Package ‘TopDom’

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Title An Efficient and Deterministic Method for Identifying Topological Domains in Genomes

Depends R (>= 3.3.0)

Imports matrixStats, grid, ggplot2, reshape2, tibble

Suggests diffobj (>= 0.1.11)

Description The 'TopDom' method identifies topological domains in genomes from Hi-C sequence data (Shin et al., 2016 <doi:10.1093/nar/gkv1505>). The authors published an implementation of their method as an R script (two different versions; also available in this package). This package originates from those original 'TopDom' R scripts and provides help pages adopted from the original 'TopDom' PDF documentation. It also provides a small number of bug fixes to the original code.

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LazyLoad TRUE

URL https://github.com/HenrikBengtsson/TopDom

BugReports https://github.com/HenrikBengtsson/TopDom/issues

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#### `countsPerRegion`

*Calculates Counts per Region in a TopDomData Object*

**Description**

Calculates Counts per Region in a TopDomData Object

**Usage**

`countsPerRegion(data, regions)`

**Arguments**

- **data** A TopDomData object.
- **regions** TopDom regions (a data.frame), e.g. domains.

**Value**

A numeric vector of length `nrow(regions)`.

**Author(s)**

Henrik Bengtsson.
ggCountHeatmap

Produce a Count Heatmap

Description
Produce a Count Heatmap

Usage
ggCountHeatmap(data, transform, colors, ...)

Arguments
- data: A TopDomData object.
- transform: A function applied to the counts prior to generating heatmap colors.
- colors: A named list to control the color scale.
- ...: Not used.

Value
A ggplot2::ggplot object.

Author(s)
Henrik Bengtsson.

See Also
See TopDom for an example.

ggDomain

Add a Topological Domain to a Count Heatmap

Description
Add a Topological Domain to a Count Heatmap

Usage
ggDomain(td, dx = NULL, delta = 0.04, vline = 0, size = 2, color = "#666666")
Arguments

- **td**: A single-row data frame.
- **dx, delta, vline**: Absolute distance to heatmap. If `dx = NULL` (default), then `dx = delta * w + vline` where `w` is the width of the domain.
- **size, color**: The thickness and color of the domain line.

Value

A `ggplot2::geom_segment` object to be added to the count heatmap.

---

**ggDomainLabel**

Add a Topological Domain Label to a Count Heatmap

Description

Add a Topological Domain Label to a Count Heatmap

Usage

```r
ggDomainLabel(
  td,
  fmt = "%s: %.2f - %.2f Mbp",
  rot = 45,
  dx = 0,
  vjust = 2.5,
  cex = 1.5
)
```

Arguments

- **td**: A single-row data frame.
- **fmt**: The `base::sprintf`-format string taking (chromosome, start, stop) as (string, numeric, numeric) input.
- **rot**: The amount of rotation in [0,360] of label.
- **dx, vjust**: The vertical adjustment of the label (relative to rotation)
- **cex**: The scale factor of the label.

Value

A `ggplot2::ggproto` object to be added to the count heatmap.
Easy Access to the Original TopDom 0.0.1 and 0.0.2 Implementations

**Description**

Easy Access to the Original TopDom 0.0.1 and 0.0.2 Implementations

**Usage**

```r
legacy(version = c("0.0.1", "0.0.2"))
```

**Arguments**

- `version`  
  A version string.

**Value**

An environment containing the legacy TopDom API.

**Examples**

```r
TopDom::legacy("0.0.2")$TopDom  
TopDom::legacy("0.0.1")$Detect.Local.Extreme
```

---

Overlap Scores Between Two Sets of Topological Domains

**Description**

Calculates Overlap Scores Between Two Sets of Topological Domains

**Usage**

```r
overlapScores(a, reference, debug = getOption("TopDom.debug", FALSE))
```

**Arguments**

- `a, reference`  
  Topological domain (TD) set `A` and TD reference set `R` both in a format as returned by `TopDom()`.

- `debug`  
  If TRUE, debug output is produced.
Details

The overlap score, \( \text{overlap}(A', r_i) \), represents how well a consecutive subset \( A' \) of topological domains (TDs) in \( A \) overlap with topological domain \( r_i \) in reference set \( R \). For each reference TD \( r_i \), the best match \( A'_{\text{max}} \) is identified, that is, the \( A' \) subset that maximize \( \text{overlap}(A', r_i) \). For exact definitions, see Page 8 in Shin et al. (2016).

Note that the overlap score is an asymmetric score, which means that \( \text{overlapScores}(a, b) \neq \text{overlapScores}(b, a) \).

Value

Returns a named list of class \text{TopDomOverlapScores}, where the names correspond to the chromosomes in domain reference set \( R \). Each of these chromosome elements contains a data.frame with fields:

- \text{chromosome} - \( D_{R,c} \) character strings
- \text{best_score} - \( D_{R,c} \) numerics in \([0, 1]\)
- \text{best_length} - \( D_{R,c} \) positive integers
- \text{best_set} - list of \( D_{R,c} \) index vectors

where \( D_{R,c} \) is the number of TDs in reference set \( R \) on chromosome \( c \). If a TD in reference \( R \) is not a “domain”, then the corresponding \text{best_score} and \text{best_length} values are \text{NA_real_} and \text{NA_integer_}, respectively, while \text{best_set} is an empty list.

Warning - This might differ not be the correct implementation

The original TopDom scripts do not provide an implementation for calculating overlap scores. Instead, the implementation of TopDom::\text{overlapScores()} is based on the textual description of overlap scores provided in Shin et al. (2016). It is not known if this is the exact same algorithm and implementation as the authors of the TopDom article used.

Author(s)

Henrik Bengtsson - based on the description in Shin et al. (2016).

References

- Shin et al., TopDom: an efficient and deterministic method for identifying topological domains in genomes, Nucleic Acids Research, 44(7): e70, April 2016. doi: 10.1093/nar/gkv1505, PMCID: PMC4838359, PMID: 26704975

See Also

TopDom.
**Examples**

```r
library(tibble)
path <- system.file("exdata", package = "TopDom", mustWork = TRUE)

## Original count data (on a subset of the bins to speed up example)
chr <- "chr19"
pathname <- file.path(path, sprintf("nij.%s.gz", chr))
data <- readHiC(pathname, chr = chr, binSize = 40e3, bins = 1:500)
print(data)

## Find topological domains using TopDom method for two window sizes
tds_5 <- TopDom(data, window.size = 5L)
tds_6 <- TopDom(data, window.size = 6L)

## Overlap scores (in both directions)
overlap_56 <- overlapScores(tds_6, reference = tds_5)
print(overlap_56)
print(as_tibble(overlap_56))

overlap_65 <- overlapScores(tds_5, reference = tds_6)
print(overlap_65)
print(as_tibble(overlap_65))
```

---

**readHiC**

*Reads Hi-C Contact Data from File*

**Description**

Reads Hi-C Contact Data from File

**Usage**

```r
readHiC(
  file,
  chr = NULL,
  binSize = NULL,
  ...,
  debug =getOption("TopDom.debug", FALSE)
)
```

**Arguments**

- `file` The pathname of a normalize Hi-C contact matrix file stored as a whitespace-delimited file. See below for details. Also a gzip-compressed file can be used.
- `chr, binSize` If the file contains a count matrix without bin annotation, the latter is created from these parameters.
- `debug` If TRUE, debug output is produced.
- `...` Arguments passed to `utils::read.table()` as-is.
Value

A list with elements bins (an N-by-4 data.frame) and counts (N-by-N matrix).

Format of HiC contact-matrix file

The contact-matrix file should be a whitespace-delimited text file with neither row names nor column names. The content should be a N-by-(3+N) table where the first three columns correspond to chr (string), from.coord (integer position), and to.coord (integer position). These column defines the genomic location of the N Hi-C bins (in order). The last N columns should contain normalized contact counts (float) such that element (r,3+c) in this table corresponds to count (r,c) in the normalized contact matrix.

If an N-by-(4+N) table, then the first column is assumed to contain an id (integer), and everything else as above.

Example:

```
chr10 0 40000 0 0 0 0 ...  
chr10 40000 80000 0 0 0 0 ...  
chr10 80000 120000 0 0 0 0 ...  
chr10 120000 160000 0 0 0 0 ...  
...  
```

See Also

TopDom.

Examples

```
path <- system.file("exdata", package = "TopDom", mustWork = TRUE)

## Original count data
chr <- "chr19"
pathname <- file.path(path, sprintf("nij.%s.gz", chr))
data <- readHiC(pathname, chr = chr, binSize = 40e3)
print(data)
str(data)
```

---

subsetByRegion  Subset a TopDomData Object by Region

Description

Subset a TopDomData Object by Region

Usage

```
subsetByRegion(data, region, margin = 1/2)
```
TopDom

Arguments

data A TopDomData object.
region A TopDom domain (a data.frame).
margin An non-negative numeric specifying the additional margin extracted around the
domain. If margin < 1, then the size of the margin is relative to the size of the
domain.

Value

A TopDomData object.

Author(s)

Henrik Bengtsson.

TopDom (Identify Topological Domains from a Hi-C Contact Matrix)

Description

Identify Topological Domains from a Hi-C Contact Matrix

Usage

TopDom(
  data,
  window.size,
  outFile = NULL,
  statFilter = TRUE,
  ...
  debug = getOption("TopDom.debug", FALSE)
)

Arguments

data A TopDomData object, or the pathname to a normalized Hi-C contact matrix
file as read by readHiC(), that specify N bins.
window.size The number of bins to extend (as a non-negative integer). Recommended range
is in 5, ..., 20.
outFile (optional) The filename without extension of the three result files optionally
produced. See details below.
statFilter (logical) Specifies whether non-significant topological-domain boundaries should
be dropped or not.
... Additional arguments passed to readHiC().
dead If TRUE, debug output is produced.
Value

A named list of class TopDom with data.frame elements binSignal, domain, and bed.

- The binSignal data frame (N-by-7) holds mean contact frequency, local extreme, and p-value for every bin. The first four columns represent basic bin information given by matrix file, such as bin id (id), chromosome(chr), start coordinate (from.coord), and end coordinate (to.coord) for each bin. The last three columns (local.ext, mean.cf, and p-value) represent computed values by the TopDom algorithm. The columns are:
  - id: Bin ID
  - chr: Chromosome
  - from.coord: Start coordinate of bin
  - to.coord: End coordinate of bin
  - local.ext:
    * -1: Local minima.
    * -0.5: Gap region.
    * 0: General bin.
    * 1: Local maxima.
  - mean.cf: Average of contact frequencies between lower and upper regions for bin \( i = 1, 2, ..., N \).
  - p-value: Computed p-value by Wilcoxon rank sum test. See Shin et al. (2016) for more details.

- The domain data frame (D-by-7): Every bin is categorized by basic building block, such as gap, domain, or boundary. Each row indicates a basic building block. The first five columns include the basic information about the block, ‘tag’ column indicates the class of the building block.
  - id: Identifier of block
  - chr: Chromosome
  - from.id: Start bin index of the block
  - from.coord: Start coordinate of the block
  - to.id: End bin index of the block
  - to.coord: End coordinate of the block
  - tag: Categorized name of the block. Three possible blocks exists:
    * gap
    * domain
    * boundary
  - size: size of the block

- The bed data frame (D-by-4) is a representation of the domain data frame in the BED file format. It has four columns:
  - chrom: The name of the chromosome.
  - chromStart: The starting position of the feature in the chromosome. The first base in a chromosome is numbered 0.
  - chromEnd: The ending position of the feature in the chromosome. The chromEnd base is not included in the feature. For example, the first 100 bases of a chromosome are defined as chromStart=0, chromEnd=100, and span the bases numbered 0-99.
- name: Defines the name of the BED line. This label is displayed to the left of the BED line in the UCSC Genome Browser window when the track is open to full display mode or directly to the left of the item in pack mode.

If argument outFile is non-NULL, then the three elements (binSignal, domain, and bed) returned are also written to tab-delimited files with file names `<outFile>.binSignal`, `<outFile>.domain`, and `<outFile>.bed`, respectively. None of the files have row names, and all but the BED file have column names.

**Windows size**

The window.size parameter is by design the only tuning parameter in the TopDom method and affects the amount of smoothing applied when calculating the TopDom bin signals. The binning window extends symmetrically downstream and upstream from the bin such that the bin signal is the average window.size^2 contact frequencies. For details, see Equation (1) and Figure 1 in Shin et al. (2016). Typically, the number of identified TDs decreases while their average lengths increase as this window-size parameter increases (Figure 2). The default is window.size = 5 (bins), which is motivated as: “Considering the previously reported minimum TD size (approx. 200 kb) (Dixon et al., 2012) and our bin size of 40 kb, w[indow.size] = 5 is a reasonable setting” (Shin et al., 2016).

**Author(s)**


**References**


**Examples**

```r
path <- system.file("exdata", package = "TopDom", mustWork = TRUE)

## Original count data (on a subset of the bins to speed up example)
chr <- "chr19"
```
TopDom-data

Data for the TopDom Package
Description

The `exdata/` folder of this package provides an example data set used in examples. The data are also used to validate the TopDom implementation toward the original TopDom scripts.

Origin

The data herein contain a tiny subset of the HiC and TopDom data used in the TopDom study (Shin et al., 2016). More precisely, it contains:

1. A TopDom file ‘mESC_5w_chr19.nij.HindIII.comb.40kb.domain’, which is part of the ‘mESC_5w_domain.zip’ file (5,504 bytes; md5 ffb19996f681a4d35d5c9944f2c44343) from the Supplementary Materials of Shin et al. (2016). These data were downloaded from the TopDom website (http://zhoulab.usc.edu/TopDom/ - now defunct).

2. A normalized HiC-count matrix file ‘nij.chr19.gz’, where the non-compressed version is part of the ‘mESC_norm.tar.gz’ file (1,305,763,679 bytes; md5 2e79d0f57463b5b7c4bf86b187086d3c) originally downloaded from the UCSD Ren Lab. It is a tab-delimited file containing a 3250-by-3250 numeric matrix non-negative decimal values. The underlying HiC sequence data is available from GSE35156 on GEO and was published part of Dixon, et al. (2012).

References


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