Package ‘RxODE’

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Imports Matrix, PreciseSums (>= 0.3), Rcpp (>= 0.12.3), brew, cli, crayon, digest, dparser (>= 0.1.8), ggforce, ggplot2, inline, magrittr, memoise, methods, mvnfast, pillar, rex, sys, units(>= 0.6-0), utils, assertthat, lotri, remotes
Description Facilities for running simulations from ordinary differential equation (ODE) models, such as pharmacometrics and other compartmental models. A compilation manager translates the ODE model into C, compiles it, and dynamically loads the object code into R for improved computational efficiency. An event table object facilitates the specification of complex dosing regimens (optional) and sampling schedules. NB: The use of this package requires both C and Fortran compilers, for details on their use with R please see Section 6.3, Appendix A, and Appendix D in the “R Administration and Installation” manual. Also the code is mostly released under GPL. The VODE and LSODA are in the public domain. The information is available in the inst/COPYRIGHTS.

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Clear/Set pipeline

Description

Clear/Set pipeline

Usage

```
.clearPipe(  
  rx = NULL,  
  inits = NULL,  
  events = NULL,  
  params = NULL,  
  iCov = NULL,  
  keep = NULL,  
  thetaMat = NULL,  
  omega = NULL,  
  sigma = NULL,  
  dfObs = NULL,  
  dfSub = NULL,  
  nSub = NULL,  
  nStud = NULL  
)
```
.rxFindPow

Arguments

\r
\begin{itemize}
\item \textbf{rx} \hspace{1cm} RxODE object
\item \textbf{inits} \hspace{1cm} a vector of initial values of the state variables (e.g., amounts in each compartment), and the order in this vector must be the same as the state variables (e.g., PK/PD compartments);
\item \textbf{events} \hspace{1cm} an eventTable object describing the input (e.g., doses) to the dynamic system and observation sampling time points (see eventTable);
\item \textbf{params} \hspace{1cm} a numeric named vector with values for every parameter in the ODE system; the names must correspond to the parameter identifiers used in the ODE specification;
\item \textbf{iCov} \hspace{1cm} A data frame of individual non-time varying covariates to combine with the params to form a parameter data.frame.
\item \textbf{keep} \hspace{1cm} Columns to keep from either the input dataset or the iCov dataset. With the iCov dataset, the column is kept once per line. For the input dataset, if any records are added to the data LOCF (Last Observation Carried forward) imputation is performed.
\item \textbf{thetaMat} \hspace{1cm} Named theta matrix.
\item \textbf{omega} \hspace{1cm} Estimate of Covariance matrix. When omega is a list, assume it is a block matrix and convert it to a full matrix for simulations.
\item \textbf{sigma} \hspace{1cm} Named sigma covariance or Cholesky decomposition of a covariance matrix. The names of the columns indicate parameters that are simulated. These are simulated for every observation in the solved system.
\item \textbf{dfObs} \hspace{1cm} Degrees of freedom to sample the unexplained variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.
\item \textbf{dfSub} \hspace{1cm} Degrees of freedom to sample the between subject variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.
\item \textbf{nSub} \hspace{1cm} Number between subject variabilities (ETAs) simulated for every realization of the parameters.
\item \textbf{nStud} \hspace{1cm} Number virtual studies to characterize uncertainty in estimated parameters.
\end{itemize}

\textbf{.rxFindPow} \hspace{1cm} \textit{Find power THETAs for appropriate scaling}

Description

Find power THETAs for appropriate scaling

Usage

\texttt{	extregistered.rxFindPow(x)}

Arguments

\begin{itemize}
\item \textbf{x} \hspace{1cm} RxODE model that can be access by rxNorm
\end{itemize}
Value

THETA numbers of $x^\theta$

Author(s)

Matthew L. Fidler

---

.rxRmPrint  

*Remove print statements*

Description

Remove print statements

Usage

.rxRmPrint(x)

Arguments

$x$  

RxODE lines to remove

Value

RxODE with print lines removed.

Author(s)

Matthew L. Fidler

---

.rxRmSens  

*Remove sensitivity equations*

Description

Remove sensitivity equations

Usage

.rxRmSens(x)

Arguments

$x$  

RxODE lines to remove
Value

- Lines with \( \frac{d}{dt}(rx_{sens\ldots}) \) removed.

Author(s)

- Matthew L. Fidler

---

**.rxSymPyJacobian**  
*Calculate the full Jacobian for a model*

---

**Description**

This expands the model to calculate the Jacobian. This requires rSymPy.

**Usage**

```
.rxSymPyJacobian(model)
```

**Arguments**

- model: RxODE family of objects

**Value**

RxODE syntax for model with Jacobian specified.

**Author(s)**

- Matthew L. Fidler

---

**.rxWinRtoolsPath**  
*Setup Rtools path*

---

**Description**

Setup Rtools path

**Usage**

```
.rxWinRtoolsPath(rm.rtools = TRUE, rm.python = TRUE, retry = FALSE)
```

**Arguments**

- rm.rtools: Remove the Rtools from the current path specs.
- rm.python: Remove Python from the current path specs.
- retry: Should you retry to find Rtools? If NA, don’t throw an error if it isn’t found.
Author(s)

Matthew L. Fidler

.add.dosing

Description

Turn on/off warnings for ID sorting.

Usage

.setWarnIdSort(warnIdSort = TRUE)

Arguments

warnIdSort Boolean for if the sorting warning is turned on or off.

Value

Nothing

Author(s)

Matthew Fidler

add.dosing

Add dosing to eventTable

Description

This adds a dosing event to the event table. This is provided for piping syntax through magrittr.

Usage

add.dosing(
  eventTable,
  dose,
  nbr.doses = 1L,
  dosing.interval = 24,
  dosing.to = 1L,
  rate = NULL,
  amount.units = NA_character_,
  start.time = 0,
  do.sampling = FALSE,
  time.units = NA_character_,
  ...
)

add.dosing

Arguments

- **eventTable**: eventTable object
- **dose**: numeric scalar, dose amount in `amount.units`;
- **nbr.doses**: integer, number of doses;
- **dosing.interval**: required numeric scalar, time between doses in `time.units`, defaults to 24 of `time.units="hours"`;
- **dosing.to**: integer, compartment the dose goes into (first compartment by default);
- **rate**: for infusions, the rate of infusion (default is `NULL`, for bolus dosing);
- **amount.units**: optional string indicating the dosing units. Defaults to `NA` to indicate as per the original EventTable definition.
- **start.time**: required dosing start time;
- **do.sampling**: logical, should observation sampling records be added at the dosing times? Defaults to `FALSE`.
- **time.units**: optional string indicating the time units. Defaults to "hours" to indicate as per the original EventTable definition.
- **...**: Other parameters passed to `et`.

Value

- eventTable with updated dosing (note the event table will be updated anyway)

Author(s)

Matthew L. Fidler
Matthew L Fidler, Wenping Wang

References


See Also

eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

```r
## Model from RxODE tutorial
mod1 <-RxODE({
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
```
V3=2.97E+02;
Kin=1;
Kout=1;
EC50=200;
C2 = centr/V2;
C3 = peri/V3;
d/dt(depot) =-KA*depot;
d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
d/dt(peri) = Q*C2 - Q*C3;
d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
});

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
et(amt=20000,ii=24,until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid,qd) %>% et(seq(0,11*24,length.out=100));
bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")
et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1,"weeks"), qd) %>%
    add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))
wkOnOff <- rxSolve(mod1, et)

plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(qd, times=4, wait=set_units(1,"weeks")) %>%
  add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))

repCycle4 <- rxSolve(mod1, et)

plot(repCycle4, C2)

---

### add.sampling

**Add sampling to eventTable**

**Description**

This adds a dosing event to the event table. This is provided for piping syntax through magrittr

**Usage**

```
add.sampling(eventTable, time, time.units = NA)
```

**Arguments**

- **eventTable**: An eventTable object
- **time**: a vector of time values (in time.units).
- **time.units**: an optional string specifying the time units. Defaults to the units specified when the EventTable was initialized.

**Value**

eventTable with updated sampling. (Note the event table will be updated even if you don’t reassign the eventTable)

**Author(s)**

Matthew L Fidler, Wenping Wang

**References**

See Also

`eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE`

Examples

```r
## Model from RxODE tutorial
mod1 <- RxODE({
    KA=2.94E-01;
    CL=1.86E+01;
    V2=4.02E+01;
    Q=1.05E+01;
    V3=2.97E+02;
    Kin=1;
    Kout=1;
    EC50=200;
    C2 = centr/V2;
    C3 = peri/V3;
    d/dt(depot) =-KA*depot;
    d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
    d/dt(peri) = Q*C2 - Q*C3;
    d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
});

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
    et(amt=10000, ii=12, until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
    et(amt=20000, ii=24, until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid, qd) %>% et(seq(0, 11*24, length.out=100));
bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
    et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")
```
et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1,"weeks"), qd) %>%
  add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))
wkOnOff <- rxSolve(mod1, et)
plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(qd, times=4, wait=set_units(1,"weeks")) %>%
  add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))
repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)

---

**as.data.table.rxEt**  
*Convert an event table to a data.table*

**Description**

Convert an event table to a data.table

**Usage**

```r
as.data.table.rxEt(x, keep.rownames = FALSE, ...)
```

**Arguments**

- **x**  
  An R object.
- **keep.rownames**  
  Default is `FALSE`. If `TRUE`, adds the input object's names as a separate column named "rn". `keep.rownames = "id"` names the column "id" instead.
- **...**  
  Additional arguments to be passed to or from other methods.
as.et  
Coerce object to data.frame

Description
Coerce object to data.frame

Usage
as.et(x, ...)

## Default S3 method:
as.et(x, ...)

Arguments
x  
Object to coerce to et.
...
Other parameters

as_tibble.rxEt  
Convert to tbl

Description
Convert to tbl

Usage
## S3 method for class 'rxEt'
as_tibble(x, ...)
as.tbl.rxEt(x, ...)

Arguments
x  
RxODE event table
...
Other arguments to as.tbl

Value
tibble
Return the RxODE coefficients

Description

This returns the parameters, state variables

Usage

```r
## S3 method for class 'RxODE'
coef(object, ...)

## S3 method for class 'RxCompilationManager'
coef(...)

## S3 method for class 'solveRxODE'
coef(object, ...)

## S3 method for class 'rxDll'
coef(...)
```

Arguments

- `object` is an RxODE object
- `...` ignored arguments

Value

a rxCoef object with the following

- `params` is a list of strings for parameters for the RxODE object
- `state` is a list of strings for the names of each state in the RxODE object.
- `ini` is the model specified default values for the parameters.
- `RxODE` is the referring RxODE object

Author(s)

Matthew L. Fidler
Sample a covariance Matrix from the Posterior Inverse Wishart distribution.

Description
Note this Inverse wishart rescaled to match the original scale of the covariance matrix.

Usage
cvPost(nu, omega, n = 1L, omegaIsChol = FALSE, returnChol = FALSE)

Arguments
- **nu**: Degrees of Freedom (Number of Observations) for covariance matrix simulation.
- **omega**: Estimate of Covariance matrix.
- **n**: Number of Matrices to sample. By default this is 1.
- **omegaIsChol**: is an indicator of if the omega matrix is in the Cholesky decomposition.
- **returnChol**: Return the Cholesky decomposition of the covariance matrix sample.

Details
If your covariance matrix is a 1x1 matrix, this uses an scaled inverse chi-squared which is equivalent to the Inverse Wishart distribution in the uni-directional case.

Value
a matrix (n=1) or a list of matrices (n > 1)

Author(s)
Matthew L.Fidler & Wenping Wang

Examples

```r
## Sample a single covariance.
draw1 <- cvPost(3, matrix(c(1,.3,.3,1),2,2))

## Sample 3 covariances
set.seed(42)
draw3 <- cvPost(3, matrix(c(1,.3,.3,1),2,2), n=3)

## Sample 3 covariances, but return the cholesky decomposition
set.seed(42)
draw3c <- cvPost(3, matrix(c(1,.3,.3,1),2,2), n=3, returnChol=TRUE)
```
et

**Event Table Function**

**Description**

Event Table Function

**Usage**

```r
et(x, ..., envir = parent.frame())
## S3 method for class 'RxODE'
et(x, ..., envir = parent.frame())
## S3 method for class 'rxSolve'
et(x, ..., envir = parent.frame())
## S3 method for class 'rxParams'
et(x, ..., envir = parent.frame())
## Default S3 method:
et(
x, ...
, time,
  amt,
  evid,
  cmt,
  ii,
  addl,
  ss,
  rate,
  dur,
  until,
  id,
  amountUnits,
  timeUnits,
  addSampling,
  envir = parent.frame(),
  by = NULL,
  length.out = NULL
)
```

**Arguments**

- `x`  
  This is the first argument supplied to the event table. This is named to allow `et` to be used in a pipe-line with arbitrary objects.
... Times or event tables. They can also be one of the named arguments below.

envir the environment in which expr is to be evaluated. May also be NULL, a list, a
data frame, a pairlist or an integer as specified to sys.call.

time Time is the time of the dose or the sampling times. This can also be unspecified
and is determined by the object type (list or numeric/integer).

amt Amount of the dose. If specified, this assumes a dosing record, instead of a
sampling record.

evid Event ID; This can be:
  • 0 An observation. This can also be specified as evid=obs
  • 1 A dose observation. This can also be specified as evid=dose
  • 2 A non-dose event. This can also be specified as evid=other.
  • 3 A reset event. A reset event resets all the compartment values to zero and
turns off all infusions. This can also be specified as evid=reset.
  • 4 Dose and reset event. This can also be specified as evid=doseReset or
evid=resetDose

cmt Compartment name or number. If a number, this is an integer starting at 1. Neg-
ative compartments are not supported (there is no way to turn off a compartment
currently). If the compartment is a name, the compartment name is changed to
the correct state/compartment number before running the simulation.
Can also specify cmt as dosing.to, dose.to, doseTo, dosingTo, and state.

ii When specifying a dose, this is the inter-dose interval for ss, addl and until
options (described below).

addl The number of additional doses at an inter-dose interval after one dose.

ss Steady state flag; It can be one of:
  • 0 This dose is not a steady state dose
  • 1 This dose is a steady state dose with the between/inter dose interval of ii
  • 2 This is a steady state dose that uses the super-position principle to allow
more complex steady states, like 10 mg in the morning and 20 mg at night,
or dosing at 8 am 12 pm and 8 pm instead of every 12 hours. Since it uses
the super positioning principle, it only makes sense when you know the
kinetics are linear.
All other values of SS are currently invalid.

rate When positive, this is the rate of infusion. Otherwise:
  • 0 No infusion is on this record.
  • -1 Rate of this record is modeled by rate(cmt) = in the RxODE model.
    You may also specify type or rate by rate=model
  • -2 Duration of this record is modeled by dur(cmt) = in the RxODE model.
    You may also specify this type of rate by dur=model or rate=dur.

When a modeled bioavailability is applied to positive rates (rate > 0), the dura-
tion of infusion is changed. This is because the data specify the rate and amount,
the only think ghat modeled bioavailability can affect is duration.
If instead you want the modeled bioavailability to increase the rate of infusion
instead of the duration of infusion, specify the dur instead or model the duration
with rate=2.
Duration of infusion. When `amt` and `dur` are specified the rate is calculated from
the two data items. When `dur` is specified instead of `rate`, the bioavailability
changes will increase rate instead of duration.

This is the time until the dosing should end. It can be an easier way to figure out
how many additional doses are needed over your sampling period.

A integer vector of IDs to add or remove from the event table. If the event table
is identical for each ID, then you may expand it to include all the IDs in this
vector. All the negative IDs in this vector will be removed.

The units for the dosing records (`amt`)

The units for the time records (`time`)

This is a boolean indicating if a sampling time should be added at the same time
as a dosing time. By default this is `FALSE`.

When there are no observations in the event table, this is the amount to increment
for the observations between `from` and `to`.

The number of observations to create if there isn’t any observations in the event
table. By default this is 200.

A new event table

Matthew L Fidler, Wenping Wang

Pharmacometric Models in R." CPT: Pharmacometrics \\& Systems Pharmacology, 5(1), 3-10. ISSN
2163-8306, <URL: http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4728294/>.

eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

```r
## Model from RxODE tutorial
mod1 <-RxODE({
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
})
```
C2 = centr/V2;
C3 = peri/V3;
d/dt(depot) =-KA*depot;
d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
d/dt(peri) = Q*C2 - Q*C3;
d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
  et(amt=10000, ii=12, until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
  et(amt=20000, ii=24, until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid, qd) %>%
  rxSolve(mod1, et)

plot(bid, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
  et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>%
  et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")

et <- seq(infusion, qd)
infusionQd <- rxSolve(mod1, et)

plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>%
  et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")

et <- seq(qd, set_units(1, "weeks"), qd) %>%
  add.sampling(set_units(seq(0, 5.5, by=0.005), weeks))
wkOnOff <- rxSolve(mod1, et)

plot(wkOnOff, C2)
## You can also repeat the cycle easily with the rep function

dd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(dd, times=4, wait=set_units(1,"weeks")) %>%
  add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))
repCycle4 <- rxSolve(mod1, et)

plot(repCycle4, C2)

---

**etExpand**

*Expand additional doses*

### Description

Expand additional doses

### Usage

```r
etExpand(et)
```

### Arguments

- **et**
  - Event table to expand additional doses for.

### Value

New event table with 'addl' doses expanded

### Author(s)

Matthew Fidler

### Examples

```r
ev <- et(amt=3,ii=24,until=240);
print(ev)
etExpand(ev) # expands event table, but doesn't modify it
print(ev)
ev$expand() ## Expands the current event table and saves it in ev
```
etRbind

Combining event tables

Description

Combining event tables

Usage

etRbind(
  ..., 
  samples = c("use", "clear"),
  waitII = c("smart", "+ii"),
  id = c("merge", "unique")
)

## S3 method for class 'rxEt'
rbind(..., deparse.level = 1)

Arguments

... The event tables and optionally time between event tables, called waiting times in this help document.
samples How to handle samples when repeating an event table. The options are:
  • "clear" Clear sampling records before combining the datasets
  • "use" Use the sampling records when combining the datasets
waitII This determines how waiting times between events are handled. The options are:
  • "smart" This "smart" handling of waiting times is the default option. In this case, if the waiting time is above the last observed inter-dose interval in the first combined event table, then the actual time between doses is given by the wait time. If it is smaller than the last observed inter-dose interval, the time between event tables is given by the inter-dose interval + the waiting time between event tables.
  • "+ii" In this case, the wait time is added to the inter-dose interval no matter the length of the wait time or inter-dose interval
id This is how rbind will handle IDs. There are two different types of options:
  • merge with id="merge", the IDs are merged together, overlapping IDs would be merged into a single event table.
  • unique with id="unique", the IDs will be renumbered so that the IDs in all the event tables are not overlapping.
deparse.level The deparse.level of a traditional rbind is ignored.
**Value**

An event table

**Author(s)**

Matthew L Fidler
Matthew L Fidler, Wenping Wang

**References**


**See Also**

eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

**Examples**

```r
## Model from RxODE tutorial
mod1 <- RxODE(
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) =-KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50*C2))*eff;
)

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
  et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
  et(amt=20000,ii=24,until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
```
et <- seq(bid,qd) %>% et(seq(0,11*24,length.out=100));
bidQd <- rxSolve(mod1, et)
plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days
## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")
et <- seq(infusion,qd)
infusionQd <- rxSolve(mod1, et)
plot(infusionQd, C2)

## 2wk-on, 1wk-off
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- seq(qd, set_units(1,"weeks"), qd) %>%
   add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))
wkOnOff <- rxSolve(mod1, et)
plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function
qd <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")
et <- etRep(qd, times=4, wait=set_units(1,"weeks")) %>%
   add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))
repCycle4 <- rxSolve(mod1, et)
plot(repCycle4, C2)

---

<table>
<thead>
<tr>
<th>etRep</th>
<th>Repeat an RxODE event table</th>
</tr>
</thead>
</table>

**Description**
Repeat an RxODE event table
etRep

Usage

etRep(
  x,
  times = 1,
  length.out = NA,
  each = NA,
  n = NULL,
  wait = 0,
  id = integer(0),
  samples = c("clear", "use"),
  waitII = c("smart", "+ii"),
  ii = 24
)

## S3 method for class 'rxEt'
rep(x, ...)

Arguments

x An RxODE event table
times Number of times to repeat the event table
length.out Invalid with RxODE event tables, will throw an error if used.
each Invalid with RxODE event tables, will throw an error if used.
n The number of times to repeat the event table. Overrides times.
wait Waiting time between each repeated event table. By default there is no waiting, or wait=0
id A integer vector of IDs to add or remove from the event table. If the event table is identical for each ID, then you may expand it to include all the IDs in this vector. All the negative IDs in this vector will be removed.
samples How to handle samples when repeating an event table. The options are:
  • "clear" Clear sampling records before combining the datasets
  • "use" Use the sampling records when combining the datasets
waitII This determines how waiting times between events are handled. The options are:
  • "smart" This "smart" handling of waiting times is the default option. In this case, if the waiting time is above the last observed inter-dose interval in the first combined event table, then the actual time between doses is given by the wait time. If it is smaller than the last observed inter-dose interval, the time between event tables is given by the inter-dose interval + the waiting time between event tables.
  • "+ii" In this case, the wait time is added to the inter-dose interval no matter the length of the wait time or inter-dose interval
ii When specifying a dose, this is the inter-dose interval for ss, add1 and until options (described below).
... Times or event tables. They can also be one of the named arguments below.
Author(s)
Matthew L Fidler, Wenping Wang

References

See Also
eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE

Examples

```r
## Model from RxODE tutorial
mod1 <- RxODE({
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) =-KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
});

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
  et(amt=10000,ii=12,until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
  et(amt=20000,ii=24,until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid,qd) %>%
  et(seq(0,11*24,length.out=100));

bidQd <- rxSolve(mod1, et)

plot(bidQd, C2)
```
etSeq

Sequence of event tables

Description

This combines a sequence of event tables.

Usage

etSeq(..., samples = c("clear", "use"), waitII = c("smart", "+ii"), ii = 24)
## S3 method for class 'rxEt'
seq(...)

### Arguments

...  The event tables and optionally time between event tables, called waiting times in this help document.

`samples`  How to handle samples when repeating an event table. The options are:
- "clear" Clear sampling records before combining the datasets
- "use" Use the sampling records when combining the datasets

`waitII`  This determines how waiting times between events are handled. The options are:
- "smart" This "smart" handling of waiting times is the default option. In this case, if the waiting time is above the last observed inter-dose interval in the first combined event table, then the actual time between doses is given by the wait time. If it is smaller than the last observed inter-dose interval, the time between event tables is given by the inter-dose interval + the waiting time between event tables.
- "+ii" In this case, the wait time is added to the inter-dose interval no matter the length of the wait time or inter-dose interval

`ii`  If there was no inter-dose intervals found in the event table, assume that the interdose interval is given by this `ii` value. By default this is 24.

### Details

This sequences all the event tables in added in the argument list ... . By default when combining the event tables the offset is at least by the last inter-dose interval in the prior event table (or `ii`). If you separate any of the event tables by a number, the event tables will be separated at least the wait time defined by that number or the last inter-dose interval.

### Author(s)

Matthew L Fidler, Wenping Wang

### References


### See Also

`eventTable, add.sampling, add.dosing, et, etRep, etRbind, RxODE`
Examples

```r
## Model from RxODE tutorial
mod1 <- RxODE(
  KA=2.94E-01;
  CL=1.86E+01;
  V2=4.02E+01;
  Q=1.05E+01;
  V3=2.97E+02;
  Kin=1;
  Kout=1;
  EC50=200;
  C2 = centr/V2;
  C3 = peri/V3;
  d/dt(depot) =-KA*depot;
  d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
  d/dt(peri) = Q*C2 - Q*C3;
  d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
)

## These are making the more complex regimens of the RxODE tutorial

## bid for 5 days
bid <- et(timeUnits="hr") %>%
  et(amt=10000, ii=12, until=set_units(5, "days"))

## qd for 5 days
qd <- et(timeUnits="hr") %>%
  et(amt=20000, ii=24, until=set_units(5, "days"))

## bid for 5 days followed by qd for 5 days
et <- seq(bid, qd) %>%
  et(seq(0, 11*24, length.out=100));

bidQd <- rxSolve(mod1, et)

plot(bidQd, C2)

## Now Infusion for 5 days followed by oral for 5 days

## note you can dose to a named compartment instead of using the compartment number
infusion <- et(timeUnits = "hr") %>%
  et(amt=10000, rate=5000, ii=24, until=set_units(5, "days"), cmt="centr")

qd <- et(timeUnits = "hr") %>%
  et(amt=10000, ii=24, until=set_units(5, "days"), cmt="depot")

et <- seq(infusion, qd)

infusionQd <- rxSolve(mod1, et)
```

plot(infusionQd, C2)

## 2wk-on, 1wk-off

d <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")

e <- seq(d, set_units(1,"weeks"), d) %>%
    add.sampling(set_units(seq(0, 5.5,by=0.005),weeks))

wkOnOff <- rxSolve(mod1, e)

plot(wkOnOff, C2)

## You can also repeat the cycle easily with the rep function

d <- et(timeUnits = "hr") %>% et(amt=10000, ii=24, until=set_units(2, "weeks"), cmt="depot")

e <- etRep(d, times=4, wait=set_units(1,"weeks")) %>%
    add.sampling(set_units(seq(0, 12.5,by=0.005),weeks))

repCycle4 <- rxSolve(mod1, e)

plot(repCycle4, C2)

---

### eventTable

**Create an event table object**

**Description**

Initializes an object of class ‘EventTable’ with methods for adding and querying dosing and observation records.

**Usage**

eventTable(amount.units = NA, time.units = NA)

**Arguments**

- **amount.units**: string denoting the amount dosing units, e.g., “mg”, “ug”. Default to NA to denote unspecified units. It could also be a solved RxODE object. In that case, eventTable(obj) returns the eventTable that was used to solve the RxODE object.

- **time.units**: string denoting the time units, e.g., “hours”, “days”. Default to “hours”.

An eventTable is an object that consists of a data.frame storing ordered timestamped events of an (unspecified) PK/PD dynamic system, units (strings) for dosing and time records, plus a list of functions to add and extract event records. Currently, events can be of two types: dosing events that represent inputs to the system and sampling time events that represent observations of the system with
‘amount.units’ and ‘time.units’, respectively. In the future, additional events may include resetting of state variables (compartments), for instance, to indicate time after “wash-out”, etc.

Value

A modified data.frame with the following accessible functions:

- **get.EventTable** returns the current event table.

- **add.dosing** adds dosing records to the event table. Its arguments are:
  - `dose`: numeric scalar, dose amount in `amount.units`;
  - `nbr.doses`: integer, number of doses;
  - `dosing.interval`: required numeric scalar, time between doses in `time.units`, defaults to 24 of `time.units="hours"`;
  - `dosing.to`: integer, compartment the dose goes into (first compartment by default);
  - `rate`: for infusions, the rate of infusion (default is `NULL` for bolus dosing);
  - `start.time`: required dosing start time;
  - `do.sampling`: logical, should observation sampling records be added at the dosing times? Defaults to `FALSE`.
  - `amount.units`: optional string indicating the dosing units. Defaults to `NA` to indicate as per the original EventTable definition.
  - `time.units`: optional string indicating the time units. Defaults to “hours” to indicate as per the original EventTable definition.

- **get.dosing** returns a data.frame of dosing records.

- **clear.dosing** clears or deletes all dosing from event table.

- **add.sampling** adds sampling time observation records to the event table. Its arguments are:
  - `time`: a vector of time values (in `time.units`).
  - `time.units`: an optional string specifying the time units. Defaults to the units specified when the EventTable was initialized.

- **get.sampling** returns a data.frame of sampled observation records.

- **clear.sampling** removes all sampling from event table.

- **get.obs.rec** returns a logical vector indicating whether each event record represents an observation or not.

- **get.nobs** returns the number of observation (not dosing) records.

- **get.units** returns a two-element character vector with the dosing and time units, respectively.

- **copy** makes a copy of the current event table. To create a copy of an event table object use `qd2 <- qd$copy()`.

- **expand** Expands the event table for multi-subject solving. This is done by `qd$expand(400)` for a 400 subject data expansion.
Author(s)
Matthew Fidler, Melissa Hallow and Wenping Wang

See Also
et, RxODE

Examples

```r
# create dosing and observation (sampling) events
# QD 50mg dosing, 5 days followed by 25mg 5 days
qd <- eventTable(amount.units = "mg", time.units = "days")
qd$add.dosing(dose=50, nbr.doses=5, dosing.interval = 1, do.sampling=FALSE)
qd$add.sampling(seq(from = 0, to = 1, by = 1/24))
qd$add.sampling(seq(from = 1, to = 5, by = 12/24))

# print(qd$get.dosing()) # table of dosing records
print(qd$get.nobs()) # number of observation (not dosing) records

# BID dosing, 5 days
bid <- eventTable("mg", "days") # only dosing
bid$add.dosing(dose=10000, nbr.doses=2*5,
   dosing.interval = 12, do.sampling=FALSE)

# Use the copy() method to create a copy (clone) of an existing
# event table (simple assignments just create a new reference to
# the same event table object (closure)).

bid.ext <- bid$copy() # three-day extension for a 2nd cohort
bid.ext$add.dosing(dose = 5000, nbr.doses = 2*3,
   start.time = 120, dosing.interval = 12, do.sampling = FALSE)

qd2 <- eventTable(amount.units="mg", time.units="days") %>%
   add.dosing(dose=50, nbr.doses=5, dosing.interval=1, do.sampling=FALSE) %>%
   add.sampling(seq(from=0, to=1, by=1 / 24)) %>%
   add.sampling(seq(from=1, to=5, by=12 / 24))

# print(qd2$get.dosing()) # table of dosing records
print(qd2$get.nobs()) # number of observation (not dosing) records

# Note that piping with %>% will update the original table.
qd3 <- qd2 %>% add.sampling(seq(from=5, to=10, by=6 / 24))
print(qd3$get.nobs())
```

**forderForceBase**

Force using base order for RxODE radix sorting

**Usage**

forderForceBase(forceBase = FALSE)

**Arguments**

forceBase boolean indicating if RxODE should use R’s `order` for radix sorting instead of `data.table`’s parallel radix sorting.

**Examples**

```r
## Not run:
forderForceBase(TRUE) # Use base 'order' for RxODE sorts
forderForceBase(FALSE) # Use base 'data.table' for RxODE sorts

## End(Not run)
```

---

**genShinyApp.template**

Generate an example (template) of a dosing regimen shiny app

**Description**

Create a complete shiny application for exploring dosing regimens given a (hardcoded) PK/PD model.

**Usage**

```r
genShinyApp.template(
  appDir = "shinyExample",
  verbose = TRUE,
  ODE.config = list(ode = "model", params = c(KA = 0.294), inits = c(eff = 1), method = "lsoda", atol = 1e-08, rtol = 1e-06)
)

write.template.server(appDir)
write.template.ui(appDir, statevars)
```
Arguments

appDir  
a string with a directory where to store the shiny app, by default is "shinyExample".
The directory appDir will be created if it does not exist.

verbose  
logical specifying whether to write messages as the shiny app is generated. Defaults to TRUE.

ODE.config  
model name compiled and list of parameters sent to `rxSolve`.

statevars  
List of statevars passed to the `write.template.ui` function. This usually isn't called directly.

A PK/PD model is defined using `RxODE`, and a set of parameters and initial values are defined. Then the appropriate R scripts for the shiny’s user interface `ui.R` and the server logic `server.R` are created in the directory `appDir`.

The function evaluates the following PK/PD model by default:

\[
\begin{align*}
C2 &= \text{centr}/V2; \\
C3 &= \text{peri}/V3; \\
d/dt(\text{depot}) &= -KA\times\text{depot}; \\
d/dt(\text{centr}) &= KA\times\text{depot} - CL\times C2 - Q\times C2 + Q\times C3; \\
d/dt(\text{peri}) &= Q\times C2 - Q\times C3; \\
d/dt(\text{eff}) &= Kin - Kout*(1-C2/(EC50+C2))*\text{eff};
\end{align*}
\]

This can be changed by the `ODE.config` parameter.

To launch the shiny app, simply issue the `runApp(appDir)` R command.

Value

None, these functions are used for their side effects.

Note

These functions create a simple, but working example of a dosing regimen simulation web application. Users may want to modify the code to experiment creating shiny applications for their specific `RxODE` models.

See Also

`RxODE`, `eventTable`, and the package `shiny` ([https://shiny.rstudio.com](https://shiny.rstudio.com)).

Examples

```r
## Not run:
# create the shiny app example (template)
genShinyApp.template(appDir = "myapp")
# run the shiny app
runApp("myapp")
## End(Not run)
```
is.rxEt  

*Check to see if this is an rxEt object.*

**Description**

Check to see if this is an rxEt object.

**Usage**

```r
is.rxEt(x)
```

**Arguments**

- `x` object to check to see if it is rxEt

If this is an rxEt object that has expired strip all rxEt information.

**Author(s)**

Matthew L.Fidler

---

is.rxSolve  

*Check to see if this is an rxSolve object.*

**Description**

Check to see if this is an rxSolve object.

**Usage**

```r
is.rxSolve(x)
```

**Arguments**

- `x` object to check to see if it is rxSolve

If this is an rxSolve object that has expired strip all rxSolve information.

**Author(s)**

Matthew L.Fidler
pillar_shaft

*pillar_shaft*

**Re export of pillar_shaft**

**Description**

Re export of pillar_shaft

**Usage**

`pillar_shaft(x, ...)`

**Arguments**

- `x`  A vector to format
- `...`  Unused, for extensibility.

pillar_shaft.rxRateDur

*Pillar shaft for rxRateDur*

**Description**

Pillar shaft for rxRateDur

**Usage**

```r
# S3 method for class 'rxRateDur'
pillar_shaft(x, ...)
```

**Arguments**

- `x`  A vector to format
- `...`  Unused, for extensibility.
print.rxCoefSolve  
Print the rxCoefSolve object

Description
This prints out the user supplied arguments for the rxCoef object

Usage
## S3 method for class 'rxCoefSolve'
print(x, ...)

Arguments
x  rxCoefSolve object
... Other (ignored) parameters.

Author(s)
Matthew L.Fidler

print.RxODE  
Print information about the RxODE object.

Description
This prints the model name and its status for being able to be solved

Usage
## S3 method for class 'RxODE'
print(x, ...)

Arguments
x  An rxode object
... Ignored parameters

Author(s)
Matthew L.Fidler
rinvchisq

Scaled Inverse Chi Squared distribution

Description

Scaled Inverse Chi Squared distribution

Usage

rinvchisq(n = 1L, nu = 1, scale = 1)

Arguments

n  Number of random samples
nu degrees of freedom of inverse chi square
scale Scale of inverse chi squared distribution (default is 1).

Value

a vector of inverse chi squared deviates.

Examples

rinvchisq(3, 4, 1) ## Scale = 1, degrees of freedom = 4
rinvchisq(2, 4, 2) ## Scale = 2, degrees of freedom = 4

rxAddReturn

Add a return statement to a function.

Description

Add a return statement to a function.

Usage

rxAddReturn(fn, ret = TRUE)

Arguments

fn Function to deparse
ret boolean stating if a return statement will be added.

Value

Function with parens removed and add a return statement.
**rxAllowUnload**  

*Allow unloading of dlls*

**Description**  
Allow unloading of dlls

**Usage**  
```
rxAllowUnload(allow)
```

**Arguments**
- **allow**  
  boolean indicating if garbage collection will unload of RxODE dlls.

**Author(s)**  
Matthew L. Fidler

**Examples**

```
# Garbage collection will not unload un-used RxODE dlls
rxAllowUnload(FALSE);

# Garbage collection will unload unused RxODE dlls
rxAllowUnload(TRUE);
```

---

**rxAssignPtr**  

*Assign pointer based on model variables*

**Description**  
Assign pointer based on model variables

**Usage**  
```
rxAssignPtr(object = NULL)
```

**Arguments**
- **object**  
  RxODE family of objects
rxC14

Setup C++14 support in windows (required for nlmixr)

Description

Setup C++14 support in windows (required for nlmixr)

Usage

rxC14()

Value

nothing

rxChain

rxChain Chain or add item to solved system of equations

Description

Add item to solved system of equations

Usage

rxChain(obj1, obj2)

## S3 method for class 'solveRxDll'
obj1 + obj2

Arguments

obj1 Solved object.
obj2 New object to be added/piped/chained to solved object.

Value

When newObject is an event table, return a new solved object with the new event table.

Author(s)

Matthew L. Fidler
**rxClean**

Cleanup anonymous DLLs by unloading them

**Description**

This cleans up any RxODE loaded DLLs

**Usage**

```r
rxClean(wd)
```

**Arguments**

- `wd` What directory should be cleaned; (DEPRECATED), this no longer does anything. This unloads all RxODE anonymous dlls.

**Value**

TRUE if successful

**Author(s)**

Matthew L. Fidler

**rxCompile**

Compile a model if needed

**Description**

This is the compilation workhorse creating the RxODE model DLL files.

**Usage**

```r
rxCompile(
  model,
  dir,
  prefix,
  extraC = NULL,
  force = FALSE,
  modName = NULL,
  package = NULL,
  ...
)
```

## S3 method for class 'character'
rxCompile(
  model,
  dir = NULL,
  prefix = NULL,
  extraC = NULL,
  force = FALSE,
  modName = NULL,
  package = NULL,
  ...
)

## S3 method for class 'rxDll'
rxCompile(model, ...)

## S3 method for class 'RxODE'
rxCompile(model, ...)

Arguments

- **model**: This is the ODE model specification. It can be:
  - a string containing the set of ordinary differential equations (ODE) and other expressions defining the changes in the dynamic system.
  - a file name where the ODE system equation is contained
  - An ODE expression enclosed in `{}` (see also the filename argument). For details, see the sections “Details” and “RxODE Syntax” below.

- **dir**: This is the model directory where the C file will be stored for compiling. If unspecified, the C code is stored in a temporary directory, then the model is compiled and moved to the current directory. Afterwards the C code is removed. If specified, the C code is stored in the specified directory and then compiled in that directory. The C code is not removed after the DLL is created in the same directory. This can be useful to debug the c-code outputs.

- **prefix**: is a string indicating the prefix to use in the C based functions. If missing, it is calculated based on file name, or md5 of parsed model.

- **extraC**: Extra c code to include in the model. This can be useful to specify functions in the model. These C functions should usually take double precision arguments, and return double precision values.

- **force**: is a boolean stating if the (re)compile should be forced if RxODE detects that the models are the same as already generated.

- **modName**: a string to be used as the model name. This string is used for naming various aspects of the computations, including generating C symbol names, dynamic libraries, etc. Therefore, it is necessary that modName consists of simple ASCII alphanumeric characters starting with a letter.

- **package**: Package name for pre-compiled binaries.

... Other arguments sent to the *rxTrans* function.
**Value**

An rxDll object that has the following components

- **dll**: DLL path
- **model**: model specification
- **.c**: A function to call C code in the correct context from the DLL using the `.C` function.
- **.call**: A function to call C code in the correct context from the DLL using the `.Call` function.
- **args**: A list of the arguments used to create the rxDll object.

**Author(s)**

Matthew L. Fidler

**See Also**

RxODE

---

**rxControl**

*Solving & Simulation of a ODE/solved system (and solving options)*

**Description**

This uses RxODE family of objects, file, or model specification to solve a ODE system.

**Usage**

```r
rxControl(
  scale = NULL,
  method = c("liblsoda", "lsoda", "dop853"),
  transitAbs = NULL,
  atol = 1e-08,
  rtol = 1e-06,
  maxsteps = 70000L,
  hmin = 0L,
  hmax = NA,
  hmaxSd = 0,
  hini = 0,
  maxordn = 12L,
  maxords = 5L,
  ...,
  cores,
  covsInterpolation = c("locf", "linear", "nocb", "midpoint"),
  addCov = FALSE,
)```
matrix = FALSE,
sigma = NULL,
sigmaDf = NULL,
sigmaLower = -Inf,
sigmaUpper = Inf,
nCoresRV = 1L,
sigmaIsChol = FALSE,
nDisplayProgress = 10000L,
amountUnits = NA_character_,
timeUnits = "hours",
stiff,
theta = NULL,
thetaLower = -Inf,
thetaUpper = Inf,
eta = NULL,
addDosing = FALSE,
stateTrim = Inf,
updateObject = FALSE,
omega = NULL,
omegaDf = NULL,
omegaIsChol = FALSE,
omegaLower = -Inf,
omegaUpper = Inf,
nSub = 1L,
thetaMat = NULL,
thetaDf = NULL,
thetaIsChol = FALSE,
nStud = 1L,
dfSub = 0,
dfObs = 0,
returnType = c("rxSolve", "matrix", "data.frame", "data.frame.TBS", "data.table",
"tbl", "tibble"),
seed = NULL,
nsim = NULL,
minSS = NULL,
maxSS = 1000L,
infSSstep = 12,
strictSS = TRUE,
params = NULL,
events = NULL,
istateReset = TRUE,
subsetNonmem = TRUE,
linLog = FALSE,
maxAtolRtolFactor = 0.1,
from = NULL,
to = NULL,
by = NULL,
length.out = NULL,
iCov = NULL,
keep = NULL,
idFactor = TRUE,
mxhinl = 0,
hmxi = 0,
warnIdSort = TRUE,
ssAtol = 1e-08,
ssRtol = 1e-06,
safeZero = TRUE
)

rxSolve(object, ...)

## Default S3 method:
rxSolve(object, params = NULL, events = NULL, inits = NULL, ...)

## S3 method for class 'rxSolve'
update(object, ...)

## S3 method for class 'RxODE'
predict(object, ...)

## S3 method for class 'rxSolve'
predict(object, ...)

## S3 method for class 'rxEt'
predict(object, ...)

## S3 method for class 'rxParams'
predict(object, ...)

## S3 method for class 'RxODE'
simulate(object, nsim = 1L, seed = NULL, ...)

## S3 method for class 'rxSolve'
simulate(object, nsim = 1L, seed = NULL, ...)

## S3 method for class 'rxParams'
simulate(object, nsim = 1L, seed = NULL, ...)

## S3 method for class 'rxSolve'
solve(a, b, ...)

## S3 method for class 'RxODE'
solve(a, b, ...)

## S3 method for class 'rxParams'
solve(a, b, ...)
## S3 method for class 'rxEt'
solve(a, b, ...)

### Arguments

- **scale** a numeric named vector with scaling for ode parameters of the system. The names must correspond to the parameter identifiers in the ODE specification. Each of the ODE variables will be divided by the scaling factor. For example `scale=c(center=2)` will divide the center ODE variable by 2.

- **method** The method for solving ODEs. Currently this supports:
  - "liblsoda" thread safe lsoda. This supports parallel thread-based solving, and ignores user Jacobian specification.
  - "lsoda" – LSODA solver. Does not support parallel thread-based solving, but allows user Jacobian specification.

- **transitAbs** boolean indicating if this is a transit compartment absorption

- **atol** a numeric absolute tolerance (1e-8 by default) used by the ODE solver to determine if a good solution has been achieved; This is also used in the solved linear model to check if prior doses do not add anything to the solution.

- **rtol** a numeric relative tolerance (1e-6 by default) used by the ODE solver to determine if a good solution has been achieved. This is also used in the solved linear model to check if prior doses do not add anything to the solution.

- **maxsteps** maximum number of (internally defined) steps allowed during one call to the solver. (5000 by default)

- **hmin** The minimum absolute step size allowed. The default value is 0.

- **hmax** The maximum absolute step size allowed. When `hmax=NA` (default), uses the average difference `(hmaxSd*sd)` in times and sampling events. When `hmax=NULL` RxODE uses the maximum difference in times in your sampling and events. The value 0 is equivalent to infinite maximum absolute step size.

- **hmaxSd** The number of standard deviations of the time difference to add to hmax. The default is 0

- **hini** The step size to be attempted on the first step. The default value is determined by the solver (when hini = 0)

- **maxordn** The maximum order to be allowed for the nonstiff (Adams) method. The default is 12. It can be between 1 and 12.

- **maxords** The maximum order to be allowed for the stiff (BDF) method. The default value is 5. This can be between 1 and 5.

- **cores** Number of cores used in parallel ODE solving. This defaults to the number or system cores determined by `rxCores` for methods that support parallel solving (ie thread-safe methods like "liblsoda").
covsInterpolation specifies the interpolation method for time-varying covariates. When solving ODEs it often samples times outside the sampling time specified in events. When this happens, the time varying covariates are interpolated. Currently this can be:

- "linear" interpolation (the default), which interpolates the covariate by solving the line between the observed covariates and extrapolating the new covariate value.
- "constant" – Last observation carried forward.
- "NOCB" – Next Observation Carried Backward. This is the same method that NONMEM uses.
- "midpoint" Last observation carried forward to midpoint; Next observation carried backward to midpoint.

addCov A boolean indicating if covariates should be added to the output matrix or data frame. By default this is disabled.

matrix A boolean indicating if a matrix should be returned instead of the RxODE’s solved object.

sigma Named sigma covariance or Cholesky decomposition of a covariance matrix. The names of the columns indicate parameters that are simulated. These are simulated for every observation in the solved system.

sigmaDf Degrees of freedom of the sigma t-distribution. By default it is equivalent to Inf, or a normal distribution.

sigmaLower Lower bounds for simulated unexplained variability (by default -Inf)

sigmaUpper Upper bounds for simulated unexplained variability (by default Inf)

nCoresRV Number of cores used for the simulation of the sigma variables. By default this is 1. This uses the package rmvn and rmvt. To reproduce the results you need to run on the same platform with the same number of cores. This is the reason this is set to be one, regardless of what the number of cores are used in threaded ODE solving.

sigmaIsChol Boolean indicating if the sigma is in the Cholesky decomposition instead of a symmetric covariance

nDisplayProgress An integer indicating the minimum number of c-based solves before a progress bar is shown. By default this is 10,000.

amountUnits This supplies the dose units of a data frame supplied instead of an event table. This is for importing the data as an RxODE event table.

timeUnits This supplies the time units of a data frame supplied instead of an event table. This is for importing the data as an RxODE event table.

stiff a logical (TRUE by default) indicating whether the ODE system is stiff or not. For stiff ODE systems (stiff = TRUE), RxODE uses the LSODA (Livermore Solver for Ordinary Differential Equations) Fortran package, which implements an automatic method switching for stiff and non-stiff problems along the integration interval, authored by Hindmarsh and Petzold (2003). For non-stiff systems (stiff = FALSE), RxODE uses DOP853, an explicit Runge-Kutta method of order 8(5, 3) of Dormand and Prince as implemented in C by Hairer and Wanner (1993).
<table>
<thead>
<tr>
<th>Parameter</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>theta</td>
<td>A vector of parameters that will be named THETA[#] and added to parameters.</td>
</tr>
<tr>
<td>thetaLower</td>
<td>Lower bounds for simulated population parameter variability (by default -Inf).</td>
</tr>
<tr>
<td>thetaUpper</td>
<td>Upper bounds for simulated population unexplained variability (by default Inf).</td>
</tr>
<tr>
<td>eta</td>
<td>A vector of parameters that will be named ETA[#] and added to parameters.</td>
</tr>
<tr>
<td>addDosing</td>
<td>Boolean indicating if the solve should add RxODE EVID and related columns. This will also include dosing information and estimates at the doses.</td>
</tr>
<tr>
<td></td>
<td>RxODE only includes estimates at the observations. (default FALSE). When addDosing is NULL, only include EVID=0 on solve and exclude any model-</td>
</tr>
<tr>
<td></td>
<td>times or EVID=2. If addDosing is NA the classic RxODE EVID events. When addDosing is TRUE add the event information in NONMEM-style format; If</td>
</tr>
<tr>
<td></td>
<td>subsetNonmem=FALSE RxODE will also extra event types (EVID) for ending infusion and modeled times:</td>
</tr>
<tr>
<td></td>
<td>• EVID=-1 When the modeled rate infusions are turned off (matches rate=-1)</td>
</tr>
<tr>
<td></td>
<td>• EVID=-2 When the modeled duration infusions are turned off (matches rate=-2)</td>
</tr>
<tr>
<td></td>
<td>• EVID=-10 When the specified rate infusions are turned off (matches rate&gt;0)</td>
</tr>
<tr>
<td></td>
<td>• EVID=-20 When the specified dur infusions are turned off (matches dur&gt;0)</td>
</tr>
<tr>
<td></td>
<td>• EVID=101,102,103,... Modeled time where 101 is the first model time, 102 is the second etc.</td>
</tr>
<tr>
<td>stateTrim</td>
<td>When amounts/concentrations in one of the states are above this value, trim them to be this value. By default Inf. Also trims to -stateTrim for</td>
</tr>
<tr>
<td></td>
<td>large negative amounts/concentrations.</td>
</tr>
<tr>
<td>updateObject</td>
<td>This is an internally used flag to update the RxODE solved object (when supplying an RxODE solved object) as well as returning a new object.</td>
</tr>
<tr>
<td></td>
<td>You probably should not modify it’s FALSE default unless you are willing to have unexpected results.</td>
</tr>
<tr>
<td>omega</td>
<td>Estimate of Covariance matrix. When omega is a list, assume it is a block matrix and convert it to a full matrix for simulations.</td>
</tr>
<tr>
<td>omegaDf</td>
<td>The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a</td>
</tr>
<tr>
<td></td>
<td>normal distribution instead of a t-distribution.</td>
</tr>
<tr>
<td>omegaIsChol</td>
<td>Indicates if the omega supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.</td>
</tr>
<tr>
<td>omegaLower</td>
<td>Lower bounds for simulated ETAs (by default -Inf).</td>
</tr>
<tr>
<td>omegaUpper</td>
<td>Upper bounds for simulated ETAs (by default Inf).</td>
</tr>
<tr>
<td>nSub</td>
<td>Number between subject variabilities (ETAs) simulated for every realization of the parameters.</td>
</tr>
<tr>
<td>thetaMat</td>
<td>Named theta matrix.</td>
</tr>
<tr>
<td>thetaDf</td>
<td>The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a</td>
</tr>
<tr>
<td></td>
<td>normal distribution instead of a t-distribution.</td>
</tr>
<tr>
<td>thetaIsChol</td>
<td>Indicates if the theta supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.</td>
</tr>
</tbody>
</table>
nStud  Number virtual studies to characterize uncertainty in estimated parameters.
dfSub  Degrees of freedom to sample the between subject variability matrix from the
dfObs  inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.
returnType  This tells what type of object is returned. The currently supported types are:
  • "rxSolve" (default) will return a reactive data frame that can change easily
    change different pieces of the solve and update the data frame. This is the
    currently standard solving method in RxODE, is used for rxSolve(object,...).
    solve(object,...),
  • "data.frame" — returns a plain, non-reactive data frame; Currently very
    slightly faster than returnType="matrix"
  • "matrix" — returns a plain matrix with column names attached to the solved
    object. This is what is used object$run as well as object$solve
  • "data.table" — returns a data.table; The data.table is created by ref-erence (ie setDt()), which should be fast.
  • "tbl" or "tibble" returns a tibble format.

seed  an object specifying if and how the random number generator should be initial-

nsim  represents the number of simulations. For RxODE, if you supply single subject

minSS  Minimum number of iterations for a steady-state dose
maxSS  Maximum number of iterations for a steady-state dose
infSSstep  Step size for determining if a constant infusion has reached steady state. By
default this is large value, 420.
strictSS  Boolean indicating if a strict steady-state is required. If a strict steady-state is

params  a numeric named vector with values for every parameter in the ODE system; the

events  an eventTable object describing the input (e.g., doses) to the dynamic system

istateReset  When TRUE, reset the ISTATE variable to 1 for lsoda and liblsoda with doses,

subsetNonmem  subset to NONMEM compatible EVIDs only. By default TRUE.
linLog  Boolean indicating if linear compartment models be calculated more accurately


maxAtolRtolFactor

The maximum atol/rtol that FOCEi and other routines may adjust to. By default 0.1

from

When there is no observations in the event table, start observations at this value. By default this is zero.

to

When there is no observations in the event table, end observations at this value. By default this is 24 + maximum dose time.

by

When there are no observations in the event table, this is the amount to increment for the observations between from and to.

length.out

The number of observations to create if there isn’t any observations in the event table. By default this is 200.

iCov

A data frame of individual non-time varying covariates to combine with the params to form a parameter data.frame.

keep

Columns to keep from either the input dataset or the iCov dataset. With the iCov dataset, the column is kept once per line. For the input dataset, if any records are added to the data LOCF (Last Observation Carried forward) imputation is performed.

idFactor

This boolean indicates if original ID values should be maintained. This changes the default sequentially ordered ID to a factor with the original ID values in the original dataset. By default this is enabled.

mxhnil

maximum number of messages printed (per problem) warning that T + H = T on a step (H = step size). This must be positive to result in a non-default value. The default value is 0 (or infinite).

hmxi

inverse of the maximum absolute value of H to be used. hmxi = 0.0 is allowed and corresponds to an infinite hmax (default). hmin and hmxi may be changed at any time, but will not take effect until the next change of H is considered. This option is only considered with method=liblsoda.

warnIdSort

Warn if the ID is not present and RxODE assumes the order of the parameters/iCov are the same as the order of the parameters in the input dataset.

ssAtol

Steady state atol convergence factor. Can be a vector based on each state.

ssRtol

Steady state rtol convergence factor. Can be a vector based on each state.

safeZero

Use safe zero divide and log routines. By default this is turned on but you may turn it off if you wish.

object

is a either a RxODE family of objects, or a file-name with a RxODE model specification, or a string with a RxODE model specification.

inits

a vector of initial values of the state variables (e.g., amounts in each compartment), and the order in this vector must be the same as the state variables (e.g., PK/PD compartments);

a

when using solve, this is equivalent to the object argument. If you specify object later in the argument list it overwrites this parameter.

b

when using solve, this is equivalent to the params argument. If you specify params as a named argument, this overwrites the output
Value

An “rxSolve” solve object that stores the solved value in a matrix with as many rows as there are sampled time points and as many columns as system variables (as defined by the ODEs and additional assignments in the RxODE model code). It also stores information about the call to allow dynamic updating of the solved object.

The operations for the object are similar to a data-frame, but expand the $ and [[""]] access operators and assignment operators to resolve based on different parameter values, initial conditions, solver parameters, or events (by updating the time variable).

You can call the `eventTable` methods on the solved object to update the event table and resolve the system of equations.

Author(s)

Matthew Fidler, Melissa Hallow and Wenping Wang

References


See Also

RxODE

rxCores

---

### Description

Get the number of cores in a system

### Usage

```r
rxCores()
```
**rxDelete**

*Delete the DLL for the model*

**Description**
This function deletes the DLL, but doesn’t delete the model information in the object.

**Usage**
rxDelete(obj)

**Arguments**
obj RxODE family of objects

**Value**
A boolean stating if the operation was successful.

**Author(s)**
Matthew L. Fidler

---

**rxDfdy**

*Jacobian and parameter derivatives*

**Description**
Return Jacobian and parameter derivatives

**Usage**
rxDfdy(obj)

**Arguments**
obj RxODE family of objects

**Value**
A list of the jacobian parameters defined in this RxODE object.

**Author(s)**
Matthew L. Fidler
**rxEvid**

*EVID formatting for tibble and other places.*

**Description**

This is to make an EVID more readable by non pharmacometricians. It displays what each means and allows it to be displayed in a tibble.

**Usage**

```r
rxEvid(x)

as.rxEvid(x)
```

```r
## S3 method for class 'single.Var'
rxEvid

c(x, ...)
```

```r
## S3 method for class 'single.Var'
rxEvid

x[...]
```

```r
## S3 method for class 'single.Var'
rxEvid

as.character(x, ...)
```

```r
## S3 method for class 'single.Var'
rxEvid

format(x, ...)
```

```r
## S3 method for class 'single.Var'
rxEvid

print(x, ...)
```

```r
## S3 method for class 'single.Var'
rxEvid

x[...]  
```

```r
## S3 method for class 'single.Var'
rxEvid

type_sum(x)
```

```r
## S3 method for class 'single.Var'
rxEvid

pillar_shaft(x, ...)
```

**Arguments**

- `x`  
  Item to be converted to a RxODE EVID specification.
- `...`  
  Other parameters

**Examples**

```r
rxEvid(1:7)
```
rxFoExpandEta  \textit{First Order Expansion of ETA}

\begin{description}
\item[Description] First Order Expansion of ETA
\item[Usage] \texttt{rxFoExpandEta(expr)}
\item[Arguments] \begin{align*}
\text{expr} & \quad \text{RxODE model}
\end{align*}
\item[Value] Return a RxODE model with first order Taylor expansion around ETA
\item[Author(s)] Matthew L. Fidler
\end{description}

rxGetRxODE  \textit{Get RxODE model from object}

\begin{description}
\item[Description] Get RxODE model from object
\item[Usage] \texttt{rxGetRxODE(obj)}
\item[Arguments] \begin{align*}
\text{obj} & \quad \text{RxODE family of objects}
\end{align*}
\end{description}
**rxHtml**

Format `rxSolve` and related objects as html.

**Description**

Format `rxSolve` and related objects as html.

**Usage**

```r
rxHtml(x, ...)
```

```r
## S3 method for class 'rxSolve'
rxHtml(x, ...)
```

**Arguments**

- `x` RxODE object
- `...` Extra arguments sent to `kable`

**Author(s)**

Matthew L. Fidler

---

**rxInv**

Invert matrix using `RcppArmadillo`.

**Description**

Invert matrix using `RcppArmadillo`.

**Usage**

```r
rxInv(matrix)
```

**Arguments**

- `matrix` matrix to be inverted.

**Value**

inverse or pseudo inverse of matrix.
### rxIsCurrent

*Checks if the RxODE object was built with the current build*

**Description**

Checks if the RxODE object was built with the current build

**Usage**

```r
rxIsCurrent(obj)
```

**Arguments**

- `obj` RxODE family of objects

**Value**

boolean indicating if this was built with current RxODE

### rxLhs

*Left handed Variables*

**Description**

This returns the model calculated variables

**Usage**

```r
rxLhs(obj)
```

**Arguments**

- `obj` RxODE family of objects

**Value**

a character vector listing the calculated parameters

**Author(s)**

Matthew L. Fidler

**See Also**

*RxODE*
**rxLock**

**Description**
Lock/unlocking of RxODE dll file

**Usage**
- `rxLock(obj)`
- `rxUnlock(obj)`

**Arguments**
- `obj` A RxODE family of objects

---

**rxNorm**

**Description**
This get the syntax preferred model for processing

**Usage**
- `rxNorm(obj, condition = NULL, removeInis, removeJac, removeSens)`

**Arguments**
- `obj` RxODE family of objects
- `condition` Character string of a logical condition to use for subsetting the normalized model. When missing, and a condition is not set via `rxCondition`, return the whole code with all the conditional settings intact. When a condition is set with `rxCondition`, use that condition.
- `removeInis` A boolean indicating if parameter initialization will be removed from the model
- `removeJac` A boolean indicating if the Jacobians will be removed.
- `removeSens` A boolean indicating if the sensitivities will be removed.

**Value**
Normalized Normal syntax (no comments)

**Author(s)**
Matthew L. Fidler
Create an ODE-based model specification

Description

Create a dynamic ODE-based model object suitably for translation into fast C code

Usage

```r
RxODE(
  model,
  modName = basename(wd),
  wd = getwd(),
  filename = NULL,
  extraC = NULL,
  debug = FALSE,
  calcJac = NULL,
  calcSens = NULL,
  collapseModel = FALSE,
  package = NULL,
  ...
)
```

Arguments

- **model**: This is the ODE model specification. It can be:
  - a string containing the set of ordinary differential equations (ODE) and other expressions defining the changes in the dynamic system.
  - a file name where the ODE system equation is contained
  - An ODE expression enclosed in `{}`

(see also the `filename` argument). For details, see the sections “Details” and “RxODE Syntax” below.

- **modName**: a string to be used as the model name. This string is used for naming various aspects of the computations, including generating C symbol names, dynamic libraries, etc. Therefore, it is necessary that `modName` consists of simple ASCII alphanumeric characters starting with a letter.

- **wd**: character string with a working directory where to create a subdirectory according to `modName`. When specified, a subdirectory named after the "modName.d" will be created and populated with a C file, a dynamic loading library, plus various other working files. If missing, the files are created (and removed) in the temporary directory, and the RxODE DLL for the model is created in the current directory named `rx_????_platform`, for example `rx_129f8f97fb94a87ca49ca8dafe691e1e_i386.dll`.

- **filename**: A file name or connection object where the ODE-based model specification resides. Only one of `model` or `filename` may be specified.
extraC Extra C code to include in the model. This can be useful to specify functions in the model. These C functions should usually take double precision arguments, and return double precision values.

debug is a boolean indicating if the executable should be compiled with verbose debugging information turned on.

calcJac boolean indicating if RxODE will calculate the Jacobian according to the specified ODEs.

calcSens boolean indicating if RxODE will calculate the sensitivities according to the specified ODEs.

collapseModel boolean indicating if RxODE will remove all LHS variables when calculating sensitivities.

package Package name for pre-compiled binaries.

... ignored arguments.

The “Rx” in the name RxODE is meant to suggest the abbreviation Rx for a medical prescription, and thus to suggest the package emphasis on pharmacometrics modeling, including pharmacokinetics (PK), pharmacodynamics (PD), disease progression, drug-disease modeling, etc.

The ODE-based model specification may be coded inside a character string or in a text file, see Section RxODE Syntax below for coding details. An internal RxODE compilation manager object translates the ODE system into C, compiles it, and dynamically loads the object code into the current R session. The call to RxODE produces an object of class RxODE which consists of a list-like structure (closure) with various member functions (see Section Value below).

For evaluating RxODE models, two types of inputs may be provided: a required set of time points for querying the state of the ODE system and an optional set of doses (input amounts). These inputs are combined into a single event table object created with the function eventTable.

Value

An object (closure) of class “RxODE” (see Chambers and Temple Lang (2001)) consisting of the following list of strings and functions:

modName the name of the model (a copy of the input argument).

model a character string holding the source model specification.

get.modelVars a function that returns a list with 3 character vectors, params, state, and lhs of variable names used in the model specification. These will be output when the model is computed (i.e., the ODE solved by integration).

solve this function solves (integrates) the ODE. This is done by passing the code to rxSolve. This is as if you called rxSolve(RxODEobject,...), but returns a matrix instead of a rxSolve object.

params: a numeric named vector with values for every parameter in the ODE system; the names must correspond to the parameter identifiers used in the ODE specification;

events: an eventTable object describing the input (e.g., doses) to the dynamic system and observation sampling time points (see eventTable);
**RxODE**

- **init**: a vector of initial values of the state variables (e.g., amounts in each compartment), and the order in this vector must be the same as the state variables (e.g., PK/PD compartments);
- **stiff**: a logical (TRUE by default) indicating whether the ODE system is stiff or not.

For stiff ODE systems (stiff = TRUE), RxODE uses the LSODA (Livermore Solver for Ordinary Differential Equations) Fortran package, which implements an automatic method switching for stiff and non-stiff problems along the integration interval, authored by Hindmarsh and Petzold (2003).

For non-stiff systems (stiff = FALSE), RxODE uses DOP853, an explicit Runge-Kutta method of order 8(5, 3) of Dormand and Prince as implemented in C by Hairer and Wanner (1993).

- **trans_abs**: a logical (FALSE by default) indicating whether to fit a transit absorption term (TODO: need further documentation and example);
- **atol**: a numeric absolute tolerance (1e-08 by default);
- **rtol**: a numeric relative tolerance (1e-06 by default).

The output of “solve” is a matrix with as many rows as there are sampled time points and as many columns as system variables (as defined by the ODEs and additional assignments in the RxODE model code).

- **isValid**: a function that (naively) checks for model validity, namely that the C object code reflects the latest model specification.
- **version**: a string with the version of the RxODE object (not the package).
- **dynLoad**: a function with one force = FALSE argument that dynamically loads the object code if needed.
- **dynUnload**: a function with no argument that unloads the model object code.
- **delete**: removes all created model files, including C and DLL files. The model object is no longer valid and should be removed, e.g., rm(m1).
- **run**: deprecated, use solve.
- **parse**: deprecated.
- **compile**: deprecated.
- **get.index**: deprecated.
- **getObj**: internal (not user callable) function.

**RxODE Syntax**

An RxODE model specification consists of one or more statements terminated by semi-colons, ‘;’, and optional comments (comments are delimited by # and an end-of-line marker). **NB:** Comments are not allowed inside statements.

A block of statements is a set of statements delimited by curly braces, ‘{ ... }’. Statements can be either assignments or conditional if statements. Assignment statements can be: (1) “simple” assignments, where the left hand is an identifier (i.e., variable), (2) special “time-derivative” assignments, where the left hand specifies the change of that variable with respect to time e.g., \( \frac{d}{dt}(depot) \), or (3) special “jacobian” assignments, where the left hand specifies the change of the ODE with respect to one of the parameters, e.g. \( \frac{df(depot)}{dy(kel)} \). The “jacobian” assignments are not required, and are only useful for very stiff differential systems.
Expressions in assignment and ‘if’ statements can be numeric or logical (no character expressions are currently supported). Numeric expressions can include the following numeric operators (‘+’, ‘-’, ‘*’, ‘/’, ‘^’), and those mathematical functions defined in the C or the R math libraries (e.g., fabs, exp, log, sin). (Notice that the modulo operator ‘%’ is currently not supported.)

Identifiers in an RxODE model specification can refer to:

- state variables in the dynamic system (e.g., compartments in a pharmacokinetics/pharmacodynamics model);
- implied input variable, t (time), podo (oral dose, for absorption models), and tlast (last time point);
- model parameters, (ka rate of absorption, CL clearance, etc.);
- pi, for the constant pi.
- others, as created by assignments as part of the model specification.

Identifiers consists of case-sensitive alphanumeric characters, plus the underscore '_' character. **NB:** the dot '.' character is **not** a valid character identifier.

The values of these variables at pre-specified time points are saved as part of the fitted/integrated/solved model (see `eventTable`, in particular its member function `add.sampling` that defines a set of time points at which to capture a snapshot of the system via the values of these variables).

The ODE specification mini-language is parsed with the help of the open source tool `dparser`, Plevyak (2015).

**Author(s)**

Melissa Hallow, Wenping Wang and Matthew Fidler

**References**


**See Also**

`eventTable`, `et`, `add.sampling`, `add.dosing`
Examples

# Step 1 - Create a model specification
ode <- "
# A 4-compartment model, 3 PK and a PD (effect) compartment
# (notice state variable names 'depot', 'centr', 'peri', 'eff')

C2 = centr/V2;
C3 = peri/V3;
d/dt(depot) = -KA*depot;
d/dt(centr) = KA*depot - CL*C2 - Q*C2 + Q*C3;
d/dt(peri) = Q*C2 - Q*C3;
d/dt(eff) = Kin - Kout*(1-C2/(EC50+C2))*eff;
"

m1 <- RxODE(model = ode)
print(m1)

# Step 2 - Create the model input as an EventTable,
# including dosing and observation (sampling) events

# QD (once daily) dosing for 5 days.
qd <- eventTable(amount.units = "ug", time.units = "hours")
qd$add.dosing(dose = 10000, nbr.doses = 5, dosing.interval = 24)

# Sample the system hourly during the first day, every 8 hours
# then after
qd$add.sampling(0:24)
qd$add.sampling(seq(from = 24+8, to = 5*24, by = 8))

# Step 3 - set starting parameter estimates and initial
# values of the state

theta <-
c(KA = .291, CL = 18.6,
 V2 = 40.2, Q = 10.5, V3 = 297.0,
 Kin = 1.0, Kout = 1.0, EC50 = 200.0)

# init state variable
inits <- c(0, 0, 0, 1);

# Step 4 - Fit the model to the data
qd.cp <- m1$solve(theta, events = qd, inits)

head(qd.cp)

# This returns a matrix. Note that you can also
# solve using name initial values. For example:
inits <- c(eff = 1);
qd.cp <- solve(m1, theta, events = qd, inits);
print(qd.cp)
plot(qd.cp)

---

**rxOptExpr**

*Optimize RxODE for computer evaluation*

**Description**

This optimizes RxODE code for computer evaluation by only calculating redundant expressions once.

**Usage**

`rxOptExpr(x)`

**Arguments**

- `x` RxODE model that can be access by `rxNorm`

**Value**

Optimized RxODE model text. The order and type lhs and state variables is maintained while the evaluation is sped up. While parameters names are maintained, their order may be modified.

**Author(s)**

Matthew L. Fidler

---

**rxOptions**

*Options for RxODE*

**Description**

This is a backend for `rxPermissive` (with `op.rx = 2`) and `rxStrict` (with `op.rx = 1`)
Usage

```r
rxOptions(
  expr,
  op.rx = NULL,
  silent = .isTestthat(),
  respect = FALSE,
  cran = FALSE,
  on.validate = FALSE
)
```

Arguments

- **expr**: Expression to evaluate in the permissive/strict environment. If unspecified, set the options for the current environment.
- **op.rx**: A numeric for strict (1) or permissive (2) syntax.
- **silent**: when true, also silence the syntax errors and interactive output (useful in testing).
- **respect**: when TRUE, respect any options that are specified. This is called at startup, but really should not be called elsewhere, otherwise the options are not changed.
- **cran**: When specified and true, run on CRAN. Otherwise it is skipped on CRAN.
- **on.validate**: When TRUE run only when validating.

Details

When `expr` is missing and `op.rx` is NULL, this displays the current RxODE options.

Author(s)

Matthew L. Fidler

---

```r
rxParams

Parameters specified by the model
```

Description

This returns the model’s parameters that are required to solve the ODE system, and can be used to pipe parameters into an RxODE solve

Usage

```r
rxParams(obj, ...)
```

## S3 method for class 'RxODE'

```r
rxParams(
  obj,
```
### S3 method for class 'rxSolve'

```r
rxParams(
  obj,
  constants = TRUE,
  ...,  
  params = NULL,
  inits = NULL,
  iCov = NULL,
  keep = NULL,
  thetaMat = NULL,
  omega = NULL,
  dfSub = NULL,
  sigma = NULL,
  dfObs = NULL,
  nSub = NULL,
  nStud = NULL
)
```

### S3 method for class 'rxEt'

```r
rxParams(
  obj,
  ...,  
  params = NULL,
  inits = NULL,
  iCov = NULL,
  keep = NULL,
  thetaMat = NULL,
  omega = NULL,
  dfSub = NULL,
  sigma = NULL,
  dfObs = NULL,
  nSub = NULL,
  nStud = NULL
)
```
rxParam(obj, ...)

Arguments

obj  RxODE family of objects
...

other arguments including scaling factors for each compartment. This includes
S# = numeric will scale a compartment # by a dividing the compartment amount
by the scale factor, like NONMEM.

constants is a boolean indicating if constants should be included in the list of parameters.
Currently RxODE parses constants into variables in case you wish to change
them without recompiling the RxODE model.

params a numeric named vector with values for every parameter in the ODE system; the
names must correspond to the parameter identifiers used in the ODE specification;

inits a vector of initial values of the state variables (e.g., amounts in each compart-
ment), and the order in this vector must be the same as the state variables (e.g.,
PK/PD compartments);

iCov A data frame of individual non-time varying covariates to combine with the
params to form a parameter data.frame.

keep Columns to keep from either the input dataset or the iCov dataset. With the iCov
dataset, the column is kept once per line. For the input dataset, if any records
are added to the data LOCF (Last Observation Carried forward) imputation is
performed.

thetaMat Named theta matrix.

omega Estimate of Covariance matrix. When omega is a list, assume it is a block matrix
and convert it to a full matrix for simulations.

dfSub Degrees of freedom to sample the between subject variability matrix from the
inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.

sigma Named sigma covariance or Cholesky decomposition of a covariance matrix.
The names of the columns indicate parameters that are simulated. These are
simulated for every observation in the solved system.

dfObs Degrees of freedom to sample the unexplained variability matrix from the in-
verse Wishart distribution (scaled) or scaled inverse chi squared distribution.

nSub Number between subject variabilities (ETAs) simulated for every realization of
the parameters.

nStud Number virtual studies to characterize uncertainty in estimated parameters.

Value

When extracting the parameters from an RxODE model, a character vector listing the parameters in
the model.

Author(s)

Matthew L. Fidler
**rxPermissive**

*Permissive or Strict RxODE syntax options*

**Description**

This sets the RxODE syntax to be permissive or strict.

**Usage**

```r
rxPermissive(
  expr,
  silent = .isTestthat(),
  respect = FALSE,
  cran = FALSE,
  on.validate = FALSE
)
```

```r
rxStrict(
  expr,
  silent = .isTestthat(),
  respect = FALSE,
  cran = FALSE,
  on.validate = FALSE
)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><code>expr</code></td>
<td>Expression to evaluate in the permissive/strict environment. If unspecified, set the options for the current environment.</td>
</tr>
<tr>
<td><code>silent</code></td>
<td>when true, also silence the syntax errors and interactive output (useful in testing).</td>
</tr>
<tr>
<td><code>respect</code></td>
<td>when TRUE, respect any options that are specified. This is called at startup, but really should not be called elsewhere, otherwise the options are not changed.</td>
</tr>
<tr>
<td><code>cran</code></td>
<td>When specified and true, run on CRAN. Otherwise it is skipped on CRAN.</td>
</tr>
<tr>
<td><code>on.validate</code></td>
<td>When TRUE run only when validating.</td>
</tr>
</tbody>
</table>

**Author(s)**

Matthew L. Fidler
**Description**

`rxProgress` sets up the progress bar

**Usage**

```r
rxProgress(num, core = 0L)
```

```r
rxTick()
```

```r
rxProgressStop(clear = TRUE)
```

```r
rxProgressAbort(error = "Aborted calculation")
```

**Arguments**

- `num`  
  Tot number of operations to track
- `core`  
  Number of cores to show. If below 1, don’t show number of cores
- `clear`  
  Boolean telling if you should clear the progress bar after completion (as if it wasn’t displayed). By default this is TRUE
- `error`  
  With `rxProgressAbort` this is the error that is displayed

**Details**

- `rxTick` is a progress bar tick
- `rxProgressStop` stop progress bar
- `rxProgressAbort` shows an abort if `rxProgressStop` wasn’t called.

**Value**

All return NULL invisibly.

**Author(s)**

Matthew L. Fidler

**Examples**

```r
f <- function()
  on.exit({rxProgressAbort()});
  rxProgress(100)
  for (i in 1:100) {
    rxTick()
    Sys.sleep(1 / 100)
  }
```
## Not run:
f();
## End(Not run)

---

### rxRateDur

**Creates a rxRateDur object**

**Description**

This is primarily to display information about rate

**Usage**

```r
rxRateDur(x)
```

```r
as.rxRateDur(x)
```

**Arguments**

- `x`: rxRateDur data

---

### rxSetIni0

**Set Initial conditions to time zero instead of the first observed/dosed time**

**Description**

Set Initial conditions to time zero instead of the first observed/dosed time

**Usage**

```r
rxSetIni0(ini0 = TRUE)
```

**Arguments**

- `ini0`: When TRUE (default), set initial conditions to time zero. Otherwise the initial conditions are the first observed time.
**rxSetProd**  
*Choose the type of product to use in RxODE. These are used in the RxODE prod blocks*

**Description**

Choose the type of product to use in RxODE. These are used in the RxODE prod blocks.

**Usage**

```
rxSetProd(type = c("long double", "double", "logify"))
```

**Arguments**

- **type**: Product to use for `prod()` in RxODE blocks.  
  - *long double* converts to long double, performs the multiplication and then converts back.  
  - *double* uses the standard double scale for multiplication.

**Value**

nothing

**Author(s)**

Matthew L. Fidler

---

**rxSetProgressBar**  
*Set timing for progress bar*

**Description**

Set timing for progress bar.

**Usage**

```
rxSetProgressBar(seconds = 1)
```

**Arguments**

- **seconds**: This sets the number of seconds that need to elapse before drawing the next segment of the progress bar. When this is zero or below this turns off the progress bar.

**Author(s)**

Matthew Fidler
rxSetSum

Choose the type of sums to use for RxODE.

Description
Choose the types of sums to use in RxODE. These are used in the RxODE sum blocks and the rxSum function.

Usage
rxSetSum(type = c("pairwise", "fsum", "kahan", "neumaier", "c"))

Arguments

- **type**: Sum type to use for rxSum and sum() in RxODE code blocks.
  - pairwise uses the pairwise sum (fast, default)
  - fsum uses Python’s fsum function (most accurate)
  - kahan uses kahan correction
  - neumaier uses Neumaier correction
  - c uses no correction, but default/native summing

Value
nothing

Author(s)
Matthew L. Fidler

rxShiny
Use Shiny to help develop an RxODE model

Description
Use Shiny to help develop an RxODE model

Usage
rxShiny(
  object,
  params = c(),
  events = NULL,
  inits = c(),
  ...,
  data = data.frame())
## S3 method for class 'rxSolve'
rxShiny(
  object,
  params = NULL,
  events = NULL,
  inits = c(),
  ..., 
  data = data.frame()
)

## Default S3 method:
rxShiny(
  object = NULL,
  params = c(),
  events = NULL,
  inits = c(),
  ..., 
  data = data.frame()
)

### Arguments

- **object**
  A RxODE family of objects. If not supplied a 2-compartment indirect effect model is used. If it is supplied, use the model associated with the RxODE object for the model exploration.

- **params**
  Initial parameters for model

- **events**
  Event information (currently ignored)

- **inits**
  Initial estimates for model

- **...**
  Other arguments passed to rxShiny. Currently doesn’t do anything.

- **data**
  Any data that you would like to plot. If the data has a time variable as well as a compartment or calculated variable that matches the RxODE model, the data will be added to the plot of a specific compartment or calculated variable.

### Value

Nothing; Starts a shiny server

### Author(s)

Zufar Mulyukov and Matthew L. Fidler
Simulate Parameters from a Theta/Omega specification

**Description**

Simulate Parameters from a Theta/Omega specification

**Usage**

```r
rxSimThetaOmega(
  params = NULL,
  omega = NULL,
  omegaDf = NULL,
  omegaLower = as.numeric(c(R_NegInf)),
  omegaUpper = as.numeric(c(R_PosInf)),
  omegaIsChol = FALSE,
  nSub = 1L,
  thetaMat = NULL,
  thetaLower = as.numeric(c(R_NegInf)),
  thetaUpper = as.numeric(c(R_PosInf)),
  thetaDf = NULL,
  thetaIsChol = FALSE,
  nStud = 1L,
  sigma = NULL,
  sigmaLower = as.numeric(c(R_NegInf)),
  sigmaUpper = as.numeric(c(R_PosInf)),
  sigmaDf = NULL,
  sigmaIsChol = FALSE,
  nCoresRV = 1L,
  nObs = 1L,
  dfSub = 0,
  dfObs = 0,
  simSubjects = TRUE
)
```

**Arguments**

- **params**: Named Vector of RxODE model parameters
- **omega**: Named omega matrix.
- **omegaDf**: The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a normal distribution instead of a t-distribution.
- **omegaLower**: Lower bounds for simulated ETAs (by default -Inf)
- **omegaUpper**: Upper bounds for simulated ETAs (by default Inf)
- **omegaIsChol**: Indicates if the omega supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.
<table>
<thead>
<tr>
<th>Parameter</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>nSub</td>
<td>Number between subject variabilities (ETAs) simulated for every realization of the parameters.</td>
</tr>
<tr>
<td>thetaMat</td>
<td>Named theta matrix.</td>
</tr>
<tr>
<td>thetaLower</td>
<td>Lower bounds for simulated population parameter variability (by default -Inf)</td>
</tr>
<tr>
<td>thetaUpper</td>
<td>Upper bounds for simulated population unexplained variability (by default Inf)</td>
</tr>
<tr>
<td>thetaDf</td>
<td>The degrees of freedom of a t-distribution for simulation. By default this is NULL which is equivalent to Inf degrees, or to simulate from a normal distribution instead of a t-distribution.</td>
</tr>
<tr>
<td>thetaIsChol</td>
<td>Indicates if the theta supplied is a Cholesky decomposed matrix instead of the traditional symmetric matrix.</td>
</tr>
<tr>
<td>nStud</td>
<td>Number virtual studies to characterize uncertainty in estimated parameters.</td>
</tr>
<tr>
<td>sigma</td>
<td>Matrix for residual variation. Adds a &quot;NA&quot; value for each of the individual parameters, residuals are updated after solve is completed.</td>
</tr>
<tr>
<td>sigmaLower</td>
<td>Lower bounds for simulated unexplained variability (by default -Inf)</td>
</tr>
<tr>
<td>sigmaUpper</td>
<td>Upper bounds for simulated unexplained variability (by default Inf)</td>
</tr>
<tr>
<td>sigmaDf</td>
<td>Degrees of freedom of the sigma t-distribution. By default it is equivalent to Inf, or a normal distribution.</td>
</tr>
<tr>
<td>sigmaIsChol</td>
<td>Boolean indicating if the sigma is in the Cholesky decomposition instead of a symmetric covariance</td>
</tr>
<tr>
<td>nCoresRV</td>
<td>Number of cores used for the simulation of the sigma variables. By default this is 1. This uses the package rmvn and rmvt. To reproduce the results you need to run on the same platform with the same number of cores. This is the reason this is set to be one, regardless of what the number of cores are used in threaded ODE solving.</td>
</tr>
<tr>
<td>nObs</td>
<td>Number of observations to simulate (with sigma matrix)</td>
</tr>
<tr>
<td>dfSub</td>
<td>Degrees of freedom to sample the between subject variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.</td>
</tr>
<tr>
<td>dfObs</td>
<td>Degrees of freedom to sample the unexplained variability matrix from the inverse Wishart distribution (scaled) or scaled inverse chi squared distribution.</td>
</tr>
<tr>
<td>simSubjects</td>
<td>boolean indicated RxODE should simulate subjects in studies (TRUE, default) or studies (FALSE)</td>
</tr>
</tbody>
</table>

**Author(s)**

Matthew L. Fidler
\textbf{rxStack} \hspace{1cm} \textit{Stack a solved object for things like ggplot}

\textbf{Description}

Stack a solved object for things like ggplot

\textbf{Usage}

\texttt{rxStack(Data, vars = NULL)}

\textbf{Arguments}

- \texttt{Data} is a RxODE object to be stacked.
- \texttt{vars} Variables to include in stacked data; By default this is all the variables when \texttt{vars} is \texttt{NULL}.

\textbf{Value}

Stacked data with \texttt{value} and \texttt{trt}, where value is the values and \texttt{trt} is the state and \texttt{lhs} variables.

\textbf{Author(s)}

Matthew Fidler

\textbf{rxState} \hspace{1cm} \textit{State variables}

\textbf{Description}

This returns the model’s compartments or states.

\textbf{Usage}

\texttt{rxState(obj = NULL, state = NULL)}

\textbf{Arguments}

- \texttt{obj} RxODE family of objects
- \texttt{state} is a string indicating the state or compartment that you would like to lookup.

\textbf{Value}

If state is missing, return a character vector of all the states.
If state is a string, return the compartment number of the named state.
Author(s)

Matthew L. Fidler

See Also

RxODE

rxSumProdModel

Recast model in terms of sum/prod

Description

Recast model in terms of sum/prod

Usage

rxSumProdModel(model, expand = FALSE, sum = TRUE, prod = TRUE)

Arguments

model RxODE model
expand Boolean indicating if the expression is expanded.
sum Use sum(...)
prod Use prod(...)

Value

model string with prod(.) and sum(.) for all these operations.

Author(s)

Matthew L. Fidler
Description

Get Omega^-1 and derivatives

Usage

```r
rxSymInvChol(
  invObjOrMatrix,
  theta = NULL,
  type = "cholOmegaInv",
  thetaNumber = 0L
)
```

Arguments

- `invObjOrMatrix` Object for inverse-type calculations. If this is a matrix, setup the object for inversion by `rxSymInvCholCreate` with the default arguments and return a reactive s3 object. Otherwise, use the inversion object to calculate the requested derivative/inverse.
- `theta` Thetas to be used for calculation. If missing (NULL), a special s3 class is created and returned to access Omega^-1 objects as needed and cache them based on the theta that is used.
- `type` The type of object. Currently the following types are supported:
  - `cholOmegaInv` gives the Cholesky decomposition of the Omega Inverse matrix.
  - `omegaInv` gives the Omega Inverse matrix.
  - `d(omegaInv)` gives the d(Omega^-1) with respect to the theta parameter specified in `thetaNumber`.
  - `d(D)` gives the d(diagonal(Omega^-1)) with respect to the theta parameter specified in the `thetaNumber` parameter.
- `thetaNumber` For types `d(omegaInv)` and `d(D)`, the theta number that the derivative is taken against. This must be positive from 1 to the number of thetas defining the Omega matrix.

Value

Matrix based on parameters or environment with all the matrixes calculated in variables omega, omegaInv, dOmega, domegaInv.

Author(s)

Matthew L. Fidler
**rxSymPyFix**  
*Fix SymPy expressions to be R parsable expressions*

**Description**  
Fix SymPy expressions to be R parsable expressions

**Usage**  
```
rxSymPyFix(var)
```

**Arguments**  
- **var**  
sympy expression

**Value**  
R valid expression

**Author(s)**  
Matthew L. Fidler

---

**rxSymPySensitivity**  
*Calculate the sensitivity equations for a model*

**Description**  
This expands the model to calculate sensitivities. This requires rSymPy.

**Usage**  
```
rxSymPySensitivity(
    model,
    calcSens,
    calcJac = FALSE,
    keepState = NULL,
    collapseModel = FALSE
)
```
**Arguments**

- **model**: RxODE family of objects
- **calcSens**: Either a logical or list of sensitivity parameters to calculate. When TRUE, calculate the sensitivities of all the known parameters. When FALSE raise an error.
- **calcJac**: A boolean that determines if the Jacobian should be calculated.
- **keepState**: State parameters to keep the sensitivities for.
- **collapseModel**: A boolean to collapse the model that each expression only depends on the unspecified parameters (instead on LHS quantities).

**Value**

Model syntax that includes the sensitivity parameters.

**Author(s)**

Matthew L. Fidler

---

**rxSymPyVersion**

Return the version of SymPy that is running

**Description**

Return the version of SymPy that is running

**Usage**

```r
rxSymPyVersion(numeric = TRUE)
```

**Arguments**

- **numeric**: boolean that specifies if the major and minor release should be a number.

**Value**

Version of sympy that is running.

**Author(s)**

Matthew L. Fidler
rxSyncOptions  

**Description**  
Accessing RxODE options via `getOption` slows down solving. This allows the options to be synced with variables.

**Usage**  

```r
rxSyncOptions()
```

**Author(s)**  
Matthew L. Fidler

---

rxTempDir  

**Description**  
Get the RxODE temporary directory

**Usage**  

```r
rxTempDir()
```

**Value**  
RxODE temporary directory.

---

rxTrans  

**Description**  
This function translates the model to C code, if needed
Usage

```r
rxTrans(
    model,
    extraC = NULL,
    modelPrefix = "",
    md5 = "",
    modName = NULL,
    modVars = FALSE,
    ...
)
```

## Default S3 method:

```r
rxTrans(
    model,
    extraC = NULL,
    modelPrefix = "",
    md5 = "",
    modName = NULL,
    modVars = FALSE,
    ...
)
```

## S3 method for class 'character'

```r
rxTrans(
    model,
    extraC = NULL,
    modelPrefix = "",
    md5 = "",
    modName = NULL,
    modVars = FALSE,
    ...
)
```

Arguments

- **model**: This is the ODE model specification. It can be:
  - a string containing the set of ordinary differential equations (ODE) and other expressions defining the changes in the dynamic system.
  - a file name where the ODE system equation is contained
  - An ODE expression enclosed in `{}`
  (see also the filename argument). For details, see the sections “Details” and “RxODE Syntax” below.

- **extraC**: Extra c code to include in the model. This can be useful to specify functions in the model. These C functions should usually take double precision arguments, and return double precision values.

- **modelPrefix**: Prefix of the model functions that will be compiled to make sure that multiple RxODE objects can coexist in the same R session.
md5  Is the md5 of the model before parsing, and is used to embed the md5 into DLL, and then provide for functions like `rxModelVars`.

modName  a string to be used as the model name. This string is used for naming various aspects of the computations, including generating C symbol names, dynamic libraries, etc. Therefore, it is necessary that `modName` consists of simple ASCII alphanumeric characters starting with a letter.

modVars  returns the model variables instead of the named vector of translated properties.

...  Ignored parameters.

Value

a named vector of translated model properties including what type of jacobian is specified, the C function prefixes, as well as the C functions names to be called through the compiled model.

Author(s)

Matthew L. Fidler

See Also

`RxODE, rxCompile`

---

**rxUnloadAll**

Unload all RxODE Dlls that are not locked for solving.

**Description**

Unload all RxODE Dlls that are not locked for solving.

**Usage**

```r
rxUnloadAll()
```

---

**rxUse**

Use model object in your package

**Description**

Use model object in your package

**Usage**

```r
rxUse(obj, overwrite = TRUE, compress = "bzip2", internal = FALSE)
```
**rxUseRadixSort**

**Arguments**

- **obj**  
  Model to save.

- **overwrite**  
  By default, `use_data()` will not overwrite existing files. If you really want to do so, set this to **`TRUE`**.

- **compress**  
  Choose the type of compression used by `save()`. Should be one of "gzip", "bzip2", or "xz".

- **internal**  
  If this is run internally. By default this is **`FALSE`**.

---

**rxUseRadixSort**  
*Use Radix Sort when possible*

**Description**

By default RxODE uses radix sort when possible.

**Usage**

```r
rxUseRadixSort(useRadix = TRUE)
```

**Arguments**

- **useRadix**  
  Use order with method = radix when appropriate. Otherwise use timsort.

**Author(s)**

Matthew Fidler

---

**rxValidate**  
*Validate RxODE*

**Description**

This allows easy validation/qualification of nlmixr by running the testing suite on your system.

**Usage**

```r
rxValidate(full = TRUE)
```

```r
rxTest(full = TRUE)
```

**Arguments**

- **full**  
  Should a full validation be performed? (By default **`TRUE`**)

**Author(s)**

Matthew L. Fidler
**rxWinPythonSetup**

*Setup Python and SymPy for windows*

**Description**

Setup Python and SymPy for windows

**Usage**

```r
rxWinPythonSetup()
```

**Author(s)**

Matthew L. Fidler

---

**rxWinSetup**

*Setup Windows components for RxODE*

**Description**

Setup Windows components for RxODE

**Usage**

```r
rxWinSetup(rm.rtools = TRUE, rm.python = TRUE)
```

**Arguments**

- `rm.rtools` Remove the Rtools from the current path specs.
- `rm.python` Remove Python from the current path specs.

**Author(s)**

Matthew L. Fidler
**summary.RxODE**

Print expanded information about the RxODE object.

**Description**

This prints the expanded information about the RxODE object.

**Usage**

```r
## S3 method for class 'RxODE'
summary(object, ...)
```

**Arguments**

- `object` : RxODE object
- `...` : Ignored parameters

**Author(s)**

Matthew L. Fidler

---

**tibble**

*type_sum function for units*

**Description**

*type_sum function for units*

**Usage**

```r
## S3 method for class 'units'
type_sum(x, ...)

## S3 method for class 'type_sum_units'
format_type_sum(x, width, ...)

## S3 method for class 'units'
piller_shaft(x, ...)

## S3 method for class 'mixed_units'
type_sum(x, ...)

## S3 method for class 'mixed_units'
piller_shaft(x, ...)

## S3 method for class 'units'
scale_type(x)
```
Arguments

x see type_sum

... see type_sum

width see type_sum
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