Package ‘TreeSearch’

May 11, 2022

**Title**  Phylogenetic Analysis with Morphological Data

**Version**  1.1.2

**License**  GPL (>= 3)

**Copyright**  Incorporates C/C++ code from Morphy Phylogenetic Library by Martin Brazeau [https://github.com/mbrazeau/MorphyLib](https://github.com/mbrazeau/MorphyLib) (GPL3)

**Description**  Reconstruct phylogenetic trees from discrete data. Inapplicable character states are handled using the algorithm of Brazeau, Guillerme and Smith (2019) [doi:10.1093/sysbio/syy083](https://doi.org/10.1093/sysbio/syy083) with the “Morphy” library, under equal or implied step weights. Contains a “shiny” user interface for interactive tree search and exploration of results, including character visualization, rogue taxon detection, tree space mapping, and cluster consensus trees (Smith 2022a, b) [doi:10.1093/sysbio/syab099](https://doi.org/10.1093/sysbio/syab099), [doi:10.1093/sysbio/syab100](https://doi.org/10.1093/sysbio/syab100). Profile Parsimony (Faith and Trueman, 2001) [doi:10.1080/10635150118627](https://doi.org/10.1080/10635150118627), Successive Approximations (Farris, 1969) [doi:10.2307/2412182](https://doi.org/10.2307/2412182) and custom optimality criteria are implemented.

**URL**  [https://ms609.github.io/TreeSearch/](https://ms609.github.io/TreeSearch/), [https://github.com/ms609/TreeSearch/](https://github.com/ms609/TreeSearch/)

**BugReports**  [https://github.com/ms609/TreeSearch/issues/](https://github.com/ms609/TreeSearch/issues/)

**Depends**  R (>= 3.5.0)

**Imports**  ape (>= 5.6), cli (>= 3.0), cluster, fastmatch (>= 1.1.3), future, promises, protoclust, Rcpp, Rdpack (>= 0.7), Rogue (>= 2.0.0), shiny (>= 1.6.0), shinyjs, stats, TreeDist (>= 2.3.0), TreeTools (>= 1.7.0),

**Suggests**  knitr, phangorn (>= 2.2.1), Quartet, rmarkdown, shinytest, spelling, testthat, vdiffrr (>= 1.0.0),

**Config/Needs/check**  callr, pkgbuild, rcmdcheck,

**Config/Needs/coverage**  covr

**Config/Needs/memcheck**  devtools

**Config/Needs/metadata**  codemeta

**Config/Needs/revdeps**  revdepcheck
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**AdditionTree**

**Description**

Generates a starting tree by adding each taxon in turn to the most parsimonious location.

**Usage**

```r
AdditionTree(dataset, concavity = Inf, constraint, sequence)
```

**Arguments**

- **dataset**: A phylogenetic data matrix of `phangorn` class `phyDat`, whose names correspond to the labels of any accompanying tree.
- **concavity**: Numeric specifying concavity constant for implied step weighting. The most appropriate value will depend on the dataset, but values around 10–15 often perform well (Goloboff et al. 2018; Smith 2019). The character string "profile" employs an approximation of profile parsimony (Faith and Trueman 2001). Set as `Inf` for equal step weights, which underperforms step weighting approaches (Goloboff et al. 2008; Goloboff et al. 2018; Goloboff and Arias 2019; Smith 2019).
- **constraint**: An object of class `phyDat`; returned trees will be perfectly compatible with each character in constraint. See `ImposeConstraint()` and vignette for further examples.
- **sequence**: Character or numeric vector listing sequence in which to add taxa. Randomized if not provided.

**Value**

`AdditionTree()` returns a tree of class `phylo`, rooted on `sequence[1]`. 
Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

See Also

Impose a constraint: `TreeTools::ImposeConstraint()`
Neighbour-joining trees: `TreeTools::NJTree();TreeTools::ConstrainedNJ()`
Other tree generation functions: `RandomMorphyTree()`

Examples

```r
data('Lobo', package = 'TreeTools')
AdditionTree(Lobo.phy, concavity = 10)
```

AllSPR

All SPR trees

Description

All SPR trees

Usage

```r
AllSPR(parent, child, nEdge, notDuplicateRoot, edgeToBreak)
```

Arguments

- **parent**: Integer vector corresponding to the first column of the edge matrix of a tree of class `phylo`, i.e. `tree$edge[, 1]`.
- **child**: Integer vector corresponding to the second column of the edge matrix of a tree of class `phylo`, i.e. `tree$edge[, 2]`.
- **nEdge**: Integer specifying the number of edges of a tree of class `phylo`, i.e. `dim(tree$edge)[1]`.
- **notDuplicateRoot**: Logical vector of length `nEdge`, specifying for each edge whether it is the second edge leading to the root (in which case its breaking will be equivalent to breaking the other root edge... except insofar as it moves the position of the root.)
- **edgeToBreak**: (optional) integer specifying the index of an edge to bisect/prune, generated randomly if not specified. Alternatively, set to `-1` to return a complete list of all trees one step from the input tree.

Value

`AllSPR()` returns a list of edge matrices for all trees one SPR rearrangement from the starting tree

Author(s)

Martin R. Smith
Description

Calculate the number of trees in which Fitch parsimony will reconstruct \( m \) steps, where \( a \) leaves are labelled with one state, and \( b \) leaves are labelled with a second state.

Usage

\[
\text{Carter1}(m, a, b)
\]
\[
\log_2 \text{Carter1}(m, a, b)
\]
\[
\log \text{Carter1}(m, a, b)
\]

Arguments

\( m \) Number of steps.
\( a, b \) Number of leaves labelled 0 and 1.

Details

Implementation of theorem 1 from Carter et al. (1990)

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

References


See Also

Other profile parsimony functions: `PrepareDataProfile()`, `StepInformation()`, `WithOneExtraStep()`, `profiles`
Examples

# The character '0 0 1 1 1'
Carter1(1, 3, 3) # Exactly one step
Carter1(2, 3, 3) # Two steps (one extra step)

# Number of trees that the character can map onto with exactly _m_ steps
# if non-parsimonious reconstructions are permitted:
cumsum(sapply(1:3, Carter1, 3, 3))

# Three steps allow the character to map onto any of the 105 six-leaf trees.

CharacterLength | Character length

Description

Homoplasy length of each character in a dataset on a specified tree.

Usage

CharacterLength(tree, dataset, compress = FALSE)

FitchSteps(tree, dataset)

FastCharacterLength(tree, dataset)

Arguments

- **tree**: A tree of class `phylo`.
- **dataset**: A phylogenetic data matrix of `phangorn` class `phyDat`, whose names correspond to the labels of any accompanying tree.
- **compress**: Logical specifying whether to retain the compression of a `phyDat` object or to return a vector specifying to each individual character, decompressed using the dataset’s index attribute.

Value

CharacterLength() returns a vector listing the contribution of each character to tree score, according to the algorithm of Brazeau et al. (2019).

Functions

- **FastCharacterLength**: Do not perform checks. Use with care: may cause erroneous results or software crash if variables are in the incorrect format.
ClusterStrings

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

References


See Also

Other tree scoring: IWScore(), MinimumLength(), MorphyTreeLength()

Examples

data('inapplicable.datasets')
dataset <- inapplicable.phyData[[12]]
tree <- TreeTools::NJTree(dataset)
CharacterLength(tree, dataset)
CharacterLength(tree, dataset, compress = TRUE)

ClusterStrings  Cluster similar strings

Description

Calculate string similarity using the Levenshtein distance and return clusters of similar strings.

Usage

ClusterStrings(x, maxCluster = 12)

Arguments

x  Character vector.
maxCluster  Integer specifying maximum number of clusters to consider.

Value

NameClusters() returns an integer assigning each element of x to a cluster, with an attribute med specifying the median string in each cluster, and silhouette reporting the silhouette coefficient of the optimal clustering. Coefficients < 0.5 indicate weak structure, and no clusters are returned. If the number of unique elements of x is less than maxCluster, all occurrences of each entry are assigned to an individual cluster.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)
ConcordantInformation

Examples
ClusterStrings(c(paste0('FirstCluster ', 1:5),
  paste0('SecondCluster.', 8:12),
  paste0('AnotherCluster_', letters[1:6])))

ConcordantInformation  Evaluate the concordance of information between a tree and a dataset

Description
Details the amount of information in a phylogenetic dataset that is consistent with a specified phylogenetic tree, and the signal:noise ratio of the character matrix implied if the tree is true.

Usage
ConcordantInformation(tree, dataset)
Evaluate(tree, dataset)
ConcordantInfo(tree, dataset)

Arguments
  tree  A tree of class phylo.
  dataset  A phylogenetic data matrix of phangorn class phyDat, whose names correspond to the labels of any accompanying tree.

Details
Presently restricted to datasets whose characters contain a maximum of two parsimony-informative states.

Value
ConcordantInformation() returns a named vector with elements:
  • informationContent: cladistic information content of dataset
  • signal, noise: amount of cladistic information that represents phylogenetic signal and noise, according to tree
  • signalToNoise: the implied signal:noise ratio of dataset
  • treeInformation: the cladistic information content of a bifurcating tree on dataset; this is the minimum amount of information necessary to resolve a bifurcating tree, assuming no duplicate information or noise
  • matrixToTree: the ratio of the cladistic information content of the matrix to the cladistic information content of the tree, a measure of the redundancy of the matrix
  • ignored: information content of characters whose signal and noise could not be calculated (too many states) and so are not included in the totals above.
congreveLamsdellMatrices

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

Examples

data(congreveLamsdellMatrices)
myMatrix <- congreveLamsdellMatrices[[10]]
ConcordantInformation(TreeTools::NJTree(myMatrix), myMatrix)

congreveLamsdellMatrices

100 simulated data matrices

Description

Contains the 100 simulated matrices generated by (Congreve and Lamsdell 2016) using a heterogeneous Markov-k model, generated from the referenceTree topology, with all branches sharing an equal length.

Usage

congreveLamsdellMatrices

Format

A list with 100 entries, each comprising a phyDat object of 55 characters for 22 taxa

Source

doi:10.5061/dryad.7dq0j

References


Examples

data('referenceTree')
data('congreveLamsdellMatrices')
TreeLength(referenceTree, congreveLamsdellMatrices[[17]], 'profile')
Consistency

Description

Consistency() calculates the so-called consistency and retention 'indices' for each character in a dataset, given a bifurcating tree. Although there is not a straightforward interpretation of these indices, they are sometimes taken as an indicator of the fit of a character to a tree. Values correlate with the number of species sampled and the distribution of taxa between character states, so are not strictly comparable between characters in which these factors differ.

Usage

Consistency(dataset, tree, compress = FALSE)

Arguments

dataset A phylogenetic data matrix of phangorn class phyDat, whose names correspond to the labels of any accompanying tree.
tree A tree of class phylo.compress Logical specifying whether to retain the compression of a phyDat object or to return a vector specifying to each individual character, decompressed using the dataset’s index attribute.

Details

#TODO: Retention index not yet implemented.

Value

Consistency() returns a named vector specifying the consistency index (ci), retention index (ri), and rescaled consistency index (rc).

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

Examples

data(inapplicable.datasets)
dataset <- inapplicable.phyData[[4]]
Consistency(dataset, TreeTools::NJTree(dataset))
cSPR

Description

cSPR() expects a tree rooted on a single tip.

Usage

cSPR(tree, whichMove = NULL)

Arguments

tree A tree of class phylo.
whichMove Integer specifying which SPR move index to perform.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

Examples

tree <- TreeTools::BalancedTree(8)

# Tree must be rooted on leaf
tree <- TreeTools::RootTree(tree, 1)

# Random rearrangement
cSPR(tree)

# Specific rearrangement
cSPR(tree, 9)

GapHandler

Description

Gaps represented by the inapplicable token can be treated as 'missing data', i.e. as equivalent to the ambiguous token ?; as an extra state, equivalent to other states such as 0 or 1; or as 'inapplicable data' using the algorithm of Brazeau, Guillerme and Smith (2019).

Usage

GapHandler(morphyObj)
Arguments

morphyObj Object of class morphy, perhaps created with PhyDat2Morphy().

Value

GapHandler() returns a character string stating how gaps are handled by morphyObj.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

See Also

Other Morphy API functions: MorphyErrorCheck(), MorphyWeights(), PhyDat2Morphy(), SingleCharMorphy(), UnloadMorphy(), is.morphyPtr(), mpl_apply_tipdata(), mpl_attach_rawdata(), mpl_attach_symbols(), mpl_delete_Morphy(), mpl_delete_rawdata(), mpl_first_down_recon(), mpl_first_up_recon(), mpl_get_charac_weight(), mpl_get_gaphandl(), mpl_get_num_charac(), mpl_get_num_internal_nodes(), mpl_get_num_taxa(), mpl_get_symbols(), mpl_init_Morphy(), mpl_new_Morphy(), mpl_second_down_recon(), mpl_second_up_recon(), mpl_set_charac_weight(), mpl_set_num_internal_nodes(), mpl_set_parsim_t(), mpl_translate_error(), mpl_update_lower_root(), mpl_update_tip(), summary.morphyPtr()

Examples

```r
morphyObj <- SingleCharMorphy('0-0', 'Extra')
GapHandler(morphyObj)
morphyObj <- UnloadMorphy(morphyObj)
```

Description

These are the datasets used to evaluate the behaviour of the inapplicable algorithm in Brazeau et al. (2019). The name of each item corresponds to the datasets listed below. Datasets are sorted into two subsets, each sorted alphabetically; the first subset comprise simpler datasets with faster processing times. inapplicable.datasets provide the data in the matrix format generated by read.nexus.data(); inapplicable.phyData are in phyDat format. inapplicable.trees lists for each dataset a sample of up to 50 trees obtained by tree search under each inapplicable treatment, named accordingly. inapplicable.citations is a named character vector specifying the source of each dataset.

Usage

inapplicable.datasets

inapplicable.phyData
inapplicable.datasets

inapplicable.trees

inapplicable.citations

Format

An object of class list of length 30.
An object of class list of length 30.
An object of class list of length 31.
An object of class character of length 30.

Source

Subset one (faster processing):


Subset two (longer processing times):


References


Examples

data('inapplicable.datasets', package = 'TreeSearch')
names(inapplicable.datasets)
is.morphyPtr  Is an object a valid Morphy object?

Description

Is an object a valid Morphy object?

Usage

is.morphyPtr(morphyObj)

Arguments

morphyObj  Object of class morphy, perhaps created with PhyDat2Morphy().

Value

is.morphyPtr() returns TRUE if morphyObj is a valid morphy pointer, FALSE otherwise.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

See Also

Other Morphy API functions: GapHandler(), MorphyErrorCheck(), MorphyWeights(), PhyDat2Morphy(), SingleCharMorphy(), UnloadMorphy(), mpl_apply_tipdata(), mpl_attach_rawdata(), mpl_attach_symbols(), mpl_delete_Morphy(), mpl_delete_rawdata(), mpl_first_down_recon(), mpl_first_up_recon(), mpl_get_charac_weight(), mpl_get_gaphandl(), mpl_get_num_charac(), mpl_get_num_internal_nodes(), mpl_get_numtaxa(), mpl_get_symbols(), mpl_init_Morphy(), mpl_new_Morphy(), mpl_second_down_recon(), mpl_second_up_recon(), mpl_set_charac_weight(), mpl_set_num_internal_nodes(), mpl_set_parsim_t(), mpl_translate_error(), mpl_update_lower_root(), mpl_update_tip(), summary.morphyPtr()

IWScore  Calculate the parsimony score of a tree given a dataset

Description

TreeLength() uses the Morphy library (Brazeau et al. 2017) to calculate a parsimony score for a tree, handling inapplicable data according to the algorithm of Brazeau et al. (2019). Trees may be scored using equal weights, implied weights (Goloboff 1993), or profile parsimony (Faith and Trueman 2001).
IWScore

Usage

IWScore(tree, dataset, concavity = 10L, ...) 
TreeLength(tree, dataset, concavity = Inf)

## S3 method for class 'phylo'
TreeLength(tree, dataset, concavity = Inf)

## S3 method for class 'numeric'
TreeLength(tree, dataset, concavity = Inf)

## S3 method for class 'list'
TreeLength(tree, dataset, concavity = Inf)

## S3 method for class 'multiPhylo'
TreeLength(tree, dataset, concavity = Inf)

Fitch(tree, dataset)

Arguments

tree A tree of class phylo, a list thereof (optionally of class multiPhylo), or an integer – in which case tree random trees will be uniformly sampled.
dataset A phylogenetic data matrix of phangorn class phyDat, whose names correspond to the labels of any accompanying tree.
concavity Determines the degree to which extra steps beyond the first are penalized. Specify a numeric value to use implied weighting (Goloboff 1993): concavity specifies \( k \) in \( k/e + k \). A value of 10 is recommended; TNT sets a default of 3, but this is too low in some circumstances (Goloboff et al. 2018; Smith 2019). Better still explore the sensitivity of results under a range of concavity values, e.g. \( k = 2 ^ {1:7} \). Specify Inf to weight each additional step equally. Specify 'profile' to employ profile parsimony (Faith and Trueman 2001).
...

Value

TreeLength() returns a numeric vector containing the score for each tree in tree.

Author(s)

Martin R. Smith (using Morphy C library, by Martin Brazeau)

References


**See Also**
- Conduct tree search using `MaximizeParsimony()` (command line), `EasyTrees()` (graphical user interface), or `TreeSearch()` (custom optimality criteria).
- See score for each character: `CharacterLength()`

Other tree scoring: `CharacterLength()`, `MinimumLength()`, `MorphyTreeLength()`

**Examples**

```r
data("inapplicable.datasets")
tree <- TreeTools::BalancedTree(inapplicable.phyData[[1]])
TreeLength(tree, inapplicable.phyData[[1]])
TreeLength(tree, inapplicable.phyData[[1]], concavity = 10)
TreeLength(tree, inapplicable.phyData[[1]], concavity = "profile")
TreeLength(5, inapplicable.phyData[[1]])
```

---

**Jackknife**

**Jackknife resampling**

**Description**

Resample trees using Jackknife resampling, i.e. removing a subset of characters.

**Usage**

```r
Jackknife(
  tree,
  dataset,
  resampleFreq = 2/3,
  InitializeData = PhyDat2Morphy,
  CleanUpData = UnloadMorphy,
)```
TreeScorer = MorphyLength,
EdgeSwapper = TBRSwap,
jackIter = 5000L,
searchIter = 4000L,
searchHits = 42L,
verbosity = 1L,
...
)

Arguments

tree  A tree of class phylo.
dataset a dataset in the format required by TreeScorer().
resampleFreq Double between 0 and 1 stating proportion of characters to resample.
InitializeData Function that sets up data object to prepare for tree search. The function will be passed the dataset parameter. Its return value will be passed to TreeScorer() and CleanUpData().
CleanUpData Function to destroy data object on function exit. The function will be passed the value returned by InitializeData().
TreeScorer function to score a given tree. The function will be passed three parameters, corresponding to the parent and child entries of a tree's edge list, and a dataset.
EdgeSwapper a function that rearranges a parent and child vector, and returns a list with modified vectors; for example SPRSwap().
jackIter Integer specifying number of jackknife iterations to conduct.
searchIter Integer specifying maximum rearrangements to perform on each bootstrap or ratchet iteration. To override this value for a single swapper function, set e.g. attr(SwapperFunction, 'searchIter') <- 99
searchHits Integer specifying maximum times to hit best score before terminating a tree search within a ratchet iteration. To override this value for a single swapper function, set e.g. attr(SwapperFunction, 'searchHits') <- 99
verbosity Numeric specifying level of detail to display in console: larger numbers provide more verbose feedback to the user.
...

Details

The function assumes that InitializeData() will return a morphy object; if this doesn’t hold for you, post a GitHub issue or e-mail the maintainer.

Value

Jackknife() returns a list of trees recovered after jackknife iterations.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)
**See Also**

- `JackLabels()`: Label nodes of a tree with jackknife supports.

Other split support functions: `JackLabels()`, `MaximizeParsimony()`.

Other custom search functions: `EdgeListSearch()`, `MorphyBootstrap()`, `SuccessiveApproximations()`.

---

### JackLabels

**Label nodes with jackknife support values**

**Description**

Label nodes with jackknife support values

**Usage**

```r
JackLabels(
  tree,
  jackTrees,
  plot = TRUE,
  add = FALSE,
  adj = 0,
  col = NULL,
  frame = "none",
  pos = 2L,
  ...
)
```

**Arguments**

- `tree`: A tree of class `phylo`.
- `jackTrees`: A list or `multiPhylo` object containing trees generated by `Jackknife()`.
- `plot`: Logical specifying whether to plot results; if `FALSE`, returns blank labels for nodes near the root that do not correspond to a unique split.
- `add`: Logical specifying whether to add the labels to an existing plot.
- `adj`, `col`, `frame`, `pos`, ...
  Parameters to pass to `nodelabels()`.

**Value**

A named vector specifying the proportion of jackknife trees consistent with each node in `tree`, as plotted. If `plot = FALSE`, blank entries are included corresponding to nodes that do not require labelling; the return value is in the value required by `phylo$node.label`.

**Author(s)**

Martin R. Smith (martin.smith@durham.ac.uk)
MaximizeParsimony

See Also

Jackknife(): Generate trees by jackknife resampling
Other split support functions: Jackknife(), MaximizeParsimony(), SiteConcordance

Examples

library('TreeTools', quietly = TRUE) # for as.phylo

# jackTrees will usually be generated with Jackknife(), but for simplicity:
jackTrees <- as.phylo(1:100, 8)

tree <- as.phylo(0, 8)
JackLabels(tree, jackTrees)

tree$node.label <- JackLabels(tree, jackTrees, plot = FALSE)

MaximizeParsimony

Find most parsimonious trees

Description

Search for most parsimonious trees using the parsimony ratchet and TBR rearrangements, treating
inapplicable data as such using the algorithm of Brazeau et al. (2019).

Tree search will be conducted from a specified or automatically-generated starting tree in order to
find a tree with an optimal parsimony score, under implied or equal weights, treating inapplicable
characters as such in order to avoid the artefacts of the standard Fitch algorithm (see Maddison
1993; Brazeau et al. 2019). Tree length is calculated using the MorphyLib C library (Brazeau et al.
2017).

Usage

MaximizeParsimony(
  dataset,
  tree,
  ratchIter = 6L,
  tbrIter = 2L,
  startIter = 2L,
  finalIter = 1L,
  maxHits = 1.8 * NTip(dataset),
  maxTime = 60,
  quickHits = 1/3,
  concavity = Inf,
  tolerance = sqrt(.Machine$double.eps),
  constraint,
  verbosity = 3L
)
MaximizeParsimony

Resample(
    dataset,
    tree,
    method = "jack",
    proportion = 2/3,
    ratchIter = 1L,
    tbrIter = 8L,
    finalIter = 3L,
    maxHits = 12L,
    concavity = Inf,
    tolerance = sqrt(.Machine$double.eps),
    constraint,
    verbosity = 2L,
    ...
)

EasyTrees()

Arguments

dataset A phylogenetic data matrix of `phangorn` class `phyDat`, whose names correspond to the labels of any accompanying tree.
tree (optional) A bifurcating tree of class `phylo`, containing only the tips listed in dataset, from which the search should begin. If unspecified, an addition tree will be generated from dataset, respecting any supplied constraint. Edge lengths are not supported and will be deleted.
ratchIter Numeric specifying number of iterations of the parsimony ratchet (Nixon 1999) to conduct.
tbrIter Numeric specifying the maximum number of TBR break points to evaluate before concluding each search. The counter is reset to zero each time tree score improves. The counter is reset to zero each time tree score improves. One ‘iteration’ comprises breaking a single branch and evaluating all possible reconnections.
startIter Numeric: an initial round of tree search with `startIter` × `tbrIter` TBR break points is conducted in order to locate a local optimum before beginning ratchet searches.
finalIter Numeric: a final round of tree search will evaluate `finalIter` × `tbrIter` TBR break points, in order to sample the final optimal neighbourhood more intensely.
maxHits Numeric specifying the maximum times that an optimal parsimony score may be hit before concluding a ratchet iteration or final search concluded.
maxTime Numeric: after `maxTime` minutes, stop tree search at the next opportunity.
quickHits Numeric: iterations on subsampled datasets will retain `quickHits` × `maxHits` trees with the best score.
concavity Numeric specifying concavity constant for implied step weighting. The most appropriate value will depend on the dataset, but values around 10–15 often perform well (Goloboff et al. 2018; Smith 2019). The character string "profile" employs an approximation of profile parsimony (Faith and Trueman 2001). Set as Inf for equal step weights, which underperforms step weighting approaches (Goloboff et al. 2008; Goloboff et al. 2018; Goloboff and Arias 2019; Smith 2019).

tolerance Numeric specifying degree of suboptimality to tolerate before rejecting a tree. The default, sqrt(.Machine$double.eps), retains trees that may be equally parsimonious but for rounding errors. Setting to larger values will include trees suboptimal by up to tolerance in search results, which may improve the accuracy of the consensus tree (at the expense of resolution) (Smith 2019).

constraint An object of class phyDat; returned trees will be perfectly compatible with each character in constraint. See ImposeConstraint() and vignette for further examples.

verbosity Integer specifying level of messaging; higher values give more detailed commentary on search progress. Set to 0 to run silently.

method Unambiguous abbreviation of jackknife or bootstrap specifying how to resample characters. Note that jackknife is considered to give more meaningful results.

proportion Numeric between 0 and 1 specifying what proportion of characters to retain under jackknife resampling.

... Additional parameters to MaximizeParsimony().

Details

Tree search commences with ratchIter iterations of the parsimony ratchet (Nixon 1999), which bootstraps the input dataset in order to escape local optima. A final round of tree bisection and reconnection (TBR) is conducted to broaden the sampling of trees.

This function can be called using the R command line / terminal, or through the 'shiny' graphical user interface app (type EasyTrees() to launch).

For detailed documentation of the 'TreeSearch' package, including full instructions for loading phylogenetic data into R and initiating and configuring tree search, see the package documentation.

Value

MaximizeParsimony() returns a list of trees with class multiPhylo. This lists all trees found during each search step that are within tolerance of the optimal score, listed in the sequence that they were first visited, and named according to the step in which they were first found; it may contain more than maxHits elements. Note that the default search parameters may need to be increased in order for these trees to be the globally optimal trees; examine the messages printed during tree search to evaluate whether the optimal score has stabilized.

The return value has the attribute firstHit, a named integer vector listing the number of optimal trees visited for the first time in each stage of the tree search. Stages are named:

- seed: starting trees;
Maximize Parsimony

- **start**: Initial TBR search;
- **ratchN**: Ratchet iteration \( N \);
- **final**: Final TBR search. The first tree hit for the first time in ratchet iteration three is named \( \text{ratch3}_1 \).

\( \text{Resample()} \) returns a \text{multiPhylo} object containing a list of trees obtained by tree search using a resampled version of dataset.

### Resampling

Note that bootstrap support is a measure of the amount of data supporting a split, rather than the amount of confidence that should be afforded the grouping. "Bootstrap support of 100% is not enough, the tree must also be correct" (Phillips et al. 2004). See discussion in Egan (2006); Wagele et al. (2009); (Simmons and Freudenstein 2011); Kumar et al. (2012).

For a discussion of suitable search parameters in resampling estimates, see Muller (2005). The user should decide whether to start each resampling from the optimal tree (which may be quicker, but result in overestimated support values as searches get stuck in local optima close to the optimal tree) or a random tree (which may take longer as more rearrangements are necessary to find an optimal tree on each iteration).

For other ways to estimate clade concordance, see \( \text{SiteConcordance()} \).

### Author(s)

**Martin R. Smith** (martin.smith@durham.ac.uk)

### References


Goloboff PA, Torres A, Arias JS (2018). “Weighted parsimony outperforms other methods of


See Also

Tree search via graphical user interface: `EasyTrees()`

Other split support functions: `JackLabels()`, `Jackknife()`, `SiteConcordance`

Examples

```r
## Only run examples in interactive R sessions
if (interactive()) {
  # launch 'shiny' point-and-click interface
  EasyTrees()

  # Here too, use the "continue search" function to ensure that tree score
  # has stabilized and a global optimum has been found
}

# Load data for analysis in R
library('TreeTools')
data('congreveLamsdellMatrices', package = 'TreeSearch')
dataset <- congreveLamsdellMatrices[[42]]
```
MinimumLength

# A very quick run for demonstration purposes
trees <- MaximizeParsimony(dataset, ratchIter = 0, startIter = 0,
                           tbrIter = 1, maxHits = 4, maxTime = 1/100,
                           concavity = 10, verbosity = 4)

names(trees)

# In actual use, be sure to check that the score has converged on a global
# optimum, conducting additional iterations and runs as necessary.

if (interactive()) {
  # Jackknife resampling
  nReplicates <- 10
  jackTrees <- replicate(nReplicates,
                         c(Resample(dataset, trees, ratchIter = 0, tbrIter = 2, startIter = 1,
                                      maxHits = 5, maxTime = 1 / 10,
                                      concavity = 10, verbosity = 0))
  )

  # In a serious analysis, more replicates would be conducted, and each
  # search would undergo more iterations.
  # Now we must decide what to do with the multiple optimal trees from
  # each replicate.

  # Treat each tree equally
  JackLabels(ape::consensus(trees), unlist(jackTrees, recursive = FALSE))

  # Take the strict consensus of all trees for each replicate
  JackLabels(ape::consensus(trees), lapply(jackTrees, ape::consensus))

  # Take a single tree from each replicate (the first; order's irrelevant)
  JackLabels(ape::consensus(trees), lapply(jackTrees, `[[`, 1))
}

# Tree search with a constraint
constraint <- MatrixToPhyDat(c(a = 1, b = 1, c = 0, d = 0, e = 0, f = 0))
characters <- MatrixToPhyDat(matrix(
  c(0, 1, 1, 0, 0,
   1, 1, 0, 0, 0), ncol = 2,
   dimnames = list(letters[1:6], NULL)))

MaximizeParsimony(characters, constraint = constraint, verbosity = 0)

---

MinimumLength

<table>
<thead>
<tr>
<th>Minimum length</th>
</tr>
</thead>
</table>

**Description**

The smallest length that a character can obtain on any tree.
Usage

MinimumLength(x, compress = FALSE)

## S3 method for class 'phyDat'
MinimumLength(x, compress = FALSE)

## S3 method for class 'numeric'
MinimumLength(x, compress = NA)

## S3 method for class 'character'
MinimumLength(x, compress = TRUE)

MinimumSteps(x)

Arguments

x
An object of class phyDat; or a string to be coerced to a phyDat object via TreeTools::StringToPhyDat(); or an integer vector listing the tokens that may be present at each tip along a single character, with each token represented as a binary digit; e.g. a value of 11 ( = 2^0 + 2^1 + 2^3) means that the tip may have tokens 0, 1 or 3.

Inapplicable tokens should be denoted with the integer 0 (not 2^0).

compress
Logical specifying whether to retain the compression of a phyDat object or to return a vector specifying to each individual character, decompressed using the dataset’s index attribute.

Value

MinimumLength() returns a vector of integers specifying the minimum number of steps that each character must contain.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

See Also

Other tree scoring: CharacterLength(), IWScore(), MorphyTreeLength()

Examples

data('inapplicable.datasets')
myPhyDat <- inapplicable.phyData[[4]]

# load your own data with
# my.PhyDat <- as.phyDat(read.nexus.data('filepath'))
# or Windows users can select a file interactively using:
# my.PhyDat <- as.phyDat(read.nexus.data(choose.files()))
class(myPhyDat) # phyDat object

# Minimum length of each character in turn
MinimumLength(myPhyDat)

# Collapse duplicate characters, per phyDat compression
MinimumLength(myPhyDat, compress = TRUE)

# Calculate length of a single character from its textual representation
MinimumLength('\{-1\}{-2\}{-3}\{2\}{2\}{3\}')

---

**MorphyBootstrap**  
**Parsimony Ratchet**

**Description**

Ratchet() uses the parsimony ratchet (Nixon 1999) to search for a more parsimonious tree using custom optimality criteria.

**Usage**

```r
MorphyBootstrap(
    edgeList,
    morphyObj,
    EdgeSwapper = NNISwap,
    maxIter,
    maxHits,
    verbosity = 1L,
    stopAtPeak = FALSE,
    stopAtPlateau = 0L,
...
)
```

```r
Ratchet(
    tree,
    dataset,
    InitializeData = PhyDat2Morphy,
    CleanUpData = UnloadMorphy,
    TreeScorer = MorphyLength,
    Bootstrapper = MorphyBootstrap,
    swappers = list(TBRSwap, SPRSwap, NNISwap),
    BootstrapSwapper = if (is.list(swappers)) swappers[[length(swappers)]] else swappers,
    returnAll = FALSE,
    stopAtScore = NULL,
    stopAtPeak = FALSE,
    stopAtPlateau = 0L,
    ratchIter = 100,
    ratchHits = 10,
)```
searchIter = 4000,
searchHits = 42,
bootstrapIter = searchIter,
bootstrapHits = searchHits,
verbosity = 1L,
suboptimal = sqrt(.Machine$double.eps),
...
)

MultiRatchet(
  tree,
  dataset,
  ratchHits = 10,
  searchIter = 500,
  searchHits = 20,
  verbosity = 0L,
  swappers = list(RootedNNISwap),
  nSearch = 10,
  stopAtScore = NULL,
  ...
)

RatchetConsensus(
  tree,
  dataset,
  ratchHits = 10,
  searchIter = 500,
  searchHits = 20,
  verbosity = 0L,
  swappers = list(RootedNNISwap),
  nSearch = 10,
  stopAtScore = NULL,
  ...
)

Arguments

edgeList a list containing the following: - vector of integers corresponding to the parent of each edge in turn - vector of integers corresponding to the child of each edge in turn - (optionally) score of the tree - (optionally, if score provided) number of times this score has been hit

morphyObj Object of class morphy, perhaps created with PhyDat2Morphy().

EdgeSwapper a function that rearranges a parent and child vector, and returns a list with modified vectors; for example SPRSwap().

maxIter Numeric specifying maximum number of iterations to perform in tree search.

maxHits Numeric specifying maximum number of hits to accomplish in tree search.

verbosity Numeric specifying level of detail to display in console: larger numbers provide more verbose feedback to the user.
stopAtPeak Logical specifying whether to terminate search once a subsequent iteration recovers a sub-optimal score. Will be overridden if a passed function has an attribute stopAtPeak set by attr(FunctionName, 'stopAtPeak') <- TRUE.

stopAtPlateau Integer. If > 0, tree search will terminate if the score has not improved after stopAtPlateau iterations. Will be overridden if a passed function has an attribute stopAtPlateau set by attr(FunctionName, 'stopAtPlateau') <- TRUE.

... further arguments to pass to TreeScorer(), e.g. dataset = .

tree A tree of class phylo.

dataset a dataset in the format required by TreeScorer().

InitializeData Function that sets up data object to prepare for tree search. The function will be passed the dataset parameter. Its return value will be passed to TreeScorer() and CleanUpData().

CleanUpData Function to destroy data object on function exit. The function will be passed the value returned by InitializeData().

TreeScorer function to score a given tree. The function will be passed three parameters, corresponding to the parent and child entries of a tree’s edge list, and a dataset.

Bootstrapper Function to perform bootstrapped rearrangements of tree. First arguments will be an edgelist and a dataset, initialized using InitializeData(). Should return a rearranged edgelist.

swappers A list of functions to use to conduct edge rearrangement during tree search. Provide functions like NNI_Swap to shuffle root position, or RootedTBRSwap if the position of the root should be retained. You may wish to use extreme swappers (such as TBR) early in the list, and a more subtle rearranger (such as NNI) later in the list to make incremental tinkerings once an almost-optimal tree has been found.

BootstrapSwapper Function such as RootedNNI_Swap to use to rearrange trees within Bootstrapper().

returnAll Set to TRUE to report all MPTs encountered during the search, perhaps to analyse consensus.

stopAtScore stop search as soon as this score is hit or beaten.

ratchIter Stop when this many ratchet iterations have been performed.

ratchHits Stop when this many ratchet iterations have found the same best score.

searchIter Integer specifying maximum rearrangements to perform on each bootstrap or ratchet iteration. To override this value for a single swapper function, set e.g. attr(SwapperFunction, 'searchIter') <- 99

searchHits Integer specifying maximum times to hit best score before terminating a tree search within a ratchet iteration. To override this value for a single swapper function, set e.g. attr(SwapperFunction, 'searchHits') <- 99

bootstrapIter Integer specifying maximum rearrangements to perform on each bootstrap iteration (default: searchIter).

bootstrapHits Integer specifying maximum times to hit best score on each bootstrap iteration (default: searchHits).
MorphyBootstrap

suboptimal retain trees that are suboptimal by this score. Defaults to a small value that will counter rounding errors.
nSearch Number of Ratchet searches to conduct (for RatchetConsensus())

Details

For usage pointers, see the vignette.

Value

MorphyBootstrap() returns a tree that is optimal under a random sampling of the original characters.
Ratchet() returns a tree modified by parsimony ratchet iterations.
MultiRatchet() returns a list of optimal trees produced by nSearch ratchet searches, from which a consensus tree can be generated using ape::consensus() or TreeTools::ConsensusWithout().

Functions

- RatchetConsensus: deprecated alias for MultiRatchet()

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

References


See Also

- Adapted from pratchet() in the phangorn package.

Other custom search functions: EdgeListSearch(), Jackknife(), SuccessiveApproximations()

Examples

data('Lobo', package = 'TreeTools')
njtree <- TreeTools::NJTree(Lobo.phy)
# Increase value of ratchIter and searchHits to do a proper search
quickResult <- Ratchet(njtree, Lobo.phy, ratchIter = 2, searchHits = 3)

# Plot result (legibly)
oldPar <- par(mar = rep(0, 4), cex = 0.75)
plot(quickResult)
par(oldPar)
MorphyWeights

Set and get the character weightings associated with a Morphy object.

Description

MorphyWeights() details the approximate and exact weights associated with characters in a Morphy object; SetMorphyWeights() edits them.

Usage

MorphyWeights(morphyObj)

SetMorphyWeights(weight, morphyObj, checkInput = TRUE)

Arguments

morphyObj Object of class morphy, perhaps created with PhyDat2Morphy().
weight A vector listing the new weights to be applied to each character
checkInput Whether to sanity-check input data before applying. Defaults to TRUE to protect
the user from crashes.

Value

MorphyWeights() returns a data frame with two named rows and one column per character pattern: row 1, approx, is a list of integers specifying the approximate (integral) weights used by MorphyLib; row 2, exact, is a list of numerics specifying the exact weights specified by the user.

SetMorphyWeights() returns the Morphy error code generated when applying weight.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

See Also

Other Morphy API functions: GapHandler(), MorphyErrorCheck(), PhyDat2Morphy(), SingleCharMorphy(), UnloadMorphy(), is.morphyPtr(), mpl_apply_tipdata(), mpl_attach_rawdata(), mpl_attach_symbols(), mpl_delete_Morphy(), mpl_delete_rawdata(), mpl_first_down_recon(), mpl_first_up_recon(), mpl_get_charac_weight(), mpl_get_gaphandl(), mpl_get_num_charac(), mpl_get_num_internal_nodes(), mpl_get_num_taxa(), mpl_get_symbols(), mpl_init_Morphy(), mpl_new_Morphy(), mpl_second_down_recon(), mpl_second_up_recon(), mpl_set_charac_weight(), mpl_set_num_internal_nodes(), mpl_set_parsim_t(), mpl_translate_error(), mpl_update_lower_root(), mpl_update_tip(), summary.morphyPtr()
Examples

```r
tokens <- matrix(c(0, 0, 0, 1, 1, 2,
                   0, 0, 0, 0, 0, 0), byrow = TRUE, nrow = 2L,
                   dimnames = list(letters[1:2], NULL))
pd <- TreeTools::MatrixToPhyDat(tokens)
morphObj <- PhyDat2Morphy(pd)
MorphyWeights(morphObj)
if (SetMorphyWeights(c(1, 1.5, 2/3), morphObj) != 0L) message("Errored")
MorphyWeights(morphObj)
morphObj <- UnloadMorphy(morphObj)
```

Description

NNI() performs a single iteration of the nearest-neighbour interchange algorithm; RootedNNI() retains the position of the root. These functions are based on equivalents in the phangorn package. cNNI() is an equivalent function coded in C, that runs much faster.

Usage

```r
NNI(tree, edgeToBreak = NULL)
cNNI(tree, edgeToBreak = NULL, whichSwitch = NULL)
NNISwap(parent, child, nTips = (length(parent)/2L) + 1L, edgeToBreak = NULL)
RootedNNI(tree, edgeToBreak = NULL)
RootedNNISwap(
  parent,
  child,
  nTips = (length(parent)/2L) + 1L,
  edgeToBreak = NULL
)
```

Arguments

- **tree**: A tree of class `phylo`.
- **edgeToBreak**: In (Rooted)NNI(), an optional integer specifying the index of an edge to bisect/prune, generated randomly if not specified. If -1, a complete list of all trees one step from the input tree will be returned. In cNNI(), an integer from zero to nEdge(tree) - nTip(tree) - 2, specifying which internal edge to break.
- **whichSwitch**: Integer from zero to one, specifying which way to re-build the broken internal edge.
parent  Integer vector corresponding to the first column of the edge matrix of a tree of class \texttt{phylo}, i.e. \texttt{tree$edge[, 1]}.

child  Integer vector corresponding to the second column of the edge matrix of a tree of class \texttt{phylo}, i.e. \texttt{tree$edge[, 2]}.

\texttt{nTips}  (optional) Number of tips.

Details

Branch lengths are not supported.

All nodes in a tree must be bifurcating; \texttt{ape::collapse.singles()} and \texttt{ape::multi2di()} may help.

Value

Returns a tree with class \texttt{phylo} (if \texttt{returnAll = FALSE}) or a set of trees, with class \texttt{multiPhylo} (if \texttt{returnAll = TRUE}).

\texttt{cNNI()} returns a tree of class \texttt{phylo}, rooted on the same leaf, on which the specified rearrangement has been conducted.

\texttt{NNISwap()} returns a list containing two elements, corresponding in turn to the rearranged parent and child parameters.

a list containing two elements, corresponding in turn to the rearranged parent and child parameters

Functions

- \texttt{NNISwap}: faster version that takes and returns parent and child parameters
- \texttt{RootedNNI}: Perform NNI rearrangement, retaining position of root
- \texttt{RootedNNISwap}: faster version that takes and returns parent and child parameters

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

References


See Also

Other tree rearrangement functions: \texttt{SPR()}, \texttt{TBR()}

Examples

tree <- TreeTools::BalancedTree(8)
# A random rearrangement
NNI(tree)
cNNI(tree)
PhyDat2Morphy

# All trees one NNI rearrangement away
NNI(tree, edgeToBreak = -1)

# Manual random sampling
cNNI(tree, sample.int(14 - 8 - 1, 1), sample.int(2, 1))

# A specified rearrangement
cNNI(tree, 0, 0)

# If a tree may not be binary, collapse nodes with
tree <- TreeTools::MakeTreeBinary(tree)

# If a tree may be improperly rooted, use
tree <- TreeTools::RootTree(tree, 1)

# If a tree may exhibit unusual node ordering, this can be addressed with
tree <- TreeTools::Preorder(tree)

PhyDat2Morphy

Initialize a Morphy object from a phyDat object

Description

Creates a new Morphy object with the same size and characters as the phyDat object. Once finished with the object, it should be destroyed using `UnloadMorphy()` to free the allocated memory.

Usage

PhyDat2Morphy(phy, gap = "inapplicable")

Arguments

phy
An object of `phangorn` class phyDat.

gap
An unambiguous abbreviation of inapplicable, ambiguous (= missing), or extra state, specifying how gaps will be handled.

Value

PhyDat2Morphy() returns a pointer to an initialized Morphy object.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)
See Also

Other Morphy API functions: GapHandler(), MorphyErrorCheck(), SingleCharMorphy(), UnloadMorphy(), is.morphyPtr(), mpl_apply_tipdata(), mpl_attach_rawdata(), mpl_attach_symbols(), mpl_delete_Morphy(), mpl_delete_rawdata(), mpl_first_down_recon(), mpl_first_up_recon(), mpl_get_charac_weight(), mpl_get_gaphandl(), mpl_get_num_charac(), mpl_get_num_internal_nodes(), mpl_get_numtaxa(), mpl_get_symbols(), mpl_init_Morphy(), mpl_new_Morphy(), mpl_second_down_recon(), mpl_second_up_recon(), mpl_set_charac_weight(), mpl_set_num_internal_nodes(), mpl_set_parsim_t(), mpl_translate_error(), mpl_update_lower_root(), mpl_update_tip(), summary.morphyPtr()

Examples

data('Lobo', package='TreeTools')
morphyObj <- PhyDat2Morphy(Lobo.phy)
# Set object to be destroyed at end of session or closure of function
# on.exit(morphyObj <- UnloadMorphy(morphyObj), add = TRUE)

# Do something with pointer
# ....

# Or, instead of on.exit, manually destroy morphy object and free memory:
morphyObj <- UnloadMorphy(morphyObj)

---

PlotCharacter 

Plot the distribution of a character on a tree

Description

Reconstructs the distribution of a character on a tree topology using the modified Fitch algorithm presented in Brazeau et al. (2019).

Usage

PlotCharacter(
  tree, 
  dataset, 
  char = 1L, 
  updateTips = FALSE, 
  plot = TRUE, 
  tokenCol = NULL, 
  ambigCol = "grey", 
  inappCol = "lightgrey", 
  ambigLty = "dotted", 
  inappLty = "dashed", 
  plainLty = par("lty"), 
  tipOffset = 1, 
  unitEdge = FALSE,
  ...
)
PlotCharacter

Arguments

- `tree`: A tree of class `phylo`.
- `dataset`: A phylogenetic data matrix of `phangorn` class `phyDat`, whose names correspond to the labels of any accompanying tree.
- `char`: Index of character to plot.
- `updateTips`: Logical; if `FALSE`, tips will be labelled with their original state in `dataset`.
- `plot`: Logical specifying whether to plot the output.
- `tokenCol`: Palette specifying colours to associate with each token in turn, in the sequence listed in `attr(dataset, 'levels')`.
- `ambiCol, ambigLty, inappCol, inappLty, plainLty`: Colours and line types to apply to ambiguous, inapplicable and applicable tokens. See the `lty` graphical parameter for details of line styles. Overrides `tokenCol`.
- `tipOffset`: Numeric: how much to offset tips from their labels.
- `unitEdge`: Logical: Should all edges be plotted with a unit length?
- ... Further arguments to pass to `plot.phylo()`.

Value

`PlotCharacter()` returns a matrix in which each row corresponds to a numbered tip or node of `tree`, and each column corresponds to a token; the tokens that might parsimoniously be present at each point on a tree are denoted with `TRUE`.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

References


Examples

```r
# Set up plotting area
oPar <- par(mar = rep(0, 4))

tree <- ape::read.tree(text = "((((((a, b), c), d), e), f), (g, (h, (i, (j, (k, l))))));")
## A character with inapplicable data
dataset <- TreeTools::StringToPhyDat("23--1??--032", tips = tree)
PlotCharacter(tree, dataset)

# Character from a real dataset
data("Lobo", package = "TreeTools")
dataset <- Lobo.phy
tree <- TreeTools::NJTree(dataset)
```
PrepareDataProfile

**Description**
Calculates profiles for each character in a dataset. Will also simplify characters, with a warning, where they are too complex for the present implementation of profile parsimony:

- inapplicable tokens will be replaced with the ambiguous token (i.e. \(-\rightarrow ?\));
- Ambiguous tokens will be treated as fully ambiguous (i.e. \(\{02\} \rightarrow ?\))
- Where more than two states are informative (i.e. unambiguously present in more than one taxon), states beyond the two most informative will be ignored.

**Usage**

```r
PrepareDataProfile(dataset)

PrepareDataIW(dataset)
```

**Arguments**

dataset dataset of class phyDat

**Value**
An object of class phyDat, with additional attributes. `PrepareDataProfile` adds the attributes:

- `info.amounts`: details the information represented by each character when subject to N additional steps.
- `informative`: logical specifying which characters contain any phylogenetic information.
- `bootstrap`: The character vector `c('info.amounts', 'split.sizes')`, indicating attributes to sample when bootstrapping the dataset (e.g. in Ratchet searches).

`PrepareDataIW` adds the attribute:

- `min.length`: The minimum number of steps that must be present in each transformation series.

**Functions**
- `PrepareDataIW`: Prepare data for implied weighting

**Author(s)**
Martin R. Smith; written with reference to `phangorn:::prepareDataFitch()`
profiles

See Also

Other profile parsimony functions: `Carter1()`, `StepInformation()`, `WithOneExtraStep()`, `profiles`

Examples

```r
data('congreveLamsdellMatrices')
dataset <- congreveLamsdellMatrices[[42]]
PrepareDataProfile(dataset)
```

---

### profiles

Empirically counted profiles for small trees

---

Description

The base 2 logarithm of the number of trees containing \( s \) steps, calculated by scoring a character on each \( n \)-leaf tree.

Usage

`profiles`

Format

A list with the structure `profiles[[number of leaves]][[number of tokens]][[tokens in smallest split]]`

The list entry returns a named numeric vector; each entry lists \( \log_2(\text{proportion of } n\text{-leaf trees with } s \text{ or fewer steps for this character}) \).

See Also

Other profile parsimony functions: `Carter1()`, `PrepareDataProfile()`, `StepInformation()`, `WithOneExtraStep()`

Examples

```r
data(profiles)
# Load profile for a character of the structure 0 0 1 1 1 1
profile3.5 <- profiles[[8]][[2]][[3]]

# Number of trees with _s_ or fewer steps on that character
TreeTools::NUnrooted(8) * 2 ^ profile3.5
```
RandomMorphyTree

Description
Random postorder tree

Usage
RandomMorphyTree(nTip)

Arguments
nTip    Integer specifying the number of tips to include in the tree (minimum 2).

Value
A list with three elements, each a vector of integers, respectively containing:
- The parent of each tip and node, in order
- The left child of each node
- The right child of each node.

See Also
Other tree generation functions: AdditionTree()

RandomTreeScore

Description
Parsimony score of random postorder tree

Usage
RandomTreeScore(morphyObj)

Arguments
morphyObj    Object of class morphy, perhaps created with PhyDat2Morphy().

Value
RandomTreeScore() returns the parsimony score of a random tree for the given Morphy object.
RearrangeEdges

Examples

tokens <- matrix(c(
  0, '-', '-', 1, 1, 2,
  0, 1, 0, 1, 2, 2,
  0, '-', '-', 0, 0, 0), byrow = TRUE, nrow = 3L, 
dimnames = list(letters[1:3], NULL))
pd <- TreeTools::MatrixToPhyDat(tokens)
morphyObj <- PhyDat2Morphy(pd)

RandomTreeScore(morphyObj)

morphyObj <- UnloadMorphy(morphyObj)

RearrangeEdges

Rearrange edges of a phylogenetic tree

Description

RearrangeEdges() performs the specified edge rearrangement on a matrix that corresponds to the edges of a phylogenetic tree, returning the score of the new tree. Will generally be called from within a tree search function.

Usage

RearrangeEdges(
  parent,
  child,
  dataset,
  TreeScorer = MorphyLength,
  EdgeSwapper,
  scoreToBeat = TreeScorer(parent, child, dataset, ...),
  iter = "?",
  hits = 0L,
  verbosity = 0L,
  ...
)

Arguments

parent
  Integer vector corresponding to the first column of the edge matrix of a tree of class phylo, i.e. tree$edge[, 1].

child
  Integer vector corresponding to the second column of the edge matrix of a tree of class phylo, i.e. tree$edge[, 2].

dataset
  Third argument to pass to TreeScorer.

TreeScorer
  function to score a given tree. The function will be passed three parameters, corresponding to the parent and child entries of a tree’s edge list, and a dataset.
EdgeSwapper a function that rearranges a parent and child vector, and returns a list with modified vectors; for example SPRSwap().

scoreToBeat Double giving score of input tree.

iter iteration number of calling function, for reporting to user only.

hits Integer giving number of times the input tree has already been hit.

verbosity Numeric specifying level of detail to display in console: larger numbers provide more verbose feedback to the user.

... further arguments to pass to TreeScorer(), e.g. dataset = .

**Details**

RearrangeTree() performs one tree rearrangement of a specified type, and returns the score of the tree (with the given dataset). It also reports the number of times that this score was hit in the current function call.

**Value**

This function returns a list with two to four elements, corresponding to a binary tree: - 1. Integer vector listing the parent node of each edge; - 2. Integer vector listing the child node of each edge; - 3. Score of the tree; - 4. Number of times that score has been hit.

**Author(s)**

Martin R. Smith

**Examples**

data('Lobo', package='TreeTools')
tree <- TreeTools::NJTree(Lobo.phy)
edge <- tree$edge
parent <- edge[, 1]
child <- edge[, 2]
dataset <- PhyDat2Morphy(Lobo.phy)
RearrangeEdges(parent, child, dataset, EdgeSwapper = RootedNNISwap)
# Remember to free memory:
dataset <- UnloadMorphy(dataset)

---

referenceTree  
**Tree topology for matrix simulation**

**Description**

The tree topology used to generate the matrices in congreveLamsdellMatrices

**Usage**

referenceTree
Format

A single phylogenetic tree saved as an object of class phylo

Source

Congreve & Lamsdell (2016); doi:10.1111/pala.12236

References


Examples

data(referenceTree)
plot(referenceTree)

---

SingleCharMorphy  

**Morphy object from single character**

Description

Morphy object from single character

Usage

SingleCharMorphy(char, gap = "inapp")

Arguments

- **char**: State of each character at each tip in turn, in a format that will be converted to a character string by `paste0(char, ';', collapse='')`.
- **gap**: An unambiguous abbreviation of inapplicable, ambiguous (= missing), or extra state, specifying how gaps will be handled.

Value

A pointer to an object of class morphyObj. Don’t forget to unload it when you’ve finished with it.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)
SiteConcordance

See Also

Score a tree: `MorphyTreeLength()`

Other Morphy API functions: `GapHandler()`, `MorphyErrorCheck()`, `MorphyWeights()`, `PhyDat2Morphy()`, `UnloadMorphy()`, `is.morphyPtr()`, `mpl_apply_tipdata()`, `mpl_attach_rawdata()`, `mpl_attach_symbols()`, `mpl_delete_Morphy()`, `mpl_delete_rawdata()`, `mpl_first_down_recon()`, `mpl_first_up_recon()`, `mpl_get_charac_weight()`, `mpl_get_gaphandl()`, `mpl_get_num_charac()`, `mpl_get_num_internal_nodes()`, `mpl_get_numtaxa()`, `mpl_get_symbols()`, `mpl_init_Morphy()`, `mpl_new_Morphy()`, `mpl_second_down_recon()`, `mpl_second_up_recon()`, `mpl_set_charac_weight()`, `mpl_set_num_internal_nodes()`, `mpl_set_parsim_t()`, `mpl_translate_error()`, `mpl_update_lower_root()`, `mpl_update_tip()`, `summary.morphyPtr()`

Examples

```r
morphyObj <- SingleCharMorphy('θ-θ', gap = 'Extra')
RandomTreeScore(morphyObj)
morphyObj <- UnloadMorphy(morphyObj)
```

SiteConcordance

*Calculate site concordance factor*

Description

The site concordance factor (Minh et al. 2020) is a measure of the strength of support that the dataset presents for a given split in a tree.

Usage

```r
QuartetConcordance(tree, dataset = NULL)
ClusteringConcordance(tree, dataset)
PhylogeneticConcordance(tree, dataset)
MutualClusteringConcordance(tree, dataset)
SharedPhylogeneticConcordance(tree, dataset)
```

Arguments

- `tree` A tree of class `phylo`.
- `dataset` A phylogenetic data matrix of `phangorn` class `phyDat`, whose names correspond to the labels of any accompanying tree.
Details

QuartetConcordance() is the proportion of quartets (sets of four leaves) that are decisive for a split which are also concordant with it. For example, a quartet with the characters 0 0 0 1 is not decisive, as all relationships between those leaves are equally parsimonious. But a quartet with characters 0 0 1 1 is decisive, and is concordant with any tree that groups the first two leaves together to the exclusion of the second.

NOTE: These functions are under development, and may be incompletely tested or change without notice. Complete documentation and discussion will follow soon.

Author(s)

Martin R. Smith (martin.smith@durham.ac.uk)

References


See Also

Other split support functions: JackLabels(), Jackknife(), MaximizeParsimony()

Examples

data('congreveLamsdellMatrices', package = 'TreeSearch')
dataset <- congreveLamsdellMatrices[[1]][, 1:20]
tree <- referenceTree
qc <- QuartetConcordance(tree, dataset)
cc <- ClusteringConcordance(tree, dataset)
pc <- PhylogeneticConcordance(tree, dataset)
spc <- SharedPhylogeneticConcordance(tree, dataset)
mcc <- MutualClusteringConcordance(tree, dataset)

oPar <- par(mar = rep(0, 4), cex = 0.8)
plot(tree)
TreeTools::LabelSplits(tree, signif(qc, 3))
TreeTools::LabelSplits(tree, signif(cc, 3))
TreeTools::LabelSplits(tree, signif(pc, 3))
par(oPar)

pairs(cbind(qc, cc, pc, spc, mcc))
Subtree pruning and rearrangement (SPR)

Description

Perform one SPR rearrangement on a tree

Usage

SPR(tree, edgeToBreak = NULL, mergeEdge = NULL)

SPRMoves(tree, edgeToBreak = integer(0))

## S3 method for class 'phylo'
SPRMoves(tree, edgeToBreak = integer(0))

## S3 method for class 'matrix'
SPRMoves(tree, edgeToBreak = integer(0))

SPRSwap(
  parent,
  child,
  nEdge = length(parent),
  nNode = nEdge/2L,
  edgeToBreak = NULL,
  mergeEdge = NULL
)

RootedSPR(tree, edgeToBreak = NULL, mergeEdge = NULL)

RootedSPRSwap(
  parent,
  child,
  nEdge = length(parent),
  nNode = nEdge/2L,
  edgeToBreak = NULL,
  mergeEdge = NULL
)

Arguments

tree A tree of class phylo.
edgeToBreak the index of an edge to bisect, generated randomly if not specified.
mergeEdge the index of an edge on which to merge the broken edge.
parent Integer vector corresponding to the first column of the edge matrix of a tree of class phylo, i.e. tree$edge[, 1].
child Integer vector corresponding to the second column of the edge matrix of a tree of class `phylo`, i.e. `tree$edge[, 2]`.
nEdge (optional) integer specifying the number of edges of a tree of class `phylo`, i.e. `dim(tree$edge)[1]`
nNode (optional) Number of nodes.

Details

Equivalent to `kSPR()` in the `phangorn` package, but faster. Note that rearrangements that only change the position of the root WILL be returned by `SPR`. If the position of the root is irrelevant (as in Fitch parsimony, for example) then this function will occasionally return a functionally equivalent topology. `RootIrrelevantSPR` will search tree space more efficiently in these cases. Branch lengths are not (yet) supported.

All nodes in a tree must be bifurcating; `ape::collapse.singles` and `ape::multi2di` may help.

Value

This function returns a tree in `phyDat` format that has undergone one `SPR` iteration.

`TBRMoves()` returns a list of all trees one `SPR` move away from `tree`, with edges and nodes in preorder, rooted on the first-labelled tip.

A list containing two elements, corresponding in turn to the rearranged parent and child parameters

A list containing two elements, corresponding in turn to the rearranged parent and child parameters

Functions

- `SPRSwap`: faster version that takes and returns parent and child parameters
- `RootedSPR`: Perform `SPR` rearrangement, retaining position of root
- `RootedSPRSwap`: faster version that takes and returns parent and child parameters

Author(s)

Martin R. Smith

References


See Also

- `RootedSPR()`: useful when the position of the root node should be retained.

Other tree rearrangement functions: `NNI()`, `TBR()`
Examples

```{r}
  tree <- ape::rtree(20, br=FALSE)
  SPR(tree)
```

**StepInformation**

*Information content of a character known to contain e steps*

**Description**

`StepInformation()` calculates the phylogenetic information content of a character `char` when `e` extra steps are present, for all possible values of `e`.

**Usage**

`StepInformation(char, ambiguousTokens = c("-", "?")))`

**Arguments**

- `char` Vector of tokens listing states for the character in question.
- `ambiguousTokens` Vector specifying which tokens, if any, correspond to the ambiguous token (`?`).

**Details**

Calculates the number of trees consistent with the character having `e` extra steps, where `e` ranges from its minimum possible value (i.e. number of different tokens minus one) to its maximum.

**Value**

`StepInformation()` returns a numeric vector detailing the amount of phylogenetic information (in bits) associated with the character when 0, 1, 2... extra steps are present. The vector is named with the total number of steps associated with each entry in the vector: for example, a character with three observed tokens must exhibit two steps, so the first entry (zero extra steps) is named 2 (two steps observed).

**Author(s)**

Martin R. Smith (martin.smith@durham.ac.uk)

**See Also**

Other profile parsimony functions: `Carter1()`, `PrepareDataProfile()`, `WithOneExtraStep()`, `profiles`
**Examples**

```r
character <- rep(c(0:3, '?', '-'), c(8, 5, 1, 1, 2, 2))
StepInformation(character)
```

---

**summary.morphyPtr**  
Details the attributes of a morphy object

---

**Description**

Details the attributes of a morphy object

**Usage**

```r
## S3 method for class 'morphyPtr'
summary(object, ...)
```

**Arguments**

- `object` A Morphy object
- `...` any other parameters...

**Value**

A list detailing the number of taxa, internal nodes, and characters and their weights.

**Author(s)**

Martin R. Smith (martin.smith@durham.ac.uk)

**See Also**

Other Morphy API functions: `GapHandler()`, `MorphyErrorCheck()`, `MorphyWeights()`, `PhyDat2Morphy()`, `SingleCharMorphy()`, `UnloadMorphy()`, `is.morphyPtr()`, `mpl_apply_tipdata()`, `mpl_attach_rawdata()`, `mpl_attach_symbols()`, `mpl_delete_Morphy()`, `mpl_delete_rawdata()`, `mpl_first_down_recon()`, `mpl_first_up_recon()`, `mpl_get_charac_weight()`, `mpl_get_gaphandl()`, `mpl_get_num_charac()`, `mpl_get_num_internal_nodes()`, `mpl_get_numtaxa()`, `mpl_get_symbols()`, `mpl_init_Morphy()`, `mpl_new_Morphy()`, `mpl_second_down_recon()`, `mpl_second_up_recon()`, `mpl_set_charac_weight()`, `mpl_set_num_internal_nodes()`, `mpl_set_parsim_t()`, `mpl_translate_error()`, `mpl_update_lower_root()`, `mpl_update_tip()`
Description

TBR performs a single random TBR iteration.

Usage

TBR(tree, edgeToBreak = NULL, mergeEdges = NULL)

TBRMoves(tree, edgeToBreak = integer(0))

## S3 method for class 'phylo'
TBRMoves(tree, edgeToBreak = integer(0))

## S3 method for class 'matrix'
TBRMoves(tree, edgeToBreak = integer(0))

TBRSwap(
  parent,
  child,
  nEdge = length(parent),
  edgeToBreak = NULL,
  mergeEdges = NULL
)

RootedTBR(tree, edgeToBreak = NULL, mergeEdges = NULL)

RootedTBRSwap(
  parent,
  child,
  nEdge = length(parent),
  edgeToBreak = NULL,
  mergeEdges = NULL
)

Arguments

tree

A bifurcating tree of class phylo, with all nodes resolved;

edgeToBreak

(optional) integer specifying the index of an edge to bisect/prune, generated randomly if not specified. Alternatively, set to -1 to return a complete list of all trees one step from the input tree.

mergeEdges

(optional) vector of length 1 or 2, listing edge(s) to be joined: In SPR, this is where the pruned subtree will be reconnected. In TBR, these edges will be reconnected (so must be on opposite sides of edgeToBreak); if only a single edge is specified, the second will be chosen at random.
parent  Integer vector corresponding to the first column of the edge matrix of a tree of class \texttt{phylo}, i.e. \texttt{tree$edge[, 1]}.

cchild  Integer vector corresponding to the second column of the edge matrix of a tree of class \texttt{phylo}, i.e. \texttt{tree$edge[, 2]}.

\texttt{nEdge}  (optional) Number of edges.

**Details**

Branch lengths are not (yet) supported.

All nodes in a tree must be bifurcating; \texttt{ape::collapse.singles} and \texttt{ape::multi2di} may help.

**Value**

This function returns a tree in \texttt{phyDat} format that has undergone one TBR iteration.

\texttt{TBRMoves()} returns a \texttt{multiPhylo} object listing all trees one TBR move away from \texttt{tree}, with edges and nodes in preorder, rooted on the first-labelled tip.

a list containing two elements, corresponding in turn to the rearranged parent and child parameters

**Functions**

- \texttt{TBRSwap}: faster version that takes and returns parent and child parameters
- \texttt{RootedTBR}: Perform TBR rearrangement, retaining position of root
- \texttt{RootedTBRswap}: faster version that takes and returns parent and child parameters

**Author(s)**

Martin R. Smith

**References**


**See Also**

\texttt{RootedTBR()}: useful when the position of the root node should be retained.

Other tree rearrangement functions: \texttt{NNI()}, \texttt{SPR()}

**Examples**

```r
{
  library('ape')
  tree <- rtree(20, br=NULL)
  TBR(tree)
}
```
UnloadMorphy

Destroy a Morphy object

Description

Destroys a previously-created Morphy object.

Usage

UnloadMorphy(morphyObj)

Arguments

morphyObj Object of class morphy, perhaps created with PhyDat2Morphy().

Details

Best practice is to call morphyObj <- UnloadMorphy(morphyObj) Failure to do so will cause a crash if UnloadMorphy() is called on an object that has already been destroyed

Value

Morphy error code, decipherable using mpl_translate_error

Author(s)

Martin R. Smith

See Also

Other Morphy API functions: GapHandler(), MorphyErrorCheck(), MorphyWeights(), PhyDat2Morphy(), SingleCharMorphy(), is.morphyPtr(), mpl_apply_tipdata(), mpl_attach_rawdata(), mpl_attach_symbols(), mpl_delete_Morphy(), mpl_delete_rawdata(), mpl_first_down_recon(), mpl_first_up_recon(), mpl_get_charac_weight(), mpl_get_gaphandl(), mpl_get_num_charac(), mpl_get_num_internal_nodes(), mpl_get_numtaxa(), mpl_get_symbols(), mpl_init_Morphy(), mpl_new_Morphy(), mpl_second_down_recon(), mpl_second_up_recon(), mpl_set_charac_weight(), mpl_set_num_internal_nodes(), mpl_set_parsim_t(), mpl_translate_error(), mpl_update_lower_root(), mpl_update_tip(), summary.morphyPtr()
**WithOneExtraStep**

| WithOneExtraStep | Number of trees with one extra step |

---

**Description**

Number of trees with one extra step

**Usage**

```
WithOneExtraStep(...)  
```

**Arguments**

```
...  Vector or series of integers specifying the number of leaves bearing each distinct non-ambiguous token.  
```

**See Also**

Other profile parsimony functions: `Carter1()`, `PrepareDataProfile()`, `StepInformation()`, `profiles`

**Examples**

```
WithOneExtraStep(1, 2, 3)  
```
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