
  end = NULL,
  channel = 1,
  wl = 512,
  hop = 0.25,
  mode = c("spec", "ceps"),
  detections = c("cepstrum", "click", "whistle"),
  detCol = c("red", "blue", "purple"),
  brightness = 0,
  contrast = 0,
  q = 0.01,
  cmap = gray.colors(64, start = 1, end = 0),
  size = 1,
  add = FALSE,
  title = NULL,
  sr = NULL,
  freqRange = NULL,
  ...
)

Arguments

x  an AcousticStudy object

evNum if x is a study, the event index number to plot. Note that this is the index in the order that they appear in the AcousticStudy object, not the actual event number. Alternatively full event names can be used

start start time of the plot, either POSIXct or seconds from the start of the event

dead end time of the plot, either POSIXct or seconds from the start of the event.

channel channel to plot

wl window length of FFT to use for creating plot

hop hop value of FFT to use for creating plot, either as a percentage of wl or number of samples

mode one of 'spec' or 'ceps' to plot either spectrogram or ceprogram

detections vector containing any of 'cepstrum', 'click', and/or 'whistle' indicating which detections to overlay on the plot

detCol vector containing colors to use for plotting detections. Order matches order of detections (default alphabetical - cepstrum, click, whistle)

brightness value from -255 to 255, positive values increase brightness, negative values decrease brightness of concatenated spectrogram image

contrast value from -255 to 255, positive values increase contrast, negative values decrease contrast of concatenated spectrogram image
plotWaveform

q lower and upper quantiles to remove for scaling concatenated spectrogram. Or if a single value, then quantiles q and 1-q will be used. Ex. if q=.01, then the bottom 1 plotting the image. This is done purely for cosmetic reasons, no output data is affected

cmap color map for the spectrogram, either a palette function or vector of colors

size size scaling to apply to plotted points and lines, as a multiple factor to the default values. Can be a vector of length equal to detections to apply different size scales

add logical flag FALSE to create a new plot (default), or TRUE to only add new detections to an existing plotGram plot with the same parameters. Can be used to add detections with different visual parameters for the same detection type (e.g. clicks with different point size and color). Typically will only work well if start and end have been set manually.

title optional title for plot, defaults to "Spectrogram" or "Cepstrogram"

sr sample rate to decimate to

freqRange frequency range to plot, in Hz for mode='spec' or milliseconds for mode='ceps'

... additional arguments to pass to points or lines

Value

nothing, just plots

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
recs <- system.file('extdata', 'Recordings', package='PAMpal')
exStudy <- updateFiles(exStudy, bin=system.file('extdata', 'Binaries', package='PAMpal'), db = system.file('extdata', 'Example.sqlite3', package='PAMpal'))
exStudy <- addRecordings(exStudy, folder=recs, log=FALSE, progress=FALSE)
# No detections will appear on plot because included recordings are heavily decimated
plotGram(exStudy)

plotWaveform  Plot Graphical Representations of Waveforms

Description

Fetches matching binary data from a single or multiple detections in an AcousticStudy object, then plot the resulting data
plotWaveform

Usage

plotWaveform(x, UID, length = NULL, sr = NULL)

plotSpectrogram(x, UID, length = NULL, sr = NULL, ...)

plotWigner(x, UID, length = NULL, sr = NULL, ...)

Arguments

x a AcousticStudy object, a list of AcousticEvent objects, or a single AcousticEvent object

UID the UID(s) of the individual detections to fetch the binary data for

length length of the waveform to use for plotting, in samples. The clip used will be centered around the maximum value of the waveform, if length is NULL (default), the entire waveform will be used. If length is greater than the stored clip, the waveform will be zero-padded to length

sr if NULL (default) will try to read sample rate from your data. If provided as a value will override sample rate in the data.

... other arguments to pass to the spectrogram or wigner functions

Details

The plotSpectrogram function uses the function specgram to plot the spectrogram, see this function for plotting options. The plotWigner function uses the function wignerTransform to plot the Wigner-Ville transform, see this function for options.

Value

Nothing, just shows plots for every channel of the waveform for each UID provided

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
plotWaveform(exStudy, 8000003)
plotSpectrogram(exStudy, 8000003)
plotWigner(exStudy, 8000003)
processPgDetections

Load and Process Detections from Pamguard

Description

Loads and processes acoustic detection data that has been run through Pamguard. Uses the binary files and database(s) contained in pps, and will group your data into events by the grouping present in the 'OfflineEvents' and 'Detection Group Localiser' tables (mode = 'db') or by the grouping specified by start and end times in the supplied grouping (mode = 'time'), or by start and end of recording files (mode = 'recording'). Will apply all processing functions in pps to the appropriate modules.

Usage

processPgDetections(
  pps,
  mode = c("db", "time", "recording"),
  id = NULL,
  grouping = NULL,
  format = c("%m/%d/%Y %H:%M:%OS", "%m-%d-%Y %H:%M:%OS",
             "%Y/%m/%d %H:%M:%OS", "%Y-%m-%d %H:%M:%OS"),
  progress = TRUE,
  verbose = TRUE,
  ...
)

Arguments

pps

a PAMpalSettings object containing the databases, binaries, and functions to use for processing data. See PAMpalSettings. Can also be an AcousticStudy object, in which case the pps slot will be used.

mode

selector for how to organize your data into events. db will organize by events based on tables in the databases. time will organize into events based on timestamps provided in grouping. recording will organize events by the start and end times of recording files found in the database. For time and recording, ALL detections between the start and end times are included, for db only selected detections are included.

id

an event name or id for this study, will default to today's date if not supplied (recommended to supply your own informative id).

grouping

For mode = 'db', the table to group events by. Either event to use the OfflineEvents table, or detGroup to use the detection group localiser module groups. For mode = 'time', this should be a data frame with three mandatory columns and 1 row for each separate event. The mandatory columns are start, end, and id. start and end should specify the start and end time of the event and must be in UTC. id should specify a unique id for each event. There are also optional columns species, db, and sr. species should provide a species ID.
if it is available. \(db\) and \(sr\) are the corresponding database and sample rate to associate with a particular event, these typically do not need to be specified as the function will attempt to automatically match them based on the times of the events and the databases. Note that \(db\) must be the full filepath to the database. If a clear match is not found then the user will be prompted to either select from a list or input the proper sample rate.

Grouping can be supplied either as a data frame or as a filepath to a csv file.

- **format**: the date format for the start and end columns in grouping if it is a csv. Times are assumed to be UTC. See details section of \texttt{strptime} for more information on how to properly format this.
- **progress**: logical flag to show progress bars
- **verbose**: logical flag to show messages
- **...**: additional arguments to pass onto to different methods

### Details

If \(mode\) is not specified, it will try to be automatically determined in the following order. 1) if a grouping data.frame or CSV is provided, then \(mode='time'\) will be used. 2) If there are labelled events present in the database, \(mode='db'\) will be used. 3) \(mode='recording'\) will be used, which should be equivalent to loading all possible data.

### Value

- an \texttt{AcousticStudy} object with one \texttt{AcousticEvent} for each event in the \texttt{events} slot, and the \texttt{PAMPalSettings} object used stored in the \texttt{pps} slot.

### Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

### Examples

```r
exPps <- new('PAMPalSettings')
exPps <- addDatabase(exPps, system.file('extdata', 'Example.sqlite3', package='PAMPal'))
exPps <- addBinaries(exPps, system.file('extdata', 'Binaries', package='PAMPal'))
exClick <- function(data) {
  standardClickCalcs(data, calibration=NULL, filterfrom_khz = 0)
}
exPps <- addFunction(exPps, exClick, module = 'ClickDetector')
exPps <- addFunction(exPps, roccaWhistleCalcs, module='WhistlesMoans')
exPps <- addFunction(exPps, standardCepstrumCalcs, module = 'Cepstrum')
# process events labelled within the Pamguard database
exStudyDb <- processPgDetections(exPps, mode='db', id='Example')
# can also give an AcousticStudy as input and it will use same functions and data
reprocess <- processPgDetections(exStudyDb, mode='db', id='Reprocess')
# process events with manually set start/end times
grp <- data.frame(start = as.POSIXct('2018-03-20 15:25:10', tz='UTC'),
  end = as.POSIXct('2018-03-20 15:25:11', tz='UTC'),
  id = 'GroupExample')
```
removeBinaries

Remove Binaries from a PAMpalSettings Object

Description

Remove a binary folder and associated files from the "binaries" slot in a PAMpalSettings object.

Usage

removeBinaries(pps, index = NULL)

Arguments

pps  
a PAMpalSettings object to remove binaries from

index  
index indicating which binary folders to remove. Can be a vector if you want to remove multiple folders. If missing user is prompted to select a folder from a list, will only show up to the first 20. You can easily remove all of the folders with a large index like 1:1000

Value

the same PAMpalSettings object as pps, with the binary folders and files associated with those folders removed from the "binaries" slot.

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

exPps <- new('PAMpalSettings')
exPps <- addBinaries(exPps, system.file('extdata', 'Binaries', package='PAMpal'))
removeBinaries(exPps, index = 1)
if(interactive()) removeBinaries(exPps)
removeCalibration

Remove a Calibration Function from a PAMpalSettings Object

Description

Remove a calibration function from the "calibration" slot of a PAMpalSettings object

Usage

removeCalibration(pps, index = NULL, module = "ClickDetector", verbose = TRUE)

Arguments

pps  a PAMpalSettings object to remove a calibration from
index  index of the calibration function to remove. If NULL, user will be prompted to select from a list. This can also be a vector to remove multiple calibration functions at once.
module  the module of the calibration function to remove, currently not needed
verbose  logical flag to show messages

Value

the same PAMpalSettings object as pps, with the calibration function removed from the "calibration" slot

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

pps <- new('PAMpalSettings')
calFile <- system.file('extdata', 'calibration.csv', package='PAMpal')
pps <- addCalibration(pps, calFile, all = TRUE, units=3)
calClick <- function(data, calibration=NULL) {
  standardClickCalcs(data, calibration=NULL)
}
pps <- addFunction(pps, calClick, module = 'ClickDetector')
pps <- applyCalibration(pps, all=TRUE)
pps
removeCalibration(pps, index=1)
removeDatabase

Remove a Database from a PAMpalSettings Object

Description

Remove a database from the "db" slot in a PAMpalSettings object.

Usage

removeDatabase(pps, index = NULL)

Arguments

pps a PAMpalSettings object to remove a database from
index index indicating which database(s) to remove. Can be a vector if you want to
remove multiple databases. If missing user is prompted to select a database
from a list, will only show up to the first 20. You can easily remove all of the
databases with a large index like 1:1000

Value

the same PAMpalSettings object as pps, with the database(s) removed from the "db" slot

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

exPps <- new('PAMpalSettings')
exPps <- addDatabase(exPps, system.file('extdata', 'Example.sqlite3', package='PAMpal'))
removeDatabase(exPps, 1)
if(interactive()) removeDatabase(exPps)

removeFunction

Remove a Function from a PAMpalSettings Object

Description

Remove a function from the "function" slot in a PAMpalSettings object.

Usage

removeFunction(pps, index = NULL)
Arguments

pps          a PAMpalSettings object to remove a function from
index        index indicating which function to move, counting from ClickDetector functions first, then WhistlesMoans functions, then Cepstrum functions. This is the same order functions appear in when examining the pps object. For example, if there are two Click functions and one Whistle function, the Whistle function would have an index of 3. If missing, user can select from a list. This can also be a vector to remove multiple functions at once.

Value

the same PAMpalSettings object as pps, with the function removed from the "functions" slot

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

Examples

```r
exPps <- new('PAMpalSettings')
exPps <- addFunction(exPps, roccaWhistleCalcs, module='WhistlesMoans')
exPps <- addFunction(exPps, standardCepstrumCalcs, module = 'Cepstrum')
removeFunction(exPps, 1)
removeFunction(exPps, 1:2)
# normally best to use interactively instead of specifying index
if(interactive()) removeFunction(exPps)
```

Description

Remove a note added with addNote

Usage

```r
removeNote(x, index)
```

Arguments

x           An AcousticStudy or AcousticEvent object
index       The index of the note to remove, order matches the output of getNotes

Value

For addNote, the same data as x, with notes added. For getNotes, a list of all notes present in x
removeSettings

Author(s)
Taiki Sakai <taiki.sakai@noaa.gov>

Examples

data(exStudy)
exStudy <- addNote(exStudy, to='study', label='Note1',
                   note='My first note for this study')
exStudy <- addNote(exStudy, to='event', evNum=1:2, label='Note2',
                   note='A note for the first two events')
exStudy
removeNote(exStudy, 1)
removeNote(exStudy, 2)
removeNote(exStudy, 3)

removeSettings	Remove Settings from a PAMpalSettings Object

Description
Remove all settings from the "settings" slot in a PAMpalSettings object.

Usage
removeSettings(pps)

Arguments
pps a PAMpalSettings object to remove settings from

Value
the same PAMpalSettings object as pps, with all settings removed from the "settings" slot

Author(s)
Taiki Sakai <taiki.sakai@noaa.gov>

Examples
exPps <- new('PAMpalSettings')
exPps <- addSettings(exPps, system.file('extdata', 'Example.xml', package='PAMpal'))
removeSettings(exPps)
roccaWhistleCalcs  
*Calculate a Set of Measurements for Whistles*

**Description**  
Calculate a set of measurements from a whistle contour. All calculations following ROCCA method from Julie and Michael Oswald, as implemented in Pamguard and detailed in Oswald et al (2007) <doi:10.1121/1.2743157>

**Usage**  
roccaWhistleCalcs(data)

**Arguments**  
- **data**  
a list that must have `freq` the whistle contour stored as a vector of FFT bin frequencies in hertz, and `time` the time in seconds at each bin.

**Value**  
A list with 50 calculated ROCCA parameters, each item in the list will only have 1 entry so that this can easily be converted to a data frame.

**Author(s)**  
Taiki Sakai <taiki.sakai@noaa.gov>

**Examples**  
```r  
data(testWhistle)  
roccaWhistleCalcs(testWhistle)  
```

runDepthReview  
*Run Echo Depth Review App*

**Description**  
Runs a Shiny app to review the slant delay and estimated depths of an `AcousticStudy` object that has been processed with `calculateEchoDepth`. App allows users to select detections that should not be included in future analysis and marks them with the tag `keepClick=FALSE`, similar to `filterEchoDepths`.

**Usage**  
runDepthReview(x)
sampleDetector

Subsample Detectors in AcousticStudy

Description
samples either a fraction or fixed number from each detector in each event of an AcousticStudy

Usage
```
sampleDetector(x, n = 1)
```

Arguments
- **x**: AcousticStudy object
- **n**: if less than 1, proportion of rows to sample from each detector. If 1 or more, the number of rows to sample from each detector.

Details
Uses `slice_sample` to do the sampling, `n` is converted either to prop or `n` based on its size. Negative values are treated the same as in `slice_sample`

Value
subsampled AcousticStudy x
setSpecies

Set the Species Classification of Events

Description

Sets the species slot of AcousticEvent objects within an AcousticStudy

Usage

setSpecies(x, method = c("pamguard", "manual", "reassign"), value, type = "id")

Arguments

x  a AcousticStudy object, a list of AcousticEvent objects, or a single AcousticEvent object
method  the method for assigning species to an event. Currently supports pamguard, which will use the 'eventType' or 'Text_Annotation' column to assign species, manual which will use value to assign species manually, or reassign which will use value to reassign an old species label to a new one
value  required only if method is set to 'manual' or 'reassign'. For 'manual', can either be a single value to assign to all events, or a vector with length equal to the number of events. Can also be a dataframe with columns event and species, in which case species will be matched to corresponding event names instead of just relying on the order. If using this, please note the prefix OE or DGL present on most event numbers (see the id slot of your events, or names(events(x))). For 'reassign', value must be a data frame with columns old and new. Any events with species id in the old column of the dataframe will get reassigned to the corresponding id in the new column.
type  the type of classification to set, this is just a label within the species slot. Default 'id' should typically not be changed since this is used by other functions
**standardCepstrumCalcs**

*Calculate a Set of Measurements from a Cepstrum Contour*

**Description**

Calculate a set of measurements from a cepstrum contour. This is currently used to measure the inter-click interval of the burst pulse type calls.

**Usage**

standardCepstrumCalcs(data)

**Arguments**

data a list that must have quefrency the "quefrency" at each cepstrum contour, sr the sample rate of the data, and time the time in seconds at each bin

**Value**

A list with inter-click interval (ici), duration (seconds), and slope of the ici.
**standardClickCalcs**

*Calculate a Set of Measurements for Clicks*

**Description**

Calculate a set of "standard" measurements for odontocete clicks

**Usage**

```r
standardClickCalcs(
  data, 
  sr_hz = "auto", 
  calibration = NULL, 
  filterfrom_khz = 10, 
  filterto_khz = NULL, 
  winLen_sec = 0.0025
)
```

**Arguments**

- **data**: a list that must have 'wave' containing the wave form as a matrix with a separate column for each channel, and 'sr' the sample rate of the data. Data can also be a Wave class object, like one created by `Wave`.
- **sr_hz**: either 'auto' (default) or the numeric value of the sample rate in hertz. If 'auto', the sample rate will be read from the 'sr' of data.
- **calibration**: a calibration function to apply to the spectrum, must be a gam. If NULL no calibration will be applied (not recommended).
- **filterfrom_khz**: frequency in khz of highpass filter to apply, or the lower bound of a bandpass filter if filterto_khz is not NULL.
- **filterto_khz**: if a bandpass filter is desired, set this as the upper bound. If only a highpass filter is desired, leave as the default NULL value.
- **winLen_sec**: length in seconds of fft window. The click wave is first shortened to this number of samples around the peak of the wave, removing a lot of the noise around the click. Following approach of JB/EG/MS.
**Details**


**Value**

A data frame with one row for each channel of click waveform

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>

**Examples**

```r
data(testClick)
standardClickCalcs(testClick)
```

---

**summariseDiveDepth**  
*Summarise Dive Depth*

**Description**

Summarise results of dive depth estimation using `calculateEchoDepth` and related functions

**Usage**

```r
summariseDiveDepth(x, hpDepthError = 1, locType = "PGTargetMotion")
```

**Arguments**

- `x`  
an `AcousticStudy` that has been processed with `calculateEchoDepth`
- `hpDepthError`  
  hydrophone depth error to use for error estimation  
- `locType`  
  name of localization, note that this function is not computing any localization, only using previously calculated

**Value**

a dataframe with columns summarising the estimated dive depth for each event in `x`

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

```r
# example not run because \link{calculateEchoDepth} must be run first,
# and it requires a large amount of data not stored in the package
## Not run:
study <- calculateEchoDepth(study, wav='path/to/wavFiles')
summariseDiveDepth(study)
## End(Not run)
```

testCeps

A fake cepstrum contour

Description

A manually created fake cepstrum contour, mimicing what the output would be from the Pamguard module and fed into the cepstrum calcs.

Usage

data(testCeps)

Format

A list with three items:

- `quefrency` a vector of the cepstrum contour quefrency values
- `time` a vector of the time values of the cepstrum contour in seconds
- `sr` the sample rate of the recording

testClick

A two-channel recording of a delphinid click

Description

An example delphinid click waveform. This is a two-channel recording of some kind of delphinid click, recorded at 500kHz. There are 800 samples recorded on each channel.

Usage

data(testClick)
**Format**

A list with two items:

- **wave**  a matrix with two columns of 800 samples, each column is a separate recording channel
- **sr**  the sample rate of the recording

**Source**

Southwest Fisheries Science Center / NMFS / NOAA

---

**testGPL**  
*A fake GPL detection*

**Description**

A manually created fake GPL contour, mimicking what the output would be from the Pamguard module and fed into the GPL calcs

**Usage**

```r
data(testGPL)
```

**Format**

A list with three items:

- **freq**  a vector of the frequency contour values in hertz
- **time**  a vector of the time values of the contour in seconds

---

**testWhistle**  
*A fake whistle contour*

**Description**

A manually created fake whistle contour reanging from 1kHz to 3.1kHz

**Usage**

```r
data(testWhistle)
```

**Format**

A list with two items:

- **freq**  a vector of the frequency contour values in hertz
- **time**  a vector of the time values of the contour in seconds
updateFiles

Update Location of Files in an AcousticStudy

Description

Updates the stored locations of binary, database, and/or recording files in the files slots of an AcousticStudy and all AcousticEvent objects within. Runs interactively to prompt users to select folders if missing files are found. Typically used after changing computers, or if original data was on an external hard drive. If any missing files are not able to be located, they will be kept in the files slot so that this function can be run again.

Usage

updateFiles(
  x,
  bin = NULL,
  db = NULL,
  recording = NULL,
  verbose = TRUE,
  check = TRUE
)

Arguments

x an AcousticStudy or AcousticEvent object
bin folder containing updated binary file locations. If NULL (default), user will be prompted to select a folder. If NA, binary files will be skipped.
db single file or folder containing updated database file locations. NULL (default), user will be prompted to select a folder. If NA, database files will be skipped.
recording folder containing updated recording file locations. If NULL (default), user will be prompted to select a folder. If NA, recording files will be skipped.
verbose logical flag to print messages about success of replacement
check logical flag to do extra checking. You do not need to set this parameter, used internally for speed purposes so certain checks are not repeated

Value

the same AcousticStudy and AcousticEvent object as x with updated file locations

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

data(exStudy)
    # files in exStudy will have paths local to package creator's computer
files(exStudy)$db
file.exists(files(exStudy)$db)
files(exStudy)$binaries
file.exists(files(exStudy)$binaries)
    # folder with example DB
    db <- system.file('extdata', package='PAMpal')
    # folder with example binaries
bin <- system.file('extdata', 'Binaries', package='PAMpal')
exStudy <- updateFiles(exStudy, db=db, bin=bin)
files(exStudy)$db
file.exists(files(exStudy)$db)
files(exStudy)$binaries
file.exists(files(exStudy)$binaries)

updatePamObject Update PAMpal S4 Object

Description

Updates older versions of PAMpal's S4 objects to stop "validObjec" warning messages

Usage

updatePamObject(x)

Arguments

x an AcousticStudy, AcousticEvent, or PAMpalSettings object

Details

As of v0.12.0 this updates any previous version's PAMpalSettings objects to have the new "settings" slot, as well as updating any PAMpalSettings objects within an AcousticStudy

Value

the same object as x with any slot changes made

Author(s)

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

```r
## Not run:
pps <- new('PAMPalSettings')
# manually breaking this S4 class, don't try this at home
attr(pps, 'settings') <- NULL
# This will now give an error
pps
pps <- updatePamObject(pps)
# Fixed!
pps

## End(Not run)
```

writeEventClips  
Create Wav Clips of Data

Description

Creates audio clips containing sounds from events or detections

Usage

```r
writeEventClips(
  x,
  buffer = c(0, 0.1),
  outDir = ".",
  mode = c("event", "detection"),
  channel = 1,
  filter = 0,
  useSample = FALSE,
  progress = TRUE,
  verbose = TRUE,
  fixLength = FALSE
)
```

parseEventClipName(file, part = c("event", "time", "UID", "channel", "UTC"))

Arguments

- **x**  
  AcousticStudy object containing data to make wav clips for

- **buffer**  
  amount before and after each event to also include in the clip, in seconds. Can either be a vector of length two specifying how much to buffer before and after (first number should be negative), or a single value if the buffer amount should be identical.

- **outDir**  
  directory to write clips to, defaults to current directory
mode  either 'event' or 'detection' specifying whether to create wav clips of entire events or individual detections
channel  channel(s) of clips to write
filter  filter to apply to wav clips before writing, values in kHz. A value of 0 applies no filter. A single value applies a highpass filter at that value. A vector of two values applies a lowpass filter if the first number is 0, or a bandpass filter if both are non-zero.
useSample  logical flag to use startSample information in binaries instead of UTC time for start of detections. This can be slightly more accurate (~1ms) but will take longer
progress  logical flag to show progress bar
verbose  logical flag to show summary messages
fixLength  logical flag to fix the output clip length to a constant value. If TRUE, then output clip length is entirely determined by the buffer value, as if the detection or event had zero length. E.g. buffer=c(-2,1) will produce clips 3 seconds long, starting 2 seconds before the detection/event start time.
file  file name to parse
part  part of file name to return

details

parseEventClipName parses the file names created to pull out event names or file start times

value

A vector of file names for the wav clips that were successfully created, any that were not able to be written will be NA. Note that currently this can only write clips with up to 2 channels. File names will be formatted as [Event or Detection]_[Id]CH[ChannelNumber(s)]_[YYYYMMDD]_[HHMMSS]_[mmm].wav

The last numbers are the start time of the file in UTC, accurate to milliseconds. The Id is either the event ID or the detection UID.

author(s)

Taiki Sakai <taiki.sakai@noaa.gov>

examples

data(exStudy)
recs <- system.file('extdata', 'Recordings', package='PAMpal')
exStudy <- addRecordings(exStudy, folder=recs, log=FALSE, progress=FALSE)
## Not run:
# not running so that no wav clips are written to disk
wavs <- writeEventClips(exStudy, outDir='WavFolder', mode='event')

## End(Not run)
**writeWignerData**

Write Wigner Transform Data of Click Detections to Disk

**Description**

Create Wigner-Ville transform data of click clips from all detections and save them to disk. A CSV file will also be written that lists all UIDs contained in the output.

**Usage**

```r
writeWignerData(
  x,
  n = 256,
  t = 300,
  outDir = ".",
  mode = "narray",
  progress = TRUE,
  ...
)
```

**Arguments**

- `x` *AcousticStudy* object containing data to make Wigner data for
- `n` number of frequency bins for Wigner transform (recommended power of 2)
- `t` number of samples to use for the click clip passed to the transform
- `outDir` directory to write data to
- `mode` specifies the kind of output that will be created, currently only supports creating NumPy arrays using the reticulate package, in future will support image creation
- `progress` logical flag to show progress bar
- `...` optional arguments to pass

**Value**

A list with two items:  
- `files` - a vector of file names for the Wigner data that were successfully created, any that were not able to be written will be NA,  
- `warnings` - a list with items containing event IDs that triggered any warnings

**Author(s)**

Taiki Sakai <taiki.sakai@noaa.gov>
Examples

```r
data(exStudy)
exStudy <- setSpecies(exStudy, method='pamguard')
## Not run:
# not running because files are written to disk
wigFiles <- writeWignerData(exStudy, outDir = 'WigFolder')

## End(Not run)
```
Index

* datasets
  - exStudy, 28
  - testCeps, 66
  - testClick, 66
  - testGPL, 67
  - testWhistle, 67
  - [],AcousticEvent,ANY,ANY,ANY-method (PAMpal.accessors), 42
  - [],AcousticStudy,ANY,ANY,ANY-method (PAMpal.accessors), 42
  - [<-,AcousticEvent,ANY,ANY,ANY-method (PAMpal.accessors), 42
  - [<-,AcousticStudy,ANY,ANY,ANY-method (PAMpal.accessors), 42
  - [[]<-,AcousticEvent,ANY,ANY,ANY-method (PAMpal.accessors), 42
  - [[]<-,AcousticStudy,ANY,ANY,ANY-method (PAMpal.accessors), 42
  - $,AcousticEvent-method (PAMpal.accessors), 42
  - $,AcousticStudy-method (PAMpal.accessors), 42
  - <$-,AcousticEvent-method (PAMpal.accessors), 42
  - <$-,AcousticStudy-method (PAMpal.accessors), 42
  - AcousticEvent-class, 3
  - AcousticStudy, 4, 5, 9, 11–17, 19, 22, 24, 26–32, 35, 37, 40–42, 46, 50–54, 58, 60–62, 65, 68–70, 72
  - AcousticStudy-class, 4
  - addAnnotation, 4
  - addBinaries, 6
  - addCalibration, 7
  - addDatabase, 8
  - addFPOD, 9
  - addFunction, 10
  - addGps, 11, 40
  - addGps,AcousticEvent-method (addGps), 11
  - addGps,AcousticStudy-method (addGps), 11
  - addGps,ANY-method (addGps), 11
  - addGps,data.frame-method (addGps), 11
  - addGps,list-method (addGps), 11
  - addHydrophoneDepth, 12
  - addMeasures, 13
  - addNote, 14, 58
  - addRecordings, 15
  - addSettings, 16
  - addWaveHeight, 17
  - ancillary (PAMpal.accessors), 42
  - ancillary,AcousticEvent-method (PAMpal.accessors), 42
  - ancillary,AcousticStudy-method (PAMpal.accessors), 42
  - ancillary<-,AcousticEvent-method (PAMpal.accessors), 42
  - ancillary<-,AcousticStudy-method (PAMpal.accessors), 42
  - applyCalibration (addCalibration), 7
  - bindStudies, 18
  - calculateAverageSpectra, 19
  - calculateEchoDepth, 21, 30, 60, 61, 65
  - calculateICI, 23, 42
  - calculateICI,AcousticEvent-method (calculateICI), 23
  - calculateICI,AcousticStudy-method (calculateICI), 23
  - calculateModuleData, 25
  - checkAnnotationData, 25
  - checkAnnotation (addAnnotation), 4
checkStudy, 26

detectors (PAMpal.accessors), 42
detectors, AcousticEvent-method (PAMpal.accessors), 42
detectors, AcousticStudy-method (PAMpal.accessors), 42
detectors <- (PAMpal.accessors), 42
detectors <-, AcousticEvent-method (PAMpal.accessors), 42
effort (PAMpal.accessors), 42
effort, AcousticStudy-method (PAMpal.accessors), 42
effort <- (PAMpal.accessors), 42
effort <-, AcousticStudy-method (PAMpal.accessors), 42
events (PAMpal.accessors), 42
events, AcousticStudy-method (PAMpal.accessors), 42
events <- (PAMpal.accessors), 42
events <-, AcousticStudy-method (PAMpal.accessors), 42
export_anomate (addAnnotation), 4
export_banter, 27
exStudy, 28

files (PAMpal.accessors), 42
files, AcousticEvent-method (PAMpal.accessors), 42
files, AcousticStudy-method (PAMpal.accessors), 42
files <- (PAMpal.accessors), 42
files <-, AcousticEvent-method (PAMpal.accessors), 42
files <-, AcousticStudy-method (PAMpal.accessors), 42
filter, 29
filter, AcousticStudy, 29
filterEchoDepths, 30, 60

getAnnotation (addAnnotation), 4
getBinaryData, 31
getCepstrumData (getDetectorData), 33
getchClickData, 42
getchClickData (getDetectorData), 33
getchClipData, 32
getchDetectorData, 33
getchFPODData (getDetectorData), 33
getchGPLData (getDetectorData), 33
getchICI (calculateICI), 23
getchMeasures, 42
getchMeasures (addMeasures), 13
getchNotes, 58
getchNotes (addNote), 14
getchWarnings, 35
getchWhistleData (getDetectorData), 33
gps (PAMpal.accessors), 42
gps, AcousticStudy-method (PAMpal.accessors), 42
gps <- (PAMpal.accessors), 42
gps <-, AcousticStudy-method (PAMpal.accessors), 42
id (PAMpal.accessors), 42
id, AcousticEvent-method (PAMpal.accessors), 42
id, AcousticStudy-method (PAMpal.accessors), 42
id <- (PAMpal.accessors), 42
id <-, AcousticEvent-method (PAMpal.accessors), 42
id <-, AcousticStudy-method (PAMpal.accessors), 42
is.AcousticEvent, 36
is.AcousticStudy, 36
is.PAMpalSettings, 36

lines, 51
loadPamguardBinaryFile, 25, 31
loadPamguardXML, 37
localizations (PAMpal.accessors), 42
localizations, AcousticEvent-method (PAMpal.accessors), 42
localizations <- (PAMpal.accessors), 42
localizations <-, AcousticEvent-method (PAMpal.accessors), 42
markAnnotated, 37
matchEnvData, AcousticEvent-method, 39
matchEnvData, AcousticStudy-method, 29, 42
matchEnvData, AcousticStudy-method (matchEnvData, AcousticEvent-method), 39
matchRecordingUrl (addAnnotation), 4
matchTimeData, 41
models (PAMpal.accessors), 42
models, AcousticStudy-method (PAMpal.accessors), 42
models<-, AcousticStudy-method (PAMpal.accessors), 42
nCepstrum (getDetectorData), 33
nClicks (getDetectorData), 33
ncToData, 40
nDetections (getDetectorData), 33
nGPL (getDetectorData), 33
nWhistles (getDetectorData), 33
PAMpal.accessors, 42
PAMpalSettings, 4, 6–10, 16–18, 47, 47, 53–59, 69
PAMpalSettings-class, 48
parseEventClipName (writeEventClips), 70
peakTrough, 65
plotDataExplorer, 49
plotGram, 49
plotSpectrogram (plotWaveform), 51
plotWaveform, 51
plotWigner (plotWaveform), 51
points, 51
pps (PAMpal.accessors), 42
pps, AcousticStudy-method (PAMpal.accessors), 42
pps<-, (PAMpal.accessors), 42
prepAnnotation (addAnnotation), 4
processPgDetections, 26, 53
removeBinaries, 55
removeCalibration, 56
removeDatabase, 57
removeFunction, 57
removeNote, 58
removeSettings, 59
roccaWhistleCalcs, 47, 60
runDepthReview, 60
sampleDetector, 61
setSpecies, 62
settings (PAMpal.accessors), 42
settings, AcousticEvent-method (PAMpal.accessors), 42
settings, AcousticStudy-method (PAMpal.accessors), 42
settings<- (PAMpal.accessors), 42
slice_sample, 61
specgram, 52
species (PAMpal.accessors), 42
species, AcousticEvent-method (PAMpal.accessors), 42
species, AcousticStudy-method (PAMpal.accessors), 42
species<- (PAMpal.accessors), 42
specgram, 52
squeeList, 18
standardCepstrumCalcs, 47, 63
standardClickCalcs, 47, 64
strptime, 54
summariseDiveDepth, 65
testCeps, 66
testClick, 66
testGPL, 67
testWhistle, 67
timeJoin (matchTimeData), 41
updateFiles, 68
updatePamObject, 69
Wave, 64
wignerTransform, 52
writeEventClips, 5, 21, 70
writeWignerData, 72