Package ‘camtrapR’

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VignetteBuilder R.rsp

SystemRequirements ExifTool (https://exiftool.org/)

Description Management of and data extraction from camera trap data in wildlife studies. The package provides a workflow for storing and sorting camera trap photos (and videos), tabulates records of species and individuals, and creates detection/non-detection matrices for occupancy and spatial capture-recapture analyses with great flexibility. In addition, it can visualise species activity data and provides simple mapping functions with GIS export.

URL https://github.com/jniedballa/camtrapR,
https://jniedballa.github.io/camtrapR/,
https://groups.google.com/forum/#!forum/camtrapr

BugReports https://groups.google.com/forum/#!forum/camtrapr

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**R topics documented:**

- camtrapR-package .............................................. 3
- activityDensity .................................................. 6
- activityHistogram .................................................. 8
- activityOverlap .................................................... 10
- activityRadial ...................................................... 13
- addCopyrightTag ................................................... 15
- addToPath .......................................................... 17
- appendSpeciesNames ............................................... 18
- cameraOperation ................................................... 20
- camtraps ............................................................. 24
- camtrapsMultiSeason ............................................... 24
- checkSpeciesIdentification ......................................... 26
- checkSpeciesNames .................................................. 29
- commOccu-class ..................................................... 30
- communityModel .................................................... 31
- createSpeciesFolders ............................................... 38
- createStationFolders ............................................... 40
- detectionHistory ................................................... 41
- detectionMaps ....................................................... 46
- exifTagNames ......................................................... 49
- filterRecordTable ................................................... 51
- fit,commOccu-method ............................................... 54
- fixDateTimeOriginal ................................................ 55
- getSpeciesImages .................................................... 56
- imageRename ......................................................... 58
- OCRdataFields ....................................................... 61
- plot_coef,commOccu-method ........................................ 63
- plot_effects,commOccu-method ...................................... 65
- predict,commOccu-method ........................................... 66
- recordTable .......................................................... 67
- recordTableIndividual .............................................. 73
- recordTableIndividualSample ........................................ 78
- recordTableIndividualSampleMultiSeason ................................ 79
camtrapR-package

Overview of the functions in the camtrapR package

Description

This package provides a streamlined workflow for processing data generated in camera trap-based wildlife studies and prepares input for further analyses, particularly in occupancy and spatial capture-recapture frameworks. It suggests a simple data structure and provides functions for managing digital camera trap photographs (and videos), generating record tables, maps of species richness and species detections and species activity diagrams. It further helps prepare subsequent analyses by creating detection/non-detection matrices for occupancy analyses, e.g. in the *unmarked* or *ubms* packages, and *capthist* objects for spatial capture-recapture analyses in the *secr* package. In addition, basic survey statistics are computed. The functions build on one another in a logical sequence. The only manual input needed is species (and individual) identification, which is achieved by moving images into species directories or by tagging images in image management software. Besides, a table holding basic information about camera trap station IDs, locations and trapping periods must be created in spreadsheet software.

Details

Image metadata (such as date and time or user-assigned tags) are extracted from the images using Phil Harvey’s ExifTool (available from [https://exiftool.org/](https://exiftool.org/)) and the information is stored in a record table. An adjustable criterion for temporal independence of records can be applied. Maps of species presence and species richness can be generated. Several functions are available for plotting single- and two-species activity patterns. Information about the camera-specific trapping periods (and periods of malfunction) are summarized into a matrix about camera trap operability. These, together with the record table, are used to generate species detection histories for occupancy and spatial capture-recapture analyses. The user has considerable freedom in generating the detection histories; sampling occasion length, beginning date and and occasion start times are adjustable. In addition, trapping effort (i.e. active trap nights per station and occasion) can be computed for use as a covariate / offset on detection probability.

User support

The camtrapR Google group is an online support and help forum for camtrapR users. You can find it here: [https://groups.google.com/forum/#!forum/camtrapr](https://groups.google.com/forum/#!forum/camtrapr).
**Image organisation and management**

The functions in this section set up a directory structure for storing camera trap images and identifying species and individuals from images. They build on one another and can be run in sequential order as needed.

- `createStationFolders`: Create camera trap station directories for raw images
- `fixDateTimeOriginal`: Fix DateTimeOriginal Exif metadata tag in Reconyx Hyperfire cameras
- `OCRdataFields`: Optical character recognition (OCR) from data fields in images
- `writeDateTimeOriginal`: Write values to DateTimeOriginal tag in image metadata
- `timeShiftImages`: Apply time shifts to JPEG images
- `imageRename`: Copy and rename images based on station ID and image creation date
- `addCopyrightTag`: Write a copyright tag into JPEG image metadata
- `appendSpeciesNames`: Add or remove species names from image filenames

**Species / individual identification**

These functions assist in species identification and prepare individual identification of animals.

- `checkSpeciesNames`: Check species names against the ITIS taxonomic database
- `createSpeciesFolders`: Create directories for species identification
- `checkSpeciesIdentification`: Consistency check on species image identification
- `getSpeciesImages`: Gather all images of a species in a new directory

**Image data extraction**

These functions use the directory structure built above (Section 'Image management workflow') and a table containing basic information about camera traps and/or stations (IDs, location, trapping period).

- `recordTable`: Create a species record table from camera trap images and videos
- `recordTableIndividual`: Create a single-species record table from camera trap images and videos with individual IDs
- `exifTagNames`: Return Exif metadata tags and tag names from JPEG images
- `addToPath`: Add the directory containing exiftool.exe to PATH temporarily (Windows only)
- `filterRecordTable`: Filter existing record table for temporal independence

**Data exploration and visualisation**

These plots are generated from the record table and the camera trap table.

- `detectionMaps`: Generate maps of species richness and species presence by station, export shapefiles
- `activityHistogram`: Single-species diel activity histograms
- `activityDensity`: Single-species diel activity kernel density estimation plots
- `activityRadial`: Single-species diel activity radial plot
- `activityOverlap`: Two-species diel activity overlap plots and estimates
Prepare occupancy and spatial capture-recapture analyses, and summarise surveys

- **cameraOperation**: Create a camera operation matrix
- **detectionHistory**: Species detection histories for occupancy analyses (single and multi-season)
- **spatialDetectionHistory**: Detection histories of individuals for spatial capture-recapture analyses
- **surveyReport**: Create a report about camera trap surveys and species detections
- **surveyDashboard**: Shiny dashboard for summarizing and analyzing camera trap survey data

Community (multi-species) occupancy models

- **communityModel**: Create a community (multi-species) occupancy model for JAGS or Nimble
- **commOccu-class**: CommOccu objects
- **fit,commOccu-method**: Fit a community (multi-species) occupancy model
- **predict,commOccu-method**: Predictions from community occupancy models
- **summary,commOccu-method**: Summarize community occupancy model
- **plot_coef**: Plot effect sizes of covariates in community occupancy model
- **plot_effects**: Plot Marginal Effects of Covariates

Sample data

- **camtraps**: Sample camera trap station information table
- **recordTableSample**: Sample species record table
- **recordTableIndividualSample**: Single-species record table with individual IDs
- **camtrapsMultiSeason**: Sample multi season camera trap station information table
- **recordTableSampleMultiSeason**: Sample multi season species record table
- **recordTableIndividualSampleMultiSeason**: Single-species multi season record table with individual IDs
- **timeShiftTable**: Sample camera trap time shift information

Vignettes

1. Organising raw camera trap images
2. Identifying species and individuals
3. Extracting Data from Camera Trapping Images and Videos
4. Data exploration and visualisation
5. Multi-species occupancy models
Author(s)
Juergen Niedballa
Maintainer: Juergen Niedballa <camtrapr@gmail.com>

References

camtrapR Google Group https://groups.google.com/forum/#!forum/camtrapr

Phil Harvey’s ExifTool https://exiftool.org/

See Also
overlap unmarked ubms secr wiqid

activityDensity

Plot kernel density estimation of single-species activity

Description
The function plots a kernel density estimation of species diel activity using function densityPlot from package overlap.

Usage
activityDensity(
  recordTable,
  species,
  allSpecies = FALSE,
  speciesCol = "Species",
  recordDateTimeCol = "DateTimeOriginal",
  recordDateTimeFormat = "ymd HMS",
  plotR = TRUE,
  writePNG = FALSE,
  plotDirectory,
  createDir = FALSE,
  pngMaxPix = 1000,
  add.rug = TRUE,
  ...
)
**activityDensity**

**Arguments**

- `recordTable` data.frame. the record table created by `recordTable`
- `species` Name of the species for which to create a kernel density plot of activity
- `allSpecies` logical. Create plots for all species in speciesCol of `recordTable`? Overrides argument `species`
- `speciesCol` character. name of the column specifying species names in `recordTable`
- `recordDateTimeCol` character. name of the column specifying date and time in `recordTable`
- `recordDateTimeFormat` character. format of column `recordDateTimeCol` in `recordTable`
- `plotR` logical. Show plots in R graphics device?
- `writePNG` logical. Create pngs of the plots?
- `plotDirectory` character. Directory in which to create png plots if `writePNG = TRUE`
- `createDir` logical. Create `plotDirectory` if `writePNG = TRUE`?
- `pngMaxPix` integer. image size of png (pixels along x-axis)
- `add.rug` logical. add a rug to the plot?
- `...` additional arguments to be passed to function `densityPlot`

**Details**

- `species` must be in the `speciesCol` of `recordTable`.
- `recordDateTimeFormat` defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". `recordDateTimeFormat` can be interpreted either by base-R via `strptime` or in `lubridate` via `parse_date_time` (argument "orders"). `lubridate` will be used if there are no "%" characters in `recordDateTimeFormat`.
- For "YYYY-MM-DD HH:MM:SS", `recordDateTimeFormat` would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see `strptime` or `parse_date_time`.

**Value**

Returns invisibly a vector of species record observation times in radians, i.e. scaled to \([0, 2\pi]\). If `allSpecies == TRUE`, all species’ vectors are returned in an invisible named list.

**Author(s)**

- Juergen Niedballa

**References**

- Mike Meredith and Martin Ridout (2018). `overlap`: Estimates of coefficient of overlapping for animal activity patterns. R package version 0.3.2. [https://CRAN.R-project.org/package=overlap](https://CRAN.R-project.org/package=overlap)
activityHistogram

Plot histogram of single-species activity

Description

The function generates a histogram of species diel activity in 1-hour intervals.

Usage

activityHistogram(
  recordTable, 
  species, 
  allSpecies = FALSE, 
  speciesCol = "Species", 
  recordDateTimeCol = "DateTimeOriginal", 
  recordDateTimeFormat = "ymd HMS", 
  plotR = TRUE, 
  writePNG = FALSE, 
  plotDirectory, 
  createDir = FALSE, 
  pngMaxPix = 1000, 
  ... 
)
Arguments

recordTable data.frame. the record table created by recordTable

species Name of the single species for which to create a histogram of activity

allSpecies logical. Create plots for all species in speciesCol of recordTable? Overrides argument species

speciesCol character. name of the column specifying species names in recordTable

recordDateTimeCol character. name of the column specifying date and time in recordTable

recordDateTimeFormat character. format of column recordDateTimeCol in recordTable

plotR logical. Show plots in R graphics device?

writePNG logical. Create pngs of the plots?

plotDirectory character. Directory in which to create png plots if writePNG = TRUE

createDir logical. Create plotDirectory?

pngMaxPix integer. image size of png (pixels along x-axis)

... additional arguments to be passed to function hist

Details

Activity is calculated from the time of day of records. The date is ignored.

recordDateTimeFormat defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". recordDateTimeFormat can be interpreted either by base-R via strptime or in lubridate via parse_date_time (argument "orders"). lubridate will be used if there are no "%" characters in recordDateTimeFormat.

For "YYYY-MM-DD HH:MM:SS", recordDateTimeFormat would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see strptime or parse_date_time.

Value

It returns invisibly a vector of species record date and time in POSIXlt format. If allSpecies == TRUE, all species’ vectors are returned in an invisible named list.

Note

If you have a sufficiently large number of records you may wish to consider using activityDensity instead. Please be aware that this function (like the other activity... function of this package) use clock time. If your survey was long enough to see changes in sunrise and sunset times, this may result in biased representations of species activity.

Author(s)

Juergen Niedballa
See Also

activityDensity, activityRadial, activityOverlap

Examples

# load record table
data(recordTableSample)

# generate activity histogram
species4activity <- "VTA"  # = Viverra tangalunga, Malay Civet

activityHistogram (recordTable = recordTableSample,
                  species = species4activity,
                  allSpecies = FALSE)

activityOverlap

Plot overlapping kernel densities of two-species activities

Description

This function plots kernel density estimates of two species’ diel activity data by calling the function overlapPlot from package overlap. It further computes the overlap coefficient $D_{hat1}$ by calling overlapEst.

Usage

activityOverlap(
  recordTable,
  speciesA,
  speciesB,
  speciesCol = "Species",
  recordDateTimeCol = "DateTimeOriginal",
  recordDateTimeFormat = "ymd HMS",
  plotR = TRUE,
  writePNG = FALSE,
  addLegend = TRUE,
  legendPosition = "topleft",
  plotDirectory,
  createDir = FALSE,
  pngMaxPix = 1000,
  add.rug = TRUE,
  overlapEstimator = c("Dhat1", "Dhat4", "Dhat5"),
  ...
)
**activityOverlap**

**Arguments**

- `recordTable`  
  data.frame. the record table created by `recordTable`
- `speciesA`  
  Name of species 1 (as found in `speciesCol` of `recordTable`)
- `speciesB`  
  Name of species 2 (as found in `speciesCol` of `recordTable`)
- `speciesCol`  
  character. name of the column specifying species names in `recordTable`
- `recordDateTimeCol`  
  character. name of the column specifying date and time in `recordTable`
- `recordDateTimeFormat`  
  character. format of column `recordDateTimeCol` in `recordTable`
- `plotR`  
  logical. Show plots in R graphics device?
- `writePNG`  
  logical. Create pngs of the plots?
- `addLegend`  
  logical. Add a legend to the plots?
- `legendPosition`  
  character. Position of the legend (keyword)
- `plotDirectory`  
  character. Directory in which to create png plots if `writePNG = TRUE`
- `createDir`  
  logical. Create `plotDirectory`?
- `pngMaxPix`  
  integer. image size of png (pixels along x-axis)
- `add.rug`  
  logical. add a rug to the plot?
- `overlapEstimator`  
  character. Which overlap estimator to return (passed on to argument `type` in `overlapEst`)
- `...`  
  additional arguments to be passed to function `overlapPlot`

**Details**

... can be graphical parameters passed on to function `overlapPlot`, e.g. `linetype`, `linewidth`, `linecol` (see example below).

`recordDateTimeFormat` defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". `recordDateTimeFormat` can be interpreted either by base-R via `strptime` or in `lubridate` via `parse_date_time` (argument "orders"). `lubridate` will be used if there are no "%" characters in `recordDateTimeFormat`.

For "YYYY-MM-DD HH:MM:SS", `recordDateTimeFormat` would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see `strptime` or `parse_date_time`.

**Value**

Returns invisibly the `data.frame` with plot coordinates returned by `overlapPlot`.

**Note**

Please be aware that the function (like the other activity... function of this package) use clock time, not solar time. If your survey was long enough to see changes in sunrise and sunset times, this may result in biased representations of species activity.
Author(s)
Juergen Niedballa

References


See Also
activityDensity
https://www.kent.ac.uk/smsas/personal/msr/overlap.html

Examples

if(requireNamespace("overlap")) {

# load record table
data(recordTableSample)

# define species of interest
speciesA_for_activity <- "VTA"  # = Viverra tangalunga, Malay Civet
speciesB_for_activity <- "PBE"  # = Prionailurus bengalensis, Leopard Cat

# create activity overlap plot (basic)
activityOverlap (recordTable = recordTableSample,
speciesA = "VTA", # = Viverra tangalunga, Malay Civet
speciesB = "PBE", # = Prionailurus bengalensis, Leopard Cat
writePNG = FALSE,
plotR = TRUE
)

# create activity overlap plot (prettier and with some overlapPlot arguments set)

activityOverlap (recordTable = recordTableSample,
speciesA = speciesA_for_activity,
speciesB = speciesB_for_activity,
writePNG = FALSE,
plotR = TRUE,
createDir = FALSE,
pngMaxPix = 1000,
linecol = c("black", "blue"),
linewidth = c(5,3),
linetype = c(1, 2),
olapcol = "darkgrey",}
activityRadial

Radial plots of single-species activity

Description

The function generates a radial plot of species diel activity using an adapted version of function radial.plot from package plotrix (without the need to install the package). Records are aggregated by hour. The number of independent events is used as input, which in turn is based on the argument minDeltaTime in recordTable.

Usage

activityRadial(
  recordTable,
  species,
  allSpecies = FALSE,
  speciesCol = "Species",
  recordDateTimeCol = "DateTimeOriginal",
  recordDateTimeFormat = "ymd HMS",
  byNumber = FALSE,
  plotR = TRUE,
  writePNG = FALSE,
  plotDirectory,
  createDir = FALSE,
  pngMaxPix = 1000,
  ...
)

Arguments

recordTable   data.frame. the record table created by recordTable
species       Name of the species for which to create an kernel density plot of activity
allSpecies    logical. Create plots for all species in speciesCol of recordTable? Overrides argument species
speciesCol    character. name of the column specifying species names in recordTable
recordDateTimeCol    character. name of the column specifying date and time in recordTable
activityRadial

recordDateTimeFormat
   character. format of column recordDateTimeCol in recordTable

byNumber
   logical. If FALSE, plot proportion of records. If TRUE, plot number of records

plotR
   logical. Show plots in R graphics device?

writePNG
   logical. Create pngs of the plots?

plotDirectory
   character. Directory in which to create png plots if writePNG = TRUE

createDir
   logical. Create plotDirectory?

pngMaxPix
   integer. image size of png (pixels along x-axis)

... additional arguments to be passed to function radial.plot

Details

radial.plot was adjusted to show a clockwise 24-hour clock face. It is recommended to set argument lwd to a value >= 2. You may also wish to add argument rp.type="p" to show a polygon instead of bars.

recordDateTimeFormat defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". recordDateTimeFormat can be interpreted either by base-R via strptime or in lubridate via parse_date_time (argument "orders"). lubridate will be used if there are no "%" characters in recordDateTimeFormat.

For "YYYY-MM-DD HH:MM:SS", recordDateTimeFormat would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see strptime or parse_date_time.

Value

Returns invisibly a data.frame containing all information needed to create the plot: radial position, lengths, hour (for labels). If allSpecies == TRUE, all species’ data frames are returned in an invisible named list.

Author(s)

Juergen Niedballa

References


See Also

activityDensity, activityHistogram, activityOverlap
### Examples

```r
# load record table
data(recordTableSample)

species4activity <- "PBE"  # = Prionailurus bengalensis, Leopard Cat

activityRadial(recordTable = recordTableSample,
                species = species4activity,
                allSpecies = FALSE,
                speciesCol = "Species",
                recordDateTimeCol = "DateTimeOriginal",
                plotR = TRUE,
                writePNG = FALSE,
                lwd = 5)

# plot type = polygon

activityRadial(recordTable = recordTableSample,
                species = species4activity,
                allSpecies = FALSE,
                speciesCol = "Species",
                recordDateTimeCol = "DateTimeOriginal",
                plotR = TRUE,
                writePNG = FALSE,
                lwd = 5,
                rp.type = "p")
```

---

#### addCopyrightTag

**Write a copyright tag into JPEG image metadata**

**Description**

This function writes a copyright tag into the copyright field of JPEG image Exif metadata. It does so recursively, so it works both for images that are sorted into subdirectories and unsorted images. Note that all images in subdirectories of `inDir` will be tagged. It is not required to run this function in the camtrapR workflow, but may be desired for data sharing or publishing.

**Usage**

```r
addCopyrightTag(
    inDir,  # directory containing the images
    copyrightTag,  # the text to write as copyright
    askFirst = TRUE)  # prompt the user before processing
```
Arguments

inDir character. Name of the directory containing camera trap images.
copyrightTag character. The tag to be written into the Exif Copyright field
askFirst logical. Ask user to confirm before execution?
keepJPG_original logical. Keep original JPG files as .JPG_original files (TRUE) or overwrite JPGs (FALSE)?
ignoreMinorErrors logical. Ignore minor errors that would cause the function to fail (set TRUE for images with bad MakerNotes, observed in Panthera V4 cameras)

Details

If askFirst = TRUE, the function will show a menu and asks the user to confirm the action before execution. Type "1" to write copyright tags and "2" to abort.

By default Exiftool creates a copy of each JPG image and preserves the original images (without the copyright tag) as .JPG_original files. Note that this behaviour will instantly double the number of images in inDir and the disk space required. If this is not desired, set keepJPG_original = FALSE.

ignoreMinorErrors is useful if copyright tags can’t be updated correctly. This can be caused by bad MakerNotes and so far was only observed in Panthera V4 cameras. In that case, set ignoreMinorErrors to TRUE. This will add the "-m" option to the Exiftool call, thereby ignoring minor errors and warnings and assigning the copyright tag regardless.

Value

An invisible list of Exiftool output.

More importantly, the specified copyright tag is written into the Copyright field of the Exif metadata of all images in inDir.

Author(s)

Juergen Niedballa

Examples

```r
## Not run:
if (Sys.which("exiftool") != "){  # only run this example if ExifTool is available
# copy sample images to temporary directory (so we don't mess around in the package directory)
wd_images_ID <- system.file(file.path("pictures", "sample_images_species_dir"),
```
package = "camtrapR"
file.copy(from = wd_images_ID, to = tempdir(), recursive = TRUE)
wd_images_ID_copy <- file.path(tempdir(), "sample_images_species_dir")

# define a sample tag
copyrightTagToAdd <- "Your Name (Your Organisation)"

# add the tag to the images
addCopyrightTag(inDir = wd_images_ID_copy,
copyrightTag = copyrightTagToAdd)

# we choose "YES", i.e., we want to add a copyright tag

# you can check the outcome with function exifTagNames

metadat <- exifTagNames(wd_images_ID_copy)
metadat [metadat$tag_name == "Copyright",]

## End(Not run)

---

**addToPath**

*Add a directory to PATH temporarily*

**Description**

Temporarily adds a directory to the environmental variable PATH for system calls from within R. This allows Windows users to store exiftool.exe anywhere on their hard drive and is useful if they cannot store the file in system directories. It is not needed on Linux or MacOS machines.

**Usage**

```r
addToPath(directory)
```

**Arguments**

- `directory` character. the directory in the file system to add to PATH (e.g. the directory containing exiftool.exe).

**Details**

Several functions within this package depend on ExifTool. Under Windows, exiftool.exe cannot be used if it is not in a directory path specified in PATH. This can be solved by adding the directory containing exiftool.exe for temporary use within the running R process. It can also be useful in other contexts besides Exiftool.
appendSpeciesNames

Value

invisible logical indicating whether directory was added to PATH successfully (in the running R process).

Note

The directories in PATH can be queried by Sys.getenv("PATH").

Author(s)

Juergen Niedballa

Examples

```r
exiftool_dir <- "C:/Path/To/Exiftool"
addToPath(directory = exiftool_dir)

# check if it has been added to PATH
grepl(exiftool_dir, Sys.getenv("PATH"))
```

appendSpeciesNames  Add or remove species names from JPEG image filenames

Description

Add or remove species names from JPEG image filenames. It makes it easier to find images of a species.

Usage

```r
appendSpeciesNames(
  inDir,
  IDfrom,
  hasCameraFolders,
  metadataSpeciesTag,
  metadataHierarchyDelimiter = "|",
  removeNames = FALSE,
  writecsv = FALSE
)
```

Arguments

- `inDir` character. Directory containing camera trap images sorted into station subdirectories (e.g. inDir/StationA/)
- `IDfrom` character. Read species ID from image metadata ("metadata") of from species directory names ("directory")?
appendSpeciesNames

hasCameraFolders
logical. Do the station subdirectories of inDir have camera-subdirectories (e.g. inDir/StationA/CameraA1; inDir/StationA/CameraA2)?

metadataSpeciesTag
character. The species ID tag name in image metadata (if IDfrom = "metadata").

metadataHierarchyDelimiter
character. The character delimiting hierarchy levels in image metadata tags in field "HierarchicalSubject". Either |" or ":".

removeNames
logical. remove appended species names?

writecsv
logical. write csv table containing old and new file names into inDir?

Details
Species names can be appended or removed from image filenames. Before running the function, you may want to run checkSpeciesIdentification to detect possible misidentifications. As an example, the function would change an image file name from "StationA__2015-05-41__20-59-59(1).JPG" to "StationA__2015-05-41__20-59-59(1)__Species Name.JPG". If species names were appended several times by accident, they can all be removed by running the function with removeNames = TRUE

Value
A data.frame containing the old and new file names and directories.

Author(s)
Juergen Niedballa

Examples

## Not run:

# copy sample images to another location (so we don't mess around in the package directory)
wd_images_ID <- system.file("pictures/sample_images_species_dir", package = "camtrapR")
file.copy(from = wd_images_ID, to = getwd(), recursive = TRUE)
wd_images_ID_copy <- file.path(getwd(), "sample_images_species_dir")

# append species names
SpecNameAppend1 <- appendSpeciesNames(inDir = wd_images_ID_copy,
                                  IDfrom = "directory",
                                  hasCameraFolders = FALSE,
                                  removeNames = FALSE,
                                  writecsv = FALSE)

SpecNameAppend1

# remove species names
SpecNameRemove1 <- appendSpeciesNames(inDir = wd_images_ID_copy,
                                   IDfrom = "directory",
                                   hasCameraFolders = FALSE,
                                   removeNames = FALSE,
                                   writecsv = FALSE)
cameraOperation

Create a camera trap station operation matrix

Description

Construct a matrix of daily camera trap station operation for use in detectionHistory and spatialDetectionHistory, where it is needed for calculating trapping effort per occasion. It is also used in surveyReport to calculate the number of trap nights during a survey. If several cameras were deployed per station, the matrix can contain camera- or station-specific trap operation information, or information about sessions during repeated surveys.

Usage

cameraOperation(
  CTtable,
  stationCol = "Station",
  cameraCol = NULL,  # optional. If empty, 1 camera per station is assumed.
  sessionCol,
  setupCol,
  retrievalCol,
  hasProblems = FALSE,
  byCamera = TRUE,
  allCamsOn = FALSE,
  camerasIndependent = FALSE,
  dateFormat = "ymd",
  occasionStartTime = 0,
  writecsv = FALSE,
  outDir = FALSE
)

Arguments

CTtable data.frame containing information about location and trapping period of camera trap stations
stationCol character. name of the column specifying Station ID in CTtable
cameraCol character. name of the column specifying Camera ID in CTtable (optional). If empty, 1 camera per station is assumed.
sessionCol character. name of the column specifying session ID in CTtable (optional). Use it for creating multi-session / multi-season detection histories (unmarked: unmarkedMultFrame; secr: capthist)

setupCol character. name of the column containing camera setup dates in CTtable

retrievalCol character. name of the column containing camera retrieval dates in CTtable

hasProblems logical. If TRUE, function will look for columns specifying malfunction periods in CTtable (naming convention: ProblemX_from and ProblemX_to, where X is a number)

byCamera logical. If TRUE, camera operation matrix is computed by camera, not by station (requires cameraCol)

allCamsOn logical. Takes effect only if cameraCol is defined and if byCamera is FALSE. If allCamsOn = TRUE, all cameras at a station need to be operational for the station to be operational (e.g. 1 camera out of 2 malfunctioning renders the station inoperational). Output values can be 1/0/NA only (all cameras at a station operational/ at least 1 camera not operational/ no camera set up). If allCamsOn = FALSE, at least 1 active camera makes a station operational.

camerasIndependent logical. Return number of active camera traps by station? Only if byCamera is FALSE and allCamsOn is FALSE. If camerasIndependent is TRUE, output values will be the number of operational cameras at a station. If camerasIndependent is FALSE, the value is 1 if at least 1 camera was operational, otherwise 0. In both cases, values are NA if no camera was set up.

dateFormat character. The format of columns setupCol and retrievalCol (and potential problem columns) in CTtable. Must be interpretable by either as.Date or the "orders" argument parse_date_time in lubridate. Can be a date or (since version 2.1) a date-time.

occasionStartTime integer. time of day (the full hour) at which to begin occasions. Replaces occasionStartTime from detectionHistory and spatialDetectionHistory.

writecsv logical. Should the camera operation matrix be saved as a .csv?

outDir character. Directory into which csv is saved

Details
cameraCol is NULL by default, meaning the function assumes there was 1 camera per station in CTtable. If more than 1 camera was deployed per station, cameraCol needs to be specified to identify individual cameras within a station. Likewise, sessionCol can be used to if camera trap stations were operated during multiple sessions / trapping seasons.

dateFormat defaults to "YYYY-MM-DD", e.g. "2014-10-31", but can be any other date format or date-time also. It can be specified either in the format required by strptime or the 'orders' argument in parse_date_time in lubridate. In the example above, "YYYY-MM-DD" would be specified as "%Y-%m-%d" in base R or "ymd" in lubridate.

Since version 2.1, dateFormat can be a date-time. That makes it possible to specify the exact time cameras were set up / retrieved / malfunctioned / worked again. This information is used to calculate the daily trapping effort more precisely on days with incomplete effort.
Previously, setup and retrieval day were counted as 1, indicating a whole day of effort on those days. Since version 2.1, setup and retrieval are assumed to have happened at 12 noon (resulting in daily effort of 0.5 instead of 1). Users can also specify the exact time cameras were set up (by providing a date-time in the setup / retrieval / problem columns). See vignette 3 for more details.

If `hasProblems` is TRUE, the function tries to find columns `ProblemX_from` and `ProblemX_to` in `CTtable`. X is a consecutive number from 1 to n, specifying periods in which a camera or station was not operational. If `hasProblems` is FALSE, cameras are assumed to have been operational uninterruptedly from setup to retrieval (see `camtraps` for details).

`allCamsOn` only has an effect if there was more than 1 camera at a station. If TRUE, for the station to be considered operational, all cameras at a station need to be operational. If FALSE, at least 1 active camera renders the station operational. Argument `camerasIndependent` defines if cameras record animals independently (it thus only has an effect if there was more than 1 camera at a station). This is the case if an observation at one camera does not increase the probability for detection at another camera (cameras face different trails at a distance of one another). Non-independence occurs if an animal is likely to trigger both cameras (as would be the case with 2 cameras facing each other).

If `camerasIndependent` is TRUE, 2 active cameras at a station will result in a station operation value of 2 in the resulting matrix, i.e., 2 independent trap days at 1 station and day. If `camerasIndependent` is FALSE, 2 active cameras will return value 1, i.e., 1 trap night at 1 station per day.

Row names depend on the input arguments and contain the station name and potentially session and camera names (if `sessionCol` and/or `cameraCol` are defined). Naming convention is (since version 1.2) **Bold** information are from the columns `stationCol`, `sessionCol` and `cameraCol` in `CTtable`:

**Station**

- `Station__SESS_SessionID`
- `Station__CAM_CameraID`
- `Station__SESS_SessionID__CAM_CameraID`

Session are designated with prefix "__SESS_", cameras with prefix "__CAM_". Therefore, these are reserved words and may not be part of station, session or camera names. Here’s what it may look like in real life:

- `Station1__SESS_2019`
- `Station1__CAM_1024152`
- `Station1__SESS_2019__CAM_1024152`

Functions `detectionHistory` and `spatialDetectionHistory` recognize these and use the information accordingly.

**Value**

A matrix. Row names always indicate Station IDs. If `sessionCol` and/or `cameraCol` are defined, they are contained in the row names also (camera ID only if `byCamera = TRUE`). Column names are dates.

Legend: NA: camera(s) not set up, 0: camera(s) not operational, 1 (or higher): number of operational camera(s) or an indicator for whether the station was operational (depending on `camerasIndependent` and `allCamsOn`).
cameraOperation

Note

Setting camerasIndependent according to the sampling situation is important for the functions detectionHistory and spatialDetectionHistory, if sampling effort (the number of active trap nights in an occasion) is to be computed and returned.

Author(s)

Juergen Niedballa

Examples

data(camtraps)

# no problems/malfunction
camop_no_problem <- cameraOperation(CTtable = camtraps,
    stationCol = "Station",
    setupCol = "Setup_date",
    retrievalCol = "Retrieval_date",
    writecsv = FALSE,
    hasProblems = FALSE,
    dateFormat = "dmy"
)

# with problems/malfunction

camop_problem <- cameraOperation(CTtable = camtraps,
    stationCol = "Station",
    setupCol = "Setup_date",
    retrievalCol = "Retrieval_date",
    writecsv = FALSE,
    hasProblems = TRUE,
    dateFormat = "dmy"
)

# with problems/malfunction / dateFormat in strptime format

camop_problem_lubridate <- cameraOperation(CTtable = camtraps,
    stationCol = "Station",
    setupCol = "Setup_date",
    retrievalCol = "Retrieval_date",
    writecsv = FALSE,
    hasProblems = TRUE,
    dateFormat = "%d/%m/%Y"
)

camop_no_problem

camop_problem

camop_problem_lubridate
camtrapsMultiSeason

---

**Description**

Sample camera trap station information

**Usage**

data(camtraps)

**Format**

A data frame with 3 rows and 7 variables

**Details**

This is a general example of how information about camera trap stations are arranged in camtrapR. It contains setup and retrieval dates and coordinates. If more than 1 camera was set up at a station (e.g. 2 cameras facing each other), a camera ID column must be added, with camera-specific information instead of station-specific information. If cameras malfunctioned repeatedly, additional pairs of problem columns can be added, e.g. "Problem2_from" and "Problem2_to" etc..

The variables are as follows:

<table>
<thead>
<tr>
<th>Variable</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Station</td>
<td>Camera trap station ID</td>
</tr>
<tr>
<td>utm_y</td>
<td>y coordinate of station (northing)</td>
</tr>
<tr>
<td>utm_x</td>
<td>x coordinate of station (easting)</td>
</tr>
<tr>
<td>Setup_date</td>
<td>camera trap setup date</td>
</tr>
<tr>
<td>Retrieval_date</td>
<td>camera trap retrieval date</td>
</tr>
<tr>
<td>Problem1_from</td>
<td>first day of camera malfunction</td>
</tr>
<tr>
<td>Problem1_to</td>
<td>last day of camera malfunction</td>
</tr>
</tbody>
</table>

**Note**

The coordinates can be in the units of any coordinate system. UTM was chosen as an example, but it could be latlong or anything else, too. capthist objects (as created by spatialDetectionHistory for spatial capture-recapture analyses) expect the unit to be meters.

---

camtrapsMultiSeason  

---

**Description**

Sample multi-season camera trap station information

---

**Description**

Example multi-season camera trap station information table
Usage

data(camtrapsMultiSeason)

Format

A data frame with 7 rows and 8 variables

Details

This is a general example of how information about camera trap stations from multiple seasons are arranged in camtrapR. It contains setup and retrieval dates, coordinates and a season identifier. If more than 1 camera was set up at a station (e.g. 2 cameras facing each other), a camera ID column must be added, with camera-specific information instead of station-specific information. If cameras malfunctioned repeatedly, additional pairs of problem columns can be added, e.g. "Problem2_from" and "Problem2_to" etc..

Note that season 2010 has an additional station (StationD). This is to simulate a situation where a station was not set up during an entire season.

The variables are as follows:

- **Station**: Camera trap station ID
- **utm_y**: y coordinate of station (northing)
- **utm_x**: x coordinate of station (easting)
- **Setup_date**: camera trap setup date
- **Retrieval_date**: camera trap retrieval date
- **Problem1_from**: first day of camera malfunction
- **Problem1_to**: last day of camera malfunction
- **session**: Identified for trapping session / season

Note

The coordinates can be in the units of any coordinate system. UTM was chosen as an example, but it could be latlong or anything else, too. capthist objects (as created by spatialDetectionHistory for spatial capture-recapture analyses) expect the unit to be meters. capthist also require session information as integer numbers starting with 1.

"Season" and "session" are used synonymously here. secr nomenclature is "session", in unmarked it is "season".

Examples

```r
# data were created with the following code:
data(camtraps)
camtraps_season2 <- camtraps

# change 2009 to 2010
camtraps_season2[, "Setup_date"] <- gsub("2009", "2010", camtraps_season2[, "Setup_date"])
camtraps_season2[, "Retrieval_date"] <- gsub("2009", "2010", camtraps_season2[, "Retrieval_date"])
```
checkSpeciesIdentification

Consistency check on species image identification

Description
This function serves 2 purposes: 1) it assesses possible misidentification of species and 2) compares double observer species identification (only if metadata tagging was used for species identification).

Usage
checkSpeciesIdentification(
inDir,  
IDfrom,  
hasCameraFolders,  
metadataSpeciesTag,  
metadataSpeciesTagToCompare,  
metadataHierarchyDelimiter = "|",  
maxDeltaTime,  
excludeSpecies,  
stationsToCheck,  
writecsv = FALSE)
)
checkSpeciesIdentification

Arguments

inDir character. Directory containing identified camera trap images sorted into station subdirectories (e.g. inDir/StationA/)

IDfrom character. Read species ID from image metadata ("metadata") of from species directory names ("directory")?

hasCameraFolders logical. Do the station directories in inDir have camera subdirectories (e.g. "inDir/StationA/Camera1" or "inDir/StationA/Camera1/Species1")?

metadataSpeciesTag character. The species ID tag name in image metadata (if IDfrom = "metadata").

metadataSpeciesTagToCompare character. A second species ID tag name in image metadata (if IDfrom = "metadata"). For comparing double observer species identification.

metadataHierarchyDelimiter character. The character delimiting hierarchy levels in image metadata tags in field "HierarchicalSubject". Either "|" or ":".

maxDeltaTime numeric. Maximum time interval between images to be returned (in seconds)

excludeSpecies character. vector of species to exclude from checks

stationsToCheck character. vector of stations to be checked (optionally)

writecsv logical. Should the resulting data.frame be saved as a .csv?

Details

Within each station, it assesses whether there are images of a species taken within a given time interval of another species. Often, it is unlikely that different species are encountered within a very short time intervals at the same location. This type of misidentification can arise easily if some images belonging to a sequence of images were accidentally moved into different species directories or tagged incorrectly.

Double observer identification may be desirable to increase reliability of species identification. The function returns conflicts in species identification between 2 observers. These conflicts can then be corrected.

Images may accidentally be misidentified by assigning wrong species tags or by moving them into wrong species directories. Imagine your cameras take sequences of images each time they are triggered and one image of the sequence is misidentified. The time difference between these images (that have different species assigned to them) will be very small, usually a few seconds. This function will return all these images for you to check if they were identified correctly.

If multiple observers identify images independently using metadata tagging, their identifications can be compared by setting metadataSpeciesTagToCompare. Conflicting or missing identifications will be reported. This feature is only available if images were identified by metadata tagging.

Species like "blank" or "team" can be ignored using excludeSpecies. If only specific stations are to be checked, stationsToCheck can be set.
Value

A list containing 2 data frames. The first contains a data frame with images file names, directories, time stamp and species ID that were taken within maxDeltaTime seconds of another species image at a particular station. The second data frame contains images with conflicting species IDs (if IDfrom = "metadata" and metadataSpeciesTagToCompare is defined).

Note

The function will not be able to find "isolated" images, i.e. images that were misidentified, but were not part of a sequence of images. Likewise, if all images of a sequence were misidentified, they cannot be found either. From version 0.99.0, the function can also handle images identified with metadata tags.

Author(s)

Juergen Niedballa

Examples

wd_images_ID <- system.file("pictures/sample_images_species_dir", package = "camtrapR")

if (Sys.which("exiftool") != ""){
  # only run this example if ExifTool is available
  check.folders <- checkSpeciesIdentification(inDir = wd_images_ID,
  IDfrom = "directory",
  hasCameraFolders = FALSE,
  maxDeltaTime = 120,
  writecsv = FALSE)

  check.folders # In the example, 2 different species were photographed within 2 minutes.
}

## Not run:
# now exclude one of these 2 species
check.folders2 <- checkSpeciesIdentification(inDir = wd_images_ID,
  IDfrom = "directory",
  hasCameraFolders = FALSE,
  maxDeltaTime = 120,
  excludeSpecies = "EGY",
  writecsv = FALSE)

check.folders2 # the data frame is empty

# now we check only one station
check.folders3 <- checkSpeciesIdentification(inDir = wd_images_ID,
  IDfrom = "directory",
  hasCameraFolders = FALSE,
  maxDeltaTime = 120,
checkSpeciesNames

stationsToCheck = "StationB",
writecsv = FALSE)

checkSpeciesNames # the data frame is empty
## End(Not run)

checkSpeciesNames

Check species names against the ITIS taxonomic database

Description

The function checks species names (common or scientific names) provided by the user with the ITIS taxonomic database (http://www.itis.gov/) via functions from the package taxize. It returns both common and scientific names, the taxon authors, taxon rank name and status, the TSN (taxonomic serial numbers) and ITIS urls.

Usage

checkSpeciesNames(speciesNames, searchtype, accepted = TRUE, ask = TRUE)

Arguments

- **speciesNames**: character. Vector of species names to check. Either common names or scientific names.
- **searchtype**: character. Type of names specified in speciesNames. One of 'scientific' or 'common'.
- **accepted**: logical. Return only accepted valid names? If TRUE, invalid names are returned as NA. Set to FALSE to return both accepted and unaccepted names.
- **ask**: logical. Should the function be run in interactive mode? If TRUE and more than one TSN is found for a species, the user is asked to choose one. If FALSE, NA is returned for multiple matches.

Details

Arguments searchtype, accepted and ask are passed on to get_tsn.

Value

A data.frame with the names supplied by the user, matching common and scientific names, taxon author and year, taxonomic rank, status, TSNs (taxonomic serial numbers) and ITIS urls.

Author(s)

Juergen Niedballa
References
http://www.itis.gov/

Examples

## Not run:

```r
species_common <- c("Leopard Cat", "moonrat")

# ask = TRUE. Multiple matches for leopard cat will cause menu to pop up asking user input.

species.names.check1 <- checkSpeciesNames(speciesNames = species_common,
                                         searchtype = "common",
                                         accepted = TRUE,
                                         ask = TRUE)

2  # we choose entry 2
species.names.check1

# ask = FALSE. Multiple matches for leopard cat will cause NA.

species.names.check2 <- checkSpeciesNames(speciesNames = species_common,
                                         searchtype = "common",
                                         accepted = TRUE,
                                         ask = FALSE)

species.names.check2

# search for scientific names

species_scientific <- c("Tragulus", "Prionailurus bengalensis")

species.names.check3 <- checkSpeciesNames(speciesNames = species_scientific,
                                         searchtype = "scientific",
                                         accepted = TRUE,
                                         ask = TRUE)

species.names.check3

## End(Not run)
```
communityModel

Description

commOccu objects

Value

commOccu object

Slots

- **modelText**: JAGS model code as a character vector (made up of code chunks, use cat() to print)
- **params**: Parameters to monitor in the model runs
- **inits_fun**: Function to create start values for the MCMC chains. It being a function ensures different values in each chain
- **data**: List with data needed to run the model (detection & effort matrices, site covariates, number of species / stations / occasions)
- **input**: Input data_list (unchanged)
- **nimble**: logical indicator for whether it is a Nimble model
- **modelFile**: Path of the text file containing the model code
- **covariate_info**: Data frame containing information about covariates. Only used internally in plot_* and predict methods
- **model**: character indicating whether it is a standard "Occupancy" model or Royle-Nichols ("RN") occupancy model

Note

The data slot is a list of model input data. While the exact content depends on function input, it can be summarized as:

- **y**: array of detection histories. Dimensions are: y[species, station, occasion]
- **effort_binary**: matrix of binary (1/0) survey effort. Only used to ensure p = 0 when effort = 0. Dimensions are station, occasion
- **site-occasion covariates**: The required content of data_list$obsCovs as named matrices with dimensions [station, occasion, covariate]
- **site covariates**: The required columns of data_list$siteCovs as named vectors (length = number of stations)
- **M**: Number of species
- **J**: Number of stations
- **maxocc**: Number of occasions

For categorical site-occasion covariates, an addition matrix containing an integer representation of the character matrix with suffix "_integer" is stored in the data slot.

---

**communityModel**: Create a community (multi-species) occupancy model for JAGS or Nimble
Description

Flexibly creates complete code and input data for community occupancy models for JAGS and Nimble (both standard occupancy models and Royle-Nichols occupancy models), and automatically sets initial values and parameters to monitor. Supports fixed and random effects of covariates on detection and occupancy probabilities, using both continuous and categorical covariates (both site and site-occasion covariates).

Optionally includes data augmentation (fully open community, or up to known maximum number of species, or no data augmentation). Allows combination of all these parameters for fast and flexible customization of community occupancy models.

Incidentally, the function can also be used to create model code and input for single-species single-season occupancy models (it is the special case of the community model with only one species). Such a model will run slower than proper single-species model JAGS code due to the additional species loop, but it is possible.

The function returns several derived quantities, e.g. species richness, Bayesian p-values (overall and by species), Freeman-Tukey residuals for actual and simulated data (by station and total). If doing data augmentation, metacommunity size and number of unseen species are returned also.

Usage

```r
communityModel(
  data_list,
  model = c("Occupancy", "RN"),
  occuCovs = list(fixed = NULL, independent = NULL, ranef = NULL),
  detCovs = list(fixed = NULL, ranef = NULL),
  detCovsObservation = list(fixed = NULL, ranef = NULL),
  speciesSiteRandomEffect = list(det = FALSE, occu = FALSE),
  intercepts = list(det = "fixed", occu = "fixed"),
  effortCov = "effort",
  richnessCategories = NULL,
  augmentation = NULL,
  modelFile = NULL,
  nimble = FALSE,
  keyword_quadratic = "._squared"
)
```

Arguments

- `data_list` list. Contains 3 slots: ylist, siteCovs, obsCovs. ylist is a list of detection histories (can be named), e.g. from `detectionHistory`. siteCovs is a data.frame with site covariates (optional). obsCovs is a list of site-occasion level covariates (e.g. site-occasion-specific effort, which is also returned by `detectionHistory`).

- `model` character. "Occupancy" for standard occupancy model, or "RN" for the occupancy model of Royle and Nichols (2003), which relates probability of detection of the species to the number of individuals available for detection at each station.

- `occuCovs` list. Up to 3 items named "fixed", "independent", and/or "ranef". Specifies fixed, independent or random effects of covariates on occupancy probability (continu-
ous or categorical covariates). Independent effects are only supported for continuous covariates.

- **detCovs**: list. Up to 3 items named "fixed", "independent", and/or "ranef". Specifies fixed, independent or random effects of covariates on detection probability (continuous or categorical covariates). Independent effects are only supported for continuous covariates.

- **detCovsObservation**: list. Up to 2 items named "fixed" and/or "ranef". Specifies fixed or random effects of observation-level covariates on detection probability (continuous or categorical covariates - categorical must be coded as character matrix).

- **speciesSiteRandomEffect**: list. Two items named "det" and "occu". If TRUE, adds a random effect of species and station. Only implemented for detection probability.

- **intercepts**: list. Two items named "det" and "occu" for detection and occupancy probability intercepts. Values can be "fixed" (= constant across species), "independent" (= independent estimates for each species), or "ranef" (= random effect of species on intercept).

- **effortCov**: character. Name of list item in data_list$obsCovs which contains effort. This does not include effort as a covariate on detection probability, but only uses NA / not NA information to create binary effort and ensure detection probabilities p are 0 when there was no effort (p will be 0 where ever effortCov is NA).

- **richnessCategories**: character. Name of categorical covariate in data_list$siteCovs for which to calculate separate richness estimates (optional). Can be useful to obtain separate richness estimates for different areas.

- **augmentation**: If NULL, no data augmentation (only use species in data_list$ylist), otherwise named list or vector with total number of (potential) species. Names: "max-known" or "full". Example: augmentation = c(maxknown = 30) or augmentation = c(full = 30)

- **modelFile**: character. Text file name to save model to

- **nimble**: logical. If TRUE, model code will be for Nimble (incompatible with JAGS). If FALSE, model code is for JAGS.

- **keyword_quadratic**: character. A suffix in covariate names in the model that indicates a covariate is a quadratic effect of another covariate which does not carry the suffix in its name (e.g. if the covariate is "elevation", the quadratic covariate would be "elevation_squared").

## Details

For examples of implementation, see Vignette 5: Multi-species occupancy models.

Fixed effects of covariates are constant across species, whereas random effect covariates differ between species. Independent effect differ between species and are independent (there is no underlying hyperdistribution). Fixed, independent and random effects are allowed for station-level detection and occupancy covariates (a.k.a. site covariates). Fixed and random effects are also allowed for station-occasion level covariates (a.k.a. observation covariates). Currently independent effects
are only supported for continuous site covariates, not categorical site covariates or observation-level covariates.

By default, random effects will be by species. It is however possible to use categorical site covariates for grouping (continuous|categorical). Furthermore, it is possible to use nested random effects of species and another categorical site covariate (so that there is a random effect of species and an additional random effect of a categorical covariate within each species).

Derived quantities returned by the model are:

- **Bpvalue**: Bayesian p-value (overall)
- **Bpvalue_species**: Bayesian p-value (by species)
- **Nsclass**: Species richness (only in JAGS models)
- **Nsclass_station**: Species richness at each sampling locations (only in JAGS models)
- **Nsclass_Covariate**: Species richness by categorical covariate (when using richnessCategories, only in JAGS models)
- **R2**: sum of Freeman-Tukey residuals of observed data within each species
- **new.R2**: sum of Freeman-Tukey residuals of simulated data within each species
- **R3**: Total sum of Freeman-Tukey residuals of observed data
- **new.R3**: Total sum of Freeman-Tukey residuals of simulated data
- **Ntotal**: Total metacommunity size (= observed species + n0)
- **n0**: Number of unseen species in metacommunity
- **omega**: Data augmentation parameter
- **w**: Metacommunity membership indicator for each species

Quantities in *italic* at the bottom are only returned in full data augmentation. Nspecies and Nspecies_Covariate are only returned in JAGS models (because Nimble models don’t explicitly return latent occupancy status z).

**Value**

commOccu object. It is an S4 class containing all information required to run the models. See commOccu-class for details.

**Parameter naming convention**

The parameter names are assembled from building blocks. The nomenclature is as follows:

<table>
<thead>
<tr>
<th>Name</th>
<th>Refers to</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>alpha</td>
<td>Submodel</td>
<td>detection submodel</td>
</tr>
<tr>
<td>beta</td>
<td>Submodel</td>
<td>occupancy submodel</td>
</tr>
<tr>
<td>0</td>
<td>Intercept</td>
<td>denotes the intercepts (alpha0, beta0)</td>
</tr>
<tr>
<td>fixed</td>
<td>Effect type</td>
<td>fixed effects (constant across species)</td>
</tr>
<tr>
<td>indep</td>
<td>Effect type</td>
<td>independent effects (separate for each species)</td>
</tr>
<tr>
<td>ranef</td>
<td>Effect type</td>
<td>random effects (of species and/or other categorical covariates)</td>
</tr>
<tr>
<td>cont</td>
<td>Covariate type</td>
<td>continuous covariates</td>
</tr>
<tr>
<td>categ</td>
<td>Covariate type</td>
<td>categorical covariates</td>
</tr>
<tr>
<td>mean</td>
<td>Hyperparameter</td>
<td>mean of random effect</td>
</tr>
<tr>
<td>sigma</td>
<td>Hyperparameter</td>
<td>standard deviation of random effect</td>
</tr>
<tr>
<td>tau</td>
<td>Hyperparameter</td>
<td>precision of random effect (used internally, not returned)</td>
</tr>
</tbody>
</table>
For example, a fixed intercept of occupancy (constant across species) is $\beta_0$, and a fixed intercept of detection probability is $\alpha_0$.

An occupancy probability intercept with a random effect of species is:
- $\beta_0$.mean community mean of the occupancy probability intercept
- $\beta_0$.sigma standard deviation of the community occupancy probability intercept.
- $\beta_0[1]$ occupancy probability intercept of species 1 (likewise for other species).

For effects of site covariates, the pattern is:
- submodel.effectType.covariateType.CovariateName.hyperparameter

For example:
- $\beta_{\text{ranef}.\text{cont}.\text{habitat}.\text{mean}}$ is the mean community effect of the continuous site covariate 'habitat' on occupancy probability.
- $\beta_{\text{ranef}.\text{cont}.\text{habitat}[1]}$ is the effect of continuous site covariate 'habitat' on occupancy probability of species 1.

Site-occasion covariates are denoted by ".obs" after the submodel, e.g.:
- $\alpha_{\text{obs}.\text{fixed}.\text{cont}.\text{effort}}$ is the fixed effect of the continuous observation-level covariate 'effort' on detection probability.

Author(s)
Juergen Niedballa

References

Examples

```r
## Not run:
data("camtraps")

# create camera operation matrix
camop_no_problem <- cameraOperation(CTtable = camtraps,
                        stationCol = "Station",
                        setupCol = "Setup_date",
                        retrievalCol = "Retrieval_date",
                        hasProblems = FALSE,
                        dateFormat = "dmy")

data("recordTableSample")

# make list of detection histories
DetHist_list <- lapply(unique(recordTableSample$Species), FUN = function(x) {  
```
```r
detectionHistory(
  recordTable = recordTableSample,
  camOp = camop_no_problem,
  stationCol = "Station",
  speciesCol = "Species",
  recordDateTimeCol = "DateTimeOriginal",
  species = x,
  occasionLength = 7,
  day = "station",
  datesAsOccasionNames = FALSE,
  includeEffort = TRUE,
  scaleEffort = TRUE,
  timeZone = "Asia/Kuala_Lumpur"
)
)

# assign species names to list items
names(DetHist_list) <- unique(recordTableSample$Species)

# extract detection histories (omit effort matrices)
ylist <- lapply(DetHist_list, FUN = function(x) x$detection_history)

# create some fake covariates for demonstration
sitecovs <- camtraps[, c(1:3)]
sitecovs$elevation <- c(300, 500, 600)

# scale numeric covariates
sitecovs[, c(2:4)] <- scale(sitecovs[, -1])

# bundle input data for communityModel
data_list <- list(ylist = ylist,
  siteCovs = sitecovs,
  obsCovs = list(effort = DetHist_list[[1]]$effort))

# create community model for JAGS
modelfile1 <- tempfile(fileext = ".txt")
mod.jags <- communityModel(data_list,
  occuCovs = list(fixed = "utm_y", ranef = "elevation"),
  detCovsObservation = list(fixed = "effort"),
  intercepts = list(det = "ranef", occu = "ranef"),
  modelFile = modelfile1)

summary(mod.jags)

# fit in JAGS
fit.jags <- fit(mod.jags,
  n.iter = 1000,
  n.burnin = 500,
  chains = 3)

summary(fit.jags)
```
# response curves (= marginal effect plots)
plot_effects(mod.jags,
    fit.jags,
    submodel = "state")
plot_effects(mod.jags,
    fit.jags,
    submodel = "det")

# effect sizes plot
plot_coef(mod.jags,
    fit.jags,
    submodel = "state")
plot_coef(mod.jags,
    fit.jags,
    submodel = "det")

# create community model for Nimble
modelfile2 <- tempfile(fileext = ".txt")
mod.nimble <- communityModel(data_list,
    occuCovs = list(fixed = "utm_x", ranef = "utm_y"),
    detCovsObservation = list(fixed = "effort"),
    intercepts = list(det = "ranef", occu = "ranef"),
    modelFile = modelfile2,
    nimble = TRUE)  # set nimble = TRUE

# load nimbleEcology package
# currently necessary to do explicitly, to avoid additional package dependencies
require(nimbleEcology)

# fit uncompiled model in Nimble
fit.nimble.uncomp <- fit(mod.nimble,
    n.iter = 10,
    chains = 1)

# fit compiled model in Nimble
fit.nimble.comp <- fit(mod.nimble,
    n.iter = 5000,
    n.burnin = 2500,
    chains = 3,
    compile = TRUE)

# parameter summary statistics
summary(fit.nimble.comp)

# response curves (= marginal effect plots)
plot_effects(mod.nimble,
    fit.nimble.comp,
    submodel = "state")
plot_effects(mod.nimble,
    fit.nimble.comp,
    submodel = "det")
```
# effect sizes plot
plot_coef(mod.nimble,
          fit.nimble.comp,
          submodel = "state")
plot_coef(mod.nimble,
          fit.nimble.comp,
          submodel = "det")

# traceplots
plot(fit.nimble.comp)

## End(Not run)
```

---

**createSpeciesFolders**  
Create species directories for species identification

### Description

This function creates species subdirectories within station directories. They can be used for species identification by manually moving images into the respective species directories. The function can also delete empty species directories (if species were not detected at sites). It is not necessary to run this function if animals will be identified by metadata tagging.

### Usage

```r
createSpeciesFolders(inDir, hasCameraFolders, species, removeFolders = FALSE)
```

### Arguments

- **inDir**: character. Directory containing camera trap images sorted into station subdirectories (e.g. `inDir/StationA/`)
- **hasCameraFolders**: logical. Do the station directories in `inDir` have camera-subdirectories (e.g. `inDir/StationA/CameraA1; inDir/StationA/CameraA2`)?
- **species**: character. names of species directories to be created in every station (or station/camera) subdirectory of `inDir`
- **removeFolders**: logical. Indicating whether to create (TRUE) or remove (FALSE) species directories.

### Details

This function should be run after `imageRename`. Empty directories can be created as containers for species identification if images are identified with the drag & drop method. After species identification is complete, empty species directories can be deleted using `removeFolders = TRUE`. The function will delete only directories which are specified in `species`. If `hasCameraFolders` was
createSpeciesFolders

set to TRUE in function imageRename, hasCameraFolders must be set to TRUE here too. Species
directories will then be created within each camera subdirectory of each station directory. if the
user wishes to identify species by metadata tagging, running this function is not needed.

Value

A data.frame with directory names and an indicator for whether directories were created or
deleted.

Author(s)

Juergen Niedballa

Examples

## Not run:

# create dummy directories for tests
# (normally, you'd use directory containing renamed, unsorted images)

# this will be used as inDir
wd_createDirTest <- file.path(getwd(), "createSpeciesFoldersTest")

# now we create 2 station subdirectories
dirs_to_create <- file.path(wd_createDirTest, c("StationA", "StationB"))
sapply(dirs_to_create, FUN = dir.create, recursive = TRUE)

# species names for which we want to create subdirectories
species <- c("Sambar Deer", "Bay Cat")

# create species subdirectories
SpecFolderCreate1 <- createSpeciesFolders (inDir = wd_createDirTest,
species = species,
hasCameraFolders = FALSE,
removeFolders = FALSE)

SpecFolderCreate1

# check if directories were created
list.dirs(wd_createDirTest)

# delete empty species directories
SpecFolderCreate2 <- createSpeciesFolders (inDir = wd_createDirTest,
species = species,
hasCameraFolders = FALSE,
removeFolders = TRUE)

SpecFolderCreate2

# check if species directories were deleted
list.dirs(wd_createDirTest)
createStationFolders  
Create camera trap station directories for raw camera trap images

Description

This function creates camera trap station directories, if needed with camera subdirectories. They can be used as an initial directory structure for storing raw camera trap images.

Usage

createStationFolders(inDir, stations, cameras, createinDir)

Arguments

inDir character. Directory in which station directories are to be created
stations character. Station IDs to be used as directory names within inDir
cameras character. Camera trap IDs to be used as subdirectory names in each station directory (optionally)
createinDir logical. If inDir does not exist, create it?

Details

The empty directories serve as containers for saving raw camera trap images. If more than 1 camera was set up at a station, specifying cameras is required in order to keep images from different cameras separate. Otherwise, generic filenames (e.g., IMG0001.JPG) from different cameras may lead to accidental overwriting of images if images from these cameras are saved in one station directory.

Value

A data.frame with station (and possibly camera) directory names and an indicator for whether they were created successfully.

Author(s)

Juergen Niedballa
Examples

```r
## Not run:

# create dummy directory for tests (this will be used as inDir)
# (normally, you'd set up an empty directory, e.g. .../myStudy/rawImages)
wd_createStationDir <- file.path(tempdir(), "createStationFoldersTest")

# now we load the sample camera trap station data frame
data(camtraps)

# create station directories in wd_createStationDir
StationFolderCreate1 <- createStationFolders (inDir = wd_createStationDir,
                                          stations = as.character(camtraps$Station),
                                          createinDir = TRUE)

StationFolderCreate1

# check if directories were created
listdirs(wd_createStationDir)

## End(Not run)
```

---

detectionHistory  

Species detection histories for occupancy analyses

Description

This function generates species detection histories that can be used in occupancy analyses, e.g. with package `unmarked`. It generates detection histories in different formats, with adjustable occasion length and occasion start time.

Usage

```r
detectionHistory(
  recordTable,  
species,  
camOp,  
output = c("binary", "count"),  
stationCol = "Station",  
speciesCol = "Species",  
recordDateTimeCol = "DateTimeOriginal",  
recordDateTimeFormat = "ymd HMS",  
occasionLength,  
minActiveDaysPerOccasion,  
maxNumberOfDays,
```
day1,
buffer,
includeEffort = TRUE,
scaleEffort = FALSE,
occasionStartTime = "deprecated",
datesAsOccasionNames = FALSE,
timeZone,
writecsv = FALSE,
outDir,
unmarkedMultFrameInput
)

Arguments

recordTable  data.frame. the record table created by recordTable
species  character. the species for which to compute the detection history
camOp  The camera operability matrix as created by cameraOperation
output  character. Return binary detections ("binary") or counts of detections ("count")
stationCol  character. name of the column specifying Station ID in recordTable
speciesCol  character. name of the column specifying species in recordTable
recordDateTimeCol  character. name of the column specifying date and time in recordTable
recordDateTimeFormat  character. Format of column recordDateTimeCol in recordTable
occasionLength  integer. occasion length in days
minActiveDaysPerOccasion  integer. minimum number of active trap days for occasions to be included (optional)
maxNumberDays  integer. maximum number of trap days per station (optional)
day1  character. When should occasions begin: station setup date ("station"), first day of survey ("survey"), a specific date (e.g. "2015-12-31")?
buffer  integer. Makes the first occasion begin a number of days after station setup. (optional)
includeEffort  logical. Compute trapping effort (number of active camera trap days per station and occasion)?
scaleEffort  logical. scale and center effort matrix to mean = 0 and sd = 1?
occasionStartTime  (DEPRECATED) integer. time of day (the full hour) at which to begin occasions. Please use argument occasionStartTime in cameraOperation instead.
datesAsOccasionNames  If day1 = "survey", occasion names in the detection history will be composed of first and last day of that occasion.
timeZone  character. Must be a value returned by OlsonNames
writecsv  logical. Should the detection history be saved as a .csv?
OutDir: character. Directory into which detection history .csv file is saved

unmarkedMultFrameInput: logical. Return input for multi-season occupancy models in unmarked (argument "y" in unmarkedMultFrame?)

Details

The function computes a species detection matrix, either as a detection-by-date or a detection-by-occasion matrix. day1 defines if each stations detection history will begin on that station’s setup day (day1 = "station") or if all station’s detection histories have a common origin (the day the first station was set up if day1 = "survey" or a fixed date if, e.g. day1 = "2015-12-31"). If day1 is a date, as.Date must be able to understand it. The most suitable format is "YYYY-MM-DD", e.g. "2015-12-31".

output is analogous to spatialDetectionHistory. It makes the function return either counts of detections during occasions, or a binary indicator for whether the species was detected.

includeEffort controls whether an additional effort matrix is computed or not. This also affects the detection matrices. If includeEffort = FALSE, all occasions in which a station was not set up or malfunctioning (NA or 0 in camOp) will result in NAs in the detection history. If includeEffort = TRUE, the record history will only contain 0 and 1, and no NAs. The effort matrix can then be included in occupancy models as a (continuous) observation covariate to estimate the effect of effort on detection probability.

The number of days that are aggregated is controlled by occasionLength. occasionStartTime will be removed from the function. It has moved to cameraOperation, to ensure daily effort is computed correctly and takes the occasion start time into account. default). This may be relevant for nocturnal animals, in which 1 whole night would be considered an occasion.

The values of stationCol in recordTable must be matched by the row names of camOp (case-insensitive), otherwise an error is raised.

recordDateTimeFormat defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". recordDateTimeFormat can be interpreted either by base-R via strptime or in lubridate via parse_date_time (argument "orders"). lubridate will be used if there are no "%" characters in recordDateTimeFormat. For "YYYY-MM-DD HH:MM:SS", recordDateTimeFormat would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see strptime or parse_date_time.

If the camera operation matrix (camOp) was created for a multi-season study (argument sessionCol in cameraOperation was set, it will be detected automatically. Output can be for unmarkedMultFrame by setting unmarkedMultFrameInput = TRUE. Each row corresponds to a site, and the columns are in season-major, occasion-minor order, e.g. season1-occasion1, season1-occasion2, etc.).

Value

Depending on the value of includeEffort and scaleEffort, a list with either 1, 2 or 3 elements. The first element is the species detection history. The second is the optional effort matrix and the third contains the effort scaling parameters.
detectionHistory

- **detection_history**: A species detection matrix.
- **effort**: A matrix giving the number of active camera trap days per station and occasion (= camera trapping effort). It is only returned if `includeEffort = TRUE`.
- **effort_scaling_parameters**: Scaling parameters of the effort matrix. It is only returned if `includeEffort` and `scaleEffort` are `TRUE`.

**Warning**

Setting `output = "count"` returns a count of detections, not individuals. We strongly advise against using it as input for models of animal abundance (such as N-Mixture models) models which use counts as input.

Please note the section about defining argument `timeZone` in the vignette on data extraction (accessible via vignette("DataExtraction") or online ([https://cran.r-project.org/package=camtrapR/vignettes/camtrapr3.pdf](https://cran.r-project.org/package=camtrapR/vignettes/camtrapr3.pdf)).

**Author(s)**

Juergen Niedballa

**Examples**

```r
# define image directory
wd_images_ID <- system.file("pictures/sample_images_species_dir", package = "camtrapR")

# load station information
data(camtraps)

# create camera operation matrix
camop_no_problem <- cameraOperation(CTtable = camtraps, 
stationCol = "Station", 
setupCol = "Setup_date", 
retrievalCol = "Retrieval_date", 
hasProblems = FALSE, 
dateFormat = "dmy"
)

## Not run:
if (Sys.which("exiftool") != ""){
  # only run this function if ExifTool is available
  recordTableSample <- recordTable(inDir = wd_images_ID, 
    IDfrom = "directory", 
    minDeltaTime = 60, 
    deltaTimeComparedTo = "lastRecord", 
    exclude = "UNID", 
    timeZone = "Asia/Kuala_Lumpur"
}
```

## End(Not run)

```r
data(recordTableSample) # load the record history, as created above

# compute detection history for a species

# without trapping effort
DetHist1 <- detectionHistory(recordTable = recordTableSample,
camOp = camop_no_problem,
stationCol = "Station",
speciesCol = "Species",
recordDateTimeCol = "DateTimeOriginal",
species = "VTA",
occasionLength = 7,
day1 = "station",
datesAsOccasionNames = FALSE,
includeEffort = FALSE,
timeZone = "Asia/Kuala_Lumpur"
)

DetHist1  # this is a list with 1 element
DetHist1$detection_history  # this is the contained detection/non-detection matrix

# with effort / using base R to define recordDateTimeFormat
DetHist2 <- detectionHistory(recordTable = recordTableSample,
camOp = camop_no_problem,
stationCol = "Station",
speciesCol = "Species",
recordDateTimeCol = "DateTimeOriginal",
species = "VTA",
occasionLength = 7,
day1 = "station",
datesAsOccasionNames = FALSE,
includeEffort = TRUE,
scaleEffort = FALSE,
timeZone = "Asia/Kuala_Lumpur"
)

DetHist2$detection_history  # detection history (alternatively, use: DetHist2[[1]])
DetHist2$effort  # effort (alternatively, use: DetHist2[[2]])

# with effort / using lubridate package to define recordDateTimeFormat
DetHist2_lub <- detectionHistory(recordTable = recordTableSample,
camOp = camop_no_problem,
stationCol = "Station",
speciesCol = "Species",
recordDateTimeCol = "DateTimeOriginal",
recordDateTimeFormat = "ymd HMS",
species = "VTA",
occasionLength = 7,
day1 = "station",
datesAsOccasionNames = FALSE,
)

DetHist2_lub$detection_history  # detection history (alternatively, use: DetHist2_lub[[1]])
DetHist2_lub$effort  # effort (alternatively, use: DetHist2_lub[[2]])
```
detectionMaps

includeEffort = TRUE,
scaleEffort = FALSE,
timeZone = "Asia/Kuala_Lumpur"
)

DetHist2_lub$detection_history # detection history (alternatively, use: DetHist2_lub[[1]])
DetHist2_lub$effort # effort (alternatively, use: DetHist2_lub[[2]])

# multi-season detection history

# load multi-season data
data(camtrapsMultiSeason)
data(recordTableSampleMultiSeason)

# multi-season camera operation matrix
camop_season <- cameraOperation(CTtable = camtrapsMultiSeason,
                          stationCol = "Station",
                          setupCol = "Setup_date",
                          sessionCol = "session",
                          retrievalCol = "Retrieval_date",
                          hasProblems = TRUE,
                          dateFormat = "dmy"
)

# multi-season detection history
DetHist_multi <- detectionHistory(recordTable = recordTableSampleMultiSeason,
                          camOp = camop_season,
                          stationCol = "Station",
                          speciesCol = "Species",
                          species = "VTA",
                          occasionLength = 10,
                          day1 = "station",
                          recordDateTimeCol = "DateTimeOriginal",
                          includeEffort = TRUE,
                          scaleEffort = FALSE,
                          timeZone = "UTC",
                          unmarkedMultFrameInput = TRUE
)

DetHist_multi

detectionMaps  Generate maps of observed species richness and species presences by station

Description

Generates maps of observed species richness and species presence by species and station. Output can be R graphics, PNG graphics or a shapefile for use in GIS software.
Usage

detectionMaps(
  CTtable, recordTable,
  Xcol, Ycol,
  backgroundPolygon, stationCol = "Station",
  speciesCol = "Species",
  speciesToShow, richnessPlot = TRUE,
  speciesPlots = TRUE, addLegend = TRUE, printLabels = FALSE,
  smallPoints, plotR = TRUE, writePNG = FALSE, plotDirectory,
  createPlotDir = FALSE, pngMaxPix = 1000, writeShapefile = FALSE,
  shapefileName, shapefileDirectory, shapefileProjection
)

Arguments

CTtable data.frame. contains station IDs and coordinates
recordTable data.frame. the record table created by recordTable
Xcol character. name of the column specifying x coordinates in CTtable
Ycol character. name of the column specifying y coordinates in CTtable
backgroundPolygon SpatialPolygons or SpatialPolygonsDataFrame. Polygon to be plotted in the background of the map (e.g. project area boundary)
stationCol character. name of the column specifying station ID in CTtable and recordTable
speciesCol character. name of the column specifying species in recordTable
speciesToShow character. Species to include in the maps. If missing, all species in recordTable will be included.
richnessPlot logical. Generate a species richness plot?
speciesPlots logical. Generate plots of all species number of independent events?
addLegend logical. Add legends to the plots?
printLabels logical. Add station labels to the plots?
smallPoints numeric. Number by which to decrease point sizes in plots (optional).
plotR logical. Create plots in R graphics device?
writePNG logical. Create PNGs of the plots?
plotDirectory character. Directory in which to save the PNGs
createPlotDir logical. Create plotDirectory?
pngMaxPix integer. number of pixels in pngs on the longer side
writeShapefile logical. Create a shapefile from the output?
shapefileName character. Name of the shapefile to be saved. If empty, a name will be generated automatically.
shapefileDirectory character. Directory in which to save the shapefile.
shapefileProjection character. A character string of projection arguments to use in the shapefile.

Details

The column name stationCol must be identical in CTtable and recordTable and station IDs must match.
Shapefile creation depends on the packages sf. Argument shapefileProjection must be a valid argument of st_crs (one of (i) character: a string accepted by GDAL, (ii) integer, a valid EPSG value (numeric), or (iii) an object of class crs. If shapefileProjection is undefined, the resulting shapefile will lack a coordinate reference system.

Value

An invisible data.frame with station coordinates, numbers of events by species at each station and total species number by station. In addition and optionally, R graphics or png image files.

Author(s)

Juergen Niedballa

References

A great resource for coordinate system information is https://spatialreference.org/. Use the Proj4 string as shapefileProjection argument.

Examples

# load station information
data(camtraps)

# load record table
data(recordTableSample)

# create maps
```r
Mapstest <- detectionMaps(CTtable = camtraps, 
recordTable = recordTableSample, 
Xcol = "utm_x", 
Ycol = "utm_y", 
stationCol = "Station", 
speciesCol = "Species", 
writePNG = FALSE, 
plotR = TRUE, 
printLabels = TRUE, 
richnessPlot = TRUE, 
addLegend = TRUE 
)
# with a polygon in the background, and for one species only
# make a dummy polygon for the background
library(sf)
Sr1 = st_polygon(list(cbind(c(521500,526500,527000, 521500, 521500), 
c(607500, 608000, 603500, 603500, 607500))))
aoi <- data.frame(name = "My AOI")
st_geometry(aoi) <- st_geometry(Sr1)
st_crs(aoi) <- 32650 # assign CRS: UTM50N
Mapstest2 <- detectionMaps(CTtable = camtraps, 
recordTable = recordTableSample, 
Xcol = "utm_x", 
Ycol = "utm_y", 
backgroundPolygon = aoi, # this was added 
speciesToShow = c("PBE", "VTA"), # this was added 
stationCol = "Station", 
speciesCol = "Species", 
writePNG = FALSE, 
plotR = TRUE, 
printLabels = TRUE, 
richnessPlot = TRUE, 
addLegend = TRUE 
)
```

---

**exifTagNames**

*Show Exif metadata of JPEG images or other image or video formats*
Description

The function will return metadata values, metadata tag names and group names of Exif metadata of JPEG images or other formats.

Usage

```r
exifTagNames(
  inDir,  
  whichSubDir = 1,  
  fileName,  
  returnMetadata = "DEPRECATED",  
  returnTagGroup = "DEPRECATED"
)
```

Arguments

- `inDir` character. Directory containing camera trap images sorted into station subdirectories (e.g. `inDir/StationA/`)
- `whichSubDir` integer or character. Either number or name of subdirectory of `inDir` in which to look for an image
- `fileName` character. A filename, either the file name of an image in `inDir` or a full path with file name (in which case `inDir` is not needed)
- `returnMetadata` deprecated and ignored
- `returnTagGroup` deprecated and ignored

Details

Many digital cameras record information such as ambient temperature or moon phase under makerspecific tag names in Exif metadata of JPEG images. In addition, many technical information are stored in Exif metadata. In order to extract those information from images and add them to the record tables created by the functions `recordTable` and `recordTableIndividual`, the tag names must be known so they can be passed to these functions via the `additionalMetadataTags` argument.

By default the function returns both metadata tag names and the metadata group they belong to (via argument `returnTagGroup`). This is helpful to unambiguously address specific metadata tags, because different groups can contain tags of identical names, which may cause problems executing the functions `recordTable` and `recordTableIndividual`. The format is "GROUP:tag", e.g. "EXIF:Flash".

Value

A data frame containing three columns: metadata tag group, tag name, and values.

Author(s)

Juergen Niedballa
filterRecordTable

Filter species record table for temporal independence

Description

Filter species record table for temporal independence

References

Phil Harvey’s ExifTool https://exiftool.org/

See Also

recordTable

Examples

## Not run:

wd_images_ID <- system.file("pictures/sample_images_species_dir", package = "camtrapR")

# specify directory, camtrapR will automatically take first image from first subdirectory
exifTagNames(inDir = wd_images_ID)

# specify subdirectory by name, camtrapR will use first image
exifTagNames(inDir = wd_images_ID,
              whichSubDir = "StationA")

# specifying fileName only (line break due to R package policy)
exifTagNames(fileName = file.path(wd_images_ID, "StationC", "TRA", "StationC__2009-05-02__00-10-00(1).JPG"))

# specify inDir and fileName
exifTagNames(inDir = wd_images_ID,
              fileName = file.path("StationC", "TRA", "StationC__2009-05-02__00-10-00(1).JPG"))

# it also works this way
exifTagNames(inDir = file.path(wd_images_ID, "StationC", "TRA"),
              fileName = "StationC__2009-05-02__00-10-00(1).JPG")

# with tagged sample images
wd_images_ID_tagged <- system.file("pictures/sample_images_indiv_tag", package = "camtrapR")
exifTagNames(inDir = wd_images_ID_tagged)

## End(Not run)
Usage

filterRecordTable(
  recordTable, 
  minDeltaTime = 0, 
  deltaTimeComparedTo, 
  speciesCol = "Species", 
  stationCol, 
  cameraCol, 
  camerasIndependent, 
  recordDateTimeCol = "DateTimeOriginal", 
  recordDateTimeFormat = "ymd HMS", 
  removeDuplicateRecords = TRUE, 
  exclude, 
  timeZone, 
  writecsv = FALSE, 
  outDir, 
  eventSummaryColumn, 
  eventSummaryFunction, 
  quiet = FALSE
)

Arguments

recordTable  data frame as created by recordTable.
minDeltaTime  integer. Time difference between records of the same species at the same station to be considered independent (in minutes)
deltaTimeComparedTo  character. For two records to be considered independent, must the second one be at least minDeltaTime minutes after the last independent record of the same species ("lastIndependentRecord"), or minDeltaTime minutes after the last record ("lastRecord")?
speciesCol  character. name of the column specifying species in recordTable
stationCol  character. Name of the camera trap station column. Assuming "Station" if undefined.
cameraCol  character. Name of the column specifying cameras in recordTable (optional).
camerasIndependent  logical. If TRUE, species records are considered to be independent between cameras at a station.
recordDateTimeCol  character. Name of the column specifying date and time in recordTable.
recordDateTimeFormat  character. Format of column recordDateTimeCol in recordTable
removeDuplicateRecords  logical. If there are several records of the same species at the same station (also same camera if cameraID is defined) at exactly the same time, show only one?
exclude  character. Vector of species names to be excluded from the record table
filterRecordTable

TimeZone character. Must be a value returned by OlsonNames.
writecsv logical. Should the record table be saved as a .csv?
outDir character. Directory to save csv to. If NULL and writecsv = TRUE, recordTable will be written to inDir.

eventSummaryColumn character. A column in the record table (e.g. from a metadata tag) by to summarise non-independent records (those within minDeltaTime of a given record) with a user-defined function (eventSummaryFunction)
eventSummaryFunction character. The function by which to summarise eventSummaryColumn of non-independent records, e.g. "sum", "max" (optional)
quiet logical. If TRUE, suppress printing of progress.

Value
A data frame containing species records and additional information about stations, date, time, filtered for temporal independence.

Author(s)
Juergen Niedballa

Examples

if (Sys.which("exiftool") != ""){    # only run example if ExifTool is available

    # set directory with camera trap images in station directories
    wd_images_ID_species <- system.file("pictures/sample_images_species_dir",
                                        package = "camtrapR")

    # create record table without temporal filtering
    rec_table <- recordTable(inDir = wd_images_ID_species,
                              IDfrom = "directory",
                              minDeltaTime = 0,
                              exclude = "UNID",
                              timeZone = "Asia/Kuala_Lumpur",
                              removeDuplicateRecords = TRUE)

    # filter for 60 minutes temporal independence
    rec_table_filt <- filterRecordTable(recordTable = rec_table,
                                          minDeltaTime = 60,
                                          stationCol = "Station",
                                          deltaTimeComparedTo = "lastIndependentRecord")

    nrow(rec_table)
    nrow(rec_table_filt)
}


fit,commOccu-method  

Fit a community (multi-species) occupancy model

Description

Convenience function for fitting community occupancy models (defined in a commOccu object) in JAGS or Nimble.

Usage

## S4 method for signature 'commOccu'
fit(
  object,
  n.iter = 100,
  thin = 1,
  n.burnin = n.iter/2,
  n.adapt = 0,
  chains = 3,
  inits = NULL,
  compile = TRUE,
  WAIC = FALSE,
  quiet = FALSE,
  ...
)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>object</td>
<td>commOccu object</td>
</tr>
<tr>
<td>n.iter</td>
<td>number of iterations to monitor</td>
</tr>
<tr>
<td>thin</td>
<td>thinning interval for monitors</td>
</tr>
<tr>
<td>n.burnin</td>
<td>burnin length. Defaults to half of n.iter.</td>
</tr>
<tr>
<td>n.adapt</td>
<td>Length of adaptive phase</td>
</tr>
<tr>
<td>chains</td>
<td>number of MCMC chains to run</td>
</tr>
<tr>
<td>inits</td>
<td>named list. Initial values to use. If NULL (default), the values from the inits function in object are used.</td>
</tr>
<tr>
<td>compile</td>
<td>logical. If Nimble model, compile model with compileNimble before running model?</td>
</tr>
<tr>
<td>WAIC</td>
<td>logical. Return WAIC (only Nimble models)</td>
</tr>
<tr>
<td>quiet</td>
<td>if TRUE messages and progress bar will be suppressed</td>
</tr>
<tr>
<td>...</td>
<td>additional arguments to pass to runMCMC (only relevant for Nimble)</td>
</tr>
</tbody>
</table>
Details

Models will be fit either in JAGS or Nimble, depending on the decision made in the `nimble` argument in `communityModel`.

For Nimble, compilation is strongly recommended for long model runs. Uncompiled models can run extremely slow. Compilation itself can take a while also, and requires that Rtools is available on the system.

This is a convenience function only which hides some of the configuration options. If you require more control over model fitting, you can run all steps individually. See vignette 5 for details.

Value

A `coda::mcmc.list`

---

Description

Some camera models don’t store the date/time information in the standard Exif metadata tag. Consequently, camtrapR cannot find that information. This function uses Exiftool to update the DateTimeOriginal metadata tag in all images within a directory to make them readable with camtrapR (and other software).

Usage

`fixDateTimeOriginal(inDir, recursive = TRUE)`

Arguments

- `inDir` character. Name of the directory containing images to be fixed
- `recursive` logical. Recursively find images in subdirectories of `inDir`?

Details

Some Reconyx Hyperfire cameras (e.g. HC500) are known to show this problem.

Value

Returns invisibly the messages returned by the Exiftool call (warnings etc.).

Warning

Please make a backup of your images before running this function.
getSpeciesImages

**Author(s)**

Juergen Niedballa

**References**

This function uses the code from:

**Examples**

```r
## Not run:
# a hypothetical example

wd_images_hyperfire <- "C:/Some/Directory"

fixDateTimeOriginal(inDir = wd_images_hyperfire,
                    recursive = TRUE)

## End(Not run)
```

---

**getSpeciesImages**  
*Collect all images of a species*

**Description**

This function will fetch all images of a particular species from all camera trap stations and copies these images to a new location. The images which are to be copied are found in one of 2 possible ways, 1) by providing an existing record table (created with `recordTable`) or 2) by reading species IDs from species directories or from metadata (calling ExifTool). Earlier in the workflow, i.e., before running this function, images should have been renamed (with `imageRename`) to give images unique file names based on station ID and date/time.

**Usage**

```r
getSpeciesImages(  
      species,  
      recordTable,  
      speciesCol = "Species",  
      stationCol = "Station",  
      inDir,  
      outDir,  
      createStationSubfolders = FALSE,  
      IDfrom,  
      metadataSpeciesTag,  
      metadataHierarchyDelimiter = "|"
    )
```
**getSpeciesImages**

**Arguments**

- **species** character. Species whose images are to be fetched.
- **recordTable** data frame. A data frame as returned by function `recordTable`. If you specify this argument, do not specify `inDir`.
- **speciesCol** character. Name of the column specifying species ID in `recordTable`. Only required if `recordTable` is defined.
- **stationCol** character. Name of the column specifying station ID in `recordTable`. Only required if `recordTable` is defined.
- **inDir** character. Directory containing identified (species level) camera trap images sorted into station subdirectories (e.g. `inDir/StationA`). If you specify this argument, do not specify `recordTable`.
- **outDir** character. Directory in which to save species images. A species subdirectory will be created in `outDir` automatically.
- **createStationSubfolders** logical. Save images in station directories within the newly created species directory in `outDir`?
- **IDfrom** character. Read species ID from image metadata ("metadata") or from species directory names ("directory")? Only required if `inDir` is defined.
- **metadataSpeciesTag** character. The species ID tag name in image metadata (if IDfrom = "metadata"). Only required if `inDir` is defined.
- **metadataHierarchyDelimiter** character. The character delimiting hierarchy levels in image metadata tags in field "HierarchicalSubject". Either "|" or ":" (if IDfrom = "metadata"). Only required if `inDir` is defined and IDfrom = "metadata".

**Details**

The function finds the images to be copied by either consulting a record table created with `recordTable` or by reading species IDs from images. The former is considerably faster because ExifTool is not called, but requires images to be in precisely the location given by the columns `Directory` and `FileName` in `recordTable`. To use this feature, provide the function with a record table in argument `recordTable`.

If you’d rather read species IDs from images within the function (to make sure all file paths are correct), images need to be in the directory structure required by the package, e.g.

```
> inDir/Station/Species
```

or

```
> inDir/Station/Camera/Species
```

if using species directories for species IDs, and

```
> inDir/Station
```

or

```
> inDir/Station/Camera
```
if reading IDs from species metadata tags. In the latter case, only station directories are needed. In any case, the argument species must match species IDs (either the speciesCol in recordTable, species directory names or species metadata tags).

Before running the function, first rename the images using function `imageRename` to provide unique file names and prevent several images from having the same name (if generic names like "IMGP0001.jpg" are used). The function will not copy images if there are duplicate filenames to prevent overwriting images unintentionally.

**Value**

A `data.frame` with old and new directories and file names and the copy status (`copy_ok`; TRUE if copying was successful, FALSE if not).

**Author(s)**

Juergen Niedballa

**Examples**

```r
## Not run:
# define image directory
wd_images_ID <- system.file("pictures/sample_images_species_dir", package = "camtrapR")
wd_images_ID_copy <- file.path(tempdir(), "sample_images_species_dir")

species_to_copy <- "VTA"  # = Viverra tangalunga, Malay Civet

specImagecopy <- getSpeciesImages(species = species_to_copy,
                                  inDir = wd_images_ID,
                                  outDir = wd_images_ID_copy,
                                  createStationSubfolders = FALSE,
                                  IDfrom = "directory"
)

## End(Not run)
```

---

**Description**

The function renames and copies raw camera trap images into a new location where they can be identified. Images are renamed with camera trap station ID, camera ID (optional), creation date and a numeric identifier for images taken within one minute of each other at a given station. Station ID and camera ID are derived from the raw image directory structure. The creation date is extracted from image metadata using ExifTool.
Usage

imageRename(
  inDir,
  outDir,
  hasCameraFolders,
  keepCameraSubfolders,
  createEmptyDirectories = FALSE,
  copyImages = FALSE,
  writecsv = FALSE
)

Arguments

inDir character. Directory containing camera trap images sorted into station subdirectories (e.g. inDir/StationA/)
outDir character. Directory into which the renamed images will be copied
hasCameraFolders logical. Do the station directories in inDir have camera subdirectories (e.g. "inDir/StationA/Camera1")?
keepCameraSubfolders logical. Should camera directories be preserved as subdirectories of outDir (e.g. "outDir/StationA/CameraA1")?
createEmptyDirectories logical. If station or camera directories are empty, should they be copied nevertheless (causing empty directories in outDir, but preserving the whole directory structure)?
copyImages logical. Copy images to outDir?
writecsv logical. Save a data frame with a summary as a .csv? The csv will be saved in outDir.

Details

Setting up the correct raw image directory structure is necessary for running the function successfully. inDir is the main directory that contains camera trap station subdirectories (e.g. inDir/StationA). If one camera was deployed per station and no camera subdirectories are used within station directories, hasCameraFolders can be set to FALSE. If more than one camera was deployed at stations, there must be subdirectories for the individual camera traps within the station directories (e.g. "inDir/StationA/CameraA1" and "inDir/StationA/CameraA2"). Even if only some stations had multiple cameras, all station will need camera subdirectories. The argument hasCameraFolders must be TRUE. Within the camera subdirectories, the directory structure is irrelevant.

Renaming of images follows the following pattern: If hasCameraFolders is TRUE, it is: "StationID__CameraID__Date__Time(Number).JPG", e.g. "StationA__CameraA1__2015-01-31__18-59-59(1).JPG". If hasCameraFolders is FALSE, it is: "StationID__Date__Time(Number).JPG", e.g. "StationA__2015-01-31__18-59-59(1).JPG".

The purpose of the number in parentheses is to prevent assigning identical file names to images taken at the same station (and camera) in the same second, as can happen if cameras take sequences
of images. It is a consecutive number given to all images taken at the same station by the same camera within one minute. The double underscore "__" in the image file names is for splitting and extracting information from file names in other functions (e.g. for retrieving camera IDs in `recordTable` if camera subdirectories are not preserved (keepCameraSubfolders = FALSE)).

The function finds all JPEG images and extracts the image timestamp from the image metadata using ExifTool and copies the images (with new file names) into `outDir`, where it will set up a directory structure based on the station IDs and, if required by `keepCameraSubfolders = TRUE`, camera IDs (e.g. `outDir/StationA/` or `outDir/StationA/CameraA1`).

copyImages can be set to FALSE to simulate the renaming and check the file names of the renamed images without copying. If you are handling large number of images (>e.g., 100,000), the function may take some time to run.

**Value**

A data.frame with original directory and file names, new directory and file names and an indicator for whether images were copied successfully.

**Author(s)**

Juergen Niedballa

**References**

Phil Harvey’s ExifTool [https://exiftool.org/](https://exiftool.org/)

**Examples**

```r
## Not run:
### "trial" run. create a table with file names after renaming, but don't copy images.

# first, find sample image directory in package directory:
wd_images_raw <- system.file("pictures/raw_images", package = "camtrapR")

# because copyImages = FALSE, outDir does not need to be defined
renaming.table <- imageRename(inDir = wd_images_raw, 
    hasCameraFolders = FALSE, 
    copyImages = FALSE, 
    writecsv = FALSE)

### a real example in which images are copied and renamed

# define raw image location
wd_images_raw <- system.file("pictures/raw_images", package = "camtrapR")
```
# define destination for renamed images
wd_images_raw_renamed <- file.path(tempdir(), "raw_images_renamed")

# now we have to define outDir because copyImages = TRUE
renaming.table2 <- imageRename(inDir = wd_images_raw, outDir = wd_images_raw_renamed, hasCameraFolders = FALSE, copyImages = TRUE, writecsv = FALSE)

# show output files
list.files(wd_images_raw_renamed, recursive = TRUE)

# output table
renaming.table2

## End(Not run)

---

**OCRdataFields**

*Optical character recognition (OCR) from data fields in digital images*

**Description**

Extracts information from the data fields in camera trap images (not the metadata). Many camera traps include data fields in camera trap images, often including date and time of images, and sometimes other information. This function extracts the information from these fields using optical character recognition provided by the package `tesseract` after reading images using the package `magick`.

**Usage**

```
OCRdataFields(inDir, geometries, invert = FALSE)
```

**Arguments**

- `inDir` character. Directory containing camera trap images (or subdirectories containing images)
- `geometries` list. A (possibly named) list of geometry strings defining the image area(s) to extract.
- `invert` logical. Invert colors in the image? Set to TRUE if text in data field is white on black background. Leave if FALSE if text is black in white background.
Details

Normally all these information should be in the image metadata. This function is meant as a last resort if image metadata are unreadable or were removed from images. OCR is not perfect and may misidentify characters, so check the output carefully.

The output of this function can be used in `writeDateTimeOriginal` to write date/time into the DateTimeOriginal tag in image metadata, making these images available for automatic processing with `recordTable` and other functions that extract image metadata.

This function reads all images in `inDir` (including subdirectories), crops them to the geometries in the "geometries" list, and performs optical character recognition (OCR) on each of these fields (leveraging the magick and tesseract packages).

Geometries are defined with `geometry_area` from `magick`. See `geometry` for details on how to specify geometries with `geometry_area`. The format is: "widthxheight+x_off+y_off", where:

- **width** width of the area of interest
- **height** height of the area of interest
- **x_off** offset from the left side of the image
- **y_off** offset from the top of the image

Units are pixels for all fields. digiKam can help in identifying the correct specification for geometries. Open the Image Editor, left-click and draw a box around the data field of interest. Ensure the entire text field is included inside the box, but nothing else. Now note two pairs of numbers at the bottom of the window, showing the offsets and box size as e.g.:

"(400, 1800) (300 x 60)"

This corresponds to the geometry values as follows:

"(x_off, y_off) (width x height)"

Using these values, you’d run:

```r
gamey_area(x_off = 400, y_off = 1800, width = 300, height = 60)
```

and receive

"300x60+400+1800"

as your geometry.

OCR in tesseract has problems with white font on black background. If that is the case in your images, set `invert` to `TRUE` to invert the image and ensure OCR uses black text on white background.

Even then, output will not be perfect. Error rates in OCR depend on multiple factors, including the text size and font type used. We don’t have control over these, so check the output carefully and edit as required.

Value

A data.frame with original directory and file names, and additional columns for the OCR data of each extracted geometry.

Author(s)

Juergen Niedballa
See Also

writeDateTimeOriginal

Examples

```r
## Not run:
# dontrun is to avoid forcing users to install additional dependencies

wd_images_OCR <- system.file("pictures/full_size_for_ocr", package = "camtrapR")

library(magick)
# define geometries
geometry1 <- geometry_area(x_off = 0, y_off = 0, width = 183, height = 37)
geometry2 <- geometry_area(x_off = 196, y_off = 0, width = 200, height = 17)
geometry3 <- geometry_area(x_off = 447, y_off = 0, width = 63, height = 17)
geometry4 <- geometry_area(x_off = 984, y_off = 0, width = 47, height = 17)
geometry5 <- geometry_area(x_off = 0, y_off = 793, width = 320, height = 17)

# combine geometries into list
geometries <- list(date = geometry1, 
                   time = geometry2, 
                   sequence_id = geometry3, 
                   temperature = geometry4, 
                   camera_model = geometry5)

df_image_data <- OCRdataFields(inDir = wd_images_OCR, 
                               geometries = geometries, 
                               invert = TRUE)

df_image_data

# note the mistake in "camera_model"
# it should be "PC850", not "PC8S00"
# date and time are correct though

## End(Not run)
```
Usage

```r
## S4 method for signature 'commOccu'
plot_coef(
  object,
  mcmc.list,
  submodel = "state",
  ordered = TRUE,
  combine = FALSE,
  outdir,
  level = c(outer = 0.95, inner = 0.75),
  colorby = "significance",
  scales = "free_y",
  community_lines = FALSE,
  ...
)
```

Arguments

- `object` commOccu object
- `mcmc.list` mcmc.list. Output of `fit` called on a `commOccu` object
- `submodel` character. Submodel to get plots for. Can be "det" or "state"
- `ordered` logical. Order species in plot by median effect (TRUE) or by species name (FALSE)
- `combine` logical. Combine multiple plots into one plot (via facets)?
- `outdir` character. Directory to save plots to (optional)
- `level` numeric. Probability mass to include in the uncertainty interval (two values named "outer" and "inner", in that order). Second value (= inner interval) will be plotted thicker.
- `colorby` character. Whether to color estimates by "significance" (of the effect estimates), or "Bayesian p-value" (of the species). Currently allows only "significance".
- `scales` character. Passed to `facet_grid`. Can be "free" to scale x axes of effect estimates independently, or "free_y" to scale all x axes identically.
- `community_lines` logical. Add faint vertical lines to the plot indicating median community effect (solid line) and its confidence interval (first value from `level`).
- `...` additional arguments for `ggsave` - only relevant if `outdir` is defined.

Details

Users who wish to create their own visualizations can use the data stored in the ggplot output. It is accessed via e.g. `output$covariate_name$data`

Value

A list of ggplot objects (one list item per covariate).
plot_effects,commOccu-method

Plot Marginal Effects of Covariates

Description

Plot marginal effect plots (= response curves if covariates are continuous) for all species in a community (multi-species) occupancy model. Takes into account species-specific intercepts (if any). Currently only supports continuous covariates, not categorical covariates.

Usage

```r
## S4 method for signature 'commOccu'
plot_effects(
  object,
  mcmc.list,
  submodel = "state",
  response = "occupancy",
  draws = 1000,
  outdir,
  level = 0.95,
  keyword_quadratic = "_squared",
  ...
)
```

Arguments

- **object**: commOccu object
- **mcmc.list**: mcmc.list. Output of `fit` called on a commOccu object
- **submodel**: character. Submodel to get plots for. Can be "det" (detection submodel) or "state" (occupancy submodel)
- **response**: character. response type on y axis. Only relevant for submodel = "state". Default is "occupancy", can be set to "abundance" for Royle-Nichols models
- **draws**: integer. Number of draws from the posterior to use when generating the plots. If fewer posterior samples than specified in `draws` are available, all posterior samples are used.
- **outdir**: character. Directory to save plots to (optional)
- **level**: numeric. Probability mass to include in the uncertainty interval.
- **keyword_quadratic**: character. A suffix in covariate names in the model that indicates a covariate is a quadratic effect of another covariate which does not carry the suffix in its name (e.g. if the covariate is "elevation", the quadratic covariate would be "elevation_squared").

... additional arguments for `ggsave` - only relevant if `outdir` is defined
Details

Users who wish to create their own visualizations can use the data stored in the ggplot output. It is accessed via e.g. `output$covariate_name$data`

Value

A list of ggplot objects (one list item per covariate).

### predict,commOccu-method

**Predictions from community occupancy models**

Description

Create (spatial) predictions of species occupancy and species richness from community occupancy models and spatial rasters or covariate data frames.

Usage

```r
## S4 method for signature 'commOccu'
predict(
  object, mcmc.list, type, draws = 1000, level = 0.95, interval = c("none", "confidence"), x, aoi = NULL, speciesSubset, batch = FALSE, seed
)
```

Arguments

- **object**: commOccu object
- **mcmc.list**: mcmc.list. Output of `fit` called on a commOccu object
- **type**: character. "psi" for species occupancy estimates, "richness" for species richness estimates, "pao" for percentage of area occupied (by species), "psi_array" for raw occupancy probabilities in an array. For Royle-Nichols models, "abundance" for species abundance, or "lambda_array" for raw species abundance estimates in an array.
- **draws**: Number of draws from the posterior to use when generating the plots. If fewer than draws are available, they are all used
- **level**: Probability mass to include in the uncertainty interval
interval  Type of interval calculation. Can be "none" or "confidence" (can be abbreviated). Calculation can be slow for type = "psi" with many cells and posterior samples.

x  SpatRaster or data.frame. Must be scaled with same parameters as site covariates used in model, and have same names.

aoi  SpatRaster with same dimensions as x (if x is a SpatRaster), indicating the area of interest (all cells with values are AOI, all NA cells are ignored). If NULL, predictions are made for all cells.

speciesSubset  species to include in richness estimates. Can be index number or species names.

batch  logical or numeric. If FALSE, all raster cells / data frame rows will be processed at once (can be memory intensive). If TRUE, computation is conducted in batches of 1000. If numeric, it is the desired batch size.

seed  numeric. Seed to use in set.seed for reproducible results (ensures that draws are identical).

Details

Processing can be very memory-intensive. If memory is insufficient, use the batch parameter. This can enable processing for higher numbers of draws or very large rasters / data frames.

Value

A SpatRaster or data.frame, depending on x. If type = "pao", a list. If type = "psi_array" or "lambda_array", a 3D-array [cell, species, draw].

recordTable  Generate a species record table from camera trap images and videos

Description

Generates a record table from camera trap images or videos. Images/videos must be sorted into station directories at least. The function can read species identification from a directory structure (Station/Species or Station/Camera/Species) or from image metadata tags.

Usage

recordTable(
inDir,
IDfrom,
cameraID,
camerasIndependent,
exclude,
minDeltaTime = 0,
deltaTimeComparedTo,
timeZone,
stationCol,
Arguments

inDir character. Directory containing station directories. It must either contain images in species subdirectories (e.g. inDir/StationA/SpeciesA) or images with species metadata tags (without species directories, e.g. inDir/StationA).

IDfrom character. Read species ID from image metadata ("metadata") of from species directory names ("directory")?

cameraID character. Where should the function look for camera IDs: 'filename', 'directory'. 'filename' requires images renamed with imageRename. 'directory' requires a camera subdirectory within station directories (station/camera/species). Can be missing.

camerasIndependent logical. If TRUE, species records are considered to be independent between cameras at a station.

exclude character. Vector of species names to be excluded from the record table

minDeltaTime integer. Time difference between records of the same species at the same station to be considered independent (in minutes)

deltaTimeComparedTo character. For two records to be considered independent, must the second one be at least minDeltaTime minutes after the last independent record of the same species ("lastIndependentRecord"), or minDeltaTime minutes after the last record ("lastRecord")?

timeZone character. Must be a value returned by OlsonNames

stationCol character. Name of the camera trap station column. Assuming "Station" if undefined.

writecsv logical. Should the record table be saved as a .csv?

outDir character. Directory to save csv to. If NULL and writecsv = TRUE, recordTable will be written to inDir.

metadataHierarchyDelimiter character. The character delimiting hierarchy levels in image metadata tags in field "HierarchicalSubject". Either "|" or ":".

metadataSpeciesTag character. In custom image metadata, the species ID tag name.
additionalMetadataTags
character. Additional camera model-specific metadata tags to be extracted. (If possible specify tag groups as returned by \texttt{exifTagNames})

removeDuplicateRecords
logical. If there are several records of the same species at the same station (also same camera if cameraID is defined) at exactly the same time, show only one?

returnFileNamesMissingTags
logical. If species are assigned with metadata and images are not tagged, return a few file names of these images as a message?

eventSummaryColumn
character. A column in the record table (e.g. from a metadata tag) by to summarise non-independent records (those within \texttt{minDeltaTime} of a given record) with a user-defined function (\texttt{eventSummaryFunction})

eventSummaryFunction
character. The function by which to summarise \texttt{eventSummaryColumn} of non-independent records, e.g. "sum", "max" (optional)

video
list. Contains information on how to handle video data (optional). See details.

**Details**

The function can handle a number of different ways of storing images, and supports species identification by moving images into species directories as well as metadata tagging. In every case, images need to be stored into station directories. If images are identified by moving them into species directories, a camera directory is optional: "Station/Species/XY.JPG" or "Station/Camera/Species/XY.JPG". Likewise, if images are identified using metadata tagging, a camera directory can be used optionally: "Station/XY.JPG" or "Station/Camera/XY.JPG".

If images are identified by metadata tagging, \texttt{metadataSpeciesTag} specifies the metadata tag group name that contains species identification tags. \texttt{metadataHierarchyDelimiter} is "|" for images tagged in DigiKam and images tagged in Adobe Bridge / Lightroom with the default settings. It is only necessary to change it if the default was changed in these programs.

\texttt{minDeltaTime} is a criterion for temporal independence of species recorded at the same station. Setting it to 0 will make the function return all records. Setting it to a higher value will remove records that were taken less than \texttt{minDeltaTime} minutes after the last record (\texttt{deltaTimeComparedTo = "lastRecord"}) or the last independent record (\texttt{deltaTimeComparedTo = "lastIndependentRecord"}).

\texttt{removeDuplicateRecords} determines whether duplicate records (identical station, species, date/time, (and camera if applicable)) are all returned (FALSE) or collapsed into a single unique record (TRUE).

\texttt{camerasIndependent} defines if the cameras at a station are to be considered independent. If TRUE, records of the same species taken by different cameras are considered independent (e.g. if they face different trails). Use FALSE if both cameras face each other and possibly TRUE).

\texttt{exclude} can be used to exclude "species" directories containing irrelevant images (e.g. "team", "blank", "unidentified"). \texttt{stationCol} can be set to match the station column name in the camera trap station table (see \texttt{camtraps}).

Many digital images contain Exif metadata tags such as "AmbientTemperature" or "MoonPhase" that can be extracted if specified in \texttt{metadataTags}. Because these are manufacturer-specific and not standardized, function \texttt{exifTagNames} provides a vector of all available tag names. Multiple names
can be specified as a character vector as: \( c(\text{Tag1, Tag2, ...}) \). The metadata tags thus extracted may be used as covariates in modelling species distributions.

\texttt{eventSummaryColumn} and \texttt{eventSummaryFunction} can be used to extract summary statistics for independent sampling events. For example, you assigned a "count" tag to your images, indicating the number of individuals in a picture. In a sequence of pictures taken within 1 minute, most pictures show one individual, but one image shows two individuals. You tagged the images accordingly (count = 1 or count = 2) and run \texttt{recordTable}. Set \texttt{eventSummaryColumn} = "count" and \texttt{eventSummaryFunction} = "max" to obtain the maximum number of count in all images within \texttt{minDeltaTime} minutes of a given record. The results is in a new column, in this example \texttt{count_max}. You can also calculate several statistics at the same time, by supplying vectors of values, e.g. \texttt{eventSummaryColumn = c("count", "count", "camera")} and \texttt{eventSummaryFunction} = \( c("\text{min", "max", "unique"}) \) to get minimum and maximum count and all unique camera IDs for that event. Note that \texttt{eventSummaryColumn} and \texttt{eventSummaryFunction} must be of same length.

Argument \texttt{video} is a named list with 2 or 4 items. 2 items (\texttt{file_formats}, \texttt{dateTimeTag}) are always required, and are sufficient if \texttt{IDfrom = "directory"}. In that case, no digiKam tags will be returned. To return digiKam tags, two additional items are required (\texttt{db_directory}, \texttt{db_filename}). This is essential when using \texttt{IDfrom = "metadata"}. When using \texttt{IDfrom = "directory"}, it is optional, but allows to extract metadata tags assigned to videos in digiKam. This workaround is necessary because digiKam tags are not written into video metadata, but are only saved in the digiKam database. So in contrast to JPG images, they can not be extracted with ExifTool. It also requires that \texttt{inDir} is in your digiKam database.

The items of argument \texttt{video} are:

- \texttt{file_formats}: The video formats to extract (include "jpg" if you want JPG image metadata)
- \texttt{dateTimeTag}: the metadata tag to extract date/time from (use \texttt{exifTagNames} to find out which tag is suitable)
- \texttt{db_directory}: The directory containing digiKam database (optional if \texttt{IDfrom = "directory"})
- \texttt{db_filename}: The digiKam database file in \texttt{db_directory} (optional if \texttt{IDfrom = "directory"})

See the examples below for for how to specify the argument \texttt{video}.

**Value**

A data frame containing species records and additional information about stations, date, time and (optionally) further metadata.

**Warning**

Custom image metadata must be organised hierarchically (tag group - tag; e.g. "Species" - "Leopard Cat"). Detailed information on how to set up and use metadata tags can be found in vignette 2: Species and Individual Identification.

Custom image metadata tags must be written to the images. The function cannot read tags from .xmp sidecar files. Make sure you set the preferences accordingly. In DigiKam, go to Settings/Configure digiKam/Metadata. There, make sure "Write to sidecar files" is unchecked.

Please note the section about defining argument \texttt{timeZone} in the vignette on data extraction (accessible via vignette("DataExtraction")) or online (https://cran.r-project.org/package=camtrapR/vignettes/camtrapr3.pdf)).
Note

The results of a number of other function will depend on the output of this function (namely on the arguments exclude for excluding species and minDeltaTime/deltaTimeComparedTo for temporal independence):

detectionMaps
detectionHistory
activityHistogram
activityDensity
activityRadial
activityOverlap
activityHistogram
surveyReport

Author(s)

Juergen Niedballa

References

Phil Harvey’s ExifTool https://exiftool.org/

Examples

```r
## Not run:  # the examples take too long to pass CRAN tests

# set directory with camera trap images in station directories
wd_images_ID_species <- system.file("pictures/sample_images_species_dir", 
packages = "camtrapR")

if (Sys.which("exiftool") !=""){
  # only run these examples if ExifTool is available

rec_table1 <- recordTable(inDir = wd_images_ID_species, 
IDfrom = "directory", 
minDeltaTime = 60, 
deltaTimeComparedTo = "lastRecord", 
writecsv = FALSE, 
additionalMetadataTags = c("EXIF:Model", "EXIF:Make")
)
# note argument additionalMetadataTags: it contains tag names as returned by function exifTagNames

rec_table2 <- recordTable(inDir = wd_images_ID_species, 
IDfrom = "directory", 
minDeltaTime = 60, 
deltaTimeComparedTo = "lastRecord",
```
exclude = "UNID",
writecsv = FALSE,
timeZone = "Asia/Kuala_Lumpur",
additionalMetadataTags = c("EXIF:Model", "EXIF:Make", "NonExistingTag"),
eventSummaryColumn = "EXIF:Make",
eventSummaryFunction = "unique"
)

# note the warning that the last tag in "additionalMetadataTags" ("NonExistingTag") was not found

any(rec_table1$Species == "UNID")    # TRUE
any(rec_table2$Species == "UNID")    # FALSE

# here’s how the removeDuplicateRecords argument works

rec_table3a <- recordTable(inDir = wd_images_ID_species,
IDfrom = "directory",
minDeltaTime = 0,
exclude = "UNID",
timeZone = "Asia/Kuala_Lumpur",
removeDuplicateRecords = FALSE)

rec_table3b <- recordTable(inDir = wd_images_ID_species,
IDfrom = "directory",
minDeltaTime = 0,
exclude = "UNID",
timeZone = "Asia/Kuala_Lumpur",
removeDuplicateRecords = TRUE)

anyDuplicated(rec_table3a[, c("Station", "Species", "DateTimeOriginal")])  # got duplicates
anyDuplicated(rec_table3b[, c("Station", "Species", "DateTimeOriginal")])  # no duplicates

# after removing duplicates, both are identical:
whichAreDuplicated <- which(duplicated(rec_table3a[,c("Station", "Species", "DateTimeOriginal")]))
all(rec_table3a[-whichAreDuplicated,] == rec_table3b)

### extracting species IDs from metadata

wd_images_ID_species_tagged <- system.file("pictures/sample_images_species_tag",
package = "camtrapR")

rec_table4 <- recordTable(inDir = wd_images_ID_species_tagged,
IDfrom = "metadata",
metadataSpeciesTag = "Species",
exclude = "unidentified")
### Including videos

Sample videos are not included in package

With videos, IDfrom = "directory", not extracting digiKam metadata

```r
rec_table4 <- recordTable(inDir = wd_images_ID_species,
                          IDfrom = "directory",
                          video = list(file_formats = c("jpg", "mp4"),
                                       dateTimeTag = "QuickTime:CreateDate")
)
```

With videos, IDfrom = "metadata", extracting digiKam metadata

```r
rec_table5 <- recordTable(inDir = wd_images_ID_species,
                          IDfrom = "metadata",
                          metadataSpeciesTag = "Species",
                          video = list(file_formats = c("jpg", "mp4", "avi", "mov"),
                                       dateTimeTag = "QuickTime:CreateDate",
                                       db_directory = "C:/Users/YourName/Pictures",
                                       db_filename = "digikam4.db")
)
```

```r
} else {
  # show function output if ExifTool is not available
  message("ExifTool is not available. Cannot test function. Loading recordTableSample instead")
  data(recordTableSample)
}
```

```r
## End(Not run)
```

---

**recordTableIndividual**  
*Generate a single-species record table with individual identification from camera trap images or videos*

**Description**

The function generates a single-species record table containing individual IDs, e.g. for (spatial) capture-recapture analyses. It prepares input for the function `spatialDetectionHistory`.

**Usage**

```r
recordTableIndividual(
  inDir,
  hasStationFolders,
  IDfrom,
  cameraID,
  camerasIndependent,
)```
Arguments

inDir character. Directory containing images of individuals. Must end with species name (e.g. ".../speciesImages/Clouded Leopard")

hasStationFolders logical. Does inDir have station subdirectories? If TRUE, station IDs will be taken from directory names. If FALSE, they will be taken from image filenames (requires images renamed with `imageRename`).

IDfrom character. Read individual ID from image metadata ("metadata") of from directory names ("directory")?

cameraID character. Should the function look for camera IDs in the image file names? If so, set to 'filename'. Requires images renamed with `imageRename`. If missing, no camera ID will be assigned and it will be assumed there was 1 camera only per station.

camerasIndependent logical. If TRUE, cameras at a station are assumed to record individuals independently. If FALSE, cameras are assumed to be non-independent (e.g. in pairs). Takes effect only if there was more than 1 camera per station and cameraID = "filename".

minDeltaTime numeric. time difference between observation of the same individual at the same station/camera to be considered independent (in minutes)

deltaTimeComparedTo character. For two records to be considered independent, must the second one be at least minDeltaTime minutes after the last independent record of the same individual ("lastIndependentRecord"), or minDeltaTime minutes after the last record ("lastRecord")?

timeZone character. Must be a value returned by `OlsonNames`

stationCol character. Name of the camera trap station column in the output table.

writecsv logical. Should the individual record table be saved as a .csv file?

outDir character. Directory to save csv file to. If NULL and writecsv = TRUE, the output csv will be written to inDir.
metadataHierarchyDelimiter
character. The character delimiting hierarchy levels in image metadata tags in field "HierarchicalSubject". Either "|" or ":".

metadataIDTag
character. In custom image metadata, the individual ID tag name.

additionalMetadataTags
character. additional camera model-specific metadata tags to be extracted. (If possible specify tag groups as returned by exifTagNames)

removeDuplicateRecords
logical. If there are several records of the same individual at the same station (also same camera if cameraID is defined) at exactly the same time, show only one?

returnFileNamesMissingTags
logical. If species are assigned with metadata and images are not tagged, return a few file names of these images as a message?

eventSummaryColumn
character. A column in the record table (e.g. from a metadata tag) by to summarise non-independent records (those within minDeltaTime of a given record) with a user-defined function (eventSummaryFunction)

eventSummaryFunction
character. The function by which to summarise eventSummaryColumn of non-independent records, e.g. "sum", "max" (optional)

video
list. Contains information on how to handle video data (optional). See details.

Details
The function can handle a number of different ways of storing images and videos. In every case, images need to be stored in a species directory first (e.g. using function getSpeciesImages). Station subdirectories are optional. Camera subdirectories are not supported. This directory structure can be created easily with function getSpeciesImages.

As with species identification, individuals can be identified in 2 different ways: by moving images into individual directories ("Species/Station/Individual/XY.JPG" or "Species/Individual/XY.JPG") or by metadata tagging (without the need for individual directories: "Species/XY.JPG" or "Species/Station/XY.JPG"). minDeltaTime is a criterion for temporal independence of records of an individual at the same station/location. Setting it to 0 will make the function return all records. camerasIndependent defines if the cameras at a station are to be considered independent (e.g. FALSE if both cameras face each other and possibly TRUE if they face different trails). stationCol is the station column name to be used in the resulting table. Station IDs are read from the station directory names if hasStationFolders = TRUE. Otherwise, the function will try to extract station IDs from the image filenames (requires images renamed with imageRename).

If individual IDs were assigned with image metadata tags, metadataIDTag must be set to the name of the metadata tag group used for individual identification. metadataHierarchyDelimiter is "|" for images tagged in DigiKam and images tagged in Adobe Bridge/ Lightroom with the default settings. Manufacturer-specific Exif metadata tags such as "AmbientTemperature" or "Moon-Phase" can be extracted if specified in additionalMetadataTags. Multiple names can be specified as a character vector as: c(Tag1, Tag2, ...). Because they are not standardized, function exifTagNames provides a vector of all available tag names. The metadata tags thus extracted may be used as individual covariates in spatial capture-recapture models.
eventSummaryColumn and eventSummaryFunction can be used to extract summary statistics for independent sampling events. For example, you assigned a "count" tag to your images, indicating the number of individuals in a picture. In a sequence of pictures taken within 1 minute, most pictures show one individual, but one image shows two individuals. You tagged the images accordingly (count = 1 or count = 2) and run recordTable. Set eventSummaryColumn = "count" and eventSummaryFunction = "max" to obtain the maximum number of count in all images within minDeltaTime minutes of a given record. The results is in a new column, in this example count_max. You can also calculate several statistics at the same time, by supplying vectors of values, e.g. eventSummaryColumn = c("count", "count", "camera") and eventSummaryFunction = c("min", "max", "unique") to get minimum and maximum count and all unique camera IDs for that event. Note that eventSummaryColumn and eventSummaryFunction must be of same length.

Argument video is analogous to recordTable, a named list with 2 or 4 items. 2 items (file_formats, dateTimeTag) are always required, and are sufficient if IDfrom = "directory". In that case, no digiKam tags will be returned. To return digiKam tags, two additional items are required (db_directory, db_filename). This is essential when using IDfrom = "metadata". When using IDfrom = "directory", it is optional, but allows to extract metadata tags assigned to videos in digiKam. This workaround is necessary because digiKam tags are not written into video metadata, but are only saved in the digiKam database. So in contrast to metadata tags assigned to JPEG images, they cannot be extracted with ExifTool. It also requires that inDir is in your digiKam database.

The items of argument video are:

- file_formats: The video formats to extract (include "jpg" if you want JPEG image metadata)
- dateTimeTag: the metadata tag to extract date/time from (use exifTagNames to find out which tag is suitable)
- db_directory: The directory containing digiKam database (optional if IDfrom = "directory")
- db_filename: The digiKam database file in db_directory (optional if IDfrom = "directory")

See the example below for how to specify the argument video.

Value

A data frame containing species records with individual IDs and additional information about stations, date, time and (optionally) further metadata.

Warning

Be sure to read the section on individual identification in the package vignette (https://CRAN.R-project.org/package=camtrapR/vignettes/camtrapr2.pdf).

If you use image metadata tags for identification, the tags must be written to the image metadata. The function cannot read tags from .xmp sidecar files. Make sure you set the preferences of your image management software accordingly. In DigiKam, go to Settings/Configure DigiKam/Metadata. There, make sure "Write to sidecar files" is unchecked.

Please note the section about defining argument timeZone in the vignette on data extraction (accessible via vignette("DataExtraction") or online (https://cran.r-project.org/package=camtrapR/vignettes/camtrapr3.pdf)).
recordTableIndividual

Author(s)
Juergen Niedballa

References
Phil Harvey's ExifTool https://exiftool.org/

Examples

## Not run: # the examples run too long to pass CRAN tests

```r
wd_images_ID_individual <- system.file("pictures/sample_images_indiv_tag/LeopardCat", package = "camtrapR")
# missing space in species = "LeopardCat" is because of CRAN package policies
# note argument additionalMetadataTags: contains tag names as returned by function exifTagNames

if (Sys.which("exiftool") != ""){
  # only run these examples if ExifTool is available
  rec_table_pbe <- recordTableIndividual(inDir = wd_images_ID_individual,
                                         minDeltaTime = 60,
                                         deltaTimeComparedTo = "lastRecord",
                                         hasStationFolders = FALSE,
                                         IDfrom = "metadata",
                                         camerasIndependent = FALSE,
                                         writecsv = FALSE,
                                         metadataIDTag = "individual",
                                         additionalMetadataTags = c("EXIF:Model", "EXIF:Make"),
                                         timeZone = "Asia/Kuala_Lumpur")
}
```

# extracting some example summary stats too
# a nonsensical example, get all unique cameras with which the event was photographed

```r
rec_table_pbe2 <- recordTableIndividual(inDir = wd_images_ID_individual,
                                         minDeltaTime = 60,
                                         deltaTimeComparedTo = "lastRecord",
                                         hasStationFolders = FALSE,
                                         IDfrom = "metadata",
                                         camerasIndependent = FALSE,
                                         writecsv = FALSE,
                                         metadataIDTag = "individual",
                                         additionalMetadataTags = c("EXIF:Model", "EXIF:Make"),
                                         timeZone = "Asia/Kuala_Lumpur",
                                         eventSummaryColumn = "EXIF:Make",
                                         eventSummaryFunction = "unique")
```

### Video example (the sample data don’t contain a video, this is hypothetical)
# with JPG, video mp4, avi, mov, ID = metadata
rec_table_ind_video <- recordTableIndividual(inDir = wd_images_ID_individual, 
hasStationFolder = FALSE, 
IDfrom = "metadata", 
metadataIDTag = "individual", 
video = list(file_formats = c("jpg", "mp4", "avi", "mov"), 
            dateTimeTag = "QuickTime:CreateDate", 
            db_directory = "C:/Users/YourName/Pictures", 
            db_filename = "digikam4.db")
}

} else {
  # show function output if ExifTool is not available
  message("ExifTool is not available. Cannot test function. Loading recordTableSample instead")
  data(recordTableSample)
}

## End(Not run)

recordTableIndividualSample

Sample single-species record table with custom metadata from camera trap images

Description

Sample single-species record table with individual IDs from the tagged sample images in the package. Generated with function `recordTableIndividual`.

Usage

data(recordTableIndividualSample)

Format

A data frame with 21 rows and 17 variables

Details

The variables are as follows:

<table>
<thead>
<tr>
<th>Variable</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Station</td>
<td>Camera trap station ID</td>
</tr>
<tr>
<td>Species</td>
<td>Species ID</td>
</tr>
<tr>
<td>Individual</td>
<td>Individual ID</td>
</tr>
<tr>
<td>DateTimeOriginal</td>
<td>Date and time as extracted from image</td>
</tr>
<tr>
<td>Date</td>
<td>record date</td>
</tr>
</tbody>
</table>
**recordTableIndividualSampleMultiSeason**

Sample single-species multi-season record table with custom metadata from camera trap images

**Description**
Sample single-species multi-season record table with individual IDs from the tagged sample images in the package. Generated with function `recordTableIndividual`, then duplicated to simulate a second year.

**Usage**

```r
data(recordTableIndividualSampleMultiSeason)
```

**Format**
A data frame with 31 rows and 17 variables

**Details**
The variables are as follows:

- **Station**: Camera trap station ID
- **Species**: Species ID
- **Individual**: Individual ID
- **DateTimeOriginal**: Date and time as extracted from image
- **Date**: record date
- **Time**: record time of day
- **delta.time.secs**: time difference to first species record at a station (seconds)
- **delta.time.mins**: time difference to first species record at a station (minutes)
- **delta.time.hours**: time difference to first species record at a station (hours)
- **delta.time.days**: time difference to first species record at a station (days)
recordTableSample

<table>
<thead>
<tr>
<th>Directory</th>
<th>Image directory</th>
</tr>
</thead>
<tbody>
<tr>
<td>FileName</td>
<td>Image filename</td>
</tr>
<tr>
<td>HierarchicalSubject</td>
<td>content of the HierarchicalSubject image metadata tag</td>
</tr>
<tr>
<td>Model</td>
<td>Camera model extracted from image metadata</td>
</tr>
<tr>
<td>Make</td>
<td>Camera make extracted from image metadata</td>
</tr>
<tr>
<td>metadata_Species</td>
<td>content of custom image metadata tag &quot;Species&quot; (see HierarchicalSubject)</td>
</tr>
<tr>
<td>metadata_individual</td>
<td>content of custom image metadata tag &quot;individual&quot; (see HierarchicalSubject)</td>
</tr>
</tbody>
</table>

Examples

# example data were created as follows:
data(recordTableIndividualSample)

recordTableIndividualSample_season2 <- recordTableIndividualSample[1:10, ]
recordTableIndividualSample_season2$DateTimeOriginal <- gsub("2009", "2010",
recordTableIndividualSample_season2$DateTimeOriginal)
recordTableIndividualSampleMultiSeason <- rbind(recordTableIndividualSample,
recordTableIndividualSample_season2)

recordTableSample

Sample species record table from camera trap images

Description

Sample species record table from camera trap images generated from the sample images in the package with the function recordTable.

Usage

data(recordTableSample)

Format

A data frame with 39 rows and 11 variables

Details

The variables are as follows:

<table>
<thead>
<tr>
<th>Station</th>
<th>Camera trap station ID</th>
</tr>
</thead>
<tbody>
<tr>
<td>Species</td>
<td>Species ID</td>
</tr>
<tr>
<td>DateTimeOriginal</td>
<td>Date and time as extracted from image</td>
</tr>
<tr>
<td>Date</td>
<td>record date</td>
</tr>
<tr>
<td>Time</td>
<td>record time of day</td>
</tr>
<tr>
<td>delta.time.secs</td>
<td>time difference to first species record at a station (seconds)</td>
</tr>
<tr>
<td>delta.time.mins</td>
<td>time difference to first species record at a station (minutes)</td>
</tr>
<tr>
<td>delta.time.hours</td>
<td>time difference to first species record at a station (hours)</td>
</tr>
</tbody>
</table>
Sample multi-season species record table from camera trap images generated from the sample images in the package with the function `recordTable`. Season 2009 is the same as `recordTableSample`, season 2010 was simulated by adding 1 year to these records.

Usage

```r
data(recordTableSampleMultiSeason)
```

Format

A data frame with 78 rows and 11 variables

Details

The variables are as follows:

- `delta.time.days`: time difference to first species record at a station (days)
- `Directory`: Image directory
- `FileName`: image filename
- `delta.time.secs`: time difference to first species record at a station (seconds)
- `delta.time.mins`: time difference to first species record at a station (minutes)
- `delta.time.hours`: time difference to first species record at a station (hours)
- `delta.time.days`: time difference to first species record at a station (days)

Examples

```r
# data were created with the following code:

data(recordTableSample)
recordTableSample_season2 <- recordTableSample
```
# substitute 2009 with 2010
recordTableSample_season2$DateTimeOriginal <- gsub("2009", "2010",
recordTableSample_season2$DateTimeOriginal)
# combine with season 2009
recordTableSampleMultiSeason <- rbind(recordTableSample, recordTableSample_season2)

spatialDetectionHistory

*Generate a capthist object for spatial capture-recapture analyses from camera-trapping data*

**Description**

This function generates spatial detection histories of individuals of a species for spatial capture-recapture analyses with package `secr`. Data are stored in a `capthist` object. The `capthist` object contains detection histories, camera-trap station location and possibly individual and station-level covariates. Detection histories can have adjustable occasion length and occasion start time (as in the function `detectionHistory`).

**Usage**

```r
spatialDetectionHistory(
  recordTableIndividual, 
  species, 
  camOp, 
  CTtable, 
  output = c("binary", "count"), 
  stationCol = "Station", 
  speciesCol = "Species", 
  sessionCol, 
  Xcol, 
  Ycol, 
  stationCovariateCols, 
  individualCol, 
  individualCovariateCols, 
  recordDateTimeCol = "DateTimeOriginal", 
  recordDateTimeFormat = "ymd HMS", 
  occasionLength, 
  minActiveDaysPerOccasion, 
  occasionStartTime = "deprecated", 
  maxNumberDays, 
  day1, 
  buffer, 
  includeEffort = TRUE, 
  scaleEffort = FALSE, 
  binaryEffort = FALSE, 
  timeZone,
)```

makeRMarkInput

Arguments

recordTableIndividual
  data.frame. the record table with individual IDs created by recordTableIndividual

species
  character. the species for which to compute the detection history

camOp
  The camera operability matrix as created by cameraOperation

CTtable
  data.frame. contains station IDs and coordinates. Same as used in cameraOperation.

output
  character. Return individual counts ("count") or binary observations ("binary")?

stationCol
  character. name of the column specifying Station ID in recordTableIndividual and CTtable

speciesCol
  character. name of the column specifying species in recordTableIndividual

sessionCol
  character. name of the column specifying session IDs, either in recordTableIndividual or in CTtable. See 'Details' for more information. Session ID values must be a sequence of integer numbers beginning with 1 (i.e., 1,2,3,...).

Xcol
  character. name of the column specifying x coordinates in CTtable

Ycol
  character. name of the column specifying y coordinates in CTtable

stationCovariateCols
  character. name of the column(s) specifying station-level covariates in CTtable

individualCol
  character. name of the column specifying individual IDs in recordTableIndividual

individualCovariateCols
  character. name of the column(s) specifying individual covariates in recordTableIndividual

recordDateTimeCol
  character. name of the column specifying date and time in recordTableIndividual

recordDateTimeFormat
  format of column recordDateTimeCol in recordTableIndividual

occasionLength
  integer. occasion length in days

minActiveDaysPerOccasion
  integer. minimum number of active trap days for occasions to be included (optional)

occasionStartTime
  (DEPRECATED) integer. time of day (the full hour) at which to begin occasions. Please use argument occasionStartTime in cameraOperation instead.

maxNumberDays
  integer. maximum number of trap days per station (optional)

day1
  character. When should occasions begin: station setup date ("station"), first day of survey ("survey"), a specific date (e.g. "2015-12-31")?

buffer
  integer. Makes the first occasion begin a number of days after station setup. (optional)

includeEffort
  logical. Include trapping effort (number of active camera trap days per station and occasion) as usage in capthist object?
scaleEffort logical. scale and center effort matrix to mean = 0 and sd = 1? Currently not used. Must be FALSE.
binaryEffort logical. Should effort be binary (1 if >1 active day per occasion, 0 otherwise)?
timeZone character. Must be a value returned by OlsonNames
makeRMarkInput logical. If FALSE, output will be a data frame for RMark. If FALSE or not specified, a secr capthist object

Details

The function creates a capthist object by combining three different objects: 1) a record table of identified individuals of a species, 2) a camera trap station table with station coordinates and 3) a camera operation matrix computed with cameraOperation. The record table must contain a column with individual IDs and optionally individual covariates. The camera trap station table must contain station coordinates and optionally station-level covariates. The camera operation matrix provides the dates stations were active or not and the number of active stations.

day1 defines if each stations detection history will begin on that station’s setup day (day1 = "station") or if all station’s detection histories have a common origin (the day the first station was set up if day1 = "survey" or a fixed date if, e.g. day1 = "2015-12-31").

includeEffort controls whether an effort matrix is computed or not. If TRUE, effort will be used for object usage information in a traps. binaryEffort makes the effort information binary. scaleEffort is currently not used and must be set to FALSE. The reason is that usage can only be either binary, or nonnegative real values, whereas scaling effort would return negative values.

The number of days that are aggregated is controlled by occasionLength. occasionStartTime will be removed from the function. It has moved to cameraOperation, to ensure daily effort is computed correctly and takes the occasion start time into account. another hour than midnight (the default). This may be relevant for nocturnal animals, in which 1 whole night would be considered an occasion.

Output can be returned as individual counts per occasion (output = "count") or as binary observation (output = "binary").

Argument sessionCol can be used to create multi-session capthist object. There are two different ways in which the argument is interpreted. It depends on whether a column with the name you specify in argument sessionCol exists in recordTableIndividual or in CTtable. If sessionCol is found in recordTableIndividual, the records will be assigned to the specified sessions, and it will be assumed that all camera trap station were used in all sessions. Alternatively, if sessionCol is found inCTtable, it will be assumed that only a subset of stations was used in each session, and the records will be assigned automatically (using the station IDs to identify which session they belong into). In both cases, session information must be provided as a sequence of integer numbers beginnign with 1, i.e., you provide the session number directly in sessionCol. See session for more information about sessions in secr.

capthist objects (as created by spatialDetectionHistory for spatial capture-recapture analyses) expect the units of coordinates (Xcol and col in CTtable) to be meters. Therefore, please use a suitable coordinate system (e.g. UTM).

recordDateTimeFormat defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". recordDateTimeFormat can be interpreted either by base-R via strptime or in lubridate via parse_date_time (argument "orders"). lubridate will be used if there are no "%" characters in recordDateTimeFormat.
For "YYYY-MM-DD HH:MM:SS", recordDateTimeFormat would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see `strptime` or `parse_date_time`.

Value

Output depends on argument `makeRMarkInput`:

```r
list("makeRMarkInput = FALSE")
   A `capthist` object
list("makeRMarkInput = TRUE")
   A data frame for use in RMark
```

Warning

Please note the section about defining argument `timeZone` in the vignette on data extraction (accessible via `vignette("DataExtraction")` or online ([https://cran.r-project.org/package=camtrapR/vignettes/camtrapr3.pdf](https://cran.r-project.org/package=camtrapR/vignettes/camtrapr3.pdf))).

Author(s)

Juergen Niedballa

See Also

`secr` `RMark`

Examples

```r
data(recordTableIndividualSample)
data(camtraps)

# create camera operation matrix (with problems/malfunction)
camop_problem <- cameraOperation(CTtable = camtraps,
       stationCol = "Station",
       setupCol = "Setup_date",
       retrievalCol = "Retrieval_date",
       writecsv = FALSE,
       hasProblems = TRUE,
       dateFormat = "dmy"
)

sdh <- spatialDetectionHistory(recordTableIndividual = recordTableIndividualSample,
       species = "LeopardCat",
       camOp = camop_problem,
       CTtable = camtraps,
       output = "binary",
       stationCol = "Station",
       speciesCol = "Species",
       Xcol = "utm_x",
)```
Ycol = "utm_y",
individualCol = "Individual",
recordDateTimeCol = "DateTimeOriginal",
recordDateTimeFormat = "ymd HMS",
occasionLength = 10,
day1 = "survey",
includeEffort = TRUE,
timeZone = "Asia/Kuala_Lumpur"
)

# missing space in species = "LeopardCat" was introduced by recordTableIndividual
# (because of CRAN package policies.
# In your data you can have spaces in your directory names)

summary(sdh)
plot(sdh, tracks = TRUE)

### multi-season capthist object
# see vignette "3. Extracting Data from Camera Trapping Images, creating occupancy & secr input"

data(camtrapsMultiSeason)
camtrapsMultiSeason$session[camtrapsMultiSeason$session == 2009] <- 1
camtrapsMultiSeason$session[camtrapsMultiSeason$session == 2010] <- 2

data(recordTableIndividualSampleMultiSeason)

# create camera operation matrix (with problems/malfunction)
camop_session <- cameraOperation(CTtable = camtrapsMultiSeason,
stationCol = "Station",
setupCol = "Setup_date",
sessionCol = "session",
retrievalCol = "Retrieval_date",
hasProblems = TRUE,
dateFormat = "dmy"
)

sdh_multi <- spatialDetectionHistory(recordTableIndividual = recordTableIndividualSampleMultiSeason,
species = "LeopardCat",
output = "binary",
camOp = camop_session,
CTtable = camtrapsMultiSeason,
stationCol = "Station",
speciesCol = "Species",
sessionCol = "session",
Xcol = "utm_x",
Ycol = "utm_y",
individualCol = "Individual",
recordDateTimeCol = "DateTimeOriginal",
recordDateTimeFormat = "ymd HMS",
occasionLength = 10,
day1 = "survey",
includeEffort = TRUE,
timeZone = "Asia/Kuala_Lumpur",
)
sumarize,commOccu-method

Summarize community occupancy model

Description

Gives an overview of the number of species, stations and occasions in a commOccu object. Also returns covariates.

Usage

## S4 method for signature 'commOccu'
summary(object, ...)

Arguments

- object: commOccu object
- ...: currently ignored

Details

The summary method is very basic and still work in progress.

Value

Model summary printed to console

__surveyDashboard__

Shiny dashboard for summarizing and analyzing camera trap survey data
surveyDashboard

Description
Open a Shiny dashboard to summarize a camera trapping survey. It provides an overview of the data, maps, can create and run occupancy models in unmarked, and plot species activity.
Maps are interactive and can be panned, zoomed and queried. Users can choose a suitable basemap in the layers button.
For occupancy models, users can flexibly customize detection histories. Then the user can specify the model structure (by selecting site covariates on detection and occupancy probability from the camera trap table) and optionally including effort. Models are fitted automatically with unmarked. Response curves (marginal effect plots), model and parameter summaries are computed and updated automatically with every change to the model. Multiple models can be compared using model selection.

Usage

```r
surveyDashboard(
  CTtable, recordTable, stationCol, cameraCol = NULL, xcol, ycol, crs, setupCol, retrievalCol, hasProblems = FALSE, CTdateFormat = "ymd", camerasIndependent, speciesCol = "Species", recordDateTimeCol = "DateTimeOriginal", recordDateTimeFormat = "ymd HMS", timeZone = "UTC", exclude = NULL)
```

Arguments

- `CTtable`: A data.frame containing the camera trap deployment information.
- `recordTable`: A data.frame containing the camera trap records.
- `stationCol`: The column name containing the camera trap station ID
- `cameraCol`: The column name containing the camera trap IDs (optional, only if 2 or more cameras per station)
- `xcol`: The column name containing the X coordinate of the camera trap station.
- `ycol`: The column name containing the Y coordinate of the camera trap station.
- `crs`: The coordinate reference system (CRS) of the camera trap data. Must be a valid argument to `st_crs`
- `setupCol`: The column name containing the camera trap deployment date (and time).
retrievalCol  The column name containing the camera trap retrieval date (optionally date-time).

hasProblems  A logical indicating whether there are periods of cameras malfunctioning

CTdateFormat  The date format of the camera trap deployment and retrieval date and time (default: "ymd").

camerasIndependent  logical. If multiple camera per station, are they independent?

speciesCol  The column name containing the species names

recordDateTimeCol  The column name containing the record date and time

recordDateTimeFormat  The date/time format of recordDateTimeCol

timeZone  Time zone of records in recordTable

exclude  Species to be excluded from the data set

Value

A Shiny dashboard for camera trap survey data.

Note

Current limitations include:
- supports only single-season data
- only single-species occupancy models
- no random or interaction effects in occupancy models
- no (spatial) model predictions yet (planned for Q4/2023)
- no support for spatial capture-recapture models (or anything related to individual IDs)
- only models in unmarked, no Bayesian models in ubms yet (likely to be included later)

Author(s)

Juergen Niedballa

Examples

```r
data("camtraps")
data("recordTableSample")

surveyDashboard(
  CTtable = camtraps,
  recordTable = recordTableSample,
  xcol = "utm_x",
  ycol = "utm_y",
  crs = "epsg:32650",  # = UTM50N (Sabah, Malaysian Borneo)
)```
surveyReport

Create a report about a camera trapping survey and species detections

Description

This function creates a report about a camera trapping survey and species records. It uses a camera trap station information table and a record table (generated with `recordTable`) as input. Output tables can be saved and a zip file for simple data sharing can be created easily.

Usage

```r
surveyReport(
  recordTable,  # data.frame containing a species record table as given by recordTable
  CTtable,      # data.frame containing information about location and trapping period of camera trap stations (equivalent to camtraps
  camOp,        # camera operation matrix created with cameraOperation
  speciesCol = "Species",  # species column
  stationCol = "Station",   # station column
  cameraCol,     # camera column
  setupCol,      # setup column
  retrievalCol,  # retrieval column
  CTDateFormat = "ymd",       # camera trap date format
  CTHasProblems = "deprecated", # data column
  recordDateTimeCol = "DateTimeOriginal", # record datetime column
  recordDateTimeFormat = "ymd HMS",   # record datetime format
  Xcol,          # x coordinate column
  Ycol,          # y coordinate column
  sinkpath,      # output directory
  makezip        # create zip file
)
```

Arguments

- `recordTable`: data.frame containing a species record table as given by `recordTable`
- `CTtable`: data.frame containing information about location and trapping period of camera trap stations (equivalent to `camtraps`)
- `camOp`: camera operation matrix created with `cameraOperation`
speciesCol character. name of the column specifying Species ID in recordTable
stationCol character. name of the column specifying Station ID in Ctable and recordTable
cameraCol character. name of the column specifying Camera ID in Ctable and recordTable
setupCol character. name of the column containing camera setup dates in Ctable
retrievalCol character. name of the column containing camera retrieval dates in Ctable
CTDateFormat character. The format of columns setupCol and retrievalCol (and potential problem columns) in Ctable. Must be interpretable by either as.Date or the "orders" argument parse_date_time in lubridate.
CTHasProblems deprecated (since version 2.1)
recordDateTimeCol character. The name of the column containing date and time of records in recordTable
recordDateTimeFormat character. The date/time format of column recordDateTimeCol in recordTable.
Xcol character. name of the column specifying x coordinates in Ctable. Used to create detection maps if makezip is TRUE. (optional)
Ycol character. name of the column specifying y coordinates in Ctable. Used to create detection maps if makezip is TRUE. (optional)
sinkpath character. The directory into which the survey report is saved (optional)
makezip logical. Create a zip file containing tables, plots and maps in sinkpath?

Details
dateFormat defaults to "YYYY-MM-DD", e.g. "2014-10-31". It can be specified either in the format required by strptime or the 'orders' argument in parse_date_time in lubridate. In the example above, "YYYY-MM-DD" would be specified as "%Y-%m-%d" or "ymd".
recordDateTimeFormat defaults to the "YYYY-MM-DD HH:MM:SS" convention, e.g. "2014-09-30 22:59:59". recordDateTimeFormat can be interpreted either by base-R via strptime or in lubridate via parse_date_time (argument "orders"). lubridate will be used if there are no "%" characters in recordDateTimeFormat.
For "YYYY-MM-DD HH:MM:SS", recordDateTimeFormat would be either "%Y-%m-%d %H:%M:%S" or "ymd HMS". For details on how to specify date and time formats in R see strptime or parse_date_time.

Note: as of version 2.1, argument CTHasProblems is deprecated and defunct. Please use camOp instead to provide information about periods of camera activity and malfunction. If camOp is not provided the legacy version of surveyReport (from camtrapR 2.0.3) will be run with a warning.

Value
An invisible list containing 5 data.frames.
survey_dates station and image date ranges, number of total and active trap days (calendar days and taking into account independent effort of multiple cameras, if applicable), number of cameras per station
species_by_station
   species numbers by station

events_by_species
   number of events and stations by species

events_by_station
   number of events for every species by station (only species that were recorded)

events_by_station2
   number of events for all species at all stations (including species that were not recorded)

The output will be saved to a .txt file if sinkpath is defined.
If makezip is TRUE, a zip file will be created in sinkpath. It contains single-species activity plots, detection maps (if Xcol and Ycol are defined), the survey report tables, the record table and the camera trap station table, and an example R script.

Author(s)
Juergen Niedballa

See Also
recordTable

Examples

data(camtraps)
data(recordTableSample)

# since version 2.1, camera operation matrix is required as input

camop_no_problem <- cameraOperation(CTtable = camtraps,
   stationCol = "Station",
   setupCol = "Setup_date",
   retrievalCol = "Retrieval_date",
   writecsv = FALSE,
   hasProblems = FALSE,
   dateFormat = "dmy"
)

reportTest <- surveyReport (recordTable = recordTableSample,
   CTtable = camtraps,
   camOp = camop_no_problem,
   speciesCol = "Species",
   stationCol = "Station",
   setupCol = "Setup_date",
   retrievalCol = "Retrieval_date",
   CTDatetimeFormat = "dmy",
   recordDateTimeCol = "DateTimeOriginal",
   recordDateTimeFormat = "ymd HMS")
class(reportTest)  # a list with
length(reportTest)  # 5 elements

reportTest[[1]]  # camera trap operation times and image date ranges
reportTest[[2]]  # number of species by station
reportTest[[3]]  # number of events and number of stations by species
reportTest[[4]]  # number of species events by station
reportTest[[5]]  # number of species events by station including 0s (non-observed species)

# with camera problems

camop_problem <- cameraOperation(CTtable = camtraps,
                                 stationCol = "Station",
                                 setupCol = "Setup_date",
                                 retrievalCol = "Retrieval_date",
                                 writecsv = FALSE,
                                 hasProblems = TRUE,
                                 dateFormat = "dmy")

reportTest_problem <- surveyReport(recordTable = recordTableSample,
                                    CTtable = camtraps,
                                    camOp = camop_problem,
                                    speciesCol = "Species",
                                    stationCol = "Station",
                                    setupCol = "Setup_date",
                                    retrievalCol = "Retrieval_date",
                                    CTDateFormat = "dmy",
                                    recordDateTimeCol = "DateTimeOriginal",
                                    recordDateTimeFormat = "ymd HMS")

reportTest_problem$survey_dates

## if camOp is missing, the legacy version (from 2.0.3) will be used:

reportTest_problem_old <- surveyReport(recordTable = recordTableSample,
                                        CTtable = camtraps,
                                        # camOp = camop_problem,
                                        speciesCol = "Species",
                                        stationCol = "Station",
                                        setupCol = "Setup_date",
                                        retrievalCol = "Retrieval_date",
                                        CTDateFormat = "dmy",
                                        recordDateTimeCol = "DateTimeOriginal",
                                        recordDateTimeFormat = "ymd HMS")

## Not run:
# run again with sinkpath defined
reportTest <- surveyReport(recordTable = recordTableSample,
                            CTtable = camtraps,
                            camOp = camop_no_problem,
timeShiftImages

### Description

Change the values of digital timestamps in image metadata using ExifTool. If date/time of images were set incorrectly, they can be corrected easily in batch mode for further analyses. Please, always make a backup of your data before using this function to avoid data loss or damage. This is because ExifTool will make a copy of your images and applies the time shifts to the copies. The file extension of the original images (.JPG) will be renamed to "JPG_original".

### Usage

```r
timeShiftImages(
inDir,
hasCameraFolders,
timeShiftTable,
stationCol,
cameraCol,
timeShiftColumn,
timeShiftSignColumn,
undo = FALSE,
ignoreMinorErrors = FALSE
)
```

### Arguments

- `inDir` character. Name of directory containing station directories with images.
- `hasCameraFolders` logical. Do the station directories in `inDir` have camera subdirectories (e.g. "inDir/StationA/Camera1")?
- `timeShiftTable` data.frame containing information about station-/camera-specific time shifts.
**timeShiftImages**

- **stationCol** character. name of the column specifying Station ID in `timeShiftTable`
- **cameraCol** character. name of the column specifying Camera ID in `timeShiftTable` (optional)
- **timeShiftColumn** character. The name of the column containing time shift values in `timeShiftTable`
- **timeShiftSignColumn** character. The name of the column with the direction of time shifts in `timeShiftTable`. Can only be "-" or "+".
- **undo** logical. Undo changes and restore the original images? Please be careful, this deletes any edited images if TRUE
- **ignoreMinorErrors** logical. Ignore minor errors that would cause the function to fail (set TRUE for images with bad MakerNotes, observed in Panthera V4 cameras)

**Details**

`timeShiftTable` is a data frame with columns for Station ID, camera ID (optional), time shift value and direction of time shift (for an example see `timeShiftTable`). Images in `inDir` must be sorted into station directories. If `hasCameraFolders = TRUE`, the function expects camera subdirectories in the station directories and will only apply time shifts to the camera subdirectories specified by `CameraCol` in `timeShiftTable`. If `hasCameraFolders = FALSE`, shifts will be applied to the whole station directory (including potential subdirectories).

The values of `timeShiftColumn` must adhere to the following pattern: "YYYY:mm:dd HH:MM:SS" ("year:month:day hour:minute:second"). Examples: "1:0:0 0:0:0" is a shift of exactly 1 year and "0:0:0 12:10:01" 12 hours and 10 minutes and 1 second. Note that stating "00" may cause problems, so use "0" instead if an entry is zero.

`timeShiftSignColumn` signifies the direction of the time shift. "+" moves image dates into the future (i.e. the image date lagged behind the actual date) and "-" moves image dates back (if the image dates were ahead of actual time).

`ExifTool` stores the original images as `.JPG_original` files in the original file location. By setting `undo = TRUE`, any JPG files in the directories specified by `timeShiftTable` will be deleted and the original JPEGs will be restored from the `JPG_original` files. Please make a backup before using `undo`.

Years can have 365 or 366 days, and months 28 to 31 days. Here is how the function handles these (from the `exiftool` help page): "The ability to shift dates by Y years, M months, etc, conflicts with the design goal of maintaining a constant shift for all time values when applying a batch shift. This is because shifting by 1 month can be equivalent to anything from 28 to 31 days, and 1 year can be 365 or 366 days, depending on the starting date. The inconsistency is handled by shifting the first tag found with the actual specified shift, then calculating the equivalent time difference in seconds for this shift and applying this difference to subsequent tags in a batch conversion."

`ignoreMinorErrors` is useful if image timestamps are not updated correctly (entries in column "n_images" of the output are ". . . files weren’t updated due to errors"). This can be caused by bad MakerNotes and so far was only observed in Panthera V4 and V6 cameras. In that case, set `ignoreMinorErrors` to `TRUE`. This will add the "-m" option to the Exiftool call, thereby ignoring minor errors and warnings and applying the time shift nevertheless.
Value

A data.frame containing the information about the processed directories and the number of images.

Author(s)

Juergen Niedballa

References

https://exiftool.org/#shift

Examples

```r
## Not run:

# copy sample images to temporary directory (so we don’t mess around in the package directory)
wd_images_ID <- system.file("pictures/sample_images_species_dir", package = "camtrapR")
file.copy(from = wd_images_ID, to = tempdir(), recursive = TRUE)
wd_images_ID_copy <- file.path(tempdir(), "sample_images_species_dir")
data(timeShiftTable)

timeshift_run <- timeShiftImages(inDir = wd_images_ID_copy, 
    timeShiftTable = timeShiftTable, 
    stationCol = "Station", 
    hasCameraFolders = FALSE, 
    timeShiftColumn = "timeshift", 
    timeShiftSignColumn = "sign", 
    undo = FALSE)

	
						
timeshift_undo <- timeShiftImages(inDir = wd_images_ID_copy, 
    timeShiftTable = timeShiftTable, 
    stationCol = "Station", 
    hasCameraFolders = FALSE, 
    timeShiftColumn = "timeshift", 
    timeShiftSignColumn = "sign", 
    undo = TRUE)

## End(Not run)
```
Sample camera trap time shift table

**Description**

Sample camera trap time shift table

**Usage**

data(timeShiftTable)

**Format**

A data frame with 2 rows and 4 variables

**Details**

If image Exif metadata timestamps are wrong systematically (e.g. because camera system time was not set after changing batteries), it can be corrected using a data.frame in the following format using function `timeShiftImages`. For details on data format, please see `timeShiftImages`. The variables are as follows:

<table>
<thead>
<tr>
<th>Variable</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Station</td>
<td>Camera trap station ID</td>
</tr>
<tr>
<td>camera</td>
<td>Camera trap ID (optional)</td>
</tr>
<tr>
<td>timeshift</td>
<td>time shift amount to be applied</td>
</tr>
<tr>
<td>sign</td>
<td>direction of time shift</td>
</tr>
</tbody>
</table>

---

**writeDateTimeOriginal**  
*Write values to DateTimeOriginal tag in image metadata*

**Description**

This function assigns values to the DateTimeOriginal tag in image’s EXIF metadata using Exiftool. It can be used when the original DateTimeOriginal values in the metadata were lost for whatever reason. In order to first read the Date/Time values from the data fields in the images, see the function `OCRdataFields`. After running OCRdataFields and checking its output you can run `writeDateTimeOriginal`.

**Usage**

`writeDateTimeOriginal(DateTimeOriginal, fileNames, parallel, overwrite = FALSE)`
Arguments

DateTimeOriginal
character. DateTimeOriginal values to write into EXIF:DateTimeOriginal tag of the files in fileNames.

fileNames
character. Full file names (including directories) of images to process.

parallel
A parallel cluster object. Specify if you wish to run this function in parallel. Provide a cluster object (output of makeCluster()) - optional.

overwrite
logical. Overwrite existing files (TRUE) or create new files while saving the original data as jpg_original files as a backup (FALSE)?

Details

The first value in DateTimeOriginal will be assigned to the first image in fileNames, and so on. Both DateTimeOriginal and fileNames can be obtained from the output of OCRdataFields. DateTimeOriginal uses the standard "YYYY-MM-DD HH:MM:SS" notation. If the values extracted via OCRdataFields are in a different format you’ll need to reformat them first. Please provide them as character. Also, before using this function, make sure that the date/time values read by OCRdataFields are correct (sometimes OCR misreads values, so check carefully).

Parallel processing is advised since the function is rather slow (due to calling Exiftool separately on every, so about 1 second per image). If you know how to batch-assign DateTimeOriginal values in one Exiftool call, please let me know.

The function only works on JPG images, not video files.

Value

Invisible NULL. The actual output is the JPG images which now have a DateTimeOriginal tag.

Author(s)

Juergen Niedballa

See Also

OCRdataFields

Examples

```r

## Not run:
# dontrun is to avoid forcing users to install additional dependencies

wd_images_OCR <- system.file("pictures/full_size_for_ocr", package = "camtrapR")

library(magick)

# define geometries
geometry1 <- geometry_area(x_off = 0, y_off = 0, width = 183, height = 37)
geometry2 <- geometry_area(x_off = 196, y_off = 0, width = 200, height = 17)
```
geometry3 <- geometry_area(x_off = 447, y_off = 0, width = 63, height = 17)
geometry4 <- geometry_area(x_off = 984, y_off = 0, width = 47, height = 17)
geometry5 <- geometry_area(x_off = 0, y_off = 793, width = 320, height = 17)

# combine geometries into list
geometries <- list(date = geometry1,
                    time = geometry2,
                    sequence_id = geometry3,
                    temperature = geometry4,
                    camera_model = geometry5)

df_image_data <- OCRdataFields(inDir = wd_images_OCR,
                                geometries = geometries,
                                invert = TRUE)

df_image_data

library(parallel)
library(lubridate)

# prepare DateTimeOriginal column (ymd_hms() automatically respects the PM indicator)
df_image_data$DateTimeOriginal <- paste(df_image_data$date, df_image_data$time)

df_image_data$DateTimeOriginal <- as.character(ymd_hms(df_image_data$DateTimeOriginal))

# create cluster (3 cores)
c1 <- makeCluster(3)

# assign new DateTimeOriginal
writeDateTimeOriginal(DateTimeOriginal = df_image_data$DateTimeOriginal,
                        fileNames = df_image_data$filename_full,
                        parallel = c1)

stopCluster(c1)

## End(Not run)
Index

* datasets
  camtraps, 24
  camtrapsMultiSeason, 24
  recordTableIndividualSample, 78
  recordTableIndividualSampleMultiSeason, 79
  recordTableSample, 80
  recordTableSampleMultiSeason, 81
  timeShiftTable, 97
  ∗ package
  camtrapR-package, 3

  activityDensity, 4, 6, 9, 10, 12, 14, 71
  activityHistogram, 4, 8, 14, 71
  activityOverlap, 4, 8, 10, 14, 71
  activityRadial, 4, 8, 10, 13, 71
  addCopyrightTag, 4, 15
  addToPath, 4, 17
  appendSpeciesNames, 4, 18
  as.Date, 43
  cameraOperation, 5, 20, 42, 43, 83, 84, 90
  camtrapR (camtrapR-package), 3
  camtrapR-package, 3
  camtraps, 5, 22, 24, 69, 90
  camtrapsMultiSeason, 5, 24
  capthist, 21, 24, 25, 82–85
  checkSpeciesIdentification, 4, 19, 26
  checkSpeciesNames, 4, 29
  commOccu-class, 5, 30
  communityModel, 5, 31, 55
  compileNimble, 54
  createSpeciesFolders, 4, 38
  createStationFolders, 4, 40
  densityPlot, 6
  detectionHistory, 5, 20, 21, 23, 32, 41, 71, 82
  detectionMaps, 4, 46, 71
  exifTagNames, 4, 49, 69, 70, 75, 76
  facet_grid, 64
  filterRecordTable, 4, 51
  fit, 64–66
  fit, commOccu-method, 5, 54
  fixDateTimeOriginal, 4, 55
  geometry, 62
  get_tsn, 29
  getSpeciesImages, 4, 56, 75
  ggsave, 64, 65
  hist, 9
  imageRename, 4, 38, 39, 56, 58, 68, 74, 75
  OCRdataFields, 4, 61, 97, 98
  OlsonNames, 42, 53, 68, 74, 84
  overlapEst, 10, 11
  overlapPlot, 10, 11
  parse_date_time, 7, 9, 11, 14, 21, 43, 84, 85, 91
  plot_coef, 5
  plot_coef (plot_coef, commOccu-method), 63
  plot_coef, commOccu-method, 63
  plot_effects, 5
  plot_effects (plot_effects, commOccu-method), 65
  plot_effects, commOccu-method, 65
  predict (predict, commOccu-method), 66
  predict, commOccu-method, 5, 66
  radial.plot, 13, 14
  recordTable, 4, 7, 9, 11, 13, 42, 47, 50–52, 56, 57, 60, 62, 67, 76, 80, 81, 90, 92
  recordTableIndividual, 4, 50, 73, 78, 79, 83
  recordTableIndividualSample, 5, 78
  recordTableIndividualSampleMultiSeason, 5, 79

100
recordTableSample, 5, 80, 81
recordTableSampleMultiSeason, 5, 81
runMCMC, 54

secr, 82
session, 84
spatialDetectionHistory, 5, 20, 21, 23–25, 43, 73, 82, 84
st_crs, 48, 88
strptime, 7, 9, 11, 14, 21, 43, 84, 85, 91
summary, commOccu-method, 5, 87
surveyDashboard, 5, 87
surveyReport, 5, 20, 71, 90

timeShiftImages, 4, 94, 97
timeShiftTable, 5, 95, 97
traps, 84

unmarked, 41
unmarkedMultFrame, 21, 43
usage, 84

writeDateTimeOriginal, 4, 62, 63, 97