Package ‘coloc’

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Type Package

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Title Colocalisation Tests of Two Genetic Traits

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Maintainer Chris Wallace <cew54@cam.ac.uk>

Description Performs the colocalisation tests described in
Giambartolomei et al (2013) <doi:10.1371/journal.pgen.1004383>,
Wallace (2020) <doi:10.1371/journal.pgen.1008720>,

License GPL

LazyLoad yes

VignetteBuilder knitr

RoxygenNote 7.1.1

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URL https://github.com/chr1swallace/coloc

BugReports https://github.com/chr1swallace/coloc/issues

Collate 'check.R' 'claudia.R' 'coloc-package.R' 'plot.R' 'private.R'

'sensitivity.R' 'split.R' 'susie.R' 'testdata.R' 'zzz.R'

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NeedsCompilation no

Author Chris Wallace [aut, cre],
Claudia Giambartolomei [aut],
Vincent Plagnol [ctb]

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| coloc-package | Colocalisation tests of two genetic traits |

**Description**

Performs the colocalisation tests described in Plagnol et al (2009) and Wallace et al (2020) and draws some plots.
**alpha_to_logbf**

**Author(s)**

Chris Wallace cew54@cam.ac.uk

---

**Description**

convert alpha matrix to log BF matrix

**Usage**

alpha_to_logbf(alpha, pi)

**Arguments**

alpha  
an L by p+1 matrix of posterior inclusion probabilities

pi  
per SNP prior probability of causality

**Value**

L by p+1 matrix of log BF

**Author(s)**

Chris Wallace

---

**approx.bf.estimates**  
Internal function, approx.bf.estimates

---

**Description**

Internal function, approx.bf.estimates

**Usage**

approx.bf.estimates(z, V, type, suffix = NULL, sdY = 1)

**Arguments**

z  
normal deviate associated with regression coefficient and its variance

V  
its variance

type  
"quant" or "cc"

suffix  
suffix to append to column names of returned data.frame

sdY  
standard deviation of the trait. If not supplied, will be estimated.
Details
Calculate approximate Bayes Factors using supplied variance of the regression coefficients

Value
data.frame containing lABF and intermediate calculations

Author(s)
Vincent Plagnol, Chris Wallace

approx.bf.p

Description
Internal function, approx.bf.p

Usage
approx.bf.p(p, f, type, N, s, suffix = NULL)

Arguments

<table>
<thead>
<tr>
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<tr>
<td>p</td>
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<td>type</td>
<td>&quot;quant&quot; or &quot;cc&quot;</td>
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<td>N</td>
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<tr>
<td>s</td>
<td>proportion of samples that are cases, ignored if type=&quot;quant&quot;</td>
</tr>
<tr>
<td>suffix</td>
<td>suffix to append to column names of returned data.frame</td>
</tr>
</tbody>
</table>

Details
Calculate approximate Bayes Factors

Value
data.frame containing lABF and intermediate calculations

Author(s)
Claudia Giambartolomei, Chris Wallace
bin2lin

binomial to linear regression conversion

Description

Convert binomial to linear regression

Usage

bin2lin(D, doplot = FALSE)

Arguments

D standard format coloc dataset
doplot plot results if TRUE - useful for debugging

Details

Estimate beta and varbeta if a linear regression had been run on a binary outcome, given log OR and their variance + MAF in controls

sets beta = cov(x,y)/var(x) varbeta = (var(y)/var(x) - cov(x,y)^2/var(x)^2)/N

Value

D, with original beta and varbeta in beta.bin, varbeta.bin, and beta and varbeta updated to linear estimates

Author(s)

Chris Wallace

check_alignment

check alignment

Description

check alignment between beta and LD

Usage

check_alignment(D, thr = 0.2, do_plot = TRUE)

check.alignment(...)

check_dataset

Arguments

- `d`  a coloc dataset
- `thr`  plot SNP pairs in absolute LD > thr
- `do_plot`  if TRUE (default) plot the diagnostic
- `...` arguments passed to check_alignment()

Value

proportion of pairs that are positive

Author(s)

Chris Wallace

check_dataset

Description

Check coloc dataset inputs for errors

Usage

check_dataset(d, suffix = "", req = c("snp"), warn.minp = 1e-06)

check.dataset(...)

Arguments

- `d` dataset to check
- `suffix` string to identify which dataset (1 or 2)
- `req` names of elements that must be present
- `warn.minp` print warning if no p value < warn.minp
- `...` arguments passed to check_dataset()

Details

Coloc is flexible, requiring perhaps only p values, or z scores, or effect estimates and standard errors, but with this flexibility, also comes difficulties describing exactly the combinations of items required.

- `pvalues` P-values for each SNP in dataset 1
- `N` Number of samples in dataset 1
- `MAF` minor allele frequency of the variants
- `beta` regression coefficient for each SNP from dataset 1
**coloc.abf**

*Fully Bayesian colocalisation analysis using Bayes Factors*

**Description**

Bayesian colocalisation analysis

**Usage**

```
coloc.abf(dataset1, dataset2, MAF = NULL, p1 = 1e-04, p2 = 1e-04, p12 = 1e-05)
```
Arguments

dataset1  a list with specifically named elements defining the dataset to be analysed. See check_dataset for details.
dataset2  as above, for dataset 2
MAF     Common minor allele frequency vector to be used for both dataset1 and dataset2, a shorthand for supplying the same vector as parts of both datasets
p1      prior probability a SNP is associated with trait 1, default 1e-4
p2      prior probability a SNP is associated with trait 2, default 1e-4
p12     prior probability a SNP is associated with both traits, default 1e-5

Details

This function calculates posterior probabilities of different causal variant configurations under the assumption of a single causal variant for each trait.

If regression coefficients and variances are available, it calculates Bayes factors for association at each SNP. If only p values are available, it uses an approximation that depends on the SNP’s MAF and ignores any uncertainty in imputation. Regression coefficients should be used if available.

Value

a list of two data.frames:

  - summary is a vector giving the number of SNPs analysed, and the posterior probabilities of H0 (no causal variant), H1 (causal variant for trait 1 only), H2 (causal variant for trait 2 only), H3 (two distinct causal variants) and H4 (one common causal variant)
  - results is an annotated version of the input data containing log Approximate Bayes Factors and intermediate calculations, and the posterior probability SNP.PP.H4 of the SNP being causal for the shared signal if H4 is true. This is only relevant if the posterior support for H4 in summary is convincing.

Author(s)

Claudia Giambartolomei, Chris Wallace

coloc.bf_bf

Coloc data through Bayes factors

Description

Colocalise two datasets represented by Bayes factors
Usage

coloc.bf_bf(
  bf1,
  bf2,
  p1 = 1e-04,
  p2 = 1e-04,
  p12 = 5e-06,
  overlap.min = 0.5,
  trim_by_posterior = TRUE
)

Arguments

bf1 named vector of BF, or matrix of BF with colnames (cols=snps, rows=signals)
bf2 named vector of BF, or matrix of BF with colnames (cols=snps, rows=signals)
p1 prior probability a SNP is associated with trait 1, default 1e-4
p2 prior probability a SNP is associated with trait 2, default 1e-4
p12 prior probability a SNP is associated with both traits, default 1e-5
overlap.min see trim_by_posterior
trim_by_posterior it is important that the signals to be colocalised are covered by adequate numbers of snps in both datasets. If TRUE, signals for which snps in common do not capture least overlap.min proportion of their posteriors support are dropped and colocalisation not attempted.

Details

This is the workhorse behind many coloc functions

Value

coloc.signals style result

Author(s)

Chris Wallace

Description

Bayesian colocalisation analysis, detailed output
Usage

```r
coloc.detail(
  dataset1,
  dataset2,
  MAF = NULL,
  p1 = 1e-04,
  p2 = 1e-04,
  p12 = 1e-05
)
```

Arguments

- `dataset1`: a list with specifically named elements defining the dataset to be analysed. See `check_dataset` for details.
- `dataset2`: as above, for dataset 2
- `MAF`: Common minor allele frequency vector to be used for both dataset1 and dataset2, a shorthand for supplying the same vector as parts of both datasets
- `p1`: prior probability a SNP is associated with trait 1, default 1e-4
- `p2`: prior probability a SNP is associated with trait 2, default 1e-4
- `p12`: prior probability a SNP is associated with both traits, default 1e-5

Details

This function replicates coloc.abf, but outputs more detail for further processing using coloc.process Intended to be called internally by coloc.signals

Value

a list of three `data.tables`:

- `summary` is a vector giving the number of SNPs analysed, and the posterior probabilities of
  H0 (no causal variant), H1 (causal variant for trait 1 only), H2 (causal variant for trait 2 only),
  H3 (two distinct causal variants) and H4 (one common causal variant)
- `df` is an annotated version of the input data containing log Approximate Bayes Factors and
  intermediate calculations, and the posterior probability SNP.PP.H4 of the SNP being causal
  for the shared signal
- `df3` is the same for all 2 SNP H3 models

Author(s)

Chris Wallace

See Also

`coloc.process`, `coloc.abf`
coloc.process

Post process a coloc.details result using masking

Description

Internal helper function

Usage

```r
coloc.process(
  obj,
  hits1 = NULL,
  hits2 = NULL,
  LD = NULL,
  r2thr = 0.01,
  p1 = 1e-04,
  p2 = 1e-04,
  p12 = 1e-06,
  LD1 = LD,
  LD2 = LD,
  mode = c("iterative", "allbutone")
)
```

Arguments

- `obj`: object returned by coloc.detail()
- `hits1`: lead snps for trait 1. If length > 1, will use masking
- `hits2`: lead snps for trait 2. If length > 1, will use masking
- `LD`: named LD matrix (for masking)
- `r2thr`: r2 threshold at which to mask
- `p1`: prior probability a SNP is associated with trait 1, default 1e-4
- `p2`: prior probability a SNP is associated with trait 2, default 1e-4
- `p12`: prior probability a SNP is associated with both traits, default 1e-5
- `LD1`: named LD matrix (for masking) for trait 1 only
- `LD2`: named LD matrix (for masking) for trait 2 only
- `mode`: either "iterative" (default) - successively condition on signals or "allbutone" - find all putative signals and condition on all but one of them in each analysis

Value

data.table of coloc results

Author(s)

Chris Wallace
coloc.signals

* Coloc with multiple signals per trait

**Description**

New coloc function, builds on coloc.abf() by allowing for multiple independent causal variants per trait through conditioning or masking.

**Usage**

```r
coloc.signals(
  dataset1,
  dataset2,
  MAF = NULL,
  LD = NULL,
  method = c("single", "cond", "mask"),
  mode = c("iterative", "allbutone"),
  p1 = 1e-04,
  p2 = 1e-04,
  p12 = NULL,
  maxhits = 3,
  r2thr = 0.01,
  pthr = 1e-06
)
```

**Arguments**

- `dataset1`: a list with specifically named elements defining the dataset to be analysed. See `check_dataset` for details.
- `dataset2`: as above, for dataset 2
- `MAF`: Common minor allele frequency vector to be used for both dataset1 and dataset2, a shorthand for supplying the same vector as parts of both datasets
- `LD`: required if method="cond". matrix of genotype correlation (ie r, not r^2) between SNPs. If dataset1 and dataset2 may have different LD, you can instead add LD=LD1 to the list of dataset1 and a different LD matrix for dataset2
- `method`: default "" means do no conditioning, should return similar to coloc.abf. if method="cond", then use conditioning to coloc multiple signals. if method="mask", use masking to coloc multiple signals. if different datasets need different methods (eg LD is only available for one of them) you can set method on a per-dataset basis by adding method="..." to the list for that dataset.
- `mode`: "iterative" or "allbutone". Easiest understood with an example. Suppose there are 3 signal SNPs detected for trait 1, A, B, C and only one for trait 2, D.
  
  Under "iterative" mode, 3 coloc will be performed:
  * trait 1 - trait 2
  * trait 1 conditioned on A - trait 2
  * trait 1 conditioned on B - trait 2
  * trait 1 conditioned on C - trait 2
* trait 1 conditioned on A+B - trait 2

Under "allbutone" mode, they would be
* trait 1 conditioned on B+C - trait 2
* trait 1 conditioned on A+C - trait 2
* trait 1 conditioned on A+B - trait 2

Only iterative mode is supported for method="mask".

The allbutone mode is optimal if the signals are known with
certainty (which they never are), because it allows each
signal to be tested without influence of the others. When
there is uncertainty, it may make sense to use iterative mode,
because the strongest signals aren't affected by conditioning
incorrectly on weaker secondary and less certain signals.

p1 prior probability a SNP is associated with trait 1, default 1e-4
p2 prior probability a SNP is associated with trait 2, default 1e-4
p12 prior probability a SNP is associated with both traits, default 1e-5
maxhits maximum number of levels to condition/mask
r2thr if masking, the threshold on r2 should be used to call two signals independent.
our experience is that this needs to be set low to avoid double calling the same
strong signal.
pthr if masking or conditioning, what p value threshold to call a secondary hit "sig-
nificant"

Value
data.table of coloc results, one row per pair of lead snps detected in each dataset

Author(s)
Chris Wallace

coloc.susie run coloc using susie to detect separate signals

description

colocalisation with multiple causal variants via SuSiE
Usage

```r
coloc.susie(
  dataset1,
  dataset2,
  back_calculate_lbf = FALSE,
  susie.args = list(),
  ...
)
```

Arguments

dataset1  
*either* a coloc-style input dataset (see `check_dataset`), or the result of running `runsusie` on such a dataset

dataset2  
*either* a coloc-style input dataset (see `check_dataset`), or the result of running `runsusie` on such a dataset

back_calculate_lbf
by default, use the log Bayes factors returned by `susie_rss`. It is also possible to back-calculate these from the posterior probabilities. It is not advised to set this to TRUE, the option exists really for testing purposes only.

susie.args
a named list of additional arguments to be passed to `runsusie`

...  
other arguments passed to `coloc.bf_bf`, in particular prior values for causal association with one trait (p1, p2) or both (p12)

Value

a list, containing elements

* summary a data.table of posterior probabilities of each global hypothesis, one row per pairwise comparison of signals from the two traits
* results a data.table of detailed results giving the posterior probability for each snp to be jointly causal for both traits assuming H4 is true. Please ignore this column if the corresponding posterior support for H4 is not high.
* priors a vector of the priors used for the analysis

Author(s)

Chris Wallace

Description

`coloc.susie_bf`  
run coloc using susie to detect separate signals

Usage

```r
coloc.susie_bf(dataset1, bf2, p1 = 1e-04, p2 = 1e-04, p12 = 5e-06, ...)
```
**Arguments**

- **dataset1**: a list with specifically named elements defining the dataset to be analysed. See `check_dataset` for details.
- **bf2**: named vector of BF, names are snp ids and will be matched to column names of susie object’s alpha
- **p1**: prior probability a SNP is associated with trait 1, default 1e-4
- **p2**: prior probability a SNP is associated with trait 2, default 1e-4
- **p12**: prior probability a SNP is associated with both traits, default 1e-5
- **...**: other arguments passed to `coloc.bf_bf`, in particular prior values for causal association with one trait (p1, p2) or both (p12)

**Value**

coloc.signals style result

**Author(s)**

Chris Wallace

---

**Description**

Simulated data to use in testing and vignettes in the coloc package

**Usage**

data(coloc_test_data)

**Format**

A four of two coloc-style datasets. Elements D1 and D2 have a single shared causal variant, and 50 SNPs. Elements D3 and D4 have 100 SNPs, one shared causal variant, and one variant unique to D3. Use these as examples of what a coloc-style dataset for a quantitative trait should look like.

**Examples**

data(coloc_test_data)

names(coloc_test_data)

str(coloc_test_data$D1)

check_dataset(coloc_test_data$D1) # should return NULL if data structure is ok
**combine.abf**

**Description**

Internal function, calculate posterior probabilities for configurations, given logABFs for each SNP and prior probs

**Usage**

```r
combine.abf(l1, l2, p1, p2, p12)
```

**Arguments**

- `l1` `merged.df$lABF.df1`
- `l2` `merged.df$lABF.df2`
- `p1` prior probability a SNP is associated with trait 1, default 1e-4
- `p2` prior probability a SNP is associated with trait 2, default 1e-4
- `p12` prior probability a SNP is associated with both traits, default 1e-5

**Value**

named numeric vector of posterior probabilities

**Author(s)**

Claudia Giambartolomei, Chris Wallace

---

**estgeno.1.ctl**

**Description**

Estimate single snp frequency distributions

**Usage**

```r
estgeno.1.ctl(f)
estgeno.1.cse(G0, b)
```

**Arguments**

- `f` MAF
- `G0` single snp frequency in controls (vector of length 3) - obtained from estgeno.1.ctl
- `b` log odds ratio
est_cond

Value
relative frequency of genotypes 0, 1, 2

Author(s)
Chris Wallace

See Also
estgeno2

data.table giving snp, beta and varbeta on remaining snps after conditioning

Author(s)
Chris Wallace
find.best.signal

*Pick out snp with most extreme Z score*

**Description**
Internal helper function

**Usage**

\[
\text{find.best.signal}(D)
\]

**Arguments**

- \(D\): standard format coloc dataset

**Value**

- \(z\) at most significant snp, named by that snp id

**Author(s)**

Chris Wallace

---

finemap.abf

*Bayesian finemapping analysis*

**Description**
Bayesian finemapping analysis

**Usage**

\[
\text{finemap.abf}(\text{dataset}, p1 = 1e^{-04})
\]

**Arguments**

- \(\text{dataset}\): a list with specifically named elements defining the dataset to be analysed. See \texttt{check_dataset} for details.
- \(p1\): prior probability a SNP is associated with the trait 1, default \(1e^{-4}\)

**Details**
This function calculates posterior probabilities of different causal variant for a single trait.
If regression coefficients and variances are available, it calculates Bayes factors for association at each SNP. If only \(p\) values are available, it uses an approximation that depends on the SNP’s MAF and ignores any uncertainty in imputation. Regression coefficients should be used if available.
Value

a data.frame:

- an annotated version of the input data containing log Approximate Bayes Factors and intermediate calculations, and the posterior probability of the SNP being causal

Author(s)

Chris Wallace

Description

This is an analogue to finemap.abf, adapted to find multiple signals where they exist, via conditioning or masking - ie a stepwise procedure

Usage

finemap.signals(
  D,
  LD = D$LD,
  method = c("single", "mask", "cond"),
  r2thr = 0.01,
  sigsnps = NULL,
  pthr = 1e-06,
  maxhits = 3
)

Arguments

- **D**: list of summary stats for a single disease, see `check_dataset`
- **LD**: matrix of signed r values (not rsq!) giving correlation between SNPs
- **method**: if method="cond", then use conditioning to coloc multiple signals. The default is mask - this is less powerful, but safer because it does not assume that the LD matrix is properly allelically aligned to estimated effect
- **r2thr**: if mask==TRUE, all snps will be masked with r2 > r2thr with any sigsnps. Otherwise ignored
- **sigsnps**: SNPs already deemed significant, to condition on or mask, expressed as a numeric vector, whose names are the snp names
- **pthr**: when p > pthr, stop successive searching
- **maxhits**: maximum depth of conditioning. procedure will stop if p > pthr OR abs(z)<zthr OR maxhits hits have been found.
- **mask**: use masking if TRUE, otherwise conditioning. defaults to TRUE
logbf_to_pp

Value

list of successively significant fine mapped SNPs, named by the SNPs

Author(s)

Chris Wallace

Description

convert logbf matrix to PP matrix

Usage

logbf_to_pp(bf, pi, last_is_null)

Arguments

bf an L by p or p+1 matrix of log Bayes factors
pi either a scalar representing the prior probability for any snp to be causal, or a full vector of per snp / null prior probabilities
last_is_null TRUE if last value of the bf vector or last column of a bf matrix relates to the null hypothesis of no association. This is standard for SuSiE results, but may not be for BF constructed in other ways.

Value

matrix of posterior probabilities, same dimensions as bf

Author(s)

Chris Wallace
### logdiff

**Description**
Internal function, logdiff

**Usage**

```r
call(diff(x, y)
```

**Arguments**

- `x`: numeric
- `y`: numeric

**Details**

This function calculates the log of the difference of the exponentiated logs taking out the max, i.e. insuring that the difference is not negative

**Value**

```
max(x) + log(exp(x - max(x,y)) - exp(y-max(x,y))
```

**Author(s)**

Chris Wallace

---

### logsum

**Description**
Internal function, logsum

**Usage**

```r
logsum(x)
```

**Arguments**

- `x`: numeric vector
map_cond

Details

This function calculates the log of the sum of the exponentiated logs taking out the max, i.e. insuring that the sum is not Inf

Value

max(x) + log(sum(exp(x - max(x))))

Author(s)

Claudia Giambartolomei

map_cond | find the next most significant SNP, conditioning on a list of sigsnps

Description

Internal helper function for finemap.signals

Usage

map_cond(D, LD, YY, sigsnps = NULL)

Arguments

D          | dataset in standard coloc format
LD         | named matrix of r
YY         | sum(y^2)
sigsnps    | names of snps to mask

Value

named numeric - Z score named by snp

Author(s)

Chris Wallace
**map_mask**

*find the next most significant SNP, masking a list of sigsnps*

**map_mask**

**Description**

Internal helper function for finemap.signals

**Usage**

map_mask(D, LD, r2thr = 0.01, sigsnps = NULL)

**Arguments**

- **D**
  - dataset in standard coloc format
- **LD**
  - named matrix of r
- **r2thr**
  - mask all snps with r2 > r2thr with any in sigsnps
- **sigsns**
  - names of snps to mask

**Value**

named numeric - Z score named by snp

**Author(s)**

Chris Wallace

---

**plot.coloc_abf**

*plot a coloc_abf object*

**Description**

plot a coloc_abf object

**Usage**

## S3 method for class 'coloc_abf'
plot(x, ...)

**Arguments**

- **x**
  - coloc_abf object to be plotted
- **...**
  - other arguments

**Value**

ggplot object
plot_dataset

Author(s)

Chris Wallace

Description

Plot a coloc structured dataset

Usage

```
plot_dataset(
  d,
  susie_obj = NULL,
  alty = NULL,
  ylab = "-log10(p)",
  show_legend = TRUE,
  color = c("dodgerblue2", "green4", "#6A3D9A", "#FF7F00", "gold1", "skyblue2",
            "#FB9A99", "palegreen2", "#CAB2D6", "#FDBF6F", "gray70", "khaki2", "maroon",
            "orchid1", "deeppink1", "blue1", "steelblue4", "darkturquoise", "green1", "yellow4",
            "yellow3", "darkorange4", "brown"),
  ...)
```

Arguments

d a coloc dataset

susie_obj optional, the output of a call to runsusie()

alty default is to plot a standard manhattan. If you wish to plot a different y value, pass it here. You may also want to change ylab to describe what you are plotting.

ylab label for y axis, default is -log10(p) and assumes you are plotting a manhattan

show_legend optional, show the legend or not. default is TRUE

color optional, specify the colours to use for each credible set when susie_obj is supplied. Default is shamelessly copied from susieR::susie_plot() so that colours will match

... other arguments passed to the base graphics plot() function

Author(s)

Chris Wallace
### Description
Print summary of a coloc.abf run

### Usage
```r
## S3 method for class 'coloc_abf'
print(x, ...)
```

### Arguments
- `x` object of class coloc_abf returned by coloc.abf() or coloc.signals()
- `...` optional arguments: "trait1" name of trait 1, "trait2" name of trait 2

### Value
- `x`, invisibly

### Author(s)
Chris Wallace

---

### Description
Internal function, process each dataset list for coloc.abf

### Usage
```r
process.dataset(d, suffix)
```

### Arguments
- `d` list
- `suffix` "df1" or "df2"

### Value
- data.frame with log(abf) or log(bf)

### Author(s)
Chris Wallace
Run susie on a single coloc-structured dataset

Description

run susie_rss storing some additional information for coloc

Usage

```r
runsusie(
  d,
  suffix = 1,
  p = NULL,
  trimz = NULL,
  r2.prune = NULL,
  maxit = 100,
  repeat_until_convergence = TRUE,
  s_init = NULL,
  ...
)
```

Arguments

d colloc dataset, must include LD (signed correlation matrix)
suffix suffix label that will be printed with any error messages
p prior probability a snp is causal (equivalent to p1 or p2 in coloc.abf). By default, this is set to NULL, upon which we will set a small null_weight to pass to susie_rss() (see vignette a06 for details why). You can override this by setting p as you would p1 or p2 in a coloc function, but note that you may miss some true signals that way. Also note that neither of these options correspond to the susie_rss() defaults, because our goal here is not fine mapping alone.
trimz used to trim datasets for development purposes
r2.prune sometimes SuSiE can return multiple signals in high LD. if you set r2.prune to a value between 0 and 1, sets with index SNPs with LD greater than r2.prune
maxit maximum number of iterations for the first run of susie_rss(). If susie_rss() does not report convergence, runs will be extended assuming repeat_until_convergence=TRUE. Most users will not need to change this default.
repeat_until_convergence keep running until susie_rss() indicates convergence. Default TRUE. If FALSE, susie_rss() will run with maxit iterations, and if not converged, runsusie() will error. Most users will not need to change this default.
s_init used internally to extend runs that haven’t converged. don’t use.
... arguments passed to susie_rss. In particular, if you want to match some coloc defaults, set
• prior_variance=0.2^2 (if a case-control trait) or (0.15/sd(Y))^2 if a quantitative trait
• estimate_prior_variance=FALSE
otherwise susie_rss will estimate the prior variance itself

Value
results of a susie_rss run, with some added dimnames

Author(s)
Chris Wallace

Examples

library(coloc)
data(coloc_test_data)
result=runsusie(coloc_test_data$D1)
summary(result)

sdY.est

| sdY.est | Estimate trait variance, internal function |

Description
Estimate trait standard deviation given vectors of variance of coefficients, MAF and sample size

Usage

sdY.est(vbeta, maf, n)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>vbeta</td>
<td>vector of variance of coefficients</td>
</tr>
<tr>
<td>maf</td>
<td>vector of MAF (same length as vbeta)</td>
</tr>
<tr>
<td>n</td>
<td>sample size</td>
</tr>
</tbody>
</table>

Details
Estimate is based on var(beta-hat) = var(Y) / (n * var(X)) var(X) = 2maf(1-maf) so we can estimate var(Y) by regressing n*var(X) against 1/var(beta)

Value
estimated standard deviation of Y

Author(s)
Chris Wallace
sensitivity  \hspace{4cm} {\textit{Prior sensitivity for coloc}}

**Description**

Shows how prior and posterior per-hypothesis probabilities change as a function of \( p_{12} \)

**Usage**

```r
sensitivity(
  obj,
  rule = "",
  dataset1 = NULL,
  dataset2 = NULL,
  npoints = 100,
  doplot = TRUE,
  plot.manhattans = TRUE,
  preserve.par = FALSE,
  row = 1
)
```

**Arguments**

- **obj**: output of coloc.detail or coloc.process
- **rule**: a decision rule. This states what values of posterior probabilities "pass" some threshold. This is a string which will be parsed and evaluated, better explained by examples. "\( H_4 > 0.5 \)" says post prob of \( H_4 \) > 0.5 is a pass. "\( H_4 > 0.9 \ \& \ H_4/H_3 > 3 \)" says post prob of \( H_4 \) must be > 0.9 AND it must be at least 3 times the post prob of \( H_3 \)."
- **dataset1**: optional the dataset1 used to run SuSiE. This will be used to make a Manhattan plot if\( \text{plot.manhattans}=\text{TRUE} \).
- **dataset2**: optional the dataset2 used to run SuSiE. This will be used to make a Manhattan plot if\( \text{plot.manhattans}=\text{TRUE} \).
- **npoints**: the number of points over which to evaluate the prior values for \( p_{12} \), equally spaced on a log scale between \( p_1*p_2 \) and \( \min(p_1,p_2) \) - these are logical limits on \( p_{12} \), but not scientifically sensible values.
- **doplot**: draw the plot. set to \text{FALSE} if you want to just evaluate the prior and posterior matrices and work with them yourself
- **plot.manhattans**: if \text{TRUE}, show Manhattans of input data
- **preserve.par**: if \text{TRUE}, do not change par() of current graphics device - this is to allow sensitivity plots to be incorporated into a larger set of plots, or to be plot one per page on a pdf, foreexample
- **row**: when coloc.signals() has been used and multiple rows are returned in the coloc summary, which row to plot
Details

Function is called mainly for plotting side effect. It draws two plots, showing how prior and posterior probabilities of each coloc hypothesis change with changing p12. A decision rule sets the values of the posterior probabilities considered acceptable, and is used to shade in green the region of the plot for which the p12 prior would give an acceptable result. The user is encouraged to consider carefully whether some prior values shown within the green shaded region are sensible before accepting the hypothesis. If no shading is shown, then no priors give rise to an accepted result.

Value

list of 3: prior matrix, posterior matrix, and a pass/fail indicator (returned invisibly)

Author(s)

Chris Wallace

Description

variance of MLE of beta for quantitative trait, assuming var(y)=1

Usage

Var.data(f, N)

Arguments

f         minor allele freq
N         sample number

Details

Internal function

Value

variance of MLE beta

Author(s)

Claudia Giambartolomei
Var.data.cc

Description
variance of MLE of beta for case-control

Usage
Var.data.cc(f, N, s)

Arguments
f minor allele freq
N sample number
s ???

Details
Internal function

Value
variance of MLE beta

Author(s)
Claudia Giambartolomei
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