Package ‘mapbayr’

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BugReports https://github.com/FelicienLL/mapbayr/issues

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add_covariates

Add covariate columns to a dataset

Description

The add_covariates() function adds an one or several covariate columns to a dataset provided as a proper data.frame or within a 'mrgsolve' model. Used in combination with adm_rows() and obs_rows(), it helps the creation of datasets in the proper format for simulations with 'mrgsolve' or parameter estimation with 'mapbayr', as explained in data_helpers.

Usage

```r
add_covariates(x, ...)
```

```r
## S3 method for class 'data.frame'
add_covariates(x, ..., covariates = list(), AOLA = FALSE, TOLA = FALSE)
```

```r
## S3 method for class 'mrgmod'
add_covariates(x, ..., covariates = list(), AOLA = NULL, TOLA = NULL)
```
Arguments

- **x**: either a data.frame or a `mrgsolve` model object
- **...**: covariates values to add to the data. For each variable, supply a vector of length 1 or with the same number of rows. Ignored if `covariates` argument is used.
- **covariates**: Covariates passed as a single list of variables. Overrides `...`.
- **AOLA, TOLA**: a logical. Should the "Amount Of Last Administration" and "Time Of Last Administration" variables be added into the dataset? Default if FALSE if `x` is a dataset, TRUE if `x` is a `mrgsolve` model where `AOLA` and `TOLA` are defined as covariates.

Value

A data.frame, or a `mrgsolve` model with a dataset in the `@args$data` slot (accessible with `get_data()`).

See Also

data_helpers

Examples

```r
# Cannot start from scratch
## Not run:
add_covariates(BW = 90, SEX = 0)
## End(Not run)

library(magrittr)
adm_rows(time = c(0, 24, 48), cmt = 1, amt = c(100, 200, 300)) %>%
  add_covariates(BW = c(90, 85, 80), SEX = 0)

# If covariates are stored in a list, use `covariates =`
adm_rows(time = c(0, 24, 48), cmt = 1, amt = c(100, 200, 300)) %>%
  add_covariates(covariates = list(BW = c(90, 85, 80), SEX = 0))

# Missing values are filled with the "next observation carried backward" rule
adm_rows(time = c(0, 24, 48), cmt = 1, amt = c(100, 200, 300)) %>%
  add_covariates(BW = c(90, 85, 80), SEX = 0) %>%
  obs_rows(time = 36, DV = .0123, cmt = 2)
# Always verify the output in case of time-varying covariates

adm_rows(time = c(0, 24, 48), amt = c(100, 200, 300), cmt = 1) %>%
  obs_rows(time = c(8, 16, 32, 40), cmt = 2, DV = runif(4)) %>%
  add_covariates(TOLA = TRUE, AOLA = TRUE) %>%
  obs_rows(time = 72, cmt = 2, DV = .123) # AOLA/TOLA re-updated afterwards

# Automatic inclusion of `TOLA`/`AOLA` if they are covariates of the model
library(mrgsolve)
model <- mcode("model", 
$PARAM @annotated @covariates
```
adm_rows

Add administration lines to a dataset

Description

The `adm_rows()` function adds one or several administration lines to a dataset provided as a proper data.frame or within a 'mrgsolve' model. Used in combination with `obs_rows()` and `add_covariates()`, it helps the creation of datasets in the proper format for simulations with 'mrgsolve' or parameter estimation with 'mapbayr', as explained in data_helpers.

Usage

```r
adm_rows(x, ...)```

## S3 method for class 'data.frame'
```r
adm_rows(
  x,
  ID = NULL,
  time = NULL,
  evid = 1L,
  cmt,
  amt,
  mdv = 1L,
  addl = NULL,
  ss = NULL,
  ii = NULL,
  rate = NULL,
  .datehour = NULL,
  ...
)
```

## S3 method for class 'missing'
```r
adm_rows(...)
```

## S3 method for class 'mrgmod'
```r
adm_rows(x, cmt = adm_cmt(x), rate = NULL, ...)
```
**Arguments**

- **x**: either a data.frame or a `mrgsolve` model object
- **...**: additional columns or arguments for `mrgsolve::ev()`
- **ID**: subject ID (default is 1)
- **time**: event time. Default is 0 if no previous events. Mind consistency with `.datehour`.
- **evid**: event identification (default is 1 for administration, 0 for observation)
- **cmt**: compartment (no default, except if [ADM] was tagged in the $CMT block in model code. See examples.)
- **amt**: dose amount (for administration records only)
- **mdv**: missing dependent value (default is 1 for administration records)
- **addl**: additional dose (optional)
- **ss**: steady-state (optional, is this dose the last of an infinity of administration? Yes, 1, or no, 0)
- **ii**: inter-dose interval (optional, use it with ss and addl)
- **rate**: rate of administration (optional, set to -2 if you model zero-order infusion. See examples.)
- **.datehour**: a object of class POSIXct, a number or a character vector that can be passed to `parse_datehour()`. Using `.datehour` will update the value of `time` in the dataset, with units in hours. Mind consistency with the `time` argument.

**Value**

a data.frame, or a `mrgsolve` model with a dataset in the @args$data slot (accessible with `get_data()`).

**See Also**

data_helpers

**Examples**

```r
# Create a dataset from scratch
adm_rows(amt = 100, cmt = 1)

# Pipe-friendly addition of administration record to a pre-existing dataset
library(magrittr)
amd_rows(amt = 100, cmt = 1) %>%
  adm_rows(time = 3, amt = 200, cmt = 1, addl = 3, ii = 1)

# Inform times using the `.datehour` argument:
amd_rows(.datehour = "2020-01-01 11:11", amt = 100, cmt = 1) %>%
amd_rows(.datehour = "2020-01-02 22:22", amt = 200, cmt = 1) %>%
amd_rows(time = 48, amt = 300, cmt = 1)

# Start from a 'mrgsolve' model
library(mrgsolve)
house() %>%
```
```r
adm_rows(amt = 100, cmt = 1) %>%
adm_rows(time = 3, amt = 200, cmt = 1, addl = 3, ii = 1) %>%
mrgsim(delta = 1)

# Default administration compartments
# Set default administration compartments in the code with `[ADM]`
model <- mcode("model","
$CMT @annotated
DEPOT : Depot [ADM]
CENTR : Central
" , compile = FALSE)
adm_cmt(model)

# Thus, no need to manually specify `cmt = 1` anymore.
model %>%
  adm_rows(amt = 100) %>%
  adm_rows(time = 3, amt = 200, addl = 3, ii = 1) %>%
  get_data()

# Automatic lines duplication if multiple depot compartments
# Automatic `rate = -2` increment if model with 0-order absorption
model <- mcode("model","
$PARAM DUR = 1.0
$CMT @annotated
DEPOT : Depot [ADM]
CENTR : Central [ADM]
$MAIN
D_CENTR = DUR ;
" , compile = FALSE)
adm_cmt(model)

model %>%
  adm_rows(amt = 100) %>%
  adm_rows(time = 3, amt = 200, addl = 3, ii = 1) %>%
  get_data()
```

---

**as.data.frame.mapbayests**

*Return the mapbay_tab as a data.frame*

---

**Description**

Return the mapbay_tab as a data.frame

**Usage**

```r
## S3 method for class 'mapbayests'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```
augment

Arguments

x A mapbayests object.
row.names, optional, ...
    passed to as.data.frame

Value

a data.frame (the mapbay_tab from estimation)

augment Compute full PK profile prediction from mapbayr estimates.

Description

Compute full PK profile prediction from mapbayr estimates.

Usage

augment(x, ...)

Arguments

x object to augment
...
    additional arguments

Value

an augmented object (depending on the object passed).

augment.mapbayests Compute full PK profile prediction from mapbayr estimates.

Description

Compute full PK profile prediction from mapbayr estimates.
Usage

```r
## S3 method for class 'mapbayests'
augment(
  x,
  data = NULL,
  start = NULL,
  end = NULL,
  delta = NULL,
  ci = FALSE,
  ci_width = 90,
  ci_method = "delta",
  ci_sims = 500,
  ...
)
```

Arguments

- **x**: A mapbayests object.
- **data**: dataset to pass to mrgsolve for simulation (default is dataset used for estimation)
- **start, end, delta**: start, end and delta of simulation time passed to mrgsim() (see details)
- **ci**: a logical. If TRUE, compute a confidence interval around the prediction (default is FALSE)
- **ci_width**: a number between 0 and 100, width of the confidence interval (default is "90" for a 90% CI)
- **ci_method**: method to compute the confidence interval. Can be "delta" (the default) to use the Delta approximation. Alternatively "simulations" for a more accurate approach, but also more time-consuming.
- **ci_sims**: number of replicates to simulate in order to derive the confidence interval (default is 500)
- **...**: additional arguments passed to mrgsim()

Details

This function is called in the background by plot() in order to simulate the full PK profile, and return a mapbayests object with an additional aug_tab data.frame inside. The latter is used with by the plot method. The time grid, for each PK profile (i.e. patient) is defaulted with the minimum time in the dataset for start and the maximum time in the dataset +20% for end. delta is a power of 10 (e.g. 0.1, 1, 10 etc...), automatically chosen to render visually appealing graphs with a reasonable computing time (about 200 time points). Additional arguments can be passed to mrgsim() through ... Note that recsort is set to 3 (see mrgsolve documentation for more details).

Value

A mapbayests object, augmented of an aug_tab data.frame.
Examples

```r
x <- ~x~ is the result of `mapbayest()`.  
# Default plot is returned by: 
# plot(x)  
# Argument passed to `plot()` are passed to `augment()` in the background:  
# plot(x, end = 240, ci = TRUE)  
# Save the augmented object if simulation time is long  
# x2 <- augment(x, ci = TRUE, ci_method = "simulations", ci_sims = 10000) %>%  
# plot(x2)
```

Description

Checks that the model respects points related exclusively to `mapbayr`. Useful at the time you wish to convert a "regular" `mrgsolve` model you used for simulation into a model to perform MAP-Bayesian estimation. Note that some elements cannot be checked:

- In `$MAIN` block, make sure that you added ETA1, ETA2... in the code. For instance: `double CL = TVCL * exp(ETA(1) + ETA1);`
- In `$OMEGA` block, make sure the order of the (diagonal) values is the same as for ETAs in `$PARAM`. For instance, if ETA1 corresponds to clearance, the first value in `$OMEGA` must be the variance of clearance.
- In `$SIGMA` block, make sure the order is respected: proportional error first, and additive error secondly.

Usage

```r
check_mapbayr_model(x, check_compile = TRUE)
```

Arguments

- `x` : model file
- `check_compile` : check if model is compiled

Value

`TRUE` (invisibly) if checks are passed, errors otherwise.

Examples

```r
library(mapbayr)  
library(mrgsolve)  
## Not run: check_mapbayr_model(house())
```
**compute_ofv**

*Compute the objective function value*

**Description**

Compute the objective function value

**Usage**

```r
compute_ofv(
  eta,
  qmod,
  sigma,
  omega_inv,
  all_cmt,
  log_transformation,
  lambda = 1,
  idvaliddata,
  idDV,
  idcmt,
  idblq = NULL,
  idlloq = NULL,
  ...
)
```

```r
do_compute_ofv(eta, argofv, ...)
```

**Arguments**

- `eta` a named vector/list of parameters
- `qmod, sigma, log_transformation, omega_inv, all_cmt, lambda` generated by `preprocess.ofv.fix`
- `idvaliddata, idDV, idcmt` generated by `preprocess.ofv.id`
- `idblq, idlloq` optionally generated by `preprocess.ofv.id`
- `...` for compatibility (not used)
- `argofv` above mentioned arguments as a list

**Details**

This function is called iteratively by the optimization function. Arguments should not be passed directly, but generated by the pre-processing functions (see `preprocess.ofv`).

**Value**

a single numeric value (the objective function value)
**Description**

Use `adm_rows()`, `obs_rows()` and `add_covariates()` to create or modify a dataset from scratch, from a pre-existing dataset, or from a dataset stored into a 'mrgsolve' model.

**Details**

Instead of importing a `.csv` file, or painfully build a data set with a call to `data.frame()` and mind how to format the data, you can pass information about:

- administrations with `adm_rows()`,
- observations with `obs_rows()`.
- covariates with `add_covariates()`.

all being called jointly with a pipe ( `%>%` or `|>`) . These functions can be used to create or modify a dataset as a proper data.frame, or to create or modify a dataset within a 'mrgsolve' model (`@args$data` slot). The latter is particularly useful in order to:

- automatically use default administration and observation compartments,
- automatically duplicate rows if there are several depot compartments,
- automatically set `rate = -2` if model has zero-order absorption pathways,
- automatically duplicate rows if concentrations of Parent drug and Metabolite are observed together,
- automatically add "Amount Of Last Administration" and "Time Of Last Administration" variables if these are covariates,
- subsequently call `mrgsim()` or `mapbayest()`.

**Examples**

```r
library(magrittr)

# First option: work with a data.frame
adm_rows(amt = 1000, cmt = 1, addl = 4, ii = 12) %>%
  obs_rows(time = c(12.3, 45.6), DV = c(.111, .222), cmt = 2) %>%
  obs_rows(time = 48, cmt = 2) %>%
  add_covariates(BW = 90, SEX = 0, TOLA = TRUE)

# You can even inform "time" using date and hours:
adm_rows(amt = 1000, cmt = 1, addl = 4, ii = 12, .datehour = "2022-01-01 11:11:11") %>%
  obs_rows(.datehour = "2022-01-02 22:22:22", DV = 0.111, cmt = 2)

# Second option: work with a dataset within a 'mrgsolve' model
mod <- exmodel(add_exdata = FALSE)
```
# call `mrgsolve::see(mod)` to see how default compartment were coded
adm_cmt(mod)
obs_cmt(mod)

mod %>%
  adm_rows(amt = 10000) %>%
  obs_rows(time = c(1.5, 4.4, 7.5, 24.6), DV = c(91.2904, 110.826, 79.384, 20.6671)) %>%
  # get_data() # for curiosity, you can extract the data set at this step
mapbayest()

deprecations

---

### Deprecated functions

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### Usage

- `mbrest(...)`
- `adm_lines(...)`
- `obs_lines(...)`

### Arguments

- `...` passed to the corresponding function

### Details

- `mbrest()` is now `mapbayest()`
- `adm_lines()` is now `adm_rows()`
- `obs_lines()` is now `obs_rows()`
An example of `mapbayests` object, corresponding to the parameter estimation of the 8 subjects from model 1. Note that the model object within is not associated to a shared object, which make some features unavailable. This object can be re-generated by executing `est001 <- mapbayest(exmodel(ID = 1:8))`.

### Usage

```r
est001
```

### Format

An object of class `mapbayests` of length 9.

### See Also

`mapbayest`

---

## `eta`

Generate a vector of "ETA"

### Description

Generate a vector of "ETA" values. If `x` is a `mrgsolve` model, these will be extracted from values defined in `$PARAM`. Otherwise, any numeric values passed to `x` and `. . .` as vector(s) or list(s) will be coerced as a single vector. Alternatively, if `x` and `. . .` are missing, generate a vector of ETA equal to `val` of length `n`.

### Usage

```r
teta(x, ..., n, val = 0)
```

### Arguments

- `x`: either a `mrgsolve` model object, or a numeric
- `. . .`: additional numeric(s)
- `n`, `val`: generate a sequence of `val` of length `n`

### Value

A single named vector of numeric
Examples

# Extract ETA from the model
mod <- exmodel()
eta(mod)

# Coerce numeric values
eta(0.1, 0.2, c(0.3, 0.4), list(0.5, 0.6))
etarunorm(4))

# Generate a sequence from scratch
eta(n = 3)
etan(3, val = 0.001)

---

exmodel_exdata  Example model and data

Description

A collection of example models and corresponding data to test and explore mapbayr.

Usage

exmodel(
  num = 1,
  add_exdata = TRUE,
  cache = TRUE,
  quiet =getOption("mrgsolve_mread_quiet", TRUE),
  ...
)
exdata(num = 1, ID = 1, clean_data = TRUE)

Arguments

num  model number (see details)
add_exdata  should data be automatically loaded with the model
cache  read the model with mrgsolve::mread_cache()
quiet  don’t print messages when compiling
...  passed to mrgsolve::mread() or mrgsolve::mread_cache()
ID  individual number to include in the data (from 1 to 8)
clean_data  remove useless columns and rows from the original data
**filter.mrgmod**

**Details**

Available models are:

- **1**: Base model. A simple monocompartmental PK model with inter-individual variability on absorption constant (KA), volume of distribution (VC) and clearance (CL). The residual error model is proportional.
- **6**: Complex absorption model. Dual 0- and 1st orders absorption phenomenons.
- **301**: Time-varying covariates. A continuous covariate (body weight "BW") and a categorical one (sex "SEX") influence the clearance parameter. In the corresponding dataset, the values randomly changes from one record to another within a single individual.
- **401**: Metabolite. The PK model of both a parent drug and its metabolite.

An example dataset of eight (simulated) individuals is available for each model. Individuals differ in terms of sampling times (sparse or rich) and dosing regimen (single or multiple dosing).

Model code and data files are stored at the location given by `system.file("exmodel", package = "mapbayr")`.

These models and data were created for the validation study of mapbayr published in CPT:Pharmacometrics & System Pharmacology. More models and full datasets can be accessed in a dedicated repository.

**Value**

`exmodel()` reads and compiles code, and returns a (mrgmod) model object. `exdata()` returns a data.frame.

**Source**

https://github.com/FelicienLL/mapbayr-CPTPSP-2021

**Examples**

```r
# Models can be loaded with data (the default), ready for parameter estimation
est <- mapbayest(exmodel())

# Number of subjects in dataset can be chosen up to 8 individuals
exdata(301, ID = c(5,8))
```

---

**filter.mrgmod**

*Filter a dataset within a mrgmod*

**Description**

Filter a dataset within a mrgmod

**Usage**

```r
## S3 method for class 'mrgmod'
filter(.data, ..., .preserve = FALSE)
```
Arguments
.data a mrgmod
... .preserve additional arguments for dplyr::filter()

Value
a mrgmod

Examples
library(magrittr)
mod <- mrgsolve::mcode("mod", "$CMT FOO", compile = FALSE)
mod %>%
  adm_rows(amt = c(100, 200, 300), cmt = 1) %>%
  filter(amt != 200) %>%
  get_data()

Description
Helpful functions to get content from a mrgmod object (i.e. data) or from a mapbayests object (data, eta, cov, param, phi).

Usage
get_data(x, ...)

## S3 method for class 'mrgmod'
get_data(x, ...)

## S3 method for class 'mapbayests'
get_data(x, ..., output = "df")

get_eta(x, ...)

## S3 method for class 'mapbayests'
get_eta(x, ..., output = NULL)

get_cov(x, ...)

## S3 method for class 'mapbayests'
get_cov(x, ..., simplify = TRUE)

get_param(x, ...)
get_x

## S3 method for class 'mapbayests'
get_param(x, ..., output = NULL, keep_ID = NULL, keep_names = NULL)

get_phi(x, ...)

## S3 method for class 'mapbayests'
get_phi(x, ...)

Arguments

- **x**: mapbayests object
- **...**: not used
- **output**: either a data.frame ("df") or a vector of numeric ("num"). Default to "num" if only one ID
- **simplify**: a logical. If TRUE (the default) and only one ID, one matrix is returned instead of a list of length 1
- **keep_ID**: a logical. By default, the ID variable is dropped if one ID in data.
- **keep_names**: a logical. By default, names are dropped if one parameter is requested, and output is not a data frame.

Value

the class of the object returned depends on the function, and on their arguments. Typically, a data.frame or a vector if the output can be reduced to one line.

Examples

# From a model object (mrgmod)
mod <- exmodel(ID = 1:2, cache = FALSE, capture = "CL")
get_data(mod)

# From an estimation object (mapbayests)
est <- mapbayest(mod)
get_data(est)
get_data(est, output = "list")

geta(est)
geta(est, output = "list")

gcov(est)
gparam(est)
gphi(est)
hist.mapbayests

Plot posterior distribution of bayesian estimates

Description
Plot posterior distribution of bayesian estimates

Usage
## S3 method for class 'mapbayests'
hist(x, select_eta = x$arg.optim$select_eta, ...)

Arguments
x            A mapbayests object.
select_eta   a vector of numeric values, the numbers of the ETAs to show (default are estimated ETAs).
...          additional arguments (not used)

Details
Use this function to plot the results of the estimations, in the form of histograms with the a priori distribution in the background. For every parameter, the inter-individual variability is displayed, as well as the percentile of the patient in the corresponding distribution (if n = 1 patient). For additional modifications, you can add extra +function(...) in order to modify the plot as a regular ggplot2 object.

Value
a ggplot object.

Examples

est <- mapbayest(exmodel(ID = 1))

# Default Method
h <- hist(est)

# Can be modified with `ggplot2`
h +
ggplot2::labs(title = "Awesome estimations")

# Select the ETAs
hist(est, select_eta = c(1,3))
mapbayest

Estimate parameters (maximum a posteriori)

Description

The main function of the mapbayr package. Performs a maximum a posteriori Bayesian estimation of parameters, from a mrgsolve model object and a dataset containing information about administrations and observed concentrations.

Usage

mapbayest(
  x,
  data = NULL,
  method = c("L-BFGS-B", "newuoa"),
  hessian = stats::optimHess,
  select_eta = NULL,
  lambda = 1,
  lloq = NULL,
  force_initial_eta = NULL,
  quantile_bound = 0.001,
  control = list(),
  check = TRUE,
  verbose = TRUE,
  progress = TRUE,
  reset = 50,
  output = NULL,
  ...
)

Arguments

x the model object
data NMTRAN-like data set
method optimization method; the default is "L-BFGS-B" (from stats::optim()), alternatively "newuoa" for minqa::newuoa()
hessian function used to compute the Hessian and variance-covariance matrix with (default is stats::optimHess, alternatively use nlmixr::nlmixrHess)
select_eta a vector of numeric values, the numbers of the ETAs to be estimated (default is NULL, all ETAs non-equal to zero)
lambda a numeric value, the weight applied to the model prior (default is 1)
lloq a numeric value, the lower limit of quantification. If not NULL, LLOQ and BLQ (below limit of quantification) variables will be added to the data. The related records will be censored with the M3 method. Ignored if LLOQ already in the data.
force_initial_eta

a vector of numeric values to start the estimation from (default to 0 for "L-BFGS-B")

quantile_bound

a numeric value representing the quantile of the normal distribution admitted to define the bounds for L-BFGS-B (default is 0.001, i.e. 0.1%)

control

a list passed to the optimizer (see stats::optim() or minqa::newuoa() documentation)

check

check model code for mapbayr specification (a logical, default is TRUE)

verbose

print a message whenever optimization is reset (a logical, default is TRUE)

progress

print a progress bar (a logical, default is TRUE)

reset

maximum allowed reset of the optimizer with new initial eta values if numerical difficulties, or with new bounds (L-BFGS-B) if estimate equal to a bound. (a numeric, default is 50)

output

if NULL (the default) a mapbayests object is returned; if df a mapbay_tab dataframe is returned

... for compatibility (not used)

Value

a mapbayests object. Basically a list containing:

- model: the model object
- arg.ofv.optim, arg.ofv.fix, arg.ofv.id: arguments passed to the optimization function. Useful for debugging but not relevant for a basic usage. Access to the data with get_data(x)
- opt.value: the original output of the optimization function
- final_eta: a list of individual vectors of final estimates. Access it with x$final_eta or get_eta(x).
- covariance: a list of individual variance-covariance matrix of estimation. Access it with x$covariance or get_cov(x).
- mapbay_tab: an output table containing the results of your estimations (data, IPRED, PRED, covariates, captured items, ETA etc...). Access it with x$mapbay_tab, as.data.frame(x) or as_tibble(x).
- information: run times and package versions.

See Also

hist.mapbayests
plot.mapbayests
use_posterior
Examples

# First, code a model
code1 <- "$PARAM ETA1 = 0, ETA2 = 0, KA = 0.5, TVCL = 1.1, TVV = 23.3
$OMEGA 0.41 0.32
$SIGMA 0.04 0
$CMT DEPOT CENT
$PK
double CL=TVCL*exp(ETA1+ETA(1));
double V=TVV*exp(ETA2+ETA(2));
$ERROR
double DV=CENT/V*(1+EPS(1))+EPS(2);
$PKMODEL ncmt = 1, depot = TRUE
$CAPTURE DV CL"

my_model <- mrgsolve::mcode("my_model", code1)

# Then, import your data
my_data <- data.frame(ID = 1, TIME = c(0, 1.1, 5.2, 12.3), EVID = c(1,0,0,0), AMT = c(500, 0,0,0),
CMT = c(1,2,2,2), DV = c(0, 15.1, 29.5, 22.3))
print(my_data)

# And estimate
my_est <- mapbayest(x = my_model, data = my_data)
print(my_est)
# see also plot(my_est) and hist(my_est)

# Use your estimation
get_eta(my_est)
get_param(my_est)
as.data.frame(my_est)
use_posterior(my_est)

obs_rows

Description

The obs_rows() function adds an one or several observation lines to a dataset provided as a proper data.frame or within a 'mrgsolve' model. Used in combination with adm_rows() and add_covariates(), it helps the creation of datasets in the proper format for simulations with 'mrgsolve' or parameter estimation with 'mapbayr', as explained in data_helpers.

Usage

obs_rows(x, ...)

## S3 method for class 'data.frame'
obs_rows(
  x,
  ID = NULL,
  time = NULL,
  evid = 0L,
  cmt,
  DV = NA_real_,
  mdv = NULL,
  .datehour = NULL,
  ...
)

## S3 method for class 'missing'
obs_rows(...)

## S3 method for class 'mrgmod'
obserows(x, cmt = NULL, DV = NA_real_, DVmet = NULL, ...)

Arguments

x either a data.frame or a 'mrgsolve' model object
...
  additional columns
ID subject ID (default is 1)
time event time. Default is 0 if no previous events. Mind consistency with .datehour.
evid event identification (default is 1 for administration, 0 for observation)
cmt compartment (no default, except if [OBS] was tagged in the $CMT block in model code. See examples.)
DV dependent value, i.e. observed concentration.
mdv missing dependent value (default is 0 a non-missing concentration value to take into account for parameter estimation, 1 otherwise)
.datehour a object of class POSIXct, a number or a character vector that can be passed to parse_datehour(). Using .datehour will update the value of time in the dataset, with units in hours. Mind consistency with the time argument.
DVmet second observation at the same time (e.g. a metabolite, "DVmet") observed jointly with parent drug ("DV"). Works only if x is a 'mrgsolve' model where two [OBS] compartments were defined (see examples)

Value

a data.frame, or a 'mrgsolve' model with a dataset in the @args$data slot (accessible with get_data()).

See Also
data_helpers
Examples

# Create a dataset from scratch
obs_rows(time = 12, DV = 0.12, cmt = 2)

# Pipe-friendly addition of observation record to a pre-existing dataset
library(magrittr)
obs_rows(time = 12, DV = 0.12, cmt = 2) %>%
  obs_rows(time = c(24, 36, 48), DV = c(0.34, 0.56, 0.78), mdv = c(0, 1, 0), cmt = 2)

# Inform times using the `.datehour` argument:
obs_rows(.datehour = "2020-01-01 11:11", DV = 0.12, cmt = 1) %>%
  obs_rows(.datehour = "2020-01-02 22:22", DV = 0.34, cmt = 1) %>%
  obs_rows(time = 48, DV = 0.56, cmt = 1)

# Start from a `mrgsolve` model
library(mrgsolve)
house() %>%
  obs_rows(time = 12, DV = 0.12, cmt = 2) %>%
  obs_rows(time = c(24, 36, 48), DV = c(0.34, 0.56, 0.78), mdv = c(0, 1, 0), cmt = 2) %>%
  mrgsim()

# Default observation compartments
# Set default observation compartments in the code with `[OBS]`
model <- mcode("model", "$CMT @annotated
DEPOT : Depot
CENTR : Central [OBS]
"
, compile = FALSE)
obs_cmt(model)

# Thus, no need to manually specify `cmt = 2` anymore.
model %>%
  obs_rows(time = 12, DV = 0.12) %>%
  obs_rows(time = c(24, 36, 48), DV = c(0.34, 0.56, 0.78), mdv = c(0, 1, 0)) %>%
  get_data()

# Automatic lines duplication if parent + metabolite defined in the model
model <- mcode("model", "$CMT @annotated
DEPOT : Depot
CENTR : Central [OBS]
PERIPH : Periph
METABO : Metabo [OBS]
"
, compile = FALSE)
obs_cmt(model)

model %>%
  obs_rows(time = 12, DV = 0.12, DVmet = 120) %>%
  obs_rows(
    time = c(24, 36, 48), DV = c(0.34, 0.56, 0.78),
    mdv = c(0, 1, 0), DVmet = c(340, 560, 780)
  ) %>%
Description
A wrapper around functions of lubridate, mainly in order to transform characters into a date-time ("POSIXct") format.

Usage

```r
parse_datehour(
  x,
  orders = getOption("mapbayr.datehour", default = c("Ymd HMS", "Ymd HM", "dmY HMS", "dmY HM"))
)
```

Arguments

- `x` a numeric or a character.
- `orders` format specification for `x`, passed to `lubridate::parse_date_time()`

Value

a POSIXct

Examples

```r
# POSIXct are returned as is.
parse_datehour(x = as.POSIXct("2022-02-02 22:22:22", tz = "UTC"))

# Numerics are passed to `lubridate::as_datetime()`.
parse_datehour(1643840542)

# Characters are passed to `lubridate::parse_date_time()`.
# The format used will be the one defined in `orders`
parse_datehour(x = "2022-02-02 22:22:22", orders = "Ymd HMS")
parse_datehour(x = "02-02-2022 22:22", orders = "dmY HM")

# By default, the following formats will be subsequently tried:
# "Ymd HMS", "Ymd HM", "dmY HMS", "dmY HM"

# Alternatively, set a format through `options(mapbayr.datehour)`.
# Convenient for the use `.datehour` in `.adm_rows()` and `.obs_rows()`.

# Following format will return NA:
adm_rows(.datehour = "22:22 02-02-2022", amt = 100, cmt = 1)
```
options(mapbayr.datehour = "H%M dmy")
adm_rows(.datehour = "22:22 02-02-2022", amt = 100, cmt = 1)
options(mapbayr.datehour = NULL)

plot.mapbayests  
Plot predictions from mapbayests object

Description
Plot predictions from mapbayests object

Usage
## S3 method for class 'mapbayests'
plot(x, ..., PREDICTION = c("IPRED", "PRED"))

Arguments
x      A mapbayests object.
...    additional arguments (passed to augment.mapbayests)
PREDICTION  plot either "IPRED", "PRED" or both.

Details
Use this function to plot the results of the estimations, in the form of concentration vs time profiles for every patient of the data set. For additional modifications, you can:

• see augment.mapbayests to modify the simulation output.
• add extra +function(...) in order to modify the plot as a regular ggplot2 object.

Value
a ggplot object.

Examples
est <- mapbayest(exmodel(ID = 1))
plot(est, end = 48) +
  ggplot2::labs(title = "Awesome prediction")
Preprocess model and data for ofv computation

Description

Functions to generate arguments passed to compute_ofv. Arguments that are fixed between individuals are created once (preprocess.ofv.fix), while others are specific of each individual (preprocess.ofv.id).

Usage

preprocess.ofv.fix(x, data, select_eta = seq_along(eta(x)), lambda = 1)

preprocess.ofv.id(x, iddata)

Arguments

x the model object
data, iddata NMTRAN-like data set. iddata is likely a dataset of one individual
select_eta numbers of the ETAs taken into account. Set the dimensions of the inversed OMEGA matrix
lambda a numeric value, the weight applied to the model prior (default is 1)

Value

A list of arguments used to compute the objective function value.

The following arguments are fixed between individuals:

- `qmod`: model object, modified to simulate without random effects and with controlled outputs
- `sigma`: a single matrix object
- `log_transformation`: a logical, whether predictions need to be log-transformed for ofv computation
- `omega_inv`: a single matrix object
- `all_cmt`: a vector of compartment numbers where observations can be expected

The following arguments differs between individuals:

- `idvaliddata`: a matrix, individual data set (with administrations and covariates), validated with `valid_data_set`
- `idDV`: a vector of (possibly log-transformed) observations
- `idcmt`: a vector of compartments where observations belong to
- `idblq,idlloq`: optional, a logical and numerical vector indicating if the observation is below the lower limit of quantification, and the LLOQ value, respectively
**Examples**

```r
mod <- exmodel(add_exdata = FALSE, compile = FALSE)
dat <- exdata(ID = c(1,4))

preprocess.ofv.fix(x = mod, data = dat)
preprocess.ofv.id(x = mod, iddata = dat[dat$ID == 1,])
preprocess.ofv.id(x = mod, iddata = dat[dat$ID == 4,])
```

---

### preprocess.optim

*Pre-process: arguments for optimization function*

#### Description

Pre-process: arguments for optimization function

#### Usage

```r
preprocess.optim(
  x,
  method = c("L-BFGS-B", "newuoa"),
  select_eta = NULL,
  control = list(),
  force_initial_eta = NULL,
  quantile_bound = 0.001
)
```

#### Arguments

- **x**: the model object
- **method**: optimization method; the default is "L-BFGS-B" (from `stat::optim()`), alternatively "newuoa" for `minqa::newuoa()`
- **select_eta**: a vector of numeric values, the numbers of the ETAs to be estimated (default is NULL, all ETAs non-equal to zero)
- **control**: a list passed to the optimizer (see `stats::optim()` or `minqa::newuoa()` documentation)
- **force_initial_eta**: a vector of numeric values to start the estimation from (default to 0 for "L-BFGS-B")
- **quantile_bound**: a numeric value representing the quantile of the normal distribution admitted to define the bounds for L-BFGS-B (default is 0.001, i.e. 0.1%)

#### Value

a list of named arguments passed to optimizer (i.e. arg.optim)
print.mapbayests  

Print a mapbayests object

Description

Print a mapbayests object

Usage

```r
## S3 method for class 'mapbayests'
print(x, ...)
```

Arguments

- `x`: A mapbayests object.
- `...`: additional arguments

Value

print the results of the estimation to the console, and returns it invisibly.

---

use_posterior  

Use posterior estimation

Description

Use posterior estimation

Usage

```r
use_posterior(
  x,
  update_omega = FALSE,
  update_cov = TRUE,
  update_eta = TRUE,
  .zero_re = NULL,
  simplify = TRUE
)
```
use_posterior

Arguments

<table>
<thead>
<tr>
<th>Name</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>x</td>
<td>A mapbayests object.</td>
</tr>
<tr>
<td>update_omega</td>
<td>Update the OMEGA matrix with the variance-covariance matrix of estimation</td>
</tr>
<tr>
<td></td>
<td>(a logical, default is FALSE).</td>
</tr>
<tr>
<td>update_cov</td>
<td>Update the values of covariates with the individual values (a logical,</td>
</tr>
<tr>
<td></td>
<td>default is TRUE).</td>
</tr>
<tr>
<td>update_eta</td>
<td>Update the values of ETA with the final estimates (a logical, default is</td>
</tr>
<tr>
<td></td>
<td>TRUE).</td>
</tr>
<tr>
<td>.zero_re</td>
<td>Set all elements of the OMEGA or SIGMA matrix to zero.</td>
</tr>
<tr>
<td></td>
<td>Default is &quot;both&quot; if update_omega is FALSE, &quot;sigma&quot; otherwise. (possible</td>
</tr>
<tr>
<td></td>
<td>values are &quot;both&quot;, &quot;sigma&quot;, &quot;omega&quot;, &quot;none&quot;)</td>
</tr>
<tr>
<td>simplify</td>
<td>a logical. If TRUE (the default) and only one ID, one mrgmod is returned</td>
</tr>
<tr>
<td></td>
<td>instead of a list of length 1</td>
</tr>
</tbody>
</table>

Details

This function takes the results of an estimation (i.e. a mapbayests object) and return a modified mrgmod in order to perform a posteriori simulations. Modifications are:

- If update_eta is TRUE, the values of ETA are updated to the estimated values (instead of 0) in $PARAM.
- If update_cov is TRUE, the covariates values are updated to the values of the individual (instead of default model values) in $PARAM.
- If update_omega is TRUE, the values of OMEGA are updated with the variance-covariance matrix of estimation (i.e. an approximation of the a posteriori distribution) instead of the inter-individual variability (i.e. the a priori distribution). Use this command in order to derive a confidence interval of concentrations that reflects the uncertainty about parameter estimation when a large number of profiles are simulated. Note that if inter-individual variability was initially defined in multiple $OMEGA blocks in the model, they will be collapsed to a single full matrix (this is irreversible).
- Depending on the values of .zero_re, the elements of $OMEGA or $SIGMA can be set to zero, whether you want to simulate one profile, or several in order to derive confidence/prediction intervals. It does not handle time-varying covariates: only the first value will be used as the individual value.

Value

a mrgmod, or a list of mrgmod if there is more than 1 ID

Examples

```r
library(magrittr)
est <- mapbayest(exmodel())
est %>%
  use_posterior() %>%
mrgsolve::ev(amt = 50000) %>%
mrgsolve::mrgsim()
```
**vs_nonmem**  

Compare results to NONMEM .phi

**Description**

Compare results to NONMEM .phi

**Usage**

```r
read_nmphi(x)
merge_phi(mapbayr_phi, nonmem_phi)
plot_phi(merged_phi, only_eta = TRUE)
summarise_phi(
  merged_phi,
  group,
  only_eta = TRUE,
  levels = c(Excellent = 0, Acceptable = 0.001, Discordant = 0.1)
)
bar_phi(summarised_phi, xaxis = NULL, facet = NULL)
```

**Arguments**

- `x`: full path to a .phi file generated by NONMEM
- `mapbayr_phi`: results of mapbayr estimations, in the form of a tibble data.frame, typically obtained from `get_phi()`
- `nonmem_phi`: results of NONMEM estimations, in the form of a tibble data.frame, typically obtained from `read_nmphi()`
- `merged_phi`: merged results of estimations, typically obtained from `merge_phi()`
- `only_eta`: filter the data with type="ETA" (a logical, default is TRUE)
- `group`: one or multiple variables to `group_by()`
- `levels`: a named vector of length 3 in order to classify the absolute differences. Default cut-offs are 0.1% and 10% in the parameters space.
- `summarised_phi`: summarized results of estimations, typically obtained from `summarise_phi()`
- `xaxis`: optional. A character value, that correspond to a variable in data, passed to the x-axis to plot multiple bars side-by-side.
- `facet`: a formula, that will be passed to `ggplot2::facet_wrap()`
Details

These functions were made to easily compare the results of mapbayr to NONMEM. For instance, it could be useful in the case of the transposition of a pre-existing NONMEM model into mapbayr. For this, you need to code your model in both mapbayr and NONMEM, and perform the MAP-Bayesian estimation on the same dataset. Ideally, the latter contains a substantial number of patients. NONMEM returns the estimations results into a .phi file.

Use `read_nmphi()` to parse the NONMEM .phi file into a convenient tibble data.frame with the columns:

- SUBJECT_NO, ID: Subject identification.
- ETA1, ETA2, ..., ETAn: Point estimates of eta.
- ETC1_1, ETC2_1, ETC2_2, ..., ETCn_n: Variance-covariance matrix of estimation.
- OBJ: objective function value

Use `get_phi()` to access to the estimations of mapbayr with the same "phi" format.

Use `merge_phi()` to combine mapbayr and NONMEM "phi files" into a single long-form data.frame with the columns:

- SUBJECT_NO, ID: Subject identification.
- variable name and its type: ETA (point estimate), VARIANCE (on-diagonal element of the matrix), COVARIANCE (off-diagonal), and OBJ.
- mapbayr and nonmem: corresponding values
- adiff: absolute difference between mapbayr and nonmem values

Use `plot_phi()` to graphically represent adiff vs variable. Alternatively, the table returned by `merge_phi()` is easy to play with in order to derive performance statistics or the graphical plot of your choice.

Use `summarise_phi()` to classify the estimation as "Excellent", "Acceptable" or "Discordant", over the whole dataset or by group.

Use `bar_phi()` to graphically represent the proportion of the aforementioned classification as bar plot.

Value

- `read_nmphi`: a tibble data.frame with a format close to the original .phi file
- `merge_phi`: a long-form tibble data.frame with results of mapbayr and NONMEM
- `summarise_phi`: a summarized tibble data.frame classifying the performance of mapbayr and NONMEM
- `plot_phi`, `bar_phi`: a ggplot2 object

Examples

```r
library(mapbayr)
nmphi <- read_nmphi(system.file("nm001", "run001.phi", package = "mapbayr"))
mapbayrphi <- get_phi(est001)
```
merged <- merge_phi(mapbayrphi, nmphi)
plot_phi(merged)

summarised <- summarise_phi(merged)
bar_phi(summarised)

# Analyse the results of multiple runs simultaneously

# Example dataset that represents 3 runs
merge3 <- dplyr::bind_rows(merged, merged, merged, .id = "RUN")
merge3$adiff <- 10 ^ runif(nrow(merge3), -8, 0)
summarised3 <- summarise_phi(merge3, group = RUN)
bar_phi(summarised3, xaxis = "RUN")

---

### x_cmt

**Read compartment options in a model**

**Description**

Read compartment options in a model

**Usage**

adm_cmt(x)

obs_cmt(x)

**Arguments**

- **x**: model object

**Details**

In a mrgsolve model, it is possible to specify options in \$CMT. If [ADM] or [OBS] are set, mapbayr will interpret these as defaults administration and observation compartments, respectively.

**Value**

A vector of compartment identified as default "administration" or "observation" compartments.

**Examples**

# Administration: Both 1st and 0-order
model <- exmodel(6, compile = FALSE)
mrgsolve::see(model)
adm_cmt(model)
# Observation: Both parent drug and metabolite
model <- exmodel(401, compile = FALSE)
mrgsolve::see(model)
obs_cmt(model)
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