Package ‘nat’

October 13, 2022

Type Package

Title NeuroAnatomy Toolbox for Analysis of 3D Image Data

Version 1.8.19


BugReports https://github.com/natverse/nat/issues

Description NeuroAnatomy Toolbox (nat) enables analysis and visualisation of 3D biological image data, especially traced neurons. Reads and writes 3D images in NRRD and 'Amira' AmiraMesh formats and reads surfaces in 'Amira' hxsurf format. Traced neurons can be imported from and written to SWC and 'Amira' LineSet and SkeletonGraph formats. These data can then be visualised in 3D via 'rgl', manipulated including applying calculated registrations, e.g. using the 'CMTK' registration suite, and analysed. There is also a simple representation for neurons that have been subjected to 3D skeletonisation but not formally traced; this allows morphological comparison between neurons including searches and clustering (via the 'nat.nblast' extension package).

Depends R (>= 2.15.1), rgl (>= 0.98.1)

Imports nabor, igraph (>= 0.7.1), methods, filehash (>= 2.3), digest, nat.utils (>= 0.4.2), plyr, yaml

Suggests Rvcg (>= 0.17), testthat, httr, XML, knitr, rmarkdown, markdown, MASS, alphashape3d, webshot2

Additional_repositories https://dmurdoch.github.io/drat

License GPL-3

LazyData yes

R topics documented:

'-neuronlist_sets.R' 'neuronlistfh.R' 'ngraph.R' 'nrrd-io.R'
'pop3d.R' 'potential_synapses.R' 'reglist.R' 'seglist.R'
'summary.R' 'utils.R' 'vaa3draw-io.R' 'vtk-io.R' 'xform.R'
'xformimage.R' 'xformpoints.R' 'zzz.R'

RoxygenNote  7.1.1
Encoding UTF-8
VignetteBuilder knitr
NeedsCompilation no
Author Gregory Jefferis [aut, cre] (<https://orcid.org/0000-0002-0587-9355>),
James Manton [aut] (<https://orcid.org/0000-0001-9260-3156>),
Dominik Krzeminski [ctb] (<https://orcid.org/0000-0003-4568-0583>)
Maintainer Gregory Jefferis <jefferis@gmail.com>
Repository CRAN
Date/Publication 2022-04-06 11:50:02 UTC

R topics documented:

- nat-package ................................................. 5
- *.dotprops .................................................. 7
- *.neuron .................................................... 8
- *.neuronlist .............................................. 9
- affmat2cmtkparams ...................................... 10
- all.equal.dotprops ...................................... 11
- all.equal.im3d ........................................... 12
- all.equal.neuron ......................................... 13
- amiratype .................................................. 14
- as.data.frame.neuronlist ............................... 15
- as.hxsurf .................................................. 16
- as.im3d ..................................................... 17
- as.mesh3d .................................................. 18
- as.neuronlist ............................................ 20
- as.neuronlist.neuronlistfh ............................. 21
- boundingbox ............................................... 21
- c.neuronlist ............................................... 23
- Cell07PNs .................................................. 24
- clampmax ................................................... 24
- cmtk.bindir ............................................... 25
- cmtk.call .................................................. 27
- cmtk.dof2mat .............................................. 28
- cmtk.extract_affine ..................................... 29
- cmtk.mat2dof .............................................. 30
- cmtk.reformatx ........................................... 31
- cmtk.statistics ........................................... 32
- cmtk.targetvolume ....................................... 34
- cmtk.version .............................................. 35
R topics documented:

- cmtkparams2affmat .................................................. 36
- cmtkreg ................................................................. 37
- cmtkreglist ............................................................ 38
- coord2ind ............................................................... 39
- dotprops ................................................................. 40
- fileformats ............................................................. 41
- find.neuron ............................................................. 44
- find.soma ................................................................. 45
- flip .......................................................... 46
- graph.nodes ............................................................. 47
- im3d .......................................................... 47
- im3d-coords ............................................................ 49
- im3d-io ................................................................. 50
- image.im3d .............................................................. 51
- imexpand.grid .......................................................... 53
- imscalebar ............................................................... 53
- imslice .......................................................... 54
- ind2coord .............................................................. 55
- intersect .............................................................. 56
- is.amiramesh ............................................................ 56
- is.fijitraces .............................................................. 57
- is.im3d ............................................................... 58
- is.neuroml ................................................................. 58
- is.neuronlist .............................................................. 59
- is.nrrd .............................................................. 59
- is.swc .......................................................... 60
- is.vaa3draw .............................................................. 61
- kcs20 .......................................................... 61
- mask .......................................................... 62
- materials .............................................................. 63
- MBL.surf .............................................................. 64
- mirror .......................................................... 64
- ndigest ............................................................ 67
- neuron .............................................................. 68
- neuronlist .............................................................. 71
- neuronlist-dataframe-methods ........................................ 72
- neuronlistth ............................................................. 75
- ngraph .............................................................. 78
- napply .............................................................. 80
- nscan .............................................................. 83
- nopen3d .............................................................. 85
- normalise_swc .......................................................... 85
- npop3d .............................................................. 86
- nrrd.voxdims ............................................................ 87
- nvertices .............................................................. 88
- nview3d .............................................................. 88
- origin .............................................................. 89
- pan3d .............................................................. 90
R topics documented:

plot.neuron .......................................................... 91
plot.neuronlist ....................................................... 93
plot3d ................................................................. 94
plot3d.boundingbox ................................................ 95
plot3d.cmtkreg ...................................................... 96
plot3d.dotprops ..................................................... 97
plot3d.hxsurf ....................................................... 98
plot3d.neuron ....................................................... 99
plot3d.neuronlist ................................................... 101
pointsinside ......................................................... 103
potential_synapses ................................................ 105
projection ............................................................ 106
prune ................................................................. 108
prune_strahler ...................................................... 109
prune_vertices ...................................................... 110
read.amiramesh .................................................... 112
read.cmtk ............................................................ 113
read.cmtkreg ........................................................ 114
read.hxsurf ........................................................ 114
read.landmarks ..................................................... 116
read.morphml ........................................................ 118
read.neuron ........................................................ 119
read.neuron.fiji ..................................................... 120
read.neuron.neuroml .............................................. 121
read.neuron.swc ................................................... 121
read.neuronlistfh .................................................. 122
read.neurons ...................................................... 123
read.nrrd ............................................................ 125
read.vaa3draw ...................................................... 126
reglist ................................................................. 127
remotesync .......................................................... 128
resample ............................................................. 129
rootpoints ........................................................... 130
scale.neuron ........................................................ 131
seglengths ........................................................... 132
seglist ............................................................... 133
seglist2swc .......................................................... 135
segmentgraph ....................................................... 136
setdiff ............................................................... 137
simplify_reglist .................................................... 138
smooth_neuron ...................................................... 138
spine ................................................................. 139
strahler_order ...................................................... 141
sub2ind .............................................................. 142
subset ............................................................... 142
subset.dotprops .................................................... 142
subset.hxsurf ...................................................... 144
subset.neuron ...................................................... 145
nat-package

Analyse 3D biological image data especially neurons

Description

nat provides tools to read, analyse, plot, transform and convert neuroanatomical data, especially representations of neurons.

Neuron Objects

At present there are 2 main representations of neuronal data:

- **neuron** objects contain one or more connected trees that make up a neuron
- **dotprops** objects can contain one (or more) neurons represented as points and tangent vectors in which the connectivity information has been discarded

The subset function has both `subset.neuron` and `subset.dotprops` methods, which can be used to keep (or reject) specified vertices within a neuron e.g. by spatial constraints. `subset.neuron` will look after the tree structure of neurons in these circumstances.

Neuron objects containing connected trees can be converted to `ngraph` objects, a lightweight wrapper around the `igraph` library’s `graph` class that preserves 3D coordinate information. This allows neurons to be manipulated based on their graph structure, e.g. by finding all nodes upstream (closer to the root) or downstream of a given node. The `as.neuron` function can convert `ngraph` objects back to neurons or selected vertex indices can be used to subset a neuron with `subset.neuron`. 
Collections of Neurons

Neurons can be collected as neuronlist objects, which contain multiple neuron or dotprops objects along with an attached dataframe of metadata. The metadata can be accessed and manipulated using the myneuronlist[i,j] notation (see neuronlist-dataframe-methods).

Neurons can be read into a neuronlist using read.neurons or written out using write.neurons with support for many of the most common formats including swc.

Metadata can be used to colour or subset the neurons during plotting (see plot3d.neuronlist and subset.neuronlist). Interactive 3D selection of neurons in a neuronlist is also possible using find.neuron (which makes use of rgl’s select3d function).

neuronlist objects also provide additional functionality to streamline arithmetic (e.g. scaling all the points in all neurons see *.neuronlist) and transformations (see Transformations section below and xform). Arbitrary functions can be applied to each individual neuron can be applied using the nlapply function, which also provides options for progress bars and simple parallelisation.

Transformations

neuron or dotprops objects can be transformed from e.g. sample to template brain space using affine or non-rigid registrations, typically calculated with the open source CMTK package available at https://www.nitrc.org/projects/cmtk/, see ?cmtk for installation details. The function xform has methods to deal with a variety of types of interest.

3D Image Data

In addition to data types defined by unstructured collections of 3D vertices such as neuron, dotprops and hxsurf objects nat provides the im3d class to handle image/density data on a regular grid. I/O is handled by read.im3d and write.im3d, which are currently implemented for the amiramesh and nrrd file formats; there is also read only access to the vaa3d raw format.

Spatial information can be queried with voxdims, boundingbox and ijkpos, xyzpos methods. You can convert between voxel data and coordinate (vertex) -based representations using the following functions:

- as.im3d The as.im3d.matrix method converts XYZ coordinates to an im3d image volume
- ind2coord Find XYZ coordinates of specified voxels of an im3d image volume
- dotprops The dotprops.im3d method converts an im3d object to a dotprops format neuron, i.e. a cloud of unconnected segments.

Surface Data

nat can read, write, transform and subset surface (mesh) objects defined by Amira’s HxSurface class. See read.hxsurf and links therein. In addition hxsurf objects can be converted to the mesh3d format, which provides a link to the rgl package and also to packages for morphometrics and sophisticated mesh manipulation such as Morpho and Rvcg.

rgl Package

nat uses the rgl package extensively for 3D visualisation. rgl’s core function is to provide interactive visualisation (usually in an X11 window depending on OpenGL - and therefore on a graphics
card or OpenGL software emulator) but recently significant functionality for static snapshots and embedding results in reports such as web pages has been added. With this in mind, Duncan Murdoch has added the rgl.useNULL option. As of nat 1.8.0, options(rgl.useNULL=TRUE) will be set before nat is loaded in non-interactive R sessions. If you want to use nat in interactive environments where X11 is not available, you may want to set options(rgl.useNULL=TRUE) manually before loading nat.

File Formats

nat supports multiple input and output data formats for the object classes. There is a registry-based mechanism which allows support for reading or writing specific file formats (see fileformats) to be plugged in to reasonably generic functions such as read.neurons. It is perfectly possible for other R packages or end users to extend the supported list of file types by registering new read/write or identification functions.

Package Options

The following options can be set to specify default behaviour.

- nat.cmtk.bindir Location of CMTK binaries. See cmtk.bindir
- nat.default.neuronlist A character string naming a neuronlist to use with the plot3d.character method
- nat.progress The default progress reporter to use with nlapply. See create_progress_bar for possible values. When unset is equivalent to special value 'auto'. To suppress altogether, use nat.progress="none".

In addition there is one read-only option:

- nat.cmtk.version which is used to store the current cmtk version when there are repeated calls to cmtk.version.

See Also

neuron, dotprops, neuronlist, nlapply, plot3d, xform, im3d, read.hxsurf, rgl which is used for visualisation, fileformats, read.neurons, cmtk.
Usage

```r
## S3 method for class 'dotprops'
 x * y

## S3 method for class 'dotprops'
 x + y

## S3 method for class 'dotprops'
 x - y

## S3 method for class 'dotprops'
 x / y
```

Arguments

- `x`: A dotprops object
- `y`: A scalar or 3-vector that will be applied to the dotprops object

Value

A new dotprops object

Description

If `x` is a 1-vector or a 3-vector, multiply xyz only. If `x` is a 4-vector, multiply xyz and diameter by that.

TODO: Figure out how to document arithmetic functions in one go.

Usage

```r
## S3 method for class 'neuron'
 n * x

## S3 method for class 'neuron'
 n + x

## S3 method for class 'neuron'
 n - x

## S3 method for class 'neuron'
 n / x
```

Arguments

- `n`: a neuron
- `x`: (a numeric vector to multiply neuron coords in neuron)
Value
modified neuron

See Also
neuron

Examples
n1<-Cell07PNs[[1]]*2
n2<-Cell07PNs[[1]]*c(2,2,2,1)
stopifnot(all.equal(n1,n2))
n3<-Cell07PNs[[1]]*c(2,2,4)

Description
If x is one number or 3-vector, multiply coordinates by that If x is a 4-vector, multiply xyz and diameter

Usage
## S3 method for class 'neuronlist'
x * y

## S3 method for class 'neuronlist'
x + y

## S3 method for class 'neuronlist'
x - y

## S3 method for class 'neuronlist'
x / y

Arguments
x a neuronlist
y (a numeric vector to multiply coords in neuronlist members)

Value
modified neuronlist
Decompose homogeneous affine matrix to CMTK registration parameters

Usage

affmat2cmtkparams(matrix, centre = c(0, 0, 0))

Arguments

matrix 4x4 homogeneous affine matrix
centre Rotation centre

Details

The version attribute of the resultant matrix marks this as compliant with CMTK>v2.4 (~ Dec 2013) when a bug in affine matrix (de)composition was fixed.

Value

5x3 matrix of CMTK registration parameters with a version attribute

See Also

Other cmtk-geometry: cmtk.dof2mat(), cmtk.mat2dof(), cmtkparams2affmat()
Description

all.equal method tailored to dotprops objects

Usage

## S3 method for class 'dotprops'
all.equal(
  target, current,
  check.attributes = FALSE,
  absoluteVectors = TRUE,
  ...)

Arguments

target, current  
dotprops objects to compare
check.attributes  
Whether to check attributes (false by default)
absoluteVectors  
Whether to check only the absolute value of eigenvectors for equality (default TRUE, see details)
...

Details

This method is required because the direction vectors are computed using an eigenvector decomposition where the sign of the eigenvector is essentially random and subject to small numerical instabilities. Therefore it does not usually make sense to check the value of vect exactly.

Examples

# equal using default
kc1=kcs20[[1]]
kc1.recalc=dotprops(kc1)
# not equal due to differences in attributes and vectors
all.equal.default(kc1.recalc, kc1)
# still not equal because of tangent vector flipping
all.equal.default(kc1.recalc, kc1, check.attributes=FALSE)
# equal using appropriate method
stopifnot(isTRUE(all.equal(kc1.recalc, kc1)))
# NB identical when recalculated on same setup from same data
stopifnot(isTRUE(all.equal.default(kc1.recalc, dotprops(kc1))))
all.equal.im3d  
Check equality on data and key attributes of im3d objects

Description

Check equality on data and key attributes of im3d objects

Usage

```r
## S3 method for class 'im3d'
all.equal(
  target,
  current,
  tolerance = 1e-06,
  attrsToCheck = c("BoundingBox"),
  attrsToCheckIfPresent = c("dim", "names", "dimnames", "x", "y", "z"),
  CheckSharedAttrsOnly = FALSE,
  ...)
```

Arguments

- `target`  
  R object.
- `current`  
  other R object, to be compared with target.
- `tolerance`  
  numeric ≥ 0. Differences smaller than `tolerance` are not reported. The default value is close to 1.5e-8.
- `attrsToCheck`  
  Which attributes in im3d should always be checked
- `attrsToCheckIfPresent`  
  Which attributes in im3d should be checked if present
- `CheckSharedAttrsOnly`  
  Logical whether to check shared attributes only (default: FALSE)
- `...`  
  additional arguments passed to `all.equal`

See Also

`all.equal`
all.equal.neuron

Check equality on key fields of neuron object

Description

Check equality on key fields of neuron object

Usage

```r
## S3 method for class 'neuron'
all.equal(
  target,
  current,
  tolerance = 1e-06,
  check.attributes = FALSE,
  fieldsToCheck = c("NumPoints", "StartPoint", "BranchPoints", "EndPoints", "NumSegs", "SegList", "d"),
  fieldsToCheckIfPresent = c("NeuronName", "nTrees", "SubTrees"),
  fieldsToExclude = character(),
  CheckSharedFieldsOnly = FALSE,
  ...
)
```

Arguments

target
  R object.

current
  other R object, to be compared with target.

tolerance
  numeric ≥ 0. Differences smaller than tolerance are not reported. The default value is close to 1.5e-8.

check.attributes
  logical indicating if the attributes of target and current (other than the names) should be compared.

fieldsToCheck
  Which fields in the neuron are always checked. The special value of NA indicates that all fields in the neurons will be compared.

fieldsToCheckIfPresent
  These fields are only checked if they are present

fieldsToExclude
  Character vector of fields to exclude from check

CheckSharedFieldsOnly
  Logical whether to check shared fields only (default: FALSE)

...
  additional arguments passed to all.equal

See Also

all.equal
Examples

```r
x <- Cell07PNs[[1]]
y <- x
y$NeuronName <- 'rhubarb'
# NOT TRUE
all.equal(x, y)
# TRUE
all.equal(x, y, fieldsToExclude='NeuronName')
```

---

**amiratype**

*Return the type of an amiramesh file on disk or a parsed header*

**Description**

Return the type of an amiramesh file on disk or a parsed header

**Usage**

```r
amiratype(x, bytes = NULL)
```

**Arguments**

- `x`: Path to files on disk or a single pre-parsed parameter list
- `bytes`: A raw vector containing at least 11 bytes from the start of the file.

**Details**

Note that when checking a file we first test if it is an amiramesh file (fast, especially when `bytes`!=NULL) before reading the header and determining content type (slow).

**Value**

character vector (NA_character_ when file invalid)

**See Also**

Other amira: `is.amiramesh()`, `read.amiramesh()`, `read.hxsurf()`, `write.hxsurf()`
as.data.frame.neuronlist

Get or set the attached data.frame of a neuronlist

Description

For as.data.frame, when there is no attached data.frame the result will be a data.frame with 0 columns but an appropriate number of rows, named by the objects in the neuronlist.

data.frame<- methods set the data frame attached to an object. At present this is only used for neuronlist objects.

Usage

```r
## S3 method for class 'neuronlist'
as.data.frame(x, row.names = names(x), optional = FALSE, ...)
data.frame(x) <- value

## S3 replacement method for class 'neuronlist'
data.frame(x) <- value
```

Arguments

- `x` neuronlist to convert
- `row.names` row names (defaults to names of objects in neuronlist, which is nearly always what you want.)
- `optional` ignored in this method
- `...` additional arguments passed to data.frame (see examples)
- `value` The new data.frame to be attached to x

Value

for as.data.frame.neuronlist, a data.frame with length(x) rows, named according to names(x) and containing the columns from the attached data.frame, when present.

for data.frame<- .neuronlist, a neuronlist with the attached data.frame.

See Also

data.frame, neuronlist
Examples

```r
head(as.data.frame(kcs20))
# add additional variables
str(as.data.frame(kcs20, i=seq(kcs20), abc=LETTERS[seq(kcs20)]))
# stop character columns being turned into factors
newdf <- as.data.frame(kcs20, i=seq(kcs20), abc=LETTERS[seq(kcs20)],
                     stringsAsFactors=FALSE)
str(newdf)
data.frame(kcs20)=newdf
```

---

`as.hxsurf`  
Convert an object to a nat.hxsurf object

Description

Convert an object to a nat.hxsurf object

Usage

```r
as.hxsurf(x, ...)  
## S3 method for class 'mesh3d'
  as.hxsurf(x, region = "Interior", col = NULL, ...)
```

Arguments

- `x`  
  A surface object

- `...`  
  Additional arguments passed to methods

- `region`  
  The default name for the surface region

- `col`  
  The surface colour (default value of NULL implies the colour specified in mesh3d object or grey when the mesh3d object has no colour.)

Details

hxsurf objects are based on the format of Amira’s surface objects (see `read.hxsurf`). They have the ability to include multiple distinct regions. However, at the moment the only method that we provide converts mesh3d objects, which can only include one region.

Value

A new surface object of class hxsurf (see `read.hxsurf`) for details.

See Also

- `as.mesh3d`

Other hxsurf:  
`as.mesh3d()`, `materials()`, `plot3d.hxsurf()`, `read.hxsurf()`, `subset.hxsurf()`, `write.hxsurf()`
Examples

tet=tetrahedron3d(col='red')
teth=as.hxsurf(tet)

plot3d(teth)

as.im3d

Convert a suitable object to an im3d object.

Description

Convert a suitable object to an im3d object.

Usage

as.im3d(x, ...)

## S3 method for class 'im3d'
as.im3d(x, ...)

## S3 method for class 'matrix'
as.im3d(x, voxdims, origin = NULL, BoundingBox = NULL, ...)

Arguments

x Object to turn into an im3d
...
voxdims Numeric vector of length 3 or an im3d compatible object (see details) completely specifying the required space.
origin the location (or centre) of the first voxel
BoundingBox Physical extent of image. See the details section of boundingbox's help for the distinction.

Details

At present the only interesting method in nat is as.im3d.matrix which can be used to convert a matrix of 3D points into a 3D volume representation. ind2coord can be used to do the reverse: convert a set of 3D coords to an im3d volume.

Other than that, this is a largely a placeholder function with the expectation that other packages may wish to provide suitable methods.

as.im3d.matrix can accept any object that can be converted to an im3d object in the voxdims argument This will completely specify the dims, voxdims, origin etc. Any value passed to those parameters will be ignored. This can be useful for producing a new im3d to match a target image on disk or a nat.templatebrains::templatebrain object. See examples.
See Also

as.mesh3d, ind2coord
as.im3d, as.im3d

Other im3d: boundingbox(), im3d-coords, im3d-io, im3d(), imexpand.grid(), imslice(), is.im3d(), mask(), origin(), projection(), threshold(), unmask(), voxdims()

Examples

## convert a list of neurons into an image volume
im=as.im3d(xyzmatrix(kcs20), voxdims=c(1, 1, 1),
BoundingBox=c(250, 410, 0, 130, 0, 120))
## Not run:
write.im3d(im, 'kc20volume.nrrd')

## use image dimensions of an image on disk
# nb note use of ReadData = FALSE so that we just fetch the dimensions of
# the target image
diskim=read.im3d("/path/to/my/image.nrrd", ReadData = FALSE)
im=as.im3d(xyzmatrix(kcs20), diskim)

## use image dimensions of JFRC2 template brain to define the image space
library(nat.flybrains)
im=as.im3d(xyzmatrix(kcs20), JFRC2)
## End(Not run)

---

as.mesh3d  Convert an object to an rgl mesh3d

Description

as.mesh3d.ashape3d converts an alphashape3d::ashape3d object into a nat/rgl compatible mesh3d surface

Note that this provides a link to the Rvcg package

Usage

## S3 method for class 'ashape3d'
as.mesh3d(x, tri_to_keep = 2L, ...)

## S3 method for class 'hxsurf'
as.mesh3d(x, Regions = NULL, material = NULL, drop = TRUE, ...)
as.mesh3d

Arguments

- **x**: Object to convert to mesh3d
- **tri_to_keep**: Which alphashape triangles to keep (expert use only - see triang entry in Value section of ashape3d docs for details.)
- **...**: Additional arguments for methods
- **Regions**: Character vector or regions to select from hxsurf object
- **material**: rgl materials such as color
- **drop**: Whether to drop unused vertices (default TRUE)

Details

An alpha shape is a generalisation of a convex hull enclosing a set of points. Unlike a convex hull, the resultant surface can be partly concave allowing the surface to more closely follow the set of points.

In this implementation, the parameter alpha is a scale factor with units of length that defines a spatial domain. When alpha is larger the alpha shape approaches the convex hull; when alpha is smaller the alpha shape has a greater number of faces / vertices i.e. it follows the points more closely.

Value

A mesh3d object which can be plotted and manipulated using rgl and nat packages.

See Also

ashape3d, mesh3d
as.mesh3d, tmesh3d, as.hxsurf, read.hxsurf

Examples

```r
library(alphashape3d)
kcs20.a=ashape3d(xyzmatrix(kcs20), alpha = 10)
plot(kcs20.a)

# convert to mesh3d
kcs20.mesh=as.mesh3d(kcs20.a)

# check that all points are inside mesh
all(pointsinside(kcs20, kcs20.mesh))
# and show that we can also use the alphashape directly
all(pointsinside(kcs20, kcs20.a))

clear3d()
wire3d(kcs20.mesh)
plot3d(kcs20, col=type, lwd=2)
```
as.neuronlist  Make a list of neurons that can be used for coordinate plotting/analysis

Description

Make a list of neurons that can be used for coordinate plotting/analysis

Usage

as.neuronlist(l, ...)

## Default S3 method:
as.neuronlist(l, df = NULL, AddClassToNeurons = TRUE, ...)

Arguments

l     An existing list or a single neuron to start a list
...
     Additional arguments passed to methods
df    the data.frame to attach with additional metadata.
AddClassToNeurons
     Whether to ensure neurons have class neuron (see details).

Details

Note that as.neuronlist can cope with both neurons and dotprops objects but AddClassToNeurons will only apply to things that look like neurons but don’t have a class of neuron.

See neuronlist details for more information.

Value

neuronlist with attr(‘df’)

See Also

is.neuronlist, is.neuron, is.dotprops
**as.neuronlist.neuronlistfh**

convert neuronlistfh to a regular (in memory) neuronlist

**Description**

close neuronlistfh to a regular (in memory) neuronlist

**Usage**

```r
## S3 method for class 'neuronlistfh'
as.neuronlist(l, ...
```

**Arguments**

- `l` An existing list or a single neuron to start a list
- `...` Additional arguments passed to methods

---

**boundingbox**

Get the bounding box of an im3d volume or other compatible object

**Description**

boundingbox.list is designed to be used on objects that contain 3D point information and for which xyzmatrix is defined.

boundingbox.shape3d is designed to be used on objects that contain 3D point information and inherit from rgl's shape3d class and for which xyzmatrix is defined. Presently this applies to mesh3d objects.

Set the bounding box of an im3d object

**Usage**

```r
boundingbox(x, ...)
```

```r
## S3 method for class 'im3d'
boundingbox(x, dims = dim(x), ...)
```

```r
## S3 method for class 'character'
boundingbox(x, ...)
```

```r
## S3 method for class 'list'
boundingbox(x, na.rm = FALSE, ...)
```

```r
## S3 method for class 'neuron'
boundingbox(x, ...)
```
boundingbox(x, na.rm = FALSE, ...)

## S3 method for class 'shape3d'
boundingbox(x, na.rm = FALSE, ...)

## Default S3 method:
boundingbox(x, dims, input = c("boundingbox", "bounds"), ...)

boundingbox(x) <- value

Arguments

x

A vector or matrix specifying a bounding box, an im3d object, any object with base class list for which xyzmatrix can extract 3D points (e.g. neurons, surfaces etc), or, for boundingbox.character, a character vector specifying a file.

...

Additional arguments for methods
dims

The number of voxels in each dimension when x is a BoundingBox matrix.

na.rm

Whether to ignore NA points (default FALSE)

input

Whether x defines the boundingbox or bounds of the image (see details).

value

The object which will provide the new boundingbox information. This can be be either an im3d object with a boundingbox or a vector or matrix defined according to boundingbox.default.

Details

The bounding box is defined as the position of the voxels at the two opposite corners of the cuboid encompassing an image, when each voxel is assumed to have a single position (sometimes thought of as its centre) and no physical extent. When written as a vector it should look like: c(x0,x1,y0,y1,z0,z1). When written as a matrix it should look like: rbind(c(x0,y0,z0),c(x1,y1,z1)) where x0,y0,z0 is the position of the origin.

Note that there are two competing definitions for the physical extent of an image that are discussed e.g. http://teem.sourceforge.net/nrrd/format.html. The definition that makes most sense depends largely on whether you think of a pixel as a little square with some defined area (and therefore a voxel as a cube with some defined volume) or you take the view that you can only define with certainty the grid points at which image data was acquired. The first view implies a physical extent which we call the bounds=dim(x) * c(dx,dy,dz); the second is defined as BoundingBox=dim(x)-1 * c(dx,dy,dz) and assumes that the extent of the image is defined by a cuboid including the sample points at the extreme corner of the grid. Amira takes this second view and this is the one we favour given our background in microscopy. If you wish to convert a bounds type definition into an im3d BoundingBox, you should pass the argument input='bounds'.

Value

A matrix with 2 rows and 3 columns with class='boundingbox' or NULL when missing.
c.neuronlist

Combine multiple neuronlists into a single list

Description

Combine multiple neuronlists into a single list

Usage

## S3 method for class 'neuronlist'
c(..., recursive = FALSE)

Arguments

... neuronlists to combine
recursive Presently ignored

Details

Uses \texttt{rbind.fill} to join any attached dataframes, so missing values are replaced with NAs.

See Also

c

Examples

stopifnot(all.equal(kcs20[1:2],c(kcs20[1],kcs20[2])))
Cell07PNs

Cell07PNs: 40 Sample Projection Neurons from Jefferis, Potter et al 2007

Description

These R lists (which have additional class neuronlist) contain 40 traced olfactory projection neurons from Jefferis, Potter et al 2007 that have been transformed onto the IS2 template brain (Cachero, Ostrovsky et al 2010).

References


See Also

head.neuronlist, with.neuronlist

Other nat-data: MBL.surf, kcs20

Examples

head(Cell07PNs)
table(with(Cell07PNs,Glomerulus))

clampmax

Return function that finds maximum of its inputs within a clamping range

Description

Return function that finds maximum of its inputs within a clamping range

Usage

clampmax(xmin, xmax, replace.infinite = NA_real_)

Arguments

xmin, xmax  clamping range. If xmax is missing xmin should be a vector of length 2.

replace.infinite

The value with which to replace non-finite values in the input vector. When codereplace.infinite=FALSE no action is taken. The default value of NA will result in e.g. Inf being mapped to NA.
Details

Note that by default infinite values in the input vector are converted to NAs before the being compared with the clampmax range.

Value

A function with signature \( f(x, \ldots, na.rm) \)

Examples

```r
## Not run:
LHMask=read.im3d(system.file('tests/testthat/testdata/nrrd/LHMask.nrrd', package='nat'))
d=unmask(rnorm(sum(LHMask),mean=5, sd=5), LHMask)
op=par(mfrow=c(1,2))
rval=image(projection(d, projfun=max))
image(projection(d, projfun=clampmax(0,10)), zlim=rval$zlim)
par(op)

## End(Not run)
```

---

cmtk.bindir  

Return path to directory containing CMTK binaries

Description

The Computational Morphometry Toolkit (CMTK) is the default image registration toolkit supported by nat. An external CMTK installation is required in order to apply CMTK registrations. This function attempts to locate the full path to the CMTK executable files and can query and set an option.

Usage

```r
cmtk.bindir(
  firstdir = getOption("nat.cmtk.bindir"),
  extradirs = c("/bin", "/usr/local/lib/cmtk/bin", "/usr/local/bin", "/opt/local/bin",
               "/opt/local/lib/cmtk/bin", "/Applications/IGSRegistrationTools/bin",
               "C:\cygwin64\usr\local\lib\cmtk\bin",
               "C:\Program Files\CMTK-3.3\CMTK\lib\cmtk\bin"),
  set = FALSE,
  check = FALSE,
  cmtktool = "gregxform"
)
```
Arguments

**firstdir**  
Character vector specifying path containing CMTK binaries or NA (see details). This defaults to `options('nat.cmtk.bindir')`.

**extradirs**  
Where to look if CMTK is not in `firstdir` or the PATH

**set**  
Whether to set `options('nat.cmtk.bindir')` with the found directory. Also check/sets cygwin path on Windows (see Installation section).

**check**  
Whether to (re)check that a path that has been set appropriately in `options(nat.cmtk.bindir='/some/path')` or now found in the PATH or alternative directories. Will throw an error on failure.

**cmtktool**  
Name of a specific cmtk tool which will be used to identify the location of all cmtk binaries.

Details

Queries `options('nat.cmtk.bindir')` if `firstdir` is not specified. If that does not contain the appropriate binaries, it will look in the system PATH for the cmtk wrapper script installed by most recent cmtk installations.

Failing that, it will look for the cmtk tool specified by `cmtktool`, first in the path and then a succession of plausible places until it finds something. Setting `options(nat.cmtk.bindir=NA)` or passing `firstdir=NA` will stop the function from trying to locate CMTK, always returning NULL unless `check=TRUE`, in which case it will error out.

Value

Character vector giving path to CMTK binary directory or NULL when this cannot be found.

Installation

It is recommended to install released CMTK versions available from the NITRC website. A bug in composition of affine transformations from CMTK parameters in the CMTK versions <2.4 series means that CMTK>=3.0 is strongly recommended. CMTK v3 registrations are not backwards compatible with CMTK v2, but CMTKv3 can correctly interpret and convert registrations from earlier versions.

On Windows, when `set=TRUE`, cmtk.bindir will also check that the cygwin bin directory is in the PATH. If it is not, then it is added for the current R session. This should solve issues with missing cygwin dlls.

See Also

`options`

Examples

```r
message(ifelse(is.null(d<-cmtk.bindir()), "CMTK not found!",
          paste("CMTK is at:".,d)))
## Not run:
# set options('nat.cmtk.bindir') according to where cmtk was found
op=options(nat.cmtk.bindir=NULL)
```
cmtk.call

Utility function to create and run calls to CMTK commandline tools

Description

cmtk.call processes arguments into a form compatible with CMTK command line tools.
cmtk.system2 actually calls a cmtk tool using a call list produced by cmtk.call

Usage

cmtk.call(
  tool,
  PROCESSED.ARGS = NULL,
  ..., 
  FINAL.ARGS = NULL,
  RETURN.TYPE = c("string", "list")
)

cmtk.system2(cmtkcall, moreargs = NULL, ...)

Arguments

tool: Name of the CMTK tool
PROCESSED.ARGS: Character vector of arguments that have already been processed by the callee.
  Placed immediately after cmtk tool.
...: Additional named arguments to be processed by (cmtk.call, see details) or passed to system2 (cmtk.system2).
FINAL.ARGS: Character vector of arguments that have already been processed by the callee.
  Placed at the end of the call after optional arguments.
RETURN.TYPE: Sets return type to a character string or list (the latter is suitable for use with system2)
cmtkcall: A list containing processed arguments prepared by cmtk.call(RETURN.TYPE="list")
moreargs: Additional arguments to add to the processed call

Details

cmtk.call processes arguments in ... as follows:

• argument names will be converted from arg.name to --arg-name
• logical vectors (which must be of length 1) will be passed on as --arg-name
• character vectors (which must be of length 1) will be passed on as --arg-name arg i.e. quoting is left up to callee.
• numeric vectors will be collapsed with commas if of length greater than 1 and then passed on unquoted e.g. target.offset=c(1,2,3) will result in --target-offset 1,2,3
cmtk.dof2mat

Convert CMTK registration to homogeneous affine matrix with dof2mat

Description
Convert CMTK registration to homogeneous affine matrix with dof2mat

Usage

cmtk.dof2mat(reg, Transpose = TRUE, version = FALSE)

Arguments

reg         Path to input registration file or 5x3 matrix of CMTK parameters.
Transpose   ouput matrix so that form on disk matches R’s convention.
version     Whether to return CMTK version string
Details

Transpose is true by default since this results in the orientation of cmtk output files matching the orientation in R. Do not change this unless you’re sure you know what you’re doing!

Value

4x4 transformation matrix

See Also

Other cmtk-commandline: cmtk.mat2dof()
Other cmtk-geometry: affmat2cmtkparams(), cmtk.mat2dof(), cmtkparams2affmat()
Use CMTK mat2dof to convert homogeneous affine matrix into CMTK registration

Description
Use CMTK mat2dof to convert homogeneous affine matrix into CMTK registration

Usage
cmtk.mat2dof(m, f = NULL, centre = NULL, Transpose = TRUE, version = FALSE)

Arguments
m Homogenous affine matrix (4x4) last row 0 0 0 1 etc
f Output file (optional)
centre Centre for rotation (optional 3-vector)
Transpose the input matrix so that it is read in as it appears on disk
version When TRUE, function returns CMTK version number of mat2dof tool

Details
If no output file is supplied, 5x3 params matrix will be returned directly. Otherwise a logical will be returned indicating success or failure at writing to disk.

Transpose is true by default since this results in an R matrix with the transpose in the fourth column being correctly interpreted by cmtk.

Value
5x3 matrix of CMTK registration parameters or logical

See Also
Other cmtk-commandline: cmtk.dof2mat()
Other cmtk-geometry: affmat2cmtparms(), cmtk.dof2mat(), cmtkparams2affmat()
Reformat an image with a CMTK registration using the reformatx tool

Description
Reformat an image with a CMTK registration using the reformatx tool

Usage
```r
cmtk.reformatx(  
  floating,  
  registrations,  
  output,  
  target,  
  mask = FALSE,  
  direction = NULL,  
  interpolation = c("linear", "nn", "cubic", "pv", "sinc-cosine", "sinc-hamming"),  
  dryrun = FALSE,  
  Verbose = TRUE,  
  MakeLock = TRUE,  
  OverWrite = c("no", "update", "yes"),  
  filesToIgnoreModTimes = NULL,  
  ...  
)
```

Arguments
- **floating**: The floating image to be reformatted
- **registrations**: One or more CMTK format registrations on disk
- **output**: The path to the output image (defaults to "<targetstem>_<floatingstem>.nrrd")
- **target**: A character vector specifying an image file on disk, an im3d object (or an object that can be coerced to im3d) or a 6- or 9-vector defining a grid in the form Nx,Ny,Nz,dX,dY,dZ,[Ox,Oy,Oz].
- **mask**: Whether to treat target as a binary mask (only reformating positive voxels)
- **direction**: Whether to transform image from sample space to reference space (called forward by CMTK) or from reference to sample space (called inverse by CMTK). Default (when NULL is forward).
- **interpolation**: What interpolation scheme to use for output image (defaults to linear - see details)
- **dryrun**: Just print command
- **Verbose**: Whether to show cmtk status messages and be verbose about file update checks. Sets command line --verbose option.
- **MakeLock**: Whether to use a lock file to allow simple parallelisation (see makelock)
**cmtk.statistics**

Calculate image statistics for a nrrd or other CMTK compatible file

---

**Description**

Calculate image statistics for a nrrd or other CMTK compatible file

---

**Details**

Note that if you are reformatting a mask then you will need to change the interpolation to "nn", since interpolating between e.g. mask levels 72 and 74 with 73 may have unintended consequences. Presently we have no way of knowing whether an image should be treated as a mask, so the interpolation must be handled manually.

**Value**

the path to the output image (whether or not it was re-created afresh) or NA_character_ if no output was possible.

**See Also**

cmtk.bindir, cmtk.call, makelock, RunCmdForNewerInput

**Examples**

```r
## Not run:
cmtk.reformatx('myimage.nrrd', target='template.nrrd',
registrations='template_myimage.list')

# get full listing of command line options
system(cmtk.call('reformatx', help=TRUE))
```

```
## End(Not run)
```
Usage

cmtk.statistics(
  f,
  mask,
  imagetype = c("greyscale", "label"),
  masktype = c("label", "binary"),
  ...
  Verbose = FALSE
)

Arguments

f Path to image file (any CMTK compatible format)
mask Optional path to a mask file
imagetype Whether image should be treated as greyscale (default) or label field.
masktype Whether mask should be treated as label field or binary mask (default label)
... Additional arguments for ctk’s statistics tool processed by cmtk.call.
Verbose Whether to show cmtk status messages and be verbose about file update checks.
  Sets command line --verbose option.

Details

When given a label mask, returns a dataframe with a row for each level of the label field.
Note that the Entropy column (sometimes H, sometimes Entropy) will always be named Entropy in the returned dataframe.

Value

data.frame describing results with the following columns when image f is of imagetype='greyscale'
(Optionally with a mask):

- MaskLevel (only present when using a mask) the integer value of the label field for this region
- min The minimum voxel value within the current region
- max The maximum voxel value within the current region
- mean The mean voxel value within the current region
- sdev The standard deviation of voxel values within the current region
- n The count of all voxel within the region (irrespective of their value)
- Entropy Information theoretic entropy of voxel value distribution within region
- sum Sum of voxel values within the region

When image f is of imagetype='label', the following results are returned:

- level The integer value of the label field for this region
- count The number of voxels in this region
- surface The surface area of this region
- volume The volume of this region
- X,Y,Z 3D coordinates of the centroid of this region
Examples

```r
## Not run:
cmtk.statistics('someneuron.nrrd', mask='neuropilregionmask.nrrd')
cmtk.statistics('somelabelfield.nrrd', imagetype='label')

## End(Not run)
```

---

cmtk.targetvolume  *Defines a target volume for a CMTK reformatx operation*

Description

cmtk.targetvolume.list is designed to cope with any user-defined class for which an as.im3d
method exists. Presently the only example in the nat.* ecosystem is nat.templatebrains::as.im3d.templatebrain.

Usage

```r
cmtk.targetvolume(target, ...)
```

## S3 method for class 'im3d'
```r
cmtk.targetvolume(target, ...)
```

## S3 method for class 'list'
```r
cmtk.targetvolume(target, ...)
```

## Default S3 method:
```r
cmtk.targetvolume(target, ...)
```

Arguments

target  A character vector specifying an image file on disk, an im3d object (or an ob-
ject that can be coerced to im3d) or a 6-or 9-vector defining a grid in the form
Nx,Ny,Nz,dX,dY,dZ,[Ox,Oy,Oz].

...  additional arguments passed to methods

Details

if the character vector specifies an amiramesh file, it will be converted to a bare im3d object and
then to an appropriate `--target-grid` specification.

Value

a character vector specifying the full cmtk reformatx `--target` or `--target-grid` argument
**cmtk.version**

### Examples

```r
## Not run:
# see https://github.com/jefferislab/nat.flybrains
library(nat.flybrains)
cmtk.targetvolume(FCWB)

## End(Not run)
```

<table>
<thead>
<tr>
<th>cmtk.version</th>
<th>Return cmtk version or test for presence of at least a specific version</th>
</tr>
</thead>
</table>

### Description

Return cmtk version or test for presence of at least a specific version

### Usage

```r
cmtk.version(minimum = NULL)
```

### Arguments

- **minimum**: If specified checks that the cmtk version

### Details

NB this function has the side effect of setting an option nat.cmtk.version the first time that it is run in the current R session.

### Value

returns numeric_version representation of CMTK version or if minimum is not NULL, returns a logical indicating whether the installed version exceeds the current version. If CMTK is not installed returns NA.

### See Also

- `cmtk.bindir`, `cmtk.dof2mat`

### Examples

```r
## Not run:
cmtk.version()
cmtk.version('3.2.2')

## End(Not run)
```
cmtkparams2affmat

Compose homogeneous affine matrix from CMTK registration parameters

Description

Compose homogeneous affine matrix from CMTK registration parameters

Usage

```r
cmtkparams2affmat(
  params = NULL,
  tx = 0,
  ty = 0,
  tz = 0,
  rx = 0,
  ry = 0,
  rz = 0,
  sx = 1,
  sy = 1,
  sz = 1,
  shx = 0,
  shy = 0,
  shz = 0,
  cx = 0,
  cy = 0,
  cz = 0,
  legacy = NA
)
```

Arguments

- `params` 5x3 matrix of CMTK registration parameters or list of length 5.
- `tx`, `ty`, `tz` Translation along x, y and z axes (default 0)
- `rx`, `ry`, `rz` Rotation about x, y and z axes (in degrees, default 0)
- `sx`, `sy`, `sz` Scale for x, y and z axes (default 1)
- `shx`, `shy`, `shz` Shear for x,y,z axes (default 0)
- `cx`, `cy`, `cz` Centre for rotation
- `legacy` Whether to assume that parameters are in the format used by CMTK <=2.4.0 (default value NA implies FALSE, see details).

Details

If the `legacy` parameter is not set explicitly, then it will be set to TRUE if params has a version attribute <2.4 or FALSE otherwise.

Translation and centre components are assumed to be in physical coordinates.
**cmtkreg**

**Value**

4x4 homogeneous affine transformation matrix

**See Also**

Other cmtk-geometry: `affmat2cmtkparams()`, `cmtk.dof2mat()`, `cmtk.mat2dof()`

---

**Description**

cmtkreg creates an object of class cmtkreg that describes one (or more) CMTK registrations. This is simply a character vector that also has class cmtkreg. as.cmtkreg converts objects to class cmtkreg, minimally just by adding an appropriate class attribute. is.cmtkreg checks if an object is a cmtk registration either by checking class (default), or inspecting file.

**Usage**

cmtkreg(x, returnDir = TRUE)

as.cmtkreg(x, ...)

## S3 method for class 'matrix'

as.cmtkreg(x, ...)

## S3 method for class 'reglist'

as.cmtkreg(x, ...)

## Default S3 method:

as.cmtkreg(x, ...)

is.cmtkreg(x, filecheck = c("none", "exists", "magic"))

**Arguments**

- **x**
  
  Path to a cmtk registration (either plain character vector or cmtkreg object)

- **returnDir**
  
  Whether to return the registration directory (default) or the actual file containing the registration

- **...**
  
  Additional arguments passed to methods. Currently ignored.

- **filecheck**
  
  Whether to check object class only (default: 'none') or find and check if registration file **exists** or check **magic** value in first line of file.
cmtkreglist

Make in-memory CMTK registration list from affine matrix or CMTK parameters

Description

Make in-memory CMTK registration list from affine matrix or CMTK parameters

Usage

cmtkreglist(x, centre = c(0, 0, 0), reference = "dummy", floating = "dummy")

Arguments

x 5x3 matrix of CMTK registration parameters OR 4x4 homogeneous affine matrix
centre Optional centre of rotation passed to affmat2cmtkparams when decomposing 4x4 affine matrix
reference, floating Path to reference and floating images.

Details

Note that this uses the modern CMTK notation of floating_study rather than model_study as used by IGSParamsToIGSRegistration (which results in an implicit inversion by CMTK tools).

Note that the reference and floating fields have no impact on the transformation encoded in the resultant .list folder and can be overridden on the command line of CMTK tools.

Value

list of class cmtkreg containing registration parameters suitable for write.cmtkreg

See Also

write.cmtkreg, affmat2cmtkparams, cmtkreg
coord2ind

**coord2ind**

*Find 1D indices into a 3D image given spatial coordinates*

---

### Description

Find 1D indices into a 3D image given spatial coordinates

### Usage

```
coord2ind(coords, ...)  # Default S3 method:
coord2ind(
    coords,
    imdims,
    voxdims = NULL,
    origin = NULL,
    aperm,
    Clamp = FALSE,
    CheckRanges = !Clamp,
    ...
)
```

### Arguments

- `coords` : spatial coordinates of image voxels.
- `...` : extra arguments passed to methods.
- `imdims` : array dimensions of 3D image OR an object for which a `as.im3d` object has been defined (see Details).
- `voxdims` : vector of 3 voxels dimensions (width, height, depth).
- `origin` : the origin of the 3D image.
- `aperm` : permutation order for axes.
- `Clamp` : ???
- `CheckRanges` : whether to check if coordinates are out of range.

### Details

`coord2ind` is designed to cope with any user-defined class for which an `as.im3d` method exists. Presently the only example in the nat.* ecosystem is `nat.templatebrains::as.im3d.templatebrain`. The existence of an `as.im3d` method implies that `voxdims,origin, and dim` functions can be called. This is the necessary information required to convert i,j,k logical indices into x,y,z spatial indices.

### See Also

`ind2coord, sub2ind, ijkpos`
Examples

coord2ind(cbind(1,2,3), imdims = c(1024,512,218),
  voxdims = c(0.622088, 0.622088, 0.622088), origin = c(0,0,0))
## Not run:
## repeat but using a templatebrain object to specify the coordinate system
library(nat.flybrains)
coord2ind(cbind(1,2,3), JFRC2)
## End(Not run)

dotprops

dotprops: Neurons as point clouds with tangent vectors (but no connectivity)

Description

dotprops makes dotprops representation from raw 3D points (extracting vertices from S3 objects
that have them)
dotprops.dotprops will default to the original value of k and copy over all attributes that are not
set by dotprops.default.
dotprops.neuronlist will run for every object in the neuronlist using napply. ... arguments
will be passed to napply in addition to the named argument OmitFailures.

Usage

is.dotprops(x)

as.dotprops(x, ...)
dotprops(x, ...)

## S3 method for class 'character'
dotprops(x, ...)

## S3 method for class 'dotprops'
dotprops(x, k = attr(x, "k"), ...)

## S3 method for class 'im3d'
dotprops(x, ...)

## S3 method for class 'neuronlist'
dotprops(x, ..., OmitFailures = NA)

## S3 method for class 'neuron'
dotprops(x, Labels = NULL, resample = NA, ...)

## Default S3 method:
dotprops(x, k = NULL, Labels = NULL, na.rm = FALSE, ...)
Arguments

- **x**: Object to be tested/converted
- **...**: Additional arguments passed to methods
- **k**: Number of nearest neighbours to use for tangent vector calculation (set to k=20 when passed NULL)
- **OmitFailures**: Whether to omit neurons for which FUN gives an error. The default value (NA) will result in nlapply stopping with an error message the moment there is an error. For other values, see details.
- **Labels**: Vector of labels for each point e.g. identifying axon vs dendrite. The default value NULL will produce class-specific default behaviour for different classes of input object, TRUE always uses labels when an incoming object has them and FALSE never uses labels.
- **resample**: When finite, a new length to which all segmented edges will be resampled. See resample.neuron.
- **na.rm**: Whether to remove NA points (default FALSE)

Details

- **k** will default to 20 nearest neighbours when unset (i.e. when it has default value of NA) unless x is a dotprops object (when the original value of k is reused).

References

The dotprops format is essentially identical to that developed in:


See Also

- nlapply

Description

fileformats returns format names, a format definition list or a table of information about the formats that match the given filter conditions.

registerformat registers a format in the io registry

getformatreader gets the function to read a file

getformatwriter gets the function to write a file
fileformats

Usage

```r
fileformats(
  format = NULL,
  ext = NULL,
  read = NULL,
  write = NULL,
  class = NULL,
  rval = c("names", "info", "all")
)
```

```r
registerformat(
  format = NULL,
  ext = format,
  read = NULL,
  write = NULL,
  magic = NULL,
  magiclen = NA_integer_,
  class = NULL
)
```

```r
getformatreader(file, class = NULL)
```

```r
getformatwriter(format = NULL, file = NULL, ext = NULL, class = NULL)
```

Arguments

- **format**: Character vector naming the format
- **ext**: Character vector of file extensions (including periods)
- **read, write**: Functions to read and write this format
- **class**: The S3 class for the format (character vector e.g. 'neuron')
- **rval**: Character vector choosing what kind of return value `fileformats` will give.
- **magic**: Function to test whether a file is of this format
- **magiclen**: Optional integer specifying maximum number of bytes required from file header to determine file’s type.
- **file**: Path to a file

Details

if a format argument is passed to `fileformats` it will be matched with partial string matching and if a unique match exists that will be returned.

getformatreader starts by reading a set number of bytes from the start off the current file and then checks using file extension and magic functions to see if it can identify the file. Presently formats are in a queue in alphabetical order, dispatching on the first match.
Value

- fileformats returns a character vector, matrix or list according to the value of rval.
- getformatreader returns a list. The reader can be accessed with $read and the format can be accessed by $format.
- getformatwriter returns a list. The writer can be accessed with $write.

getformatwriter output file

If getformatwriter is passed a file argument, it will be processed based on the registered fileformats information and the ext argument to give a final output path in the $file element of the returned list.

If ext='someext' getformatwriter will use the specified extension to overwrite the default value returned by fileformats.

If ext=NULL, the default, and file='somefilename.someext' then file will be untouched and ext will be set to 'someext' (overriding the value returned by fileformats).

If file='somefile_without_extension' then the supplied or calculated extension will be appended to file.

If ext=NA then the input file name will not be touched (even if it has no extension at all).

Note that if ext=NULL or ext=NA, then only the specified format or, failing that, the file extension will be used to query the fileformats database for a match.

See write.neuron for code to make this discussion more concrete.

See Also

write.neuron

Examples

# information about the currently registered file formats
fileformats(rval='info')
## Not run:
registerformat("swc",read=read.swc,write=read.swc,magic=is.swc,magiclen=10,
 class='neuron')

## End(Not run)
swc=tempfile(fileext = '.swc')
write.neuron(Cell07PNs[[1]], swc)
stopifnot(isTRUE(getformatreader(swc)$format=='swc'))
unlink(swc)
find.neuron  

Find neurons within a 3D selection box (usually drawn in rgl window)

Description

Find neurons within a 3D selection box (usually drawn in rgl window)

Usage

```r
find.neuron(
  sel3dfun = select3d(),
  indices = names(db),
  db = getOption("nat.default.neuronlist"),
  threshold = 0,
  invert = FALSE,
  rval = c("names", "data.frame", "neuronlist")
)
```

Arguments

- `sel3dfun`: A `select3d` style function to indicate if points are within region
- `indices`: Names of neurons to search (defaults to all neurons in list)
- `db`: neuronlist to search. Can also be a character vector naming the neuronlist. Defaults to `options('nat.default.neuronlist')`.
- `threshold`: More than this many points must be present in region
- `invert`: Whether to return neurons outside the selection box (default `FALSE`)
- `rval`: What to return (character vector, default='names')

Details

Uses `subset.neuronlist`, so can work on dotprops or neuron lists.

Value

Character vector of names of selected neurons, neuronlist, or `data.frame` of attached metadata according to the value of `rval`.

See Also

`select3d, find.soma, subset.neuronlist`
Examples

## Not run:
plot3d(kcs20)
# draw a 3D selection e.g. around tip of vertical lobe when ready
find.neuron(db=kcs20)
# would return 9 neurons
# make a standalone selection function
vertical_lobe=select3d()
find.neuron(vertical_lobe, db=kcs20)
# use base::Negate function to invert the selection function
# i.e. choose neurons that do not overlap the selection region
find.neuron(Negate(vertical_lobe), db=kcs20)

## End(Not run)

find.soma Find neurons with soma inside 3D selection box (usually drawn in rgl window)

Description

Find neurons with soma inside 3D selection box (usually drawn in rgl window)

Usage

find.soma(
  sel3dfun = select3d(),
  indices = names(db),
  db = getOption("nat.default.neuronlist"),
  invert = FALSE,
  rval = c("names", "neuronlist", "data.frame")
)

Arguments

- **sel3dfun**: A `select3d` style function to indicate if points are within region
- **indices**: Names of neurons to search (defaults to all neurons in list)
- **db**: neuronlist to search. Can also be a character vector naming the neuronlist. Defaults to options('nat.default.neuronlist').
- **invert**: Whether to return neurons outside the selection box (default FALSE)
- **rval**: What to return (character vector, default='names')

Details

Can work on neuronlists containing neuron objects or neuronlists whose attached data.frame contains soma positions specified in columns called X,Y,Z.
Value

Character vector of names of selected neurons

See Also

select3d, subset.neuronlist, find.neuron

---

flip

Flip an array, matrix or vector about an axis

Description

Flip an array, matrix or vector about an axis

Usage

flip(x, ...)

## S3 method for class 'array'
flip(x, flipdim = "X", ...)

Arguments

x Object to flip

... Additional arguments for methods

flipdim Character vector or 1-indexed integer indicating array dimension along which flip will occur. Characters X, Y, Z map onto dimensions 1, 2, 3.

Details

Note that dimensions 1 and 2 for R matrices will be rows and columns, respectively, which does not map easily onto the intuition of a 2D image matrix where the X axis would typically be thought of as running from left to right on the page and the Y axis would run from top to bottom.
**graph.nodes**

Return root, end, or branchpoints of an igraph object

**Description**

Return root, end, or branchpoints of an igraph object

**Usage**

```r
graph.nodes(
  x,
  type = c("root", "end", "branch"),
  original.ids = "label",
  exclude.isolated = TRUE
)
```

**Arguments**

- **x**
  An igraph object
- **type**
  one of root, end (which includes root) or branch
- **original.ids**
  Use named attribute to return original vertex ids (when available). Set to FALSE when this is not desired.
- **exclude.isolated**
  Do not count isolated vertices as root points (default)

**Details**

Note that the graph must be directed in order to return a root point

**im3d**

Construct an im3d object representing 3D image data, densities etc

**Description**

im3d objects consist of a data array with attributes defining the spatial positions at which the voxels are located. There should always be a `BoundingBox` attribute which defines the physical extent of the volume in the same manner as the Amira 3D visualisation and analysis software. This corresponds to the `node` centers option in the NRRD format.
Usage

```r
im3d(
  x = numeric(0),
  dims = NULL,
  voxdims = NULL,
  origin = NULL,
  BoundingBox = NULL,
  bounds = NULL,
  ...
)
```

Arguments

- **x**: The object to turn into an im3d
- **dims**: The dimensions of the image array either as an integer vector or as an im3d object, whose attributes will provide defaults for `dims`, `origin`, `BoundingBox`, `bounds` arguments. The default (`dims=NULL`) will result in `dims` being set to `x` if `x` is an im3d object or `dim(x)` otherwise.
- **voxdims**: The voxel dimensions
- **origin**: the location (or centre) of the first voxel
- **BoundingBox, bounds**: Physical extent of image. See the details section of `boundingbox`'s help for the distinction.
- **...**: Additional attributes such as units or materials

Details

We follow Amira’s convention of setting the bounding box equal to voxel dimension (rather than 0) for any dimension with only 1 voxel.

Value

An array with additional class im3d

See Also

Other im3d: `as.im3d()`, `boundingbox()`, `im3d-coords`, `im3d-io`, `imexpand.grid()`, `imslice()`, `is.im3d()`, `mask()`, `origin()`, `projection()`, `threshold()`, `unmask()`, `voxdims()`
**im3d-coords**

Interconvert pixel and physical coordinates

**Description**

`xyzpos` converts pixel coordinates to physical coordinates

`ijkpos` converts physical coordinates to pixel coordinates

**Usage**

```r
xyzpos(d, ijk)

ijkpos(d, xyz, roundToNearestPixel = TRUE)
```

**Arguments**

- `d` An `im3d` object defining a physical space
- `ijk` an Nx3 matrix of pixel coordinates (1-indexed)
- `xyz` Nx3 matrix of physical coordinates
- `roundToNearestPixel` Whether to round calculated pixel coordinates to nearest integer value (i.e. nearest pixel). default: TRUE

**Value**

Nx3 matrix of physical or pixel coordinates

**See Also**

- `ind2coord`
- Other `im3d`: `as.im3d()`, `boundingbox()`, `im3d-io`, `im3d()`, `imexpand.grid()`, `imslice()`, `is.im3d()`, `mask()`, `origin()`, `projection()`, `threshold()`, `unmask()`, `voxdims()`

**Examples**

```r
# make an empty im3d
d = im3d(dim = c(20, 30, 40), origin = c(10, 20, 30), voxdims = c(1, 2, 3))
# check round trip for origin
stopifnot(all.equal(ijkpos(d, xyzpos(d, c(1, 1, 1))), c(1, 1, 1)))
```
Description

Read/Write calibrated 3D blocks of image data

Usage

\[
\text{read.im3d(}
\text{  file,}
\text{  ReadData = TRUE,}
\text{  SimplifyAttributes = FALSE,}
\text{  ReadByteAsRaw = FALSE,}
\text{  ...}
\text{)}
\]

\[
\text{write.im3d(x, file, format = NULL, ...)}
\]

Arguments

- **file**: Character vector describing a single file
- **ReadData**: Whether to read the data itself or return metadata only. Default: TRUE
- **SimplifyAttributes**: Whether to read byte values as R raw arrays. These occupy 1/4 memory but arithmetic is less convenient. (default: FALSE)
- **x**: The image data to write (an im3d, or capable of being interpreted as such)
- **format**: Character vector specifying an image format (e.g. "nrrd", "amira\_mesh"). Optional, since the format will normally be inferred from the file extension. See \text{getformatwriter} for details.

Details

Currently only nrrd and amira formats are implemented. Furthermore implementing a registry to allow extension to arbitrary formats remains a TODO item.

The core attributes of an im3d object are BoundingBox, origin, x, y, z where x, y, z are the locations of samples in the x, y and z image axes (which are assumed to be orthogonsl).

Value

For read.im3d an objecting inheriting from base array and im3d classes.
See Also

read.nrrd, read.amrimesh
write.nrrd, getformatwriter

Other im3d: as.im3d(), boundingbox(), im3d-coords.im3d(), imexpand.grid(), imslice(),
is.im3d(), mask(), origin(), projection(), threshold(), unmask(), voxdims()

Examples

## Not run:
# read attributes of vaa3d raw file
read.im3d("L1DS1_crop_straight.raw", ReadData = F, chan=2)

## End(Not run)
Arguments

x  The im3d object containing the data to be plotted (NAs are allowed).
xlim, ylim  ranges for the plotted x and y values, defaulting to the BoundingBox of x.
zlim  the minimum and maximum z values for which colors should be plotted, defaulting to the range of the finite values of z. Each of the given colors will be used to color an equispaced interval of this range. The midpoints of the intervals cover the range, so that values just outside the range will be plotted.
plotdims  Which dimensions of 3D im3d object to plot (character vector). Defaults to c('x','y')
flipdims  Which dimensions to flip (character vector). Defaults to flipping y.
filled.contour  Whether to use a filled.contour plot instead of a regular image plot.
asp  Whether to have a a square aspect ratio (logical, default: FALSE)
axes  Whether to plot axes (default: FALSE)
xlab, ylab  each a character string giving the labels for the x and y axis. Default to the 'call names' of x or y, or to "" if these were unspecified.
nlevels  The number of colour levels in z
levels  The levels at which to break z values
color.palette  The colour palette from which col will be selected.
col  a list of colors such as that generated by rainbow, heat.colors, topo.colors, terrain.colors or similar functions.
useRaster  Whether to use rasterImage to plot images as a bitmap (much faster for large images). default useRaster=FALSE checks dev.capabilities to see if raster images are supported.
...  graphical parameters for plot or image may also be passed as arguments to this function.

Value

A list with elements:

• zlim  The z (intensity limits)
• nlevels.actual  The actual number of plotted levels
• nlevels.orig  The requested number of plotted levels
• levels  The chosen levels
• colors  A character vector of colours

Examples

## Not run:
LHMask=read.im3d(system.file('tests/testthat/testdata/nrrd/LHMask.nrrd',package='nat'))
image(imslice(LHMask,10), asp=TRUE)
# useRaster is appreciably quicker in most cases
image(imslice(LHMask,10), asp=TRUE, useRaster=TRUE)

## End(Not run)
imexpand.grid

Convert locations of im3d voxel grid into XYZ coordinates

Description

Convert locations of im3d voxel grid into XYZ coordinates

Usage

imexpand.grid(d)

Arguments

d An im3d object

Value

Nx3 matrix of image coordinates

See Also

expand.grid

Other im3d: as.im3d(), boundingbox(), im3d-coords.im3d-io, im3d(), imslice(), is.im3d(), mask(), origin(), projection(), threshold(), unmask(), voxdims()

Examples

d=im3d(dim=c(2,3,2),origin=c(10,20,30),voxdims=c(1,2,3))
imexpand.grid(d)

imscalebar

Make a scalebar to accompany an image.im3d plot

Description

Make a scalebar to accompany an image.im3d plot

Usage

imscalebar(
levels, 
col, 
nlevels = NULL, 
zlim = NULL, 
horizontal = TRUE, 
lab = "Density")
mar = c(4, 2, 2, 2) + 0.1,
border = NULL,
...
)

Arguments

levels The levels at which z values were cut or a list returned by \texttt{image.im3d}
col The plotted colours for each level
nlevels The number of colour levels (inferred from levels when NULL)
zlim The limits of the plotted z (intensity) values of the image
horizontal Whether to make a horizontal or vertical scalebar (default: TRUE)
lab The (single) axis label for the scale bar (default: Density)
mar The margins for this plot
border Color for rectangle border (see \texttt{rect}'s border argument for details).
... Additional arguments for \texttt{plot}

Examples

```r
## Not run:
LHMask = read.im3d(system.file("tests/testthat/testdata/nrrd/LHMask.nrrd", package = "nat"))
op = par(no.readonly = TRUE)
layout(matrix(c(1, 2), ncol = 2L), widths = c(1, 0.2))
rval = image(imslice(LHMask, 10), asp = TRUE)
imscalebar(rval)
par(op)
## End(Not run)
```

\textbf{imslice} \hspace{1cm} Slice out a 3D subarray (or 2d matrix) from a 3D image array

**Description**

Slice out a 3D subarray (or 2d matrix) from a 3D image array

**Usage**

\texttt{imslice(x, slice, slicedim = "z", drop = TRUE)}

**Arguments**

\begin{description}
\item \texttt{x} An im3d objet
\item \texttt{slice} Indices defining the slices to keep
\item \texttt{slicedim} Character vector or integer defining axis from which slices will be removed.
\item \texttt{drop} Whether singleton dimensions will be dropped (default: TRUE) converting 3D array to 2d matrix.
\end{description}
**ind2coord**

Find XYZ coords corresponding to 1D indices into a 3D image

### Description

If you have an image-like object and you want to turn it into a matrix of 3D coords then you need ind2coord. For the reverse operation we offer as.im3d.matrix which allows you to turn a matrix of 3D coordinates into an im3d image object.

### Usage

```r
ind2coord(inds, ...)  
```

### Arguments

- **inds** indices into an image array (either 1D, for which `dims` must be present, or a logical array).
- **...** extra arguments passed to methods.
- **dims** dimensions of 3D image array.
- **voxdims** vector of 3 voxel dimensions (width, height, depth).
- **origin** the origin.

### See Also

- coord2ind, sub2ind, xyzpos, as.im3d.matrix

---

**Details**

Note the sample locations stored in the x,y,z attributes will be updated appropriately. FIXME: Should we also update bounding box?

**See Also**

Other im3d: `as.im3d()`, `boundingbox()`, `im3d-coords`, `im3d-io`, `im3d()`, `imexpand.grid()`, `is.im3d()`, `mask()`, `origin()`, `projection()`, `threshold()`, `unmask()`, `voxdims()`
intersect  
Find the intersection of two collections of objects

Description
Find the intersection of two collections of objects

Usage
intersect(x, y, ...)

## Default S3 method:
intersect(x, y, ...)

## S3 method for class 'neuronlist'
intersect(x, y, ...)

Arguments
x the first collection to consider.
y the second collection to consider.
... additional arguments passed to methods

Details
Note that intersect.default calls base::intersect to ensure consistent behaviour for regular vectors.

Value
A collection of the same mode as x that contains all elements of x that are also present in y.

See Also
intersect

is.amiramesh  
Check if file is amiramesh format

Description
Check if file is amiramesh format

Usage
is.amiramesh(f = NULL, bytes = NULL)
is.fijitraces

Arguments

f  Path to one or more files to be tested or an array of raw bytes, for one file only.
bytes  optional raw vector of at least 11 bytes from the start of a single file (used in preference to reading file f).

Details

Tries to be as fast as possible by reading only first 11 bytes and checking if they equal to "# AmiraMesh" or (deprecated) "# HyperMesh".

Value

logical

See Also

Other amira: amiratype(), read.amiramesh(), read.hxsurf(), write.hxsurf()
is.im3d  
*Test if an object is of class im3d*

**Description**

Test if an object is of class im3d

**Usage**

```r
is.im3d(x)
```

**Arguments**

- **x**: Object to test

**Value**

logical

**See Also**

Other im3d: `as.im3d()`, `boundingbox()`, `im3d-coords`, `im3d-io`, `im3d()`, `imexpand.grid()`, `imslice()`, `mask()`, `origin()`, `projection()`, `threshold()`, `unmask()`, `voxdims()`

---

is.neuroml  
*Check whether a file is in NeuroML format*

**Description**

This will check a file on disk to see if it is in NeuroML format. Some prechecks (optionally taking place on a supplied raw vector of bytes) should weed out nearly all true negatives and identify many true positives without having to read/parsing the file header.

**Usage**

```r
is.neuroml(f, bytes = NULL)
```

**Arguments**

- **f**: path to a file on disk
- **bytes**: optional raw vector of bytes used for prechecks
is.neuronlist

Test objects of neuronlist class to store multiple neurons

Description

Tests if object is a neuronlist.

Usage

is.neuronlist(x)

Arguments

x the object to test

Details

is.neuronlist uses a relaxed definition to cope with older lists of neurons that do not have a class attribute of neuronlist.

Value

A logical indicating whether the object is a neuronlist.

See Also

Other neuronlist: *neuronlist(), neuronlist-dataframe-methods, neuronlistfh(), neuronlist(), nlapply(), read.neurons(), write.neurons()

is.nrrd

Check if a file is a NRRD file

Description

Check if a file is a NRRD file

Usage

is.nrrd(f = NULL, bytes = NULL, ReturnVersion = FALSE, TrustSuffix = FALSE)

Arguments

f A character vector specifying the path or a raw vector with at least 8 bytes.
bytes optional raw vector of at least 8 bytes from the start of a single file (used in preference to reading file f).
ReturnVersion Whether to return the version of the nrrd format in which the file is encoded (1-5).
TrustSuffix Whether to trust that a file ending in .nrrd or .nhdr is a NRRD
Details

Note that multiple files can be checked when a character vector of length > 1 is provided, but only one file can be checked when a raw byte array is provided.

Description

Test if a file is an SWC format neuron

Usage

is.swc(f, TrustSuffix = TRUE)

Arguments

f Path to one or more files
TrustSuffix Whether to trust that a file ending in .nrrd or .nhdr is a NRRD

Details

Note that this test is somewhat expensive compared with the other file tests since SWC files do not have a consistent magic value. It therefore often has to read and parse the first few lines of the file in order to determine whether they are consistent with the SWC format.

Value

logical value

See Also

read.neuron
is.vaa3draw

Description

Check if a file is in the raw image format used by Hanchuan Peng’s Vaa3D

Usage

is.vaa3draw(f, bytes = NULL)

Arguments

f A character vector specifying the path or a raw vector (see bytes).
bytes optional raw vector of at least 24 bytes from the start of a single file (used in preference to reading file f).

Details

Note that multiple files can be checked when a character vector of length > 1 is provided, but only one file can be checked when a raw byte array is provided.

kcs20

List of 20 Kenyon Cells from Chiang et al 2011 converted to dotprops objects

Description

This R list (which has additional class neuronlist) contains 20 skeletonized Drosophila Kenyon cells as dotprops objects. Original data is due to Chiang et al. 2011, who have generously shared their raw data at http://flycircuit.tw. Image registration and further processing was carried out by Greg Jefferis.

References


See Also

head.neuronlist, with.neuronlist, plot3d.neuronlist, plot3d.dotprops, dotprops

Other nat-data: Cell07PNs, MBL.surf
Examples

head(kcs20)
table(with(kcs20, type))
nopen3d()
# see plot3d.neuronlist documentation for more details

plot3d(kcs20, col=type)

---

**mask**

Mask an object, typically to produce a copy with some values zeroed out.

**Description**

Mask an object, typically to produce a copy with some values zeroed out.

**Usage**

```r
mask(x, ...)  
## S3 method for class 'im3d'
mask(x, mask, levels = NULL, rval = c("im3d", "values"), invert = FALSE, ...)
```

**Arguments**

- `x`: Object to be masked.
- `...`: Additional arguments passed to methods.
- `mask`: An `im3d` object, an array or a vector with dimensions compatible with `x`.
- `levels`: Optional numeric vector of pixel values or character vector defining named materials.
- `rval`: Whether to return an `im3d` object based on `x` or just the values from `x` matching the mask.
- `invert`: Whether to invert the voxel selection (default `FALSE`).

**Details**

Note that `mask.im3d` passes `...` arguments on to `im3d`.

**Value**

An object with attributes matching `x` and elements with value `as.vector(TRUE, mode=mode)` i.e. `TRUE`, `1`, `0x01` and `as.vector(FALSE, mode=mode)` i.e. `FALSE`, `0`, `0x00` as appropriate.

A copy of `x` with
See Also

Other im3d: `as.im3d()`, `boundingbox()`, `im3d-coords`, `im3d-io`, `im3d()`, `imexpand.grid()`, `imslice()`, `is.im3d()`, `origin()`, `projection()`, `threshold()`, `unmask()`, `voxdims()`

Examples

```r
x = im3d(array(rnorm(1000), dim = c(10, 10, 10)), BoundingBox = c(20, 200, 100, 200, 200, 300))
m = array(1:5, dim = c(10, 10, 10))
image(x[, , 1])
image(mask(x, mask = m, levels = 1)[, , 1])
image(mask(x, mask = m, levels = 1:2)[, , 1])
```

---

**materials**  
*Extract or set the materials for an object*

**Description**

`materials.character` will read the materials from an im3d compatible image file on disk.

`materials.hxsurf` will extract the materials from an hxsurf object

**Usage**

```r
materials(x, ...)
```

## Default S3 method:

```r
materials(x, ...)
```

## S3 method for class 'character'

```r
materials(x, ...)
```

## S3 method for class 'hxsurf'

```r
materials(x, ...)
```

**Arguments**

- `x`  
  An object in memory or, for `materials.character`, an image on disk.

- `...`  
  Additional parameters passed to methods (presently ignored)

**Details**

Note that the id column will be the 1-indexed order that the material appears in the `surf$Region` list for `hxsurf` objects and the 0-indexed mask values for an image.

Presently only amiramesh images are supported since they have a standardised way of encoding labels, whereas nrrds would have to use key-value pairs according to some ad hoc convention.
Value

A data.frame with columns name, id, col

See Also

Other hxsurf: as.hxsurf(), as.mesh3d(), plot3d.hxsurf(), read.hxsurf(), subset.hxsurf(), write.hxsurf()

---

MBL.surf

Surface object (hxsurf) for the left mushroom body in FCWB template space

Description

This surface object is in the same space as the 20 Kenyon cells in kcs20.

See Also

hxsurf

Other nat-data: Cell07PNs, kcs20

Examples

plot3d(kcs20)
plot3d(MBL.surf, alpha=0.3)

## Not run:
## originally generated as follows
library(nat.flybrains)
MBL.surf=subset(FCWBNP.surf, "MB.*_L", drop = T)

## End(Not run)

---

mirror

Mirror 3D object about a given axis, optionally using a warping registration

Description

mirroring with a warping registration can be used to account e.g. for the asymmetry between brain hemispheres.
mirror.character handles images on disk
mirror

Usage

mirror(x, ...)

## S3 method for class 'character'
mirror(x, output, mirrorAxisSize = NULL, target = x, ...)

## Default S3 method:
mirror(
  x,
  mirrorAxisSize,
  mirrorAxis = c("X", "Y", "Z"),
  warpfile = NULL,
  transform = c("warp", "affine", "flip"),
  ...
)

## S3 method for class 'neuronlist'
mirror(x, subset = NULL, OmitFailures = NA, ...)

Arguments

x Object with 3D points (with named cols X,Y,Z) or path to image on disk.

... additional arguments passed to methods or eventually to xform

output Path to the output image

mirrorAxisSize A single number specifying the size of the axis to mirror or a 2 vector (recommended) or 2x3 matrix specifying the boundingbox (see details).

target Path to the image defining the target grid (defaults to the input image - hard to see when this would not be wanted).

mirrorAxis Axis to mirror (default "X"). Can also be an integer in range 1:3.

warpfile Optional registration or reglist to be applied after the simple mirroring. It is called warpfile for historical reasons, since it is normally the path to a CMTK registration that specifies a non-rigid transformation to correct asymmetries in an image.

transform whether to use warp (default) or affine component of registration, or simply flip about midplane of axis.

subset For mirror.neuronlist indices (character/logical/integer) that specify a subset of the members of x to be transformed.

OmitFailures Whether to omit neurons for which FUN gives an error. The default value (NA) will result in nlapply stopping with an error message the moment there is an error. For other values, see details.

Details

The mirrorAxisSize argument can be specified in 3 ways for the x axis with extreme values, x0+x1:
• a single number equal to x0+x1
• a 2-vector c(x0, x1) (recommended)
• the boundingbox for the 3D data to be mirrored: the relevant axis specified by mirrorAxis will be extracted.

This function is agnostic re node vs cell data, but for node data BoundingBox should be supplied while for cell, it should be bounds. See boundingbox for details of BoundingBox vs bounds. See nlapply for details of the subset and OmitFailures arguments.

Value
Object with transformed points

See Also
xform, boundingbox
nlapply

Examples
nopen3d()
x=Cell07PNs[[1]]
mx=mirror(x,168)

plot3d(x,col='red')
plot3d(mx,col='green')

# also works with dotprops objects
clear3d()
y=kcs20[[1]]
my=mirror(y,mirrorAxisSize=564.2532,transform='flip')

plot3d(y, col='red')
plot3d(my, col='green')

## Not run:
## Example with an image
# note that we must specify an output image (obviously) but that as a
# convenience mirror calculates the mirrorAxisSize for us
mirror('myimage.nrrd', output='myimage-mirrored.nrrd',
   warpfile='myimage_mirror.list')

# Simple flip along a different axis
mirror('myimage.nrrd', output='myimage-flipped.nrrd', mirrorAxis="Y",
   transform='flip')

## End(Not run)
Description

The *normalised* digest should exclude any fields or attributes irrelevant to the core contents of the object (e.g. timestamps, absolute location of the input files on disk etc). In theory then, this value should be constant for the same data regardless of the particular machine on which the digest is being computed.

Usage

```r
ndigest(x, ...)
## S3 method for class 'neuronlistfh'
ndigest(x, ...)
## S3 method for class 'dotprops'
ndigest(x, absoluteVectors = TRUE, ...)
## S3 method for class 'neuron'
ndigest(
x,
  fieldsToExclude = c("InputFileName", "CreatedAt", "NodeName", "InputFileStat",
   "InputFileMD5"),
  ...
)
```

Arguments

`x` Object for which a normalised digest will be computed.

`...` Additional arguments passed to methods and then on to `digest`

`absoluteVectors` Whether to check only the absolute value of eigenvectors for equality (default TRUE, see details)

`fieldsToExclude` Character vector naming the neuron fields to exclude

Details

`ndigest.neuronlistfh` only considers the keyfilemap and df (metadata data.frame) when computing the hash value. See `neuronlistfh` for the significance of these two fields.

`ndigest.dotprops` ignores any mtime or file attributes. It also converts tangent vectors to absolute values (when `absoluteVectors=TRUE`) because the direction vectors are computed using an eigenvector decomposition where the sign of the eigenvector is essentially random and subject to
small numerical instabilities. Therefore it does not usually make sense to rely on the value of vect exactly.

`ndigest.neuron` ignores the following fields:

- `InputFileName`
- `CreatedAt`
- `NodeName`
- `InputFileStat`
- `InputFileMD5`

**Value**

A character string containing the digest of the supplied object computed by `digest`.

**See Also**

- `digest`
- `all.equal.dotprops`
- `all.equal.neuron`

**Examples**

```r
stopifnot(all.equal(ndigest(kcs20[[1]]), "4c045b0343938259cd9986494fc1c2b0")
```

---

**neuron**

**neuron: class to represent traced neurons**

**Description**

`neuron` makes a neuron object from appropriate variables.

- `is.neuron` will check if an object looks like a neuron.
- `as.neuron` will convert a suitable object to a neuron
- `as.neuron.data.frame` expects a block of SWC format data
- `as.neuron.ngraph` converts a graph (typically an `ngraph` object) to a neuron
- `as.neuron.igraph` will convert an `igraph` compatible `igraph` object into a neuron.
- `as.neuron.default` will add class "neuron" to a neuron-like object.
Usage

```
neuron(
  d,
  NumPoints = nrow(d),
  StartPoint,
  BranchPoints = integer(),
  EndPoints,
  SegList,
  SubTrees = NULL,
  InputFileName = NULL,
  NeuronName = NULL,
  ..., 
  MD5 = TRUE
)
```

```
is.neuron(x, Strict = FALSE)
```

```
as.neuron(x, ...)
```

```
## S3 method for class 'data.frame'
as.neuron(x, ...)
```

```
## S3 method for class 'ngraph'
as.neuron(x, vertexData = NULL, origin = NULL, Verbose = FALSE, ...)
```

```
## S3 method for class 'igraph'
as.neuron(x, ...)
```

```
## Default S3 method:
as.neuron(x, ...)
```

Arguments

- **d**: matrix of vertices and associated data in SWC format
- **NumPoints**: Number of points in master subtree
- **StartPoint, BranchPoints, EndPoints**: Nodes of the neuron
- **SegList**: List where each element contains the vertex indices for a single segments of the neuron, starting at root.
- **SubTrees**: List of SegLists where a neuron has multiple unconnected trees (e.g. because the soma is not part of the graph, or because the neuronal arbour has been cut.)
- **InputFileName**: Character vector with path to input file
- **NeuronName**: Character vector containing name of neuron or a function with one argument (the full path) which returns the name. The default (NULL) sets NeuronName to the file name without the file extension.
Additional fields to be included in neuron. Note that if these include Create Dt, NodeName, InputFileStat or InputFileMD5, they will override fields of that name that are calculated automatically.

**MD5**
Logical indicating whether to calculate MD5 hash of input

**x**
A neuron or other object to test/convert

**Strict**
Whether to check class of neuron or use a more relaxed definition based on object being a list with a SegList component.

**vertexData**
A dataframe with SWC fields especially X,Y,Z,W,PointNo, Parent.

**origin**
Root vertex, matched against labels (aka PointNo) when available (see details)

**Verbose**
Whether to be verbose (default: FALSE)

**Details**
neuron objects consist of a list containing multiple fields describing the 3D location and connectivity of points in a traced neuron. The critical fields of a neuron, n, are n$d which contains a dataframe in SWC format and n$SegList which contains a representation of the neuron’s topology used for most internal calculations. For historical reasons, n$SegList is limited to a single fully-connected tree. If the tree contains multiple unconnected subtrees, then these are stored in n$SubTrees and nTrees will be >1; the "master" subtree (typically the one with the most points) will then be stored in n$SegList and n$NumPoints will refer to the number of points in that subtree, not the whole neuron.

StartPoint, BranchPoints, EndPoints are indices matching the rows of the vertices in d not arbitrary point numbers typically encoded in d$PointNo.

Columns will be ordered c('PointNo', 'Label', 'X', 'Y', 'Z', 'W', 'Parent')

Uses a depth first search on the tree to reorder using the given origin.

When the graph contains multiple subgraphs, only one will be chosen as the master tree and used to construct the SegList of the resultant neuron. However all subgraphs will be listed in the SubTrees element of the neuron and nTrees will be set appropriately.

When the graph vertices have a label attribute derived from PointNo, the origin is assumed to be specified with respect to the vertex labels rather than the raw vertex ids.

**Value**
A list with elements: (NumPoints, StartPoint, BranchPoints, EndPoints, nTrees, NumSegs, SegList, [SubTrees]) NB SubTrees will only be present when nTrees>1.

**See Also**
neuronlist
graph.dfs, as.seglist

Other neuron: ngraph(), plot.neuron(), potential_synapses(), prune(), resample(), rootpoints(), spine(), subset.neuron()
Examples

## See help for functions listed in See Also for more detailed examples
## Basic properties
# a sample neuron
n = Cell07PNs[[1]]
# inspect its internal structure
str(n)
# summary of 3D points
summary(xyzmatrix(n))
# identify 3d location of endpoints
xyzmatrix(n)[endpoints(n),]

## Other methods
# plot
plot(n)
# all methods for neuron objects
methods(class = 'neuron')

## Neurons as graphs
# convert to graph and find longest paths by number of nodes
ng=as.ngraph(n)
hist(igraph::distances(ng))
# ... or in distances microns
ngw=as.ngraph(n, weights=TRUE)
hist(igraph::distances(ngw))

# converting back and forth between neurons and graphs
g=as.ngraph(Cell07PNs[[1]])
gstem=igraph::induced.subgraph(g, 1:10)
# this is fine
plot(gstem)
plot(as.neuron(gstem))

# but if you had an undirected graph
ug=igraph::as.undirected(gstem)
# you get a warning because there is no explicit origin for the graph
as.neuron(ug)

# If you need finer control of the conversion process
gstem2=as.ngraph(ug, root = 10)
plot(gstem2)
plot(as.neuron(gstem2))

neuronlist

Create a neuronlist from zero or more neurons

Description

neuronlist objects consist of a list of neuron objects (usually of class neuron or dotprops) along with an optional attached dataframe containing information about the neurons. neuronlist objects
can be indexed using their name or the number of the neuron like a regular list. Both the list itself and the attached data.frame must have the same unique (row)names. If the [ operator is used to index the list, the attached dataframe will also be subsetted.

It is perfectly acceptable not to pass any parameters, generating an empty neuronlist

Usage

neuronlist(..., DATAFRAME = NULL)

Arguments

... objects to be turned into a list
DATAFRAME an optional data.frame to attach to the neuronlist containing information about each neuron.

Value

A new neuronlist object.

See Also

as.data.frame.neuronlist, neuronlist-dataframe-methods, neuron, dotprops

Other neuronlist: *.neuronlist(), is.neuronlist(), neuronlist-dataframe-methods, neuronlistfh(), nlapply(), read.neurons(), write.neurons()

Examples

# generate an empty neuronlist
nl=neuronlist()
# slice an existing neuronlist with regular indexing
kcs5=kcs20[1:5]

# extract a single neuron from a neuronlist
n1=Cell07PNs[[1]]

# list all methods for neuronlist objects
methods(class='neuronlist')
Description

`.neuronlist` and `<-.neuronlist` behave like the corresponding base methods (`.data.frame`, `<-.data.frame`) allowing extraction or replacement of parts of the `data.frame` attached to the `neuronlist`.

droplevels Remove redundant factor levels in `data.frame` attached to `neuronlist`  
with Evaluate expression in the context of `data.frame` attached to a `neuronlist`  
head Return the first part of `data.frame` attached to `neuronlist`  
tail Return the last part of `data.frame` attached to `neuronlist`

Usage

```r
## S3 method for class 'neuronlist'
x[i, j, drop]
## S3 replacement method for class 'neuronlist'
x[i, j] <- value
## S3 method for class 'neuronlist'
droplevels(x, except = NULL, ...)
## S3 method for class 'neuronlist'
with(data, expr, ...)
## S3 method for class 'neuronlist'
head(x, ...)
## S3 method for class 'neuronlist'
tail(x, ...)
```

Arguments

- `x` A `neuronlist` object
- `i, j` elements to extract or replace. Numeric or character or, for `[` only, empty. Numeric values are coerced to integer as if by `as.integer`. See `.[data.frame]` for details.
- `drop` logical. If `TRUE` the result is coerced to the lowest possible dimension. The default is to drop if only one column is left, but `not` to drop if only one row is left.
- `value` A suitable replacement value: it will be repeated a whole number of times if necessary and it may be coerced: see the Coercion section. If `NULL`, deletes the column if a single column is selected.
- `except` indices of columns from which `not` to drop levels
- `...` Further arguments passed to default methods (and usually ignored)
- `data` A `neuronlist` object
- `expr` The expression to evaluate
Value

the attached dataframe with levels dropped (NB not the neuronlist)

See Also

[.data.frame, @seealso [<- .data.frame
droplevels
with
head
tail

Other neuronlist: *neuronlist(), is.neuronlist(), neuronlistfh(), neuronlist(), nlapply(),
read.neurons(), write.neurons()

Examples

## treat kcs20 as data.frame
kcs20[1, ]
kcs20[1:3, ]
kcs20[, 1:4]
kcs20[, 'soma_side']
# alternative to as.data.frame(kcs20)
kcs20[, ]

## can also set columns
kcs13=kcs20[1:3]
kcs13,['side']=as.character(kcs13,['soma_side'])
head(kcs13)
# or parts of columns
kcs13[1,'soma_side']='R'
kcs13['FruMARCM-M001205_seg002','soma_side']='L'
# remove a column
kcs13,['side']=NULL
all.equal(kcs13, kcs20[1:3])

# can even replace the whole data.frame like this
kcs13[,]=kcs13[,]
all.equal(kcs13, kcs20[1:3])

## get row/column names of attached data.frame
# (unfortunately implementing ncol/nrow is challenging)
rownames(kcs20)
colnames(kcs20)
neuronlistfh

neuronlistfh - List of neurons loaded on demand from disk or remote website

Description

neuronlistfh objects consist of a list of neuron objects along with an optional attached dataframe containing information about the neurons. In contrast to neuronlist objects the neurons are not present in memory but are instead dynamically loaded from disk as required. neuronlistfh objects also inherit from neuronlist and therefore any appropriate methods e.g. plot3d.neuronlist can also be used on neuronlistfh objects.

neuronlistfh constructs a neuronlistfh object from a filehash, data.frame and keyfilemap. End users will not typically use this function to make a neuronlistfh. They will usually read them using read.neuronlistfh and sometimes create them by using as.neuronlistfh on a neuronlist object.

is.neuronlistfh test if an object is a neuronlistfh

as.neuronlistfh generic function to convert an object to neuronlistfh

as.neuronlistfh.neuronlist converts a regular neuronlist to one backed by a filehash object with an on disk representation

Usage

neuronlistfh(db, df, keyfilemap, hashmap = 1000L)

is.neuronlistfh(nl)

as.neuronlistfh(x, df, ...)

## S3 method for class 'neuronlist'
as.neuronlistfh(
x,
  df = attr(x, "df"),
  dbdir = NULL,
  dbClass = c("RDS", "RDS2"),
  remote = NULL,
  WriteObjects = c("yes", "no", "missing"),
  ...
)

Arguments

db a filehash object that manages an on disk database of neuron objects. See Implementation details.

df Optional dataframe, where each row describes one neuron
keyfilemap  A named character vector in which the elements are filenames on disk (managed by the filehash object) and the names are the keys used in R to refer to the neuron objects. Note that the keyfilemap defines the order of objects in the neuronlist and will be used to reorder the dataframe if necessary.

hashmap  A logical indicating whether to add a hashed environment for rapid object lookup by name or an integer or an integer defining a threshold number of objects when this will happen (see Implementation details).

nl  Object to test

x  Object to convert

...  Additional arguments for methods, eventually passed to neuronlistfh() constructor.

dbdire  The path to the underlying filehash database on disk. By convention this should be a path whose final element is 'data'

dbClass  The filehash database class. Defaults to RDS.

remote  The url pointing to a remote repository containing files for each neuron.

WriteObjects  Whether to write objects to disk. Missing implies that existing objects will not be overwritten. Default "yes".

Value

a neuronlistfh object which is a character vector with classes neuronlistfh, neuronlist and attributes db, df. See Implementation details.

Implementation details

neuronlistfh objects are a hybrid between regular neuronlist objects that organise data and meta-data for collections of neurons and a backing filehash object. Instead of keeping objects in memory, they are always loaded from disk. Although this sounds like it might be slow, for nearly all practical purposes (e.g. plotting neurons) the time to read the neuron from disk is small compared with the time to plot the neuron; the OS will cache repeated reads of the same file. The benefits in memory and startup time (<1s vs 100s for our 16,000 neuron database) are vital for collections of 1000s of neurons e.g. for dynamic report generation using knitr or for users with <8Gb RAM or running 32 bit R.

neuronlistfh objects include:

- attr("keyfilemap") A named character vector that determines the ordering of objects in the neuronlist and translates keys in R to filenames on disk. For objects created by as.neuronlistfh the filenames will be the md5 hash of the object as calculated using digest. This design means that the same key can be used to refer to multiple distinct objects on disk. Objects are effectively versioned by their contents. So if an updated neuronlistfh object is posted to a website and then fetched by a user it will result in the automated download of any updated objects to which it refers.

- attr("db") The backing database - typically of class filehashRDS. This manages the loading of objects from disk.

- attr(x, "df") The data.frame of metadata which can be used to select and plot neurons. See neuronlist for examples.
neuronlistfh

- attr(x, "hashmap") (Optional) a hashed environment which can be used for rapid lookup using key names (rather than numeric/logical indices). There is a space potential to pay for this redundant lookup method, but it is normally worth while given that the dataframe object is typically considerably larger. To give some numbers, the additional environment might occupy ~1 time from 0.5 ms to 1us. Having located the object, on my machine it can take as little as 0.1ms to load from disk, so these savings are relevant.

Presently only backing objects which extend the filehash class are supported (although in theory other backing objects could be added). These include:

- filehash RDS
- filehash RDS2 (experimental)

We have also implemented a simple remote access protocol (currently only for the RDS format). This allows a neuronlistfh object to be read from a url and downloaded to a local path. Subsequent attempts to access neurons stored in this list will result in automated download of the requested neuron to the local cache.

An alternative backend, the experimental RDS2 format is supported (available at https://github.com/jefferis/filehash). This is likely to be the most effective for large (5,000-500,000) collections of neurons, especially when using network filesystems (nfs, afp) which are typically very slow at listing large directories.

Note that objects are stored in a filehash, which by definition does not have any ordering of its elements. However neuronlist objects (like lists) do have an ordering. Therefore the names of a neuronlistfh object are not necessarily the same as the result of calling names() on the underlying filehash object.

See Also

filehash-class

Other neuronlistfh: [.neuronlistfh(), read.neuronlistfh(), remotesync(), write.neuronlistfh()]

Other neuronlist: *.neuronlist(), is.neuronlist(), neuronlist-dataframe-methods, neuronlist(), nlapply(), read.neurons(), write.neurons()

Examples

```r
## Not run:
kcnl=read.neuronlistfh(’http://jefferislab.org/si/blast/flycircuit/kcs20.rds’,
’path/to/my/project/folder’)  
# this will automatically download the neurons from the web the first time
# it is run
plot3d(kcnl)

## End(Not run)
## Not run:
## Not run:
# create neuronlistfh object backed by filehash with one file per neuron
# by convention we create a subfolder called data in which the objects live
kcs20fh=as.neuronlistfh(kcs20, dbdir=’/path/to/my/kcdb/data’)  
plot3d(subset(kcs20fh, type==’gamma’))

# ... and, again by convention, save the neuronlistfh object next to filehash
# backing database
```
```
write.neuronlistfh(kcs20fh, file='"/path/to/my/kcdb/kcdb.rds"

# in a new session
read.neuronlistfh("/path/to/my/kcdb/kcdb.rds")
plot3d(subset(kcs20fh, type='"gamma")

## End(Not run)
```

---

**ngraph**  

**ngraph**: a graph to encode a neuron’s connectivity

### Description

The `ngraph` class contains a (completely general) graph representation of a neuron’s connectivity in an `igraph` object. It may additionally contain vertex label or position data. See details.

`ngraph()` creates an `ngraph` from edge and vertex information.

`as.ngraph` converts an object to an `ngraph`.

`as.ngraph.dataframe` constructs `ngraph` from a `data.frame` containing SWC format data.

`as.ngraph.neuron` constructs `ngraph` from a neuron.

### Usage

```r
ngraph(
el, vertexlabels, xyz = NULL, diam = NULL, directed = TRUE, weights = FALSE, vertex.attributes = NULL, graph.attributes = NULL
)
as.ngraph(x, ...)  
```

#### Arguments

- **el**: A two-column matrix (start, end) defining edges. **start** means closer to the root (soma) of the neuron.
- **vertexlabels**: Integer labels for graph - the edge list is specified using these labels.
xyz 3D coordinates of vertices (optional, Nx3 matrix, or Nx4 matrix when 4th column is assumed to be diameter)
diam Diameter of neuron at each vertex (optional)
directed Whether the resultant graph should be directed (default TRUE)
weights Logical value indicating whether edge weights defined by the 3D distance between points should be added to graph (default FALSE) or a numeric vector of weights.
vertex.attributes, graph.attributes List of named attributes to be added to the graph. The elements of vertex.attributes must be vectors whose length is compatible with the number of elements in the graph. See set.vertex.attribute for details.
x Object to convert (see method descriptions)
... Arguments passed to methods
method Whether to use the swc data (x$d) or the seglist to define neuronal connectivity to generate graph.

Details

Note that the as.ngraph.neuron method always keeps the original vertex labels (a.k.a. PointNo) as read in from the original file.

Value

an igraph object with additional class ngraph, having a vertex for each entry in vertexlabels, each vertex having a label attribute. All vertices are included whether connected or not.

Connectivity

We make the following assumptions about neurons coming in

- They have an integer vertex label that need not start from 1 and that may have gaps
- The edge list which defines connectivity specifies edges using pairs of vertex labels, _not_ raw vertex ids.

We make no attempt to determine the root points at this stage.

The raw vertex ids in the graph will be in the order of vertexlabels and can therefore be used to index a block of vertex coordinates. The vertexlabels will be stored using the vertex attribute label.

When the graph is directed (default) the edges will be from the root to the other tips of the neuron.

Morphology

The morphology of the neuron is encoded by the combination of connectivity information (i.e. the graph) and spatial data encoded as the 3D position and diameter of each vertex. Position information is stored as vertex attributes X, Y, and Z.
See Also

igraph, set.vertex.attribute, subset.neuron for example of graph-based manipulation of a neuron.

Other neuron: neuron(), plot.neuron(), potential_synapses(), prune(), resample(), rootpoints(), spine(), subset.neuron()

Examples

g=as.ngraph(Cell07PNs[[1]])
library(igraph)
# check that vertex attributes of graph match X position
all.equal(V(g)$X, Cell07PNs[[1]]$d$X)

nlapply

lapply and mapply for neuronlists (with optional parallelisation)

Description

versions of lapply and mapply that look after the class and attached dataframe of neuronlist objects.
nlapply can apply a function to only a subset of elements in the input neuronlist. Internally nlapply uses plyr::llply thereby enabling progress bars and simple parallelisation (see plyr section and examples).

Usage

nlapply(
  X,
  FUN,
  ..., 
  subset = NULL,
  OmitFailures = NA,
  .progress = getOption("nat.progress", default = "auto")
)

nmapply(
  FUN,
  X,
  ..., 
  MoreArgs = NULL,
  SIMPLIFY = FALSE,
  USE.NAMES = TRUE,
  subset = NULL,
  OmitFailures = NA
)
Arguments

- **X**: A neuronlist
- **FUN**: Function to be applied to each element of X
- **...**: Additional arguments for FUN (see details)
- **subset**: Character, numeric or logical vector specifying on which subset of X the function FUN should be applied. Elements outside the subset are passed through unmodified.
- **OmitFailures**: Whether to omit neurons for which FUN gives an error. The default value (NA) will result in nlapply stopping with an error message the moment there is an error. For other values, see details.
- **.progress**: Character vector specifying the type of progress bar (see create_progress_bar for options.) The default value of "auto" shows a progress bar in interactive use when there are >=10 elements in X. The default value can be overridden for the current session by setting the value of options(nat.progressbar) (see examples).
- **MoreArgs**: a list of other arguments to FUN.
- **SIMPLIFY**: logical or character string; attempt to reduce the result to a vector, matrix or higher dimensional array; see the simplify argument of sapply.
- **USE.NAMES**: logical; use the names of the first ...argument, or if that is an unnamed character vector, use that vector as the names.

Details

When OmitFailures is not NA, FUN will be wrapped in a call to try to ensure that failure for any single neuron does not abort the nlapply/nmapply call. When OmitFailures=TRUE the resultant neuronlist will be subsetted down to return values for which FUN evaluated successfully. When OmitFailures=FALSE, "try-error" objects will be left in place. In either of the last 2 cases error messages will not be printed because the call is wrapped as try(expr, silent=TRUE).

Value

A neuronlist

plyr

The arguments of most interest from plyr are:

- **.inform** set to TRUE to give more informative error messages that should indicate which neurons are failing for a given applied function.
- **.progress** set to "text" for a basic progress bar
- **.parallel** set to TRUE for parallelisation after registering a parallel backend (see below).
- **.paropts** Additional arguments for parallel computation. See lapply for details.

Before using parallel code within an R session you must register a suitable parallel backend. The simplest example is the multicore option provided by the doMC package that is suitable for a spreading computational load across multiple cores on a single machine. An example is provided below.
Note that the progress bar and parallel options cannot be used at the same time. You may want to start a potentially long-running job with the progress bar option and then abort and re-run with `.parallel=TRUE` if it looks likely to take a very long time.

See Also

lapply
mapply

Other neuronlist: *.neuronlist(), is.neuronlist(), neuronlist-dataframe-methods, neuronlistfh(), neuronlist(), read.neurons(), write.neurons()

Examples

```r
## nlapply example
kcs.reduced=nlapply(kcs20,function(x) subset(x,sample(nrow(x$points),50)))
open3d()
plot3d(kcs.reduced,col='red', lwd=2)
plot3d(kcs20,col='grey')
rgl.close()

## Not run:
# example of using plyr's .inform argument for debugging error conditions
xx=nlapply(Cell07PNs, prune_strahler)
# oh dear there was an error, let's get some details about the neuron
# that caused the problem
xx=nlapply(Cell07PNs, prune_strahler, .inform=TRUE)

## End(Not run)

## Not run:
## nlapply example with plyr
## dotprops.neuronlist uses nlapply under the hood
## the .progress and .parallel arguments are passed straight to
## system.time(d1<-dotprops(kcs20,resample=1,k=5,.progress='text'))
## plyr+parallel
library(doMC)
# can also specify cores e.g. registerDoMC(cores=4)
registerDoMC()
system.time(d2<-dotprops(kcs20,resample=1,k=5,.parallel=TRUE))
stopifnot(all.equal(d1,d2))

## End(Not run)

## nmapply example
# flip first neuron in X, second in Y and 3rd in Z
xyzflip=nmapply(mirror, kcs20[1:3], mirrorAxis = c("X","Y","Z"),
mirrorAxisSize=c(400,20,30))
open3d()
plot3d(kcs20[1:3])
plot3d(xyzflip)
rgl.close()
```
nlscan

Scan through a set of neurons, individually plotting each one in 3D

Description

Can also choose to select specific neurons along the way and navigate forwards and backwards.

Usage

nlscan(
  neurons,
  db = NULL,
  col = "red",
  Verbose = T,
  Wait = T,
  sleep = 0.1,
  extrafun = NULL,
  selected_file = NULL,
  selected_col = "green",
  yaml = TRUE,
  ...
)

Arguments

neurons a neuronlist object or a character vector of names of neurons to plot from the neuronlist specified by db.
db A neuronlist to use as the source of objects to plot. If NULL, the default, will use the neuronlist specified by options('nat.default.neuronlist')
col the color with which to plot the neurons (default 'red').
Verbose logical indicating that info about each selected neuron should be printed (default TRUE).
Wait logical indicating that there should be a pause between each displayed neuron.
sleep time to pause between each displayed neuron when Wait=TRUE.
extrafun an optional function called when each neuron is plotted, with two arguments: the current neuron name and the current selected neurons.

selected_file an optional path to a yaml file that already contains a selection.

selected_col the color in which selected neurons (such as those specified in selected_file) should be plotted.

yaml a logical indicating that selections should be saved to disk in (human-readable) yaml rather than (machine-readable) rda format.

... extra arguments to pass to plot3d.

Value

A character vector of names of any selected neurons, of length 0 if none selected.

See Also

plot3d.character, plot3d.neuronlist

Examples

```r
## Not run:
# scan a neuronlist
nlscan(kcs20)

# using neuron names
nlscan(names(kcs20), db=kcs20)
# equivalently using a default neuron list
options(nat.default.neuronlist='kcs20')
nlscan(names(kcs20))

## End(Not run)
# scan without waiting
nlscan(kcs20[1:4], Wait=FALSE, sleep=0)
## Not run:
# could select e.g. the gamma neurons with unbranched axons
gammas=nlscan(kcs20)
clear3d()
plot3d(kcs20[gammas])

# plot surface model of brain first
# nb depends on package only available on github
devtools::install_github(username = "jefferislab/nat.flybrains")
library(nat.flybrains)
plot3d(FCWB)
# could select e.g. the gamma neurons with unbranched axons
gammas=nlscan(kcs20)
clear3d()
plot3d(kcs20[gammas])

## End(Not run)
```
nopen3d  
*Open customised rgl window*

**Description**

Pan with right button (Ctrl+click), zoom with middle (Alt/Meta+click) button. Defaults to a white background and orthogonal projection (FOV=0)

**Usage**

nopen3d(bgcol = "white", FOV = 0, ...)

**Arguments**

- `bgcol` background colour
- `FOV` field of view
- `...` additional options passed to open3d

**Details**

Note that sometimes (parts of) objects seem to disappear after panning and zooming. See help for `pan3d`.

**Value**

current rgl device

**See Also**

open3d, pan3d

---

**normalise_swc**  
*Normalise an SWC format block of neuron morphology data*

**Description**

Normalise an SWC format block of neuron morphology data

**Usage**

```r
normalise_swc(
  x,
  requiredColumns = c("PointNo", "Label", "X", "Y", "Z", "W", "Parent"),
  ifMissing = c("usedefaults", "warning", "stop"),
  includeExtraCols = TRUE,
  defaultValue = list(PointNo = seq.int(nrow(x)), Label = 2L, X = NA_real_, Y = NA_real_, Z = NA_real_, W = NA_real_, Parent = NA_integer_)
)
```
Arguments

x: A data.frame containing neuron morphology data
requiredColumns: Character vector naming columns we should have
ifMissing: What to do if x is missing a required column
includeExtraCols: Whether to include any extra columns include in codex
defaultValue: A list containing default values to use for any missing columns

Details

Note that row.names of the resultant data.frame will be set to NULL so that they have completely standard values.

Value

A data.frame containing the normalised block of SWC data with standard columns in standard order.

See Also

\texttt{as.neuron.data.frame, seglist2swc}

\begin{verbatim}
npop3d
Remove plotted neurons or other 3D objects
\end{verbatim}

Description

The normal usage will not specify \texttt{x} in which case the last neurons plotted by \texttt{plot3d.neuronlist} or any of its friends will be removed.

Usage

\texttt{npop3d(x, slow = FALSE, type = "shapes")}

Arguments

x: rgl ids of objects to remove
slow: Whether to remove neurons one by one (slowly) default: FALSE
type: Type of objects to remove see \texttt{pop3d}.

See Also

\texttt{pop3d, plot3d.neuronlist}
nrrd.voxdims

Return voxel dimensions (by default absolute voxel dimensions)

Description

Return voxel dimensions (by default absolute voxel dimensions)

Usage

nrrd.voxdims(file, ReturnAbsoluteDims = TRUE)

Arguments

file path to nrrd/nhdr file or a list containing a nrrd header
ReturnAbsoluteDims Defaults to returning absolute value of dims even if there are any negative space directions

Details

NB Can handle off diagonal terms in space directions matrix, BUT assumes that space direction vectors are orthogonal.

Will produce a warning if no valid dimensions can be found.

Value

numeric vector of voxel dimensions (NA_real_ when missing) of length equal to the image dimension.

Author(s)

jefferis

See Also

read.nrrd.header
nvertices

Find the number of vertices in an object (or each element of a neuronlist)

Description

Find the number of vertices in an object (or each element of a neuronlist)

Usage

nvertices(x, ...)

## Default S3 method:
nvertices(x, ...)

## S3 method for class 'neuronlist'
nvertices(x, ...)

Arguments

x An object with 3d vertices (e.g. neuron, surface etc)

... Additional arguments passed to methods (currently ignored)

Value

an integer number of vertices (or a vector of length equal to a neuronlist)

Examples

nvertices(Cell07PNs[[1]])
nvertices(kcs20)

nview3d

Set the 3D viewpoint of an RGL window using anatomical terms

Description

Set the 3D viewpoint of an RGL window using anatomical terms

Usage

nview3d(
  viewpoint = c("frontal", "anterior", "dorsal", "ventral", "posterior", "left",
              "right", "oblique_right", "oblique_left"),
  FOV = 0,
  extramat = NULL,
  ...
)

**origin**

Return the space origin of a 3D image object

**Description**

Defined as the first coordinates (x,y,z) of the bounding box, which in turn matches the nrrd definition of the location of the "centre" of the first voxel.

**Usage**

```r
origin(x, ...)
```

**Arguments**

- `x` Object for which origin should be returned. See boundingbox.
- `...` Additional arguments passed to boundingbox

**See Also**

Other im3d: `as.im3d()`, `boundingbox()`, `im3d-coords`, `im3d-io`, `im3d()`, `imexpand.grid()`, `imslice()`, `is.im3d()`, `mask()`, `projection()`, `threshold()`, `unmask()`, `voxdims()`
Some useful extensions / changes to rgl defaults

Description

Set up pan call back for current rgl device

Usage

pan3d(button)

Arguments

button Integer from 1 to 3 indicating mouse button

Details

Copied verbatim from ?rgl.setMouseCallbacks for rgl version 0.92.892 Mouse button 2 is right and button 3 is middle (accessed by meta/alt key)

Note that sometimes (parts of) objects seem to disappear after panning and zooming. The example in rgl.setMouseCallbacks from which this is copied includes a note that "this doesn’t play well with rescaling"

Author(s)

Duncan Murdoch

See Also

rgl.setMouseCallbacks

Examples

## Not run:
open3d()
pan3d(2)

## End(Not run)
plot.neuron

Plot a 2D projection of a neuron

Description

Plot a 2D projection of a neuron

Usage

## S3 method for class 'neuron'
plot(
  x,
  WithLine = TRUE,
  WithNodes = TRUE,
  WithAllPoints = FALSE,
  WithText = FALSE,
  PlotSubTrees = TRUE,
  soma = FALSE,
  PlotAxes = c("XY", "YZ", "XZ", "ZY"),
  axes = TRUE,
  asp = 1,
  main = x$NeuronName,
  sub = NULL,
  xlim = NULL,
  ylim = NULL,
  AxisDirections = c(1, -1, 1),
  add = FALSE,
  col = NULL,
  PointAlpha = 1,
  tck = NA,
  lwd = par("lwd"),
  boundingbox = NULL,
  ...
)

Arguments

x
WithLine
WithNodes
WithAllPoints
WithText
PlotSubTrees
soma

a neuron to plot.
whether to plot lines for all segments in neuron.
whether points should only be drawn for nodes (branch/end points)
whether points should be drawn for all points in neuron.
whether to label plotted points with their id.
Whether to plot all sub trees when the neuron is not fully connected.
Whether to plot a circle at neuron’s origin representing the soma. Either a logical value or a numeric indicating the radius (default FALSE). When soma=TRUE the radius is hard coded to 2.
PlotAxes  the axes for the plot.

axes     whether axes should be drawn.

asp      the y/x aspect ratio, see plot.window.

main     the title for the plot

sub      sub title for the plot

xlim     limits for the horizontal axis (see also boundingbox)

ylim     limits for the vertical axis (see also boundingbox)

AxisDirections  the directions for the axes. By default, R uses the bottom-left for the origin, whilst most graphics software uses the top-left. The default value of c(1, -1, 1) makes the produced plot consistent with the latter.

add      Whether the plot should be superimposed on one already present (default: FALSE).

col      the color in which to draw the lines between nodes.

PointAlpha  the value of alpha to use in plotting the nodes.

tck      length of tick mark as fraction of plotting region (negative number is outside graph, positive number is inside, 0 suppresses ticks, 1 creates gridlines).

lwd      line width relative to the default (default=1).

boundingbox  A 2 x 3 matrix (ideally of class boundingbox) that enables the plot axis limits to be set without worrying about axis selection or reversal (see details)

...  additional arguments passed to plot

Details

This functions sets the axis ranges based on the chosen PlotAxes and the range of the data in x. It is still possible to use PlotAxes in combination with a boundingbox, for example to set the range of a plot of a number of objects.

nat assumes the default axis convention used in biological imaging, where the origin of the y axis is the top rather than the bottom of the plot. This is achieved by reversing the y axis of the 2D plot when the second data axis is the Y axis of the 3D data. Other settings can be achieved by modifying the AxisDirections argument.

Value

list of plotted points (invisibly)

See Also

plot3d.neuron

Other neuron: neuron().ngraph().potential_synapses().prune().resample().rootpoints().spine().subset.neuron()
Examples

# Draw first example neuron
plot(Cell07PNs[1])
# Overlay second example neuron
plot(Cell07PNs[2], add=TRUE)
# Clear the current plot and draw the third neuron from a different view
plot(Cell07PNs[3], PlotAxes="YZ")
# Just plot the end points for the fourth example neuron
plot(Cell07PNs[4], WithNodes=False)
# Plot with soma (of default radius)
plot(Cell07PNs[4], WithNodes=False, soma=TRUE)
# Plot with soma of defined radius
plot(Cell07PNs[4], WithNodes=False, soma=1.25)

plot.neuronlist 2D plots of the elements in a neuronlist, optionally using a subset expression

Description

2D plots of the elements in a neuronlist, optionally using a subset expression

Usage

## S3 method for class 'neuronlist'
plot(
  x, 
  subset = NULL, 
  col = NULL, 
  colpal = rainbow, 
  add = NULL, 
  boundingbox = NULL, 
  ..., 
  SUBSTITUTE = TRUE 
)

Arguments

x a neuron list or, for plot3d.character, a character vector of neuron names. The default neuronlist used by plot3d.character can be set by using options(nat.default.neuronlist=)
  See ?nat for details.
subset Expression evaluating to logical mask for neurons. See details.
col An expression specifying a colour evaluated in the context of the dataframe attached to nl (after any subsetting). See details.
colpal A vector of colours or a function that generates colours
plot3d

add Logical specifying whether to add data to an existing plot or make a new one. The default value of NULL creates a new plot with the first neuron in the neuron-list and then adds the remaining neurons.

boundingbox A 2 x 3 matrix (ideally of class boundingbox) that enables the plot axis limits to be set without worrying about axis selection or reversal (see details)

... options passed on to plot (such as colours, line width etc)

SUBSTITUTE Whether to substitute the expressions passed as arguments subset and col. Default: TRUE. For expert use only, when calling from another function.

Details

The col and subset parameters are evaluated in the context of the dataframe attribute of the neuron-list. If col evaluates to a factor and colpal is a named vector then colours will be assigned by matching factor levels against the named elements of colpal. If there is one unnamed level, this will be used as catch-all default value (see examples).

If col evaluates to a factor and colpal is a function then it will be used to generate colours with the same number of levels as are used in col.

Value

list of values of plot with subsetted dataframe as attribute 'df'

See Also

nat-package, plot3d.neuronlist

Examples

# plot 4 cells
plot(Cell07PNs[1:4])
# modify some default plot arguments
plot(Cell07PNs[1:4], ylim=c(140,75), main='First 4 neurons')
# plot one class of neurons in red and all the others in grey
plot(Cell07PNs, col=Glomerulus, colpal=c(DA1='red', 'grey'), WithNodes=FALSE)
# subset operation
plot(Cell07PNs, subset=Glomerulus%in%c("DA1", "DP1m"), col=Glomerulus,
    ylim=c(140,75), WithNodes=FALSE)

plot3d

plot3d methods for different nat objects

Description

These methods enable nat objects including neuronlists and dotprops objects to be plotted in 3D. See the help for each individual method for details along with the help for the generic in the rgl package.
plot3d.boundingbox

See Also

plot3d, plot3d.boundingbox, plot3d.character, plot3d.cmtkreg, plot3d.dotprops, plot3d.hxsurf, plot3d.neuron, plot3d.neuronlist

Examples

# all known plot3d methods
methods("plot3d")

# up to date list of all plot3d methods in this package
intersect(methods("plot3d"), ls(asNamespace("nat")))

plot3d.boundingbox  Plot a bounding box in 3D

Description

Plot a bounding box in 3D

Usage

## S3 method for class 'boundingbox'
plot3d(x, ...)

Arguments

x      the boundingbox object to plot.
...
additional arguments to pass to segments3d.

Value

A list of RGL object IDs.

See Also

boundingbox

Examples

# find the bounding box of all the neurons in a list
boundingbox(kcs20)
boundingbox(kcs20[1:3])

# plot those neurons
plot3d(kcs20)
# ... with their bounding box
plot3d(boundingbox(kcs20))
plot3d(kcs20)
# plot bounding box (in matching colours) for each neuron
# NB makes use of nlapply/neuronlist in slightly unsual context -
# plot3d.neuronlist can cope with lists containing anything with
# a valid plot3d method.
plot3d(nlapply(kcs20,boundingbox))

---

plot3d.cmtkreg  
Plot the domain of a CMTK registration

Description

Plot the domain of a CMTK registration

Usage

## S3 method for class 'cmtkreg'
plot3d(x, ...)

Arguments

x  A cmtk registration (the path to the registration folder on disk) or the resulting
of reading one in with read.cmtkreg.

...  Additional arguments passed to plot3d

See Also

cmtkreg, read.cmtkreg, plot3d

Examples

## Not run:
testdatadir=system.file("tests/testthat/testdata/cmtk", package="nat")
regpath=file.path(testdatadir, 'FCWB_JFRC2_01_warp_level-01.list/')
plot3d(cmtkreg(regpath))

# or read registration into memory if you want to work with it
reg=read.cmtkreg(regpath)
# nb calling plot3d.cmtkreg directly (rather than using the generic plot3d)
# is considered bad style but read.cmtkreg returns a plain list
# so method dispatch will fail
nat:::plot3d.cmtkreg(reg)

## End(Not run)
3D plots of dotprops objects using rgl package

Usage

## S3 method for class 'dotprops'
plot3d(
  x,
  scalevecs = 1,
  alpharange = NULL,
  color = "black",
  PlotPoints = FALSE,
  PlotVectors = TRUE,
  UseAlpha = FALSE,
  ...
)

Arguments

- **x**: A dotprops object
- **scalevecs**: Factor by which to scale unit vectors (numeric, default: 1.0)
- **alpharange**: Restrict plotting to points with alpha values in this range to plot (default: null => all points). See dotprops for definition of alpha.
- **color**: Character or numeric vector specifying colours for points/vectors. See details.
- **PlotPoints**, **PlotVectors**: Whether to plot points and/or tangent vectors (logical, default: tangent vectors only)
- **UseAlpha**: Whether to scale tangent vector length by the value of alpha
- **...**: Additional arguments passed to points3d and/or segments3d

Details

Tangent vectors are plotted by segments3d and centered on the relevant point. Points are plotted by points3d.

color will be recycled by points3d and segments3d. However in the special case that color has length equal to the number of points in x, then it will be duplicated before being passed to segments3d so that the result is that each vector is coloured uniformly according to color (since segments3d expects 2 colours for each line segment, blending them if they are different).

Value

invisible list of results of rgl plotting commands
plot3d.hxsurf

Plot amira surface objects in 3D using rgl

Description

Plot amira surface objects in 3D using rgl

Usage

## S3 method for class 'hxsurf'
plot3d(x, materials = NULL, col = NULL, ...)

Arguments

x An hxsurf surface object

materials Character vector or regex naming materials to plot (defaults to all materials in x). See subset.hxsurf.

col Character vector specifying colors for the materials, or a function that will be called with the number of materials to plot. When NULL (default) will use material colours defined in Amira (if available), or rainbow otherwise.

... Additional arguments passed to

See Also

read.hxsurf

Other hxsurf: as.hxsurf(), as.mesh3d(), materials(), read.hxsurf(), subset.hxsurf(), write.hxsurf()
Examples

plot3d(kcs20)
plot3d(MBL.surf)

# plot only vertical lobe
clear3d()
plot3d(MBL.surf, materials="VL", alpha=0.3)

# everything except vertical lobe
clear3d()
plot3d(MBL.surf, alpha=0.3,
       materials=grep("VL", MBL.surf$RegionList, value = TRUE, invert = TRUE))

plot3d.neuron

Plot neurons in 3D using rgl library

Description

Plot neurons in 3D using rgl library

Usage

## S3 method for class 'neuron'
plot3d(
  x,
  WithLine = TRUE,
  NeuronNames = FALSE,
  WithNodes = TRUE,
  WithAllPoints = FALSE,
  WithText = FALSE,
  PlotSubTrees = TRUE,
  add = TRUE,
  col = NULL,
  soma = FALSE,
  ...
)

Arguments

x A neuron to plot

WithLine Whether to plot lines for all segments in neuron

NeuronNames Logical indicating whether to label the neuron in the plot using the NeuronName field or a character vector of names.

WithNodes Whether to plot dots for branch and end points

WithAllPoints Whether to plot dots for all points in the neuron
WithText  Whether to label plotted points with their numeric id (see details)
PlotSubTrees  Whether to plot all sub trees when the neuron is not fully connected.
add  Whether to add the neuron to existing rgl plot rather than clearing the scene (default TRUE)
col  Colour specification (see rgl materials)
soma  Whether to plot a sphere at neuron’s origin representing the soma. Either a logical value or a numeric indicating the radius (default FALSE). When soma=TRUE the radius is hard coded to 2.
...  Additional arguments passed to rgl::lines3d

Details

Note that when WithText=TRUE, the numeric identifiers plotted are raw indices into the x$d array of the neuron, not the values of the PointNo column.

Value

list of rgl plotting ids (invisibly) separated into lines, points, texts according to plot element. See rgl::plot3d for details.

See Also

plot3d.neuronlist, plot3d.dotprops, nat::plot3d, rgl::plot3d

Examples

# A new plot would have been opened if required
open3d()
plot3d(Cell07PNs[[1]], col='red')
plot3d(Cell07PNs[[2]], col='green')

# clear the current plot
clear3d()
plot3d(Cell07PNs[[2]], col='blue', add=FALSE)
# plot the number of all nodes
clear3d()
plot3d(Cell07PNs[[2]], col='red', WithText=TRUE, add=FALSE)
# include cell bodies
plot3d(Cell07PNs[3:4], col='red', soma=TRUE)
plot3d(Cell07PNs[5], col='red', soma=3)
rgl.close()
plot3d.neuronlist  3D plots of the elements in a neuronlist, optionally using a subset expression

Description

plot3d.character is a convenience method intended for exploratory work on the command line.

Usage

## S3 method for class 'neuronlist'
plot3d(
  x,
  subset = NULL,
  col = NULL,
  colpal = rainbow,
  skipRedraw = ifelse(interactive(), 200L, TRUE),
  WithNodes = FALSE,
  soma = FALSE,
  ..., 
  SUBSTITUTE = TRUE
)

## S3 method for class 'character'
plot3d(x, db = NULL, ...)

Arguments

x  a neuron list or, for plot3d.character, a character vector of neuron names. The default neuronlist used by plot3d.character can be set by using options(nat.default.neuronlist=...). See ?nat for details. nat-package.

subset  Expression evaluating to logical mask for neurons. See details.

col  An expression specifying a colour evaluated in the context of the dataframe attached to nl (after any subsetting). See details.

colpal  A vector of colours or a function that generates colours

skipRedraw  When plotting more than this many (default 200) neurons skip redraw for individual neurons (this is much faster for large number of neurons). Can also accept logical values TRUE (always skip) FALSE (never skip).

WithNodes  Whether to plot points for end/branch points. Default: FALSE.

soma  Whether to plot a sphere at neuron’s origin representing the soma. Either a logical value or a numeric indicating the radius (default FALSE). When soma=TRUE the radius is hard coded to 2.

...  options passed on to plot3d (such as colours, line width etc)

SUBSTITUTE  Whether to substitute the expressions passed as arguments subset and col. Default: TRUE. For expert use only, when calling from another function.
db A neuronlist to use as the source of objects to plot. If NULL, the default, will use the neuronlist specified by `options('nat.default.neuronlist')`

**Details**

The col and subset parameters are evaluated in the context of the dataframe attribute of the neuronlist. If col evaluates to a factor and colpal is a named vector then colours will be assigned by matching factor levels against the named elements of colpal. If there is one unnamed level, this will be used as catch-all default value (see examples).

If col evaluates to a factor and colpal is a function then it will be used to generate colours with the same number of levels as are used in col.

WithNodes is FALSE by default when using `plot3d.neuronlist` but remains TRUE by default when plotting single neurons with `plot3d.neuron`. This is because the nodes quickly make plots with multiple neurons rather busy.

When soma is TRUE or a vector of numeric values (recycled as appropriate), the values are used to plot cell bodies. For neurons the values are passed to `plot3d.neuron` for neurons. In contrast dotprops objects still need special handling. There must be columns called X, Y, Z in the data.frame attached to x, that are then used directly by code in `plot3d.neuronlist`.

Whenever `plot3d.neuronlist` is called, it will add an entry to an environment `.plot3d` in `nat` that stores the ids of all the plotted shapes (neurons, cell bodies) so that they can then be removed by a call to `npop3d`.

`plot3d.character` will check if `options('nat.default.neuronlist')` has been set and then use x as an identifier to find a neuron in that neuronlist.

**Value**

list of values of `plot3d` with subsetted dataframe as attribute 'df'

**See Also**

`nat-package`

**Examples**

```r
open3d()
plot3d(kcs20,type=='gamma',col='green')
clear3d()
plot3d(kcs20,col=type)
plot3d(Cell07PNs,Glomerulus=='DA1',col='red')
plot3d(Cell07PNs,Glomerulus=='VA1d',col='green')
# Note use of default colour for non DA1 neurons
plot3d(Cell07PNs,col=Glomerulus, colpal=c(DA1='red', 'grey'))
# a subset expression
plot3d(Cell07PNs,Glomerulus%in%c('DA1','VA1d'),
      col=c("red","green")[factor(Glomerulus)])
# the same but not specifying colours explicitly
plot3d(Cell07PNs,Glomerulus%in%c('DA1','VA1d'),col=Glomerulus)
```
## Not run:
## more complex colouring strategies for a larger neuron set
# see https://github.com/jefferis/frulhns for details
library(frulhns)
# notice the sexually dimorphic projection patterns for these neurons
plot3d(jkn,cluster=='aSP-f' & shortGenotype=='JK1029',
   col=sex,colpal=c(male='green',female='magenta'))

## colour neurons of a class by input resistance
jkn.aspg=subset(jkn, cluster=='aSP-g')
# NB this comes in as a factor
Ri=with(jkn.aspg,as.numeric(as.character(Ri..GOhm.)))
# the matlab jet palette
jet.colors=colorRampPalette(c(navy,cyan,yellow,red))
plot3d(jkn.aspg,col=cut(Ri,20),colpal=jet.colors)

## End(Not run)

pointsinside

---

**pointsinside**

*Find which points of an object are inside a surface*

**Description**

Find which points of an object are inside a surface

**Usage**

```r
pointsinside(x, surf, ...)
```

## Default S3 method:

```r
pointsinside(x, surf, ..., rval = c("logical", "distance", "mesh3d"))
```

**Arguments**

- `x` an object with 3D points.
- `surf` The reference surface - either a `mesh3d` object or any object that can be converted using `as.mesh3d` including `hxsurf` and `asShape3d` objects.
- `...` additional arguments for methods, eventually passed to `as.mesh3d`.
- `rval` what to return.

**Details**

Note that `hxsurf` surface objects will be converted to `mesh3d` before being passed to `Rvcg::vcgCloseKD`, so if you are testing repeatedly against the same surface, it may make sense to pre-convert.

`pointsinside` depends on the face normals for each face pointing out of the object (see example). The face normals are defined by the order of the three vertices making up a triangular face. You can flip the face normal for a face by permuting the vertices (i.e. 1,2,3 -> 1,3,2). If you find for a
given surface that points are outside when you expect them to be inside then the face normals are probably all the wrong way round. You can invert them yourself or use the Morpho::invertFaces function to fix this.

If you find that some points but not all points are not behaving as you would expect, then it may be that some faces are not coherently oriented. The Rvcg::vcgClean function can sometimes be used to correct the orientation of the faces. Fixing more problematic cases may be possible by generating a new surface using alphashape3d::ashape3d (see examples).

Value

A vector of logical values or distances (positive inside, negative outside) equal to the number of points in x or the mesh3d object returned by Rvcg::vcgCloseKD.

Examples

```r
# check if the vertices in these neurons are inside the mushroom body calyx
# surface object
inout = pointsInside(kcs20, surf = subset(MBL.surf, "MB_CA_L"))
table(inout)

# be a bit more lenient and include points less than 5 microns from surface
MBCAL = subset(MBL.surf, "MB_CA_L")
inout5 = pointsInside(kcs20, surf = MBCAL, rval = 'distance') > -5
table(inout5)

# show which points are in or out
# Hmm seems like there are a few red points in the vertical lobe
# that are well outside the calyx
points3d(xyzmatrix(kcs20), col = ifelse(inout5, 'red', 'black'))
plot3d(MBL.surf, alpha = .3)

# Let's try to make an alphashape for the mesh to clean it up
library(alphashape3d)
MBCAL.as = ashape3d(xyzmatrix(MBCAL), alpha = 10)
# Plotting the points, we can see that is much better behaved
points3d(xyzmatrix(kcs20),
  col = ifelse(pointsInside(kcs20, MBCAL.as), 'red', 'black'))
```

```r
## Not run:
# Show the face normals for a surface
if(require('Morpho')) {
  # convert to a mesh3d object used by rgl and Morpho package
  MBCAL.mesh = as.mesh3d(subset(MBL.surf, "MB_CA_L"))
  fn = facenormals(MBCAL.mesh)
  wire3d(MBCAL.mesh)
  # show that the normals point out of the object
  plotNormals(fn, long = 5, col = 'red')
}

# invert the faces of the mesh and show that normals point in
MBCAL.inv = invertFaces(MBCAL.mesh)
```
potential_synapses

Calculate number of potential synapses between two neurons

Description
This implements the method of Stepanyants and Chklovskii

Usage
potential_synapses(a, b, s, ...)

## S3 method for class 'neuronlist'
potential_synapses(a, b, s, ...)

## S3 method for class 'neuron'
potential_synapses(
  a,
  b,
  s,
  sigma = s,
  bounds,
  method = c("direct", "approx"),
  ...
)

## S3 method for class 'dotprops'
potential_synapses(
  a,
  b,
  s,
  sigma = s,
  seglength = 1,
  bounds = NULL,
  method = c("direct", "approx"),
  ...
)

Arguments

a, b  neurons or neuronlists
s  the approach distance to consider a potential synapse
...  Additional arguments passed to methods (see details)
projection

sigma  the smoothing parameter in the approximate method (see details)
bounds  Optional bounding box to restrict comparison
method  Whether to use the direct or approximate method (see details)
seglength  how long to consider each distance between points.

details

Note that potential_synapses.neuronlist uses nlapply to process its first argument (a). This enables progress bars, robustness to errors and simple parallel execution. See the nlapply examples for further details of these arguments in action.

For this reason if you have two neuronlists of unequal sizes, it is recommended to put the larger one in argument a.

references


see also

Other neuron: neuron(), ngraph(), plot.neuron(), prune(), resample(), rootpoints(), spine(), subset.neuron()

examples

potential_synapses(Cell07PNs[1], Cell07PNs[1:3], s=2)
## Not run:
# if you have many neurons to calculate you should get a progress bar
potential_synapses(Cell07PNs[1:10], Cell07PNs[11:20], s=2)

# you can also use parallel execution, here over 7 cores
# doMC::registerDoMC(7)
potential_synapses(Cell07PNs[1:10], Cell07PNs[11:20], s=2, .parallel=TRUE)

## End(Not run)

projection  Make 2D (orthogonal) projection of 3D image data

description

Make 2D (orthogonal) projection of 3D image data
projection

Usage

projection(
  a,
  projdim = "z",
  projfun = c("integrate", "mean", "sum"),
  na.rm = T,
  mask = NULL,
  ...
)

Arguments

a Array of image data (im3d format)
projdim The image dimension down which to project
projfun The function that collapses each vector of image data down to a single pixel. Can be a character vector naming a function or a function. See details.
na.rm Logical indicating whether to ignore NA values in the image data when calculating function results. default: TRUE
mask A mask with the same extent as the image.
... Additional arguments for projfun

Details

Note that projfun must have an argument na.rm like the S3 Summary groupGeneric functions such as sum, min etc.

Note also that the BoundingBox of a 2d projection is not well-defined for the axis along which the projection was made. Presently both the evaluation location and the BoundingBox extremes are set to 0 after a projection is made but FIXME this is not completely satisfactory. Perhaps defining this to be NA or the midpoint of the orginal axis would be better justified.

See Also

groupGeneric, clampmax

Other im3d: as.im3d(), boundingbox(), im3d-coords, im3d-io, im3d(), imexpand.grid(), imslice(), is.im3d(), mask(), origin(), threshold(), unmask(), voxdims()

Examples

## Not run:
LHMask=read.im3d(system.file("tests/testthat/testdata/nrrd/LHMask.nrrd",package='nat'))
d=unmask(rnorm(sum(LHMask),mean=5,sd=5),LHMask)
op=par(mfrow=c(1,2))
rval=image(projection(d,projfun=max))
image(projection(d,projfun=clampmax(0,10)),zlim=rval$zlim)
par(op)

## End(Not run)
## Not run:
LHMask=read.im3d(system.file('tests/testthat/testdata/nrrd/LHMask.nrrd',package='nat'))
image(projection(LHMask),asp=TRUE)
## End(Not run)

---

prune

### prune

**prune**

**prune an object by removing points near (or far) from a target object**

**Description**

prune an object by removing points near (or far) from a target object

**Usage**

prune(x, target, ...)

## S3 method for class 'neuron'
prune(x, target, ...)

## S3 method for class 'dotprops'
prune(x, target, ...)

## S3 method for class 'neuronlist'
prune(x, target, ...)

## Default S3 method:
prune(x, target, maxdist, keep = c("near", "far"), return.indices = FALSE, ...)

**Arguments**

- **x**  
The object to prune. (e.g. dotprops object, see details)
- **target**  
Another object with 3D points that will determine which points in x are kept.
- **...**  
Additional arguments for methods (eventually passed to prune.default)
- **maxdist**  
The threshold distance for keeping points
- **keep**  
Whether to keep points in x that are near or far from the target
- **return.indices**  
Whether to return the indices that pass the test rather than the 3D object/points  
(default FALSE)

**Details**

prune.neuron depends on a more basic function prune_vertices and is also related to subset.neuron.
prune_strahler

Prune a neuron by removing segments with a given Strahler order

Description

Prune a neuron by removing segments with a given Strahler order

Usage

prune_strahler(x, orderstoprune = 1:2, ...)

Arguments

x A neuron
orderstoprune Integer indices of which Strahler orders to prune - defaults to the lowest two orders (1:2)
... Additional arguments passed to as.neuron.data.frame

See Also

prune_strahler, spine, prune_vertices
subset.neuron
subset.dotprops

Other neuron: neuron(), ngraph(), plot.neuron(), potential_synapses(), resample(), rootpoints(), spine(), subset.neuron()

Examples

## prune single neurons
plot3d(kcs20[[1]], col='blue')
plot3d(kcs20[[2]], col='red')

# prune neuron 2 down to points that are close to neuron 1
neuron2_close=prune(kcs20[[2]], target=kcs20[[1]], maxdist=10)
plot3d(neuron2_close, col='cyan', lwd=3)

neuron2_far=prune(kcs20[[2]], target=kcs20[[1]], maxdist=10, keep='far')
plot3d(neuron2_far, col='magenta', lwd=3)

## Prune a neuron with a neuronlist
pruned=prune(kcs20[[11]], kcs20[setdiff(1:20, 11)], maxdist=8)
plot3d(pruned, col='red', lwd=3)
plot3d(kcs20[[11]], col='green', lwd=3)
plot3d(kcs20, col='grey')
prune_vertices

Value

The pruned neuron

See Also

strahler_order, spine, for finding the longest path in a neuron, pruned for subsetting dotprops style neurons by spatial proximity, as.neuron.data.frame, which is used to generate the new neuron.

Examples

```r
x=Cell07PNs[[1]]
pruned12=prune_strahler(x)
pruned1=prune_strahler(x, 1)
plot(x)
plot(pruned1, lwd=3, col='blue', add=TRUE)
plot(pruned12, lwd=3, col='red', add=TRUE)
```

prune_vertices

Prune selected vertices or edges from a neuron

Description

prune_vertices removes vertices from a neuron

prune_edges removes edges (and any unreferenced vertices)

Usage

```r
prune_vertices(x, verticestoprune, invert = FALSE, ...)
```

```r
prune_edges(x, edges, invert = FALSE, ...)
```

Arguments

- **x**  
  A neuron to prune. This can be any object that can be converted by as.ngraph — see details.

- **verticestoprune**  
  An integer vector describing which vertices to remove.

- **invert**  
  Whether to keep vertices rather than dropping them (default FALSE).

- **edges**  
  The edges to remove. One of i) an Nx2 matrix, each row specifying a single edge defined by its raw edge id, ii) an integer vector defining a path of raw vertex ids or iii) an igraph.es edge sequence — see details and the P and path arguments of igraph::E.
**prune_vertices**

**Details**

These are relatively low-level functions and you will probably want to use `subset.neuron` or `prune.neuron` and friends in many cases.

Note that `prune_vertices` and `prune_edges` both use raw vertex ids to specify the vertices/edges to be removed. If you want to use the id in the PointNo field, then you must translate yourself (see examples).

Both `prune_vertices` and `prune_edges` first convert their input `x` to the ngraph representation of the neuron before removing points. The input `x` can therefore be in any form compatible with `as.ngraph` including an existing ngraph. There is an additional requirement that the input must be compatible with `xyzmatrix` if `invert=TRUE`.

Note that the edges argument of `prune_edges` must specify a path traversing a set of vertices in a valid order. However if the input is a matrix or vector the direction of each individual edge in this path is ignored. So if your neuron has edges 2->1 2->3 3->4 then an edge sequence 1:3 would successfully delete 2 edges.

**Value**

A pruned neuron

**See Also**

`as.neuron.ngraph`, `subset.neuron`, `prune.neuron`

**Examples**

```r
n = prune_vertices(Cell07PNs[[1]], 1:25)  # original neuron
plot(Cell07PNs[[1]])                        # with pruned neuron superimposed
plot(n, col='green', lwd=3, add=TRUE)

# use the PointNo field (= the original id from an SWC file)

n2 = prune_vertices(n, match(26:30, n$d$PointNo))
y = prune_edges(Cell07PNs[[1]], edges=1:25)

# remove the spine of a neuron
spine_ids = spine(Cell07PNs[[1]], rval='ids')
pruned = prune_edges(Cell07PNs[[1]], spine_ids)

# NB this is subtly different from this, which removes vertices along the
# spine *even* if they are part of an edge that is outside the spine.
pruned2 = prune_vertices(Cell07PNs[[1]], spine_ids)
```
**Description**

Read AmiraMesh data in binary or ascii format

Read the header of an amiramesh file

**Usage**

```r
read.amiramesh(file,
sections = NULL,
header = FALSE,
simplify = TRUE,
endian = NULL,
ReadByteAsRaw = FALSE,
Verbose = FALSE)

read.amiramesh.header(file, Parse = TRUE, Verbose = FALSE)
```

**Arguments**

- `file` Name of file (or connection) to read
- `sections` character vector containing names of sections
- `header` Whether to include the full unprocessed text header as an attribute of the returned list.
- `simplify` If there is only one datablock in file do not return wrapped in a list (default: TRUE).
- `endian` Whether multibyte data types should be treated as big or little endian. Default of NULL checks file or uses `.Platform$endian`
- `ReadByteAsRaw` Logical specifying whether to read 8 bit data as an R `raw` vector rather than integer vector (default: FALSE).
- `Verbose` Print status messages
- `Parse` Logical indicating whether to parse header (default: TRUE)

**Details**

Reading byte data as raw arrays requires 1/4 memory but complicates arithmetic.

`read.amiramesh.header` will open a connection if file is a character vector and close it when finished reading.
Value

list of named data chunks

See Also

readBin, .Platform

Other amira: amiratype(), is.amiramesh(), read.hxsurf(), write.hxsurf()

---

read.cmtk

Read CMTK TypedStream file to a list in memory

Description

This function is primarily of developer interest. End users will typically want to use more specialised functions for reading registrations and landmarks.

Usage

read.cmtk(con, CheckLabel = TRUE)

Arguments

con Path to (optionally gzipped) file or (open) connection.
CheckLabel Check, fix and warn for invalid or duplicate labels (default TRUE)

Details

This is the default format used by CMTK for registration, studylist, landmarks and image files. Although this is largely a generic function, there is special handling of the coefficients and active members of the spline warp component of a CMTK nonrigid registrartion.

Note that if an open connection is passed to read.cmtk the version number of the CMTK TypedStream will not be checked or recorded.

See Also

Other cmtk-io: cmtk.extract_affine(), read.cmtkreg(), write.cmtkreg(), write.cmtk()
read.cmtkreg  
Read a CMTK format registration

Description
Read a CMTK format registration

Usage
read.cmtkreg(filename, ReturnRegistrationOnly = FALSE, ...)

Arguments
filename  Path to a CMTK registration file
ReturnRegistrationOnly  When FALSE (default) will not attempt to extract the registration element from the registration file.
...  Additional arguments passed to read.cmtk

See Also
Other cmtk-io: cmtk.extract_affine(), read.cmtk(), write.cmtkreg(), write.cmtk()

read.hxsurf  
Read Amira surface (aka HxSurface or HyperSurface) files into hxsurf object

Description
Read Amira surface (aka HxSurface or HyperSurface) files into hxsurf object

Usage
read.hxsurf(
  filename,
  RegionNames = NULL,
  RegionChoice = "both",
  FallbackRegionCol = "grey",
  Verbose = FALSE
)
Arguments

filename Character vector defining path to file
RegionNames Character vector specifying which regions should be read from file. Default value of NULL => all regions.
RegionChoice Whether the Inner or Outer material, or both (default), should define the material of the patch. See details.
FallbackRegionCol Colour to set regions when no colour is defined
Verbose Print status messages during parsing when TRUE

Details

Note that when RegionChoice="both" or RegionChoice=c("Inner", "Outer") both polygons in inner and outer regions will be added to named regions. To understand the significance of this, consider two adjacent regions, A and B, with a shared surface. For the polygons in both A and B, Amira will have a patch with (say) InnerRegion A and OuterRegion B. This avoids duplication in the file. However, it might be convenient to add these polygons to both regions when we read them into R, so that regions A and B in our R object are both closed surfaces. To achieve this when RegionChoice="both", read.hxsurf adds these polygons to region B (as well as region A) but swaps the order of the vertices defining the polygon to ensure that the surface directionality is correct.

As a rule of thumb, stick with RegionChoice="both". If you get more regions than you wanted, then try switching to RegionChoice="Inner" or RegionChoice="Outer".

Value

A list with S3 class hxsurf with elements

- Vertices A data.frame with columns X, Y, Z, PointNo
- Regions A list with 3 column data.frames specifying triplets of vertices for each region (with reference to PointNo column in Vertices element)
- RegionList Character vector of region names (should match names of Regions element)
- RegionColourList Character vector specifying default colour to plot each region in R’s rgb format

See Also

plot3d.hxsurf, rgb

Other amira: amiratyp(), is.amiramesh(), read.amiramesh(), write.hxsurf()

Other hxsurf: as.hxsurf(), as.mesh3d(), materials(), plot3d.hxsurf(), subset.hxsurf(), write.hxsurf()
Examples

```r
## Not run:
read.hxsurf("my.surf", RegionChoice="both")

## End(Not run)
```

### read.landmarks

**Generic functions to read/write landmarks in any supported format**

**Description**

Generic functions to read/write landmarks in any supported format

**Usage**

```r
read.landmarks(f, ...)

write.landmarks(
  x,  
  file,  
  format = "amiralandmarks",  
  ext = NULL,  
  Force = FALSE,  
  MakeDir = TRUE,  
  ...)  
)
```

**Arguments**

- `f` Path to a file (can also be a URL)
- `...` Additional arguments passed on to format specific functions
- `x` The landmarks object to write. Can also be a plain matrix or data.frame.
- `file` The path to the output file. If this does not end in an extension like .landmarksAscii, then one will be added based on the value of the `ext` argument.
- `format` Character vector specifying output format. Defaults to "amiralandmarks". Partial matching is used (e.g. amira is sufficient).
- `ext` Optional character vector specifying a new or non-standard extension to use for output file, including the period (e.g. ext='.am'). When ext=NULL, the default, the default extension for the selected `format` will be added if `f` does not have an extension. When ext=NA, the extension will not be modified and no extension will be appended if `f` does not have one.
- `Force` Whether to overwrite an existing file
- `MakeDir` Whether to create directory implied by `file` argument.
Details

Presently the supported formats are

- Amira
- CMTK
- Fiji (see https://imagej.net/plugins/name-landmarks-and-register)

See examples section for how to produce a listing of all currently available formats with fileformats.

Value

for read.landmarks a matrix or list of additional class landmarks, where the rownames specify the names of each landmark if available.

For write.landmarks the path to the written file, invisibly.

Paired landmarks

Only the amiralandmarks format supports the use of paired landmarks

See Also

fileformats

Examples

```r
## Listing of supported fileformats for landmarks
fileformats(class = 'landmarks', rval = "info")

## round trip tests
m=matrix(rnorm(6), ncol=3)
rownames(m)=c("nose", "ear")
f=write.landmarks(m, file='knee', format='cmtk')
read.landmarks(f)

# write in amira format which does not support named landmarks
f2=write.landmarks(m, file='knee', format='amira')
read.landmarks(f2)

# clean up
unlink(c(f,f2))
```
read.morphml  

Return parsed XML or R list versions of a NeuroML file

Description

read.morphml is designed to expose the full details of the morphology information in a NeuroML file either as a parsed XML structure processed by the XML package or as an extensively processed R list object. To obtain a neuron object use read.neuron.neuroml.

Usage

read.morphml(f, ..., ReturnXML = FALSE)

Arguments

- **f**: Path to a file on disk or a remote URL (see xmlParse for details).
- **...**: Additional arguments passed to xmlParse
- **ReturnXML**: Whether to return a parsed XML tree (when ReturnXML=TRUE) or a more extensively processed R list object when ReturnXML=FALSE, the default.

Details

NeuroML files consist of an XML tree containing one more or more cells. Each cell contains a tree of segments defining the basic connectivity/position and an optional tree cables defining attributes on groups of segments (e.g. a name, whether they are axon/dendrite/soma etc).

read.morphml will either provide the parsed XML tree which you can query using XPath statements or a more heavily processed version which provides as much information as possible from the segments and cables trees in two R data.frames. The latter option will inevitably drop some information, but will probably be more convenient for most purposes.

Value

Either an R list of S3 class containing one morphml_cell object for every cell in the NeuroML document or an object of class XMLDocument when ReturnXML=TRUE.

References

https://neuroml.org/specifications

See Also

link[XML]{xmlParse}, read.neuron.neuroml
Description
Read a single neuron from a file

Usage
read.neuron(f, format = NULL, class = c("neuron", "ngraph"), ...)

Arguments
f          Path to file. This can be a URL, in which case the file is downloaded to a temporary location before reading.
format     The file format of the neuron. When format=NULL, the default, read.neuron will infer the file format from the extension or file header (aka magic) using the fileformats registry.
class      The class of the returned object - presently either "neuron" or "ngraph"
...        additional arguments passed to format-specific readers

Details
This function will handle neuron and dotprops objects saved in R .rds or .rda format by default. Additional file formats can be registered using fileformats.

At the moment the following formats are supported using file readers already included with the nat package:

- **swc** See read.neuron.swc. SWC files can also return an ngraph object containing the neuron structure in a (permissive) general graph format that also contains the 3D positions for each vertex.
- **neuroml** See read.neuron.neuroml
- **fijitraces** See read.neuron.fiji. The file format used by the Simple Neurite Tracer plugin of Fiji/ImageJ.
- **hxlineset,hxskel** Two distinct fileformats used by Amira. hxlineset is the generic one, hxskel is used by the hxskeletonize extension of Schmitt and Evers (see refs).
- **rda,rds** Native R cross-platform binary formats (see load, readRDS). Note that RDS only contains a single unnamed neuron, whereas rda contains one or more named neurons.

References
See Also

write.neuron, read.neurons, fileformats

Examples

## Not run:
# note that we override the default NeuronName field
n = read.neuron(system.file("tests/testthat/testdata","neuron","EBT7R.CNG.swc",package='nat'),
   NeuronName="EBT7R")
# use a function to set the NeuronName field
n3 = read.neuron(system.file("tests/testthat/testdata","neuron","EBT7R.CNG.swc",package='nat'),
   NeuronName=function(x) sub("\..*","",x))
# show the currently registered file formats that we can read
fileformats(class='neuron', read=TRUE)

## End(Not run)

---

read.neuron.fiji  Read a neuron saved by Fiji's Simple Neurite Tracer Plugin

Description

Read a neuron saved by Fiji's Simple Neurite Tracer Plugin

Usage

read.neuron.fiji(f, ..., simplify = TRUE, Verbose = FALSE)

Arguments

- **f**: Path to a file
- **...**: Additional arguments passed to xmlParse.
- **simplify**: Whether to return a single neuron as a neuron object rather than a neuronlist of length 1.
- **Verbose**: Whether to print status messages during parsing.

Details

This is an XML based format so parsing it depends on installation of the suggested XML package.

References

**read.neuron.neuroml**  
*Read one or more neurons from a NeuroML v1 file*

**Description**

Read one or more neurons from a NeuroML v1 file

**Usage**

```
read.neuron.neuroml(f, ..., AlwaysReturnNeuronList = FALSE)
```

**Arguments**

- `f`  
  Path to a NeuroML format XML file
- `...`  
  Additional arguments passed to `read.morphml` (and on to `xmlParse`)
- `AlwaysReturnNeuronList`  
  See Value section (default FALSE)

**Value**

When the XML file contains only 1 cell and `AlwaysReturnNeuronList=FALSE`, a `neuron` object, otherwise a `neuronlist` containing one or more neurons.

**References**

https://neuroml.org/specifications

**See Also**

- `read.morphml`

---

**read.neuron.swc**  
*Read a neuron in swc file format*

**Description**

`read.neuron.swc` reads an SWC file on disk into a fully parsed `neuron` representation.  
`read.ngraph.swc` reads an SWC file on disk into the more generic (and forgiving) `ngraph` representation which provides a bridge to the `igraph` library.

**Usage**

```
read.neuron.swc(f, ...)
read.ngraph.swc(f, weights = FALSE, directed = TRUE, ...)
```
Arguments

- `f` path to file
- `...` Additional arguments. `read.neuron.swc` passes these to `as.neuron` and then on to `neuron`. `read.neuron.swc` passes them to `ngraph`.
- `weights` Logical value indicating whether edge weights defined by the 3D distance between points should be added to graph (default `FALSE`) or a numeric vector of weights.
- `directed` Whether the resultant graph should be directed (default `TRUE`)

Details

These functions will accept SWC neurons with multiple trees and arbitrary point index order. However only `read.ngraph.swc` will accept SWC files with cycles.

These functions would normally be called from `read.neuron(s)` rather than used directly.

SWC Format

According to [http://www.neuronland.org/NLMorphologyConverter/MorphologyFormats/SWC/Spec.html](http://www.neuronland.org/NLMorphologyConverter/MorphologyFormats/SWC/Spec.html) SWC file format has a radius not a diameter specification

See Also

- `is.swc`

---

**read.neuronlistfh**  
*Read a local, or remote, neuronlistfh object saved to a file.*

Description

Read a local, or remote, neuronlistfh object saved to a file.

Usage

```r
read.neuronlistfh(file, localdir = NULL, update = FALSE, ...)
```

Arguments

- `file` The file path of the neuronlistfh object. Can be local, or remote (via http or ftp).
- `localdir` If the file is to be fetched from a remote location, this is the folder in which downloaded RDS file will be saved. The default value of NULL will save to a folder in the current R sessions temporary folder. See details.
- `update` Whether to update local copy of neuronlistfh (default: `FALSE`, see details)
- `...` Extra arguments to pass to `download.file`. 
read.neurons

Details

When reading a remote neuronlistfh object, it is downloaded and cached to localdir. If there is already a cached file at the appropriate location and update=TRUE then the md5sums are checked and the downloaded file will be copied on top of the original copy if they are different; if update=FALSE, the default, then no action will be taken. After downloading a remote neuronlistfh object, a check is made for the existence of the data directory that will be used to individual objects. If this does not exist it will be created.

Note also that there is a strict convention for the layout of the files on disk. The neuronlistfh object will be saved in R’s RDS format and will be placed next to a folder called data which will contain the data objects, also saved in RDS format. For example if myneurons.rds is downloaded to localdir="\path\to\localdir" the resultant file layout will be as follows:

- \path\to\localdir\myneurons.rds
- \path\to\localdir\data\2f88e16c4f21bfc0b3a8288c05bd0
- \path\to\localdir\data\5b58e04ee35f3bcbc023fb7836c842e
- \path\to\localdir\data\... etc

Given this arrangement, the data directory should always be at a fixed location with respect to the saved neuronlistfh object and this is enforced on download and the default behaviour on read and write. However it does remain possible (if not recommended) to site the neuronlistfh and filehash database directory in different relative locations; if the neuronlistfh object specified by file does not have a filehash database with a valid dir slot and there is no ‘data’ directory adjacent to the neuronlistfh object, an error will result.

See Also

Other neuronlistfh: [.neuronlistfh(), neuronlistfh(), remotesync(), write.neuronlistfh()]

Description

Read one or more neurons from file to a neuronlist in memory

Usage

read.neurons(
  paths,
  pattern = NULL,
  neuronnames = basename,
  format = NULL,
  nl = NULL,
  df = NULL,
  OmitFailures = TRUE,
  SortOnUpdate = FALSE,
  ...
)


Arguments

paths  Paths to neuron input files or a directory containing neurons or a neuronlistfh object, or a zip archive containing multiple neurons.

pattern If paths is a directory, regex that file names must match.

neuronnames Character vector or function that specifies neuron names. See details.

format File format for neuron (see read.neuron)

nl An existing neuronlist to be updated (see details)

df Optional data frame containing information about each neuron

OmitFailures Omit failures (when TRUE) or leave an NA value in the list

SortOnUpdate When nl!=NULL the resultant neuronlist will be sorted so that neurons are ordered according to the value of the paths argument.

... Additional arguments to passed to read.neuron methods

Details

This function will cope with the same set of file formats offered by read.neuron.

If the paths argument specifies a (single) directory then all files in that directory will be read unless an optional regex pattern is also specified. Similarly, if paths specifies a zip archive, all neurons within the archive will be loaded.

neuronnames must specify a unique set of names that will be used as the names of the neurons in the resultant neuronlist. If neuronnames is a a function then this will be applied to the path to each neuron. The default value is the function basename which results in each neuron being named for the input file from which it was read.

The optional data frame (df) detailing each neuron should have rownames that match the names of each neuron. It would also make sense if the same key was present in a column of the data frame. If the data frame contains more rows than neurons, the superfluous rows are dropped with a warning. If the data frame is missing rows for some neurons an error is generated. If SortOnUpdate is TRUE then updating an existing neuronlist should result in a new neuronlist with ordering identical to reading all neurons from scratch.

Value

neuronlist object containing the neurons

See Also

read.neuron, write.neurons, fileformats

Other neuronlist: *.neuronlist(), is.neuronlist(), neuronlist-dataframe-methods, neuronlistfh(), neuronlist(), nlapply(), write.neurons()

Examples

## Not run:
## Read C. elegans neurons from OpenWorm github repository
vds=paste0("VD", 1:13)
vdurls=paste0("https://raw.githubusercontent.com/openworm/CElegansNeuroML/","103d500e066125688aa7ac5eac7e9b2bb4490561/CElegans/generatedNeuroML/",vds,".morph.xml")
vdnl=read.neurons(vdurls, neuronnames=vds)
plot3d(vdnl)

## The same, but this time add some metadata to neuronlist
# fetch table of worm neurons from wormbase
library(rvest)
nlurl="http://wormatlas.org/neurons/Individual%20Neurons/Neuronframeset.html"
wormneurons = html_table(html(nlurl), fill=TRUE)[4]
vddf=subset(wormneurons, Neuron%in%vds)
rownames(vddf)=vddf$Neuron
# attach metadata to neuronlist
vdnl=read.neurons(vdurls, neuronnames=vds, df=vdddf)
# use metadata to plot a subset of neurons
clear3d()
plot3d(vdnl, grepl("P[1-6].app", Lineage))

## End(Not run)

---

**read.nrrd**  
*Read nrrd file into an array in memory*

### Description

Read nrrd file into an array in memory  
Read the (text) header of a NRRD format file

### Usage

```r
read.nrrd(
  file,
  origin = NULL,
  ReadData = TRUE,
  AttachFullHeader = TRUE,
  Verbose = FALSE,
  ReadByteAsRaw = c("unsigned", "all", "none")
)
```

```
read.nrrd.header(file, Verbose = FALSE)
```

### Arguments

- **file**  
  Path to a nrrd (or a connection for `read.nrrd.header`)
- **origin**  
  Add a user specified origin (x,y,z) to the returned object
- **ReadData**  
  When FALSE just return attributes (i.e. the nrrd header)
read.vaa3draw

AttachFullHeader

Include the full nrrd header as an attribute of the returned object (default TRUE)

Verbose

Status messages while reading

ReadByteAsRaw

Either a character vector or a logical vector specifying when R should read 8 bit
data as an R raw vector rather than integer vector.

Details

read.nrrd reads data into a raw array. If you wish to generate a im3d object that includes spatial
calibration (but is limited to representing 3D data) then you should use read.im3d.

ReadByteAsRaw=unsigned (the default) only reads unsigned byte data as a raw array. This saves
quite a bit of space and still allows data to be used for logical indexing.

Value

An array object, optionally with attributes from the nrrd header.
A list with elements for the key nrrd header fields

See Also

write.nrrd, read.im3d

read.vaa3draw Read Vaa3d format image data

Description

Read Vaa3d format image data

Usage

read.vaa3draw(f, ReadData = TRUE, Verbose = FALSE, ReadByteAsRaw = FALSE)

Arguments

f Path to image to read
ReadData Whether to read in data or just parse header
Verbose Whether to print status messages
ReadByteAsRaw Can reduce memory footprint by reading 8 bit data as a raw rather than 4 byte
integers.
**Description**

A reglist is read as a set of transformations to be applied sequentially starting with the first element, then applying the second transformation to the result of the first and so on. Each individual transformation is considered to map data from the sample (floating/moving) space to the reference (fixed/template) space.

Each transformation may have an attribute "swap" indicating that the natural direction of the transformation should be swapped (i.e. inverted). This can be done trivially in the case of affine transformations, expensively for others such as CMTK registrations (see `cmtkreg`) and not at all for others. Note that the term ‘swap’ is used to avoid a direct equivalence with inversion - many registration tools use the term *inverse* for directions that one might naively think of as as the natural direction of the transformation (see `xformpoints.cmtkreg` for discussion).

`invert_reglist` inverts a reglist object

`c.reglist` combines multiple reglists into a single reglist.

**Usage**

```r
reglist(..., swap = NULL)
invert_reglist(x)
## S3 method for class 'reglist'
c(..., recursive = FALSE)
```

**Arguments**

- `...` One or more transformations/reglists to combine
- `swap` A vector of the same length as `...` indicating whether the direction of each transformation should be swapped (i.e. mapping reference -> sample).
- `x` A reglist object to invert
- `recursive` Presently ignored

**Details**

The swap argument is provided as a convenience, but an attribute 'swap' can also be set directly on each registration.

**Inversion**

`invert_reglist` takes a minimal approach to inversion. It reverses the order of the individual elements of the registration and tags each of them with a swap attribute (or changes the value of the attribute if it already exists).
See Also

xform
c

Examples

I=diag(4)
S=I
diag(S)=c(1, 2, 3, 1)
rl=reglist(S, I)
rli=invert_reglist(rl)

## We can check the inversion by simplifying
m=simplify_reglist(rl)[[1]]
mi=simplify_reglist(rl)[[1]]
# NB solve will invert a homogeneous affine matrix
all.equal(m, solve(mi))
I=diag(4)
S=I
diag(S)=c(1, 2, 3, 1)
rl=reglist(S, I)
rl2=c(rl, 'path/to/my/reg.list')
rl3=c(reglist('path/to/my/reg.list'), rl)

remotesync Synchronise a remote object

Description

Synchronise a remote object

Usage

remotesync(
  x,
  remote = attr(x, "remote"),
  download.missing = TRUE,
  delete.extra = FALSE,
  ...
)

## S3 method for class 'neuronlistfh'
remotesync(
  x,
  remote = attr(x, "remote"),
  download.missing = FALSE,
  delete.extra = FALSE,
  indices = NULL,
resample

update.object = TRUE,
...
)

Arguments

x 
Object to synchronise with a remote URL
remote 
The remote URL to update from
download.missing 
Whether to download missing objects (default TRUE)
delete.extra 
Whether to delete objects (default TRUE)
indices 
Character vector naming neurons to update (default indices=NULL implies all neurons).
update.object 
Whether to update the neuronlistfh object itself on disk (default TRUE). Note that this assumes that the neuronlistfh object has not been renamed after it was downloaded.

Value

The updated neuronlistfh object (invisibly)

See Also

Other neuronlistfh: [.neuronlistfh(), neuronlistfh(), read.neuronlistfh(), write.neuronlistfh()]

Examples

## Not run:
kcs20=read.neuronlistfh('http://flybrain.mrc-lmb.cam.ac.uk/si/nblast/flycircuit/kcs20.rds')
# update object from the web
kcs20=remotesync(kcs20)
# download all neurons with significant innervation of the vertical lobe
mbvl_neurons=subset(kcs20, (MB_VL_R+MB_VL_L)>200, rval='names')
kcs20=remotesync(kcs20, indices=mbvl_neurons, download.missing=TRUE)
## End(Not run)

resample

Resample an object with a new spacing

Description

Resample an object with a new spacing
resample a neuron with a new spacing
Usage

resample(x, ...)

## S3 method for class 'neuron'
resample(x, stepsize, ...)

Arguments

x An object to resample

... Additional arguments passed to methods

stepsize The new spacing along the tracing

Details

resample.neuron Floating point columns including X,Y,Z,W will be interpolated using linear interpolation, while integer or factor columns will be interpolated using constant interpolation. See approx for details.

See Also

approx, seglengths

Other neuron: neuron(), ngraph(), plot.neuron(), potential_synapses(), prune(), rootpoints(), spine(), subset.neuron()

---

rootpoints Return the root or branch points of a neuron or graph

Description

A neuron may have multiple subtrees and therefore multiple roots

Usage

rootpoints(x, ...)

## Default S3 method:
rootpoints(x, ...)

## S3 method for class 'neuron'
rootpoints(x, subtrees = 1, ...)

## S3 method for class 'igraph'
rootpoints(x, ...)

branchpoints(x, ...)
## Default S3 method:
branchpoints(x, ...)

## S3 method for class 'neuron'
branchpoints(x, subtrees = 1, ...)

## S3 method for class 'igraph'
branchpoints(x, ...)

epipoints(x, ...)

## S3 method for class 'neuron'
epipoints(x, subtrees = 1, ...)

## S3 method for class 'igraph'
epipoints(x, ...)

## Default S3 method:
epipoints(x, ...)

Arguments

- **x**: Neuron or other object which might have roots
- **...**: Further arguments passed to methods
- **subtrees**: Integer index of the fully connected subtree in `x$SubTrees`. Only applicable when a neuron consists of multiple unconnected subtrees.

Details

`branchpoints.neuron` returns a list if more than one subtree is specified

Value

Integer point number of root/branch point

See Also

Other neuron: `neuron()`, `ngraph()`, `plot.neuron()`, `potential_synapses()`, `prune()`, `resample()`, `spine()`, `subset.neuron()`

---

**scale.neuron**

*Scale and centre neuron 3D coordinates*

Description

note that `scale.dotprops` recalculates the tangent vectors after scaling the 3D coords. See `dotprops` for details.
Usage

## S3 method for class 'neuron'
scale(x, center = TRUE, scale = TRUE)

## S3 method for class 'dotprops'
scale(x, center = TRUE, scale = TRUE)

Arguments

x  
A neuron

center  
3-vector to subtract from x,y,z coords

scale  
3-vector used to divide x,y,z coords

Details

If scale=FALSE, the neuron will be rescaled to unit sd in each axis. If center=FALSE, the neuron will be centred around the axis means. See base::scale.default for additional details.

Value

neuron with scaled coordinates

See Also

scale.default, *.neuron

Examples

n1.scaledown=scale(Cell07PNs[[1]],scale=c(2,2,3))
n1.scaleup=scale(Cell07PNs[[1]],scale=1/c(2,2,3))

seglengths

Calculate length of all segments in neuron

Description

Calculate length of all segments in neuron

Usage

seglengths(x, all = FALSE, flatten = TRUE, sumsegment = TRUE)
seglist makes a seglist object from a list of integer vectors of raw vertex ids. As a convenience if a vector of numeric ids are passed these are assumed to specify a neuron with 1 segment.

as.seglist.neuron will extract the seglist from a neuron, optionally extracting all subtrees (all=TRUE) and (in this case) flattening the list into a single hierarchy when flatten=TRUE. n.b. when all=TRUE but flatten=FALSE the result will always be a list of seglist objects (even if the neuron has only one subtree i.e. is fully connected).

as.seglist.igraph will convert a fully connected acyclic ngraph or igraph object into a seglist consisting of exactly one subtree.
Usage

seglist(...)
as.seglist(x, ...)

## S3 method for class 'neuron'
as.seglist(x, all = FALSE, flatten = FALSE, ...)

## S3 method for class 'igraph'
as.seglist(x, origin = NULL, Verbose = FALSE, ...)

Arguments

... for seglist integer vectors to convert to a seglist
x object passed to be converted to seglist
all Whether to include segments from all subtrees
flatten When all=TRUE flatten the lists of lists into a one-level list.
origin The origin of the tree (see details)
Verbose Whether to print progress updates to console (default FALSE)

Details

see neuron for further information about seglists.

If the graph vertices have vid attributes, typically defining the original vertex ids of a graph that was then decomposed into subgraphs, then the origin is assumed to refer to one of these vids not a raw vertex id of the current graph. The returned seglist will also contain these original vertex ids.

The head of the first segment in the seglist will be the origin.

Value

A list with additional class seglist.
a list with one entry for each unbranched segment.

See Also

neuron
ngraph,igraph

Examples

sl=seglist(c(1:2),c(2:6))
Recalculate Neurons’s SWCData using SegList and point information

Description

Uses the SegList field (indices into point array) to recalculate point numbers and parent points for SWC data field (d).

Usage

seglist2swc(x, d, RecalculateParents = TRUE, ...)

Arguments

x  Neuron containing both the SegList and d fields or a plain seglist

 d  SWC data block (only expected if x is a SegList)

RecalculateParents  Whether to recalculate parent points (default T)

...  Additional arguments passed to normalise_swc

Details

If any columns are missing then they are set to default values by normalise_swc. In particular

- PointNo integer 1:npoints
- Label = 0 (unknown)
- W NA_real

Note that each numeric entry in the incoming SegList is a raw index into the block of vertex data defined by d.

Value

A neuron if x was a neuron otherwise dataframe of swc data

See Also

as.neuron.data.frame, normalise_swc, neuron
Return a simplified segment graph for a neuron

Usage

```r
segmentgraph(
  x,
  weights = TRUE,
  segids = FALSE,
  exclude.isolated = FALSE,
  include.xyz = FALSE,
  reverse.edges = FALSE
)
```

Arguments

- `x`: neuron
- `weights`: Whether to include the original segment lengths as edge weights in the graph.
- `segids`: Whether to include the integer segment ids as an edge attribute in the graph
- `exclude.isolated`: Whether to eliminate isolated nodes
- `include.xyz`: Whether to include 3D location as vertex attribute
- `reverse.edges`: Whether to reverse the direction of each edge in the output graph to point towards (rather than away from) the root (default FALSE)

Details

The resultant graph will contain all branch and endpoints of the original neuron. This will be constructed from the SegList field, or where present, the SubTrees field (containing multiple SegLists for each isolated graph in the neuron). Each edge in the output graph will match one segment in the original SegList.

Value

igraph object containing only nodes of neuron keeping original labels (`x$d$PointNo` => `V(g)$label`) and vertex indices (1:nrow(`x$d`) => `V(g)$vid`).

Examples

```r
sg = segmentgraph(Cell07PNs[[1]])
str(sg)
library(igraph)
plot(sg, edge.arrow.size=.4, vertex.size=10)
```
setdiff

Find the (asymmetric) difference between two collections of objects

Description

Find the (asymmetric) difference between two collections of objects

Usage

setdiff(x, y, ...)

## Default S3 method:
setdiff(x, y, ...)

## S3 method for class 'neuronlist'
setdiff(x, y, ...)

Arguments

x the first collection to consider.
y the second collection to consider.
... additional arguments passed to methods

Details

Note that setdiff.default calls base::setdiff to ensure consistent behaviour for regular vectors.

As a convenience setdiff.neuronlist allows y, the second collection, to be a character vector of names.

Value

A collection of the same mode as x that contains all elements of x that are not present in y.

See Also

setdiff
simplify_reglist  
Simplify a registration list

Description
Simplify a registration list

Usage
simplify_reglist(reg, as.cmtk = NULL)

Arguments
reg  
A registration list (reglist) containing one or more transformations.

as.cmtk  
Whether to convert to a vector of CMTK format registrations (see cmtkreg). The default value of as.cmtk=NULL converts all registrations to CMTK if any one registration is in CMTK format (thus enabling them to be applied by CMTK tools in a single call). See details.

Details
This function
• inverts any affine matrices with attribute "swap"
• collapses multiple affine matrices into a single affine
• optionally converts all registrations to CMTK on disk registrations when possible.

Note that if any of the registrations are in CMTK format, the default behaviour is to try to convert all of the other registrations into CMTK format to enable them to be passed to CMTK in a single command. If as.cmtk=TRUE then there will be an error if this is not possible.

See Also
reglist, xform, cmtkreg

smooth_neuron  
Smooth the 3D coordinates of a neuron skeleton

Description
smooth_neuron smooths a neuron.

Usage
smooth_neuron(n, method = c("gauss", "spline"), ...)
smooth_segment_gauss(xyz, sigma, ...)
spine 139

Arguments

n  Neuron to smooth
method  Smoothing method
...  Additional parameters passed to segment smoothing functions
xyz  A block of 3D coordinates defining an unbranched segment
sigma  The standard deviation of the Gaussian smoothing kernel (which has the same spatial units as the object being smoothed)

Value

A new neuron with smoothed 3d coordinates

Examples

ns=smooth_neuron(Cell07PNs[[1]], sigma=2)
# plot in 2D zooming in on axon terminals
plot(Cell07PNs[[1]], col='grey', xlim=c(260,290), ylim=c(115,90))
plot(ns, col='red', add=TRUE)

# 3D plot
plot3d(Cell07PNs[[1]], col='grey')
plot3d(ns, col='red')

spine  Compute the longest path (aka spine or backbone) of a neuron

Description

Compute the longest path (aka spine or backbone) of a neuron

Usage

spine(
  n,
  UseStartPoint = FALSE,
  SpatialWeights = TRUE,
  invert = FALSE,
  rval = c("neuron", "length", "ids")
)
Arguments

- **n**: the neuron to consider.
- **UseStartPoint**: Whether to use the StartPoint of the neuron (often the soma) as the starting point of the returned spine.
- **SpatialWeights**: logical indicating whether spatial distances (default) should be used to weight segments instead of weighting each edge equally.
- **invert**: When `invert=TRUE` the spine is pruned away instead of being selected. This is only valid when `rval='neuron'` or `rval='ids'`.
- **rval**: Character vector indicating the return type, one of 'neuron', 'length' or 'ids'. See `Value` section.

Value

Either

- a neuron object corresponding to the longest path
- the length of the longest path (when `rval='length'`) or
- an integer vector of raw point indices (when `rval='ids'`).

See Also

- `diameter`, `shortest.paths`, `prune_strahler` for removing lower order branches from a neuron,
- `prune` for removing parts of a neuron by spatial criteria.

Other neuron: `neuron()`, `ngraph()`, `plot.neuron()`, `potential_synapses()`, `prune()`, `resample()`, `rootpoints()`, `subset.neuron()`

Examples

```r
pn.spine=spine(Cell07PNs[[1]])
plot3d(Cell07PNs[[1]])
plot3d(pn.spine, lwd=4, col='black')

# just extract length
spine(Cell07PNs[[1]], rval='length')
# same result since StartPoint is included in longest path
spine(Cell07PNs[[1]], rval='length', UseStartPoint=TRUE)

# extract everything but the spine
antispine=spine(Cell07PNs[[1]], invert=TRUE)
plot3d(Cell07PNs[[1]])
plot3d(antispine, lwd=4, col='red')
```
**strahler_order**

**Find the Strahler order of each point in a neuron**

---

**Description**

The Strahler order will be 1 for each tip segment and then 1 + the maximum of the Strahler order of each parent segment for internal segments. Branch points will have the Strahler order of the closest segment to the root of which they are part.

**Usage**

\[ \text{strahler_order}(x) \]

**Arguments**

- \( x \): A neuron

**Details**

It is vital that the root of the neuron is valid since this determines the flow direction for calculation of the Strahler order. At present the function is not defined for neurons with multiple subtrees.

Internally, this function uses `segmentgraph` to find a reduced segmentgraph for the neuron.

**Value**

A list containing

- points Vector of integer Strahler orders for each point in the neuron
- segments Vector of integer Strahler orders for each segment in the neuron

**References**


**See Also**

`prune_strahler`, a `segmentgraph` (a form of `ngraph`) representation is used to calculate the Strahler order.
sub2ind  
*Find 1D index given n-dimensional indices*

**Description**

Emulates the MATLAB function sub2ind.

**Usage**

```r
sub2ind(dims, indices)
```

**Arguments**

- `dims` vector of dimensions of object to index into.
- `indices` vector of n-dimensional indices.

---

**subset**  
*Subset methods for different nat objects*

**Description**

These methods enable subsets of some nat objects including neurons and neuronlists to be obtained. See the help for each individual method for details.

**See Also**

- `subset.neuron`, `subset.dotprops`, `subset.hxsurf`, `subset.neuronlist`

---

**subset.dotprops**  
*Subset points in dotprops object that match given conditions*

**Description**

Subset points in dotprops object that match given conditions

**Usage**

```r
## S3 method for class 'dotprops'
subset(x, subset, invert = FALSE, ...)
```
subset.dotprops

Arguments

x        A dotprops object
subset   A subset of points defined by indices, an expression or a function (see Details)
invert   Whether to invert the subset criteria - a convenience when selecting by function or indices.
...      Additional parameters (currently ignored)

Details

subset defines either logical or numeric indices, in which case these are simply applied to the matrices that define the points, vect fields of the dotprops object etc OR a function (which is called with the 3D points array and returns T/F. OR an expression vector).

Value

subsetted dotprops object

See Also

prune.dotprops, subset.neuron

Examples

## subset using indices ...
dp=kcs20[[10]]
dp1=subset(dp, 1:50)

# ... or an expression
dp2=subset(dp, alpha>0.7)
front=subset(dp, points[,,'Z']<40)
# use a helper function
between=function(x, lower, upper) x>=lower & x<=upper
middle=subset(dp, between(points[,,'Z'], 40, 60))

# plot results in 3D
plot3d(front, col='red')
plot3d(middle, col='green')
plot3d(dp, col='blue')

## Not run:

## subset using an selection function
s3d=select3d()
dp1=subset(dp, s3d(points))
# special case of previous version
dp2=subset(dp, s3d)
# keep the points that were removed from dp2
dp2.not=subset(dp, s3d, invert=TRUE)
# (another way of doing the same thing)
dp2.not=subset(dp, Negate(s3d))
stopifnot(all.equal(dp1, dp2))
dp2=subset(dp, alpha>0.5 & s3d(pointd))
dp3=subset(dp, 1:10)

## subset each dotprops object in a whole neuronlist
plot3d(kcs20)
s3d=select3d()
kcs20.partial = nlapply(kcs20, subset, s3d)
clear3d()
plot3d(kcs20.partial, col='red')
plot3d(kcs20, col='grey')

## End(Not run)

subset.hxsurf  Subset hxsurf object to specified regions

Description
Subset hxsurf object to specified regions

Usage
## S3 method for class 'hxsurf'
subset(x, subset = NULL, drop = TRUE, rval = c("hxsurf", "names"), ...)

Arguments
x  A dotprops object
subset  Character vector specifying regions to keep. Interpreted as regex if of length 1
and no fixed match.
drop  Whether to drop unused vertices after subsetting (default: TRUE)
rval  Whether to return a new hxsurf object or just the names of the matching regions
...  Additional parameters (currently ignored)

Value
subsetted hxsurf object

See Also
Other hxsurf: as.hxsurf(), as.mesh3d(), materials(), plot3d.hxsurf(), read.hxsurf(),
write.hxsurf()
Examples

# plot only vertical lobe
vertical_lobe = subset(MBL.surf, "VL")

plot3d(vertical_lobe, alpha=0.3)
plot3d(kcs20)

# there is also a shortcut for this
clear3d()
plot3d(MBL.surf, "VL", alpha=0.3)

subset.neuron

Subset neuron by keeping only vertices that match given conditions

Description

Subset neuron by keeping only vertices that match given conditions

Usage

## S3 method for class 'neuron'
subset(x, subset, invert = FALSE, ...)

Arguments

x
A neuron object

subset
A subset of points defined by indices, an expression, or a function (see Details)

invert
Whether to invert the subset criteria - a convenience when selecting by function or indices.

...
Additional parameters (passsed on to prune_vertices)

Details

subset defines which vertices of the neuron to keep and is one of

- logical or numeric indices, in which case these are simply used to index the vertices in the order of the data.frame x$d. Note that any NA values are ignored.
- a function (which is called with the 3D points array and returns T/F vector)
- an expression evaluated in the context of the x$d data.frame containing the SWC specification of the points and connectivity of the neuron. This can therefore refer e.g. to the X,Y,Z location of vertices in the neuron.

Value

subsetted neuron
See Also

`prune.neuron`, `prune_vertices`, `subset.dotprops`

Other neuron: `neuron()`, `ngraph()`, `plot.neuron()`, `potential_synapses()`, `prune()`, `resample()`, `rootpoints()`, `spine()`

Examples

```
n=Cell07PNs[[1]]
# keep vertices if their X location is > 2000
n1=subset(n, X>200)
# diameter of neurite >1
n2=subset(n, W>1)
# first 50 nodes
n3=subset(n, 1:50)
# everything but first 50 nodes
n4=subset(n, 1:50, invert=TRUE)

## subset neuron by graph structure
# first plot neuron and show the point that we will use to divide the neuron
n=Cell07PNs[[1]]
plot(n)
# this neuron has a tag defining a point at which the neuron enters a brain
# region (AxonLHEP = Axon Lateral Horn Entry Point)
points(t(xyzmatrix(n)[n$AxonLHEP, 1:2]), pch=19, cex=2.5)

# now find the points downstream (distal) of that with respect to the root
ng=as.ngraph(n)
# use a depth first search
distal_points=igraph::graph.dfs(ng, root=n$AxonLHEP, unreachable=FALSE, neimode='out')$order
distal_tree=subset(n, distal_points)
plot(distal_tree, add=TRUE, col='red', lwd=2)

# Find proximal tree as well
# nb this does not include the AxonLHEP itself as defined here
proximal_points=setdiff(igraph::V(ng), distal_points)
proximal_tree=subset(n, proximal_points)
plot(proximal_tree, add=TRUE, col='blue', lwd=2)

## Not run:
## subset using interactively defined spatial regions
plot3d(n)
# you can save this select3d object using save or saveRDS functions
# for future non-interactive use
s3d=select3d()
n4=subset(n, s3d(xyzmatrix(n)))
# special case of previous version
n5=subset(n, s3d)
stopifnot(all.equal(n4,n5))
# keep the points that were removed from n1
n4.not=subset(n,Negate(s3d))
# vertices with x position > 100 and inside the selector function
```
n6 = subset(n, X > 100 & s3d(X, Y, Z))

## subset each neuron object in a whole neuronlist
n10 = Cell07PNs[1:10]
plot3d(n10, lwd=0.5, col='grey')
n10.crop = nlapply(n10, subset, X > 250)
plot3d(n10.crop, col='red')

## subset a neuron using a surface
library(nat.flybrains)
# extract left lateral horn surface and convert to mesh3d
lh = as.mesh3d(subset(IS2NP.surf, "LH_L"))
# subset neuron with this surface
x = subset(Cell07PNs[[1]], function(x) pointsinside(x, lh))
shade3d(lh, alpha=0.3)
plot3d(x, lwd=3, col='blue')
# Now find the parts of the neuron outside the surface
y = subset(Cell07PNs[[1]], function(x) Negate(pointsinside)(x, lh))
plot3d(y, col='red', lwd=2)

## End(Not run)

### subset.neuronlist

Subset neuronlist returning either new neuronlist or names of chosen neurons

#### Description

Subset neuronlist returning either new neuronlist or names of chosen neurons

#### Usage

```r
## S3 method for class 'neuronlist'
subset(
x, subset, filterfun, rval = c("neuronlist", "names", "data.frame"), ...
)
```

#### Arguments

- **x**
a neuronlist

- **subset**
  An expression that can be evaluated in the context of the dataframe attached to the neuronlist. See details.

- **filterfun**
  a function which can be applied to each neuron returning TRUE when that neuron should be included in the return list.

- **rval**
  What to return (character vector, default='neuronlist')

- **...**
  additional arguments passed to filterfun
Details

The subset expression should evaluate to one of

- character vector of names
- logical vector
- vector of numeric indices

Any missing names are dropped with a warning. The filterfun expression is wrapped in a try. Neurons returning an error will be dropped with a warning.

You may also be interested in `find.neuron`, which enables objects in a neuronlist to be subsetted by a 3D selection box. In addition `subset.neuron`, `subset.dotprops` methods exist: these are used to remove points from neurons (rather than to remove neurons from neuronlists).

Value

A neuronlist, character vector of names or the attached data.frame according to the value of `rval`

See Also

`neuronlist`, `find.neuron`, `subset.data.frame`, `subset.neuron`, `subset.dotprops`

Examples

dalpns=subset(Cell07PNs,Glomerulus=='DA1')
with(dalpns,stopifnot(all(Glomerulus=='DA1')))
gammas=subset(kcs20,type=='gamma')
with(gammas,stopifnot(all(type=='gamma')))
# define a function that checks whether a neuron has points in a region in
# space, specifically the tip of the mushroom body alpha' lobe
aptip<-function(x) {xyz=xyzmatrix(x);any(xyz[,,'X']>350 & xyz[,,'Y']<40)}
# this should identify the alpha'/beta' kenyon cells only
apbps=subset(kcs20,filterfun=aptip)

head(apbps)
# combine global variables with dataframe columns
odds=rep(c(TRUE,TRUE),10)
stopifnot(all.equal(subset(kcs20,type=='gamma' & odds),

subset(kcs20,type=='gamma' & rep(c(TRUE,TRUE),10))))
## Not run:
# make a 3D selection function using interactive rgl::select3d() function
s3d=select3d()
# Apply a 3D search function to the first 100 neurons in the neuronlist dataset
subset(dps[1:100],filterfun=function(x) {sum(s3d(xyzmatrix(x)))>0},

rval='names')
# combine a search by metadata, neuropil location and 3D location
subset(dps, Gender=='M' & rAL>1000, function(x) sum(s3d(x))>0, rval='name')
# The same but specifying indices directly, which can be considerably faster
# when neuronlist is huge and memory is in short supply
subset(dps, names(dps)[1:100],filterfun=function(x) {sum(s3d(xyzmatrix(x)))>0},

rval='names')
## End(Not run)

### summary.neuronlist

**Summary statistics for neurons (e.g. cable length, number of nodes)**

### Description

summary.neuronlist computes tree statistics for all the neurons in a neuronlist object

summary.neuron computes statistics for individual neurons

summary.dotprops computes statistics for individual neurons in dotprops format. Note the veclength argument.

### Usage

```r
## S3 method for class 'neuronlist'
summary(object, ..., include.attached.dataframe = FALSE)

## S3 method for class 'neuron'
summary(object, ...)

## S3 method for class 'dotprops'
summary(object, veclength = 1, ...)
```

### Arguments

- **object**
  The neuron or neuronlist to summarise

- **...**
  For summary.neuronlist additional arguments passed on to summary methods for individual neurons

- **include.attached.dataframe**
  Whether to include the neuronlists attached metadata in the returned data.frame.

- **veclength**
  The vector length to assume for each segment so that a cable length estimate can be made.

### Value

A data.frame summarising the tree properties of the neuron with columns

- root
- nodes
- segments
- branchpoints
- endpoints
- cable.length
### Examples

```r
# summary for a whole neuronlist
summary(Cell07PNs)
# including the attached data.frame with additional metadata
head(summary(Cell07PNs, include.attached.dataframe = FALSE))
# for a single regular format neuron
summary(Cell07PNs[[1]])
# for a single dotprops format neuron
summary(kcs20[[1]])
# specify a different estimate for the cable length associated with a single
# point in the neuron
summary(kcs20[[1]], veclength=1.2)
```

---

#### Description

Threshold an object, typically to produce a mask

#### Usage

```r
threshold(x, ...)  
```

#### Arguments

- `x`  
  Object to be thresholded
- `...`  
  Additional arguments passed to methods
- `threshold`  
  Either a numeric value that pixels must exceed in order to be included in the mask or a logical vector defining foreground pixels.
- `mode`  
  The storage mode of the resultant object (see `vector`)

#### Details

Note that `threshold.im3d` passes ...arguments on to `im3d`
Value

an object with attributes matching x and elements with value as.vector(TRUE, mode=mode) i.e. TRUE, 1, 0x01 and as.vector(FALSE, mode=mode) i.e. FALSE, 0, 0x00 as appropriate.

See Also

Other im3d: as.im3d(), boundingbox(), im3d-coords, im3d-io, im3d(), imexpand.grid(), imslice(), is.im3d(), mask(), origin(), projection(), unmask(), voxdims()

Examples

x=im3d(rnorm(1000), dims=c(10,10,10), BoundingBox=c(20,200,100,200,200,300))
stopifnot(all.equal(threshold(x, 0), threshold(x, x>0)))

union

Find the union of two collections of objects

Description

Find the union of two collections of objects

Usage

union(x, y, ...)

## Default S3 method:
union(x, y, ...)

## S3 method for class 'neuronlist'
union(x, y, ...)

Arguments

x the first collection to consider.
y the second collection to consider.
... additional arguments passed to methods

Details

Note that union.default calls base::union to ensure consistent behaviour for regular vectors.

Value

A collection of the same mode as x that contains all unique elements of x and y.

See Also

union
unmask

**Description**

Make im3d image array containing values at locations defined by a mask

**Usage**

```r
unmask(
  x,
  mask,
  default = NA,
  attributes. = attributes(mask),
  copyAttributes = TRUE
)
```

**Arguments**

- `x` the data to place on a regular grid
- `mask` An im3d regular image array where non-zero voxels are the selected element.
- `default` Value for regions outside the mask (default: NA)
- `attributes.` Attributes to set on new object. Defaults to attributes of `mask`
- `copyAttributes` Whether to copy over attributes (including `dim`) from the mask to the returned object. default: TRUE

**Details**

The values in `x` will be placed into a grid defined by the dimensions of the `mask` in the order defined by the standard R linear subscripting of arrays (see e.g. `arrayInd`).

**Value**

A new im3d object with attributes/dimensions defined by `mask` and values from `x`. If `copyAttributes` is FALSE, then it will have mode of `x` and length of `mask` but no other attributes.

**See Also**

Other im3d: `as.im3d()`, `boundingbox()`, `im3d-coords`, `im3d-io`, `im3d()`, `imexpand.grid()`, `imslice()`, `is.im3d()`, `mask()`, `origin()`, `projection()`, `threshold()`, `voxdims()`
Examples

```r
## Not run:
# read in a mask
LHMask=read.im3d(system.file('tests/testthat/testdata/nrrd/LHMask.nrrd', package='nat'))
# pick out all the non zero values
inmask=LHMask[LHMask!=0]
# fill the non-zero elements of the mask with a vector that iterates over the
# values 0:9
stripes=unmask(seq(inmask)%%10, LHMask)
# make an image from one slice of that result array
image(imslice(stripes,11), asp=TRUE)

## End(Not run)
```

---

**voxdims**

*Return voxel dimensions of an object*

**Description**

This would properly be thought of as the voxel spacing when voxels are assumed not to have a physical extent (only a location).

**Usage**

```r
testfoox <- voxdims(x, ...)
```

**Arguments**

- `x` An `im3d` object with associated voxel dimensions, a path to or a 2 x 3 BoundingBox matrix.
- `...` Additional arguments for methods
- `dims` The number of voxels in each dimension when `x` is a BoundingBox matrix.

**Details**

We follow Amira’s convention of returning a voxel dimension equal to the bounding box size (rather than 0) for any dimension with only 1 voxel.
write.amiramesh

Value

A numeric vector of length 3, NA when missing.

See Also

boundingbox

Other im3d: as.im3d(), boundingbox(), im3d-coords, im3d-io, im3d(), imexpand.grid(), imslice(), is.im3d(), mask(), origin(), projection(), threshold(), unmask()

write.amiramesh  Write a 3D data object to an amiramesh format file

Description

Write a 3D data object to an amiramesh format file

Usage

write.amiramesh(
  x,        # The image data to write (an im3d, or capable of being interpreted as such)
  file,     # Character vector describing a single file
  enc = c("binary", "raw", "text", "hxzip"),
  dtype = c("float", "byte", "short", "ushort", "int", "double"),
  endian = .Platform$endian,
  WriteNrrdHeader = FALSE
)

Arguments

x          # The image data to write (an im3d, or capable of being interpreted as such)
file       # Character vector describing a single file
enc        # Encoding of the data. NB "raw" and "binary" are synonyms.
dtype      # Data type to write to disk
endian     # Endianness of data block. Defaults to current value of .Platform$endian.
WriteNrrdHeader # Whether to write a separate detached nrrd header next to the amiramesh file allowing it to be opened by a NRRD reader. See details.

Details

Note that only 'raw' or 'text' format data can accommodate a detached NRRD format header since Amira’s HxZip format is subtly different from NRRD’s gzip encoding. There is a full description of the detached NRRD format in the help for write.nrrd.
write.cmtk

Write a suitable list to a CMTK TypedStream file on disk

Description

This is probably only of interest to developers. End users will probably wish to use more specific functions such as write.cmtkreg for writing out registrations.

Usage

write.cmtk(l, con, gzip = FALSE, version = NA_character_)

Arguments

- **l**: Appropriately formatted list
- **con**: A character string specifying a path or a connection
- **gzip**: Whether to gzip output file (default FALSE)
- **version**: TYPEDSTREAM version number, defaults to "1.1" if not specified in the version attribute of l.

Details

NB a version specified on the command line overrides one encoded as an attribute in the input list.

See Also

Other cmtk-io: cmtk.extract_affine(), read.cmtkreg(), read.cmtk(), write.cmtkreg()
write.cmtkreg  Write out CMTK registration list to folder

Description
Write out CMTK registration list to folder

Usage
write.cmtkreg(reglist, foldername, version = "2.4")

Arguments
- reglist: List specifying CMTK registration parameters
- foldername: Path to registration folder (usually ending in .list)
- version: CMTK version for registration (default 2.4)

Details
Note that transformation in the forward direction (i.e. sample->ref) e.g. as calculated from a set of landmarks where set 1 is the sample is considered an inverse transformation by the IGS software. So in order to use such a transformation as an initial affine with the registration command the switch --initial-inverse must be used specifying the folder name created by this function.

CMTK v2.4 fixed a long-standing bug in affine (de)composition to CMTK params. This resulted in a non-backwards compatible change marked by writing the TYPEDSTREAM as version 2.4. The R code in this package implements both the new and old compose/decompose functions, using the new by default.

See Also
Other cmtk-io: cmtk.extract_affine(), read.cmtkreg(), read.cmtk(), write.cmtk()

write.hxsurf  Write Amira surface (aka HxSurface or HyperSurface) into .surf file.

Description
Write Amira surface (aka HxSurface or HyperSurface) into .surf file.

Usage
write.hxsurf(surf, filename)
Arguments

surf     hxsurf object to write to file.
filename character vector defining path to file.

Value

NULL or integer status from close.

See Also

plot3d.hxsurf, read.hxsurf, rgb

Other amira: amiratype(), is.amiramesh(), read.amiramesh(), read.hxsurf()

Other hxsurf: as.hxsurf(), as.mesh3d(), materials(), plot3d.hxsurf(), read.hxsurf(),
subset.hxsurf()

write.neuron  Write out a neuron in any of the file formats we know about

Description

If file is not specified the neuron’s InputFileName field will be checked (for a dotprops object it
will be the 'file' attribute). If this is missing there will be an error. If dir is specified it will be
combined with basename(file). If file is specified but format is not, it will be inferred from file’s
extension.

Usage

write.neuron(
  n,
  file = NULL,
  dir = NULL,
  format = NULL,
  ext = NULL,
  Force = FALSE,
  MakeDir = TRUE,
  ...
)

Arguments

n          A neuron
file       Path to output file
dir        Path to directory (this will replace dirname(file) if specified)
format     Unique abbreviation of one of the registered file formats for neurons including
            'swc', 'hxlineset', 'hxskel'
write.neuronlistfh

Write out a neuronlistfh object to an RDS file

Description

Write out a neuronlistfh object to an RDS file

Usage

write.neuronlistfh(x, file = attr(x, "file"), overwrite = FALSE, ...)
Arguments

- `x`: The neuronlistfh object to write out
- `file`: Path where the file will be written (see details)
- `overwrite`: Whether to overwrite an existing file
- `...`: Additional parameters passed to saveRDS

Details

This function writes the main neuronlistfh object to disk, but makes no attempt to touch/verify the associated object files.

If `file` is not specified, then the function will first check if `x` has a 'file' attribute. If that does not exist, then `attr(x, 'db')@dir`, the backing filehash database directory, is inspected. The save path file will then be constructed by taking the directory one up from the database directory and using the name of the neuronlistfh object with the suffix '.rds'. e.g. `write.neuronlistfh(kcs20)` with `db` directory '/my/path/dps/data' will be saved as '/my/path/dps/kcs20.rds'

Note that if `x` has a 'file' attribute (set by `read.neuronlistfh`) then this will be removed before the file is saved (since the file attribute must be set on read to ensure that we know exactly which file on disk was the source of the object in memory).

See Also

- saveRDS

Other neuronlistfh: [.neuronlistfh(), neuronlistfh(), read.neuronlistfh(), remotesync()]

Description

Write neurons from a neuronlist object to individual files, or a zip archive

Usage

```r
write.neurons(
  nl, 
  dir, 
  format = NULL, 
  subdir = NULL, 
  INDICES = names(nl), 
  files = NULL, 
  Force = FALSE, 
  ...
)
```
Arguments

- **nl**: neuronlist object
- **dir**: directory to write neurons, or path to zip archive (see Details).
- **format**: Unique abbreviation of one of the registered file formats for neurons including 'swc', 'hxlineset', 'hxskel'
- **subdir**: String naming field in neuron that specifies a subdirectory OR expression to evaluate in the context of neuronlist’s df attribute
- **INDICES**: Character vector of the names of a subset of neurons in neuronlist to write.
- **files**: Character vector or expression specifying output filenames. See examples and write.neuron for details.
- **Force**: Whether to overwrite an existing file
- **...**: Additional arguments passed to write.neuron

Details

See write.neuron for details of how to specify the file format/extension/name of the output files and how to establish what output file formats are available. A zip archive of files can be written by specifying a value of dir that ends in .zip.

Value

the path to the output file(s), absolute when this is a zip file.

Author(s)

jefferis

See Also

write.neuron, read.neurons, fileformats

Other neuronlist: *.neuronlist(), is.neuronlist(), neuronlist-dataframe-methods, neuronlistfh(), neuronlist(), nlapply(), read.neurons()

Examples

```R
## Not run:
# write some neurons in swc format
write.neurons(Cell07PNs, dir="testwn", format='swc')
# write some neurons in Amira hxlineset format
write.neurons(Cell07PNs, dir="testwn", format='hxlineset')

# organise new files in directory hierarchy by glomerulus and Scored.By field
write.neurons(Cell07PNs,dir="testwn",
  subdir=file.path(Glomerulus,Scored.By),format='hxlineset')
# ensure that the neurons are named according to neuronlist names
write.neurons(Cell07PNs, dir="testwn", files=names(Cell07PNs),
  subdir=file.path(Glomerulus,Scored.By),format='hxlineset')
# only write a subset
```
write.neurons(subset(Cell07PNs, Scored.By="ACH"), dir="testwn2", subdir=Glomerulus, format='hxlineset')
# The same, but likely faster for big neuron lists
write.neurons(Cell07PNs, dir="testwn3", INDICES=subset(Cell07PNs, Scored.By="ACH", rval='names'), subdir=Glomerulus, format='hxlineset')
# set file name explicitly using a field in data.frame
write.neurons(subset(Cell07PNs, Scored.By="ACH"), dir="testwn4", subdir=Glomerulus, files=paste0(ID, '.am'), format='hxlineset')

## End(Not run)

write.nrrd

Write data and metadata to NRRD file or create a detached NRRD (nhdr) file.

**Description**

write.nrrd writes an array, vector or im3d object to a NRRD file. When x is an im3d object, appropriate spatial calibration fields are added to the header.

write.nrrd.header writes a nrrd header file.

write.nrrd.header.for.file makes a detached NRRD (nhdr) file that points at another image file on disk, making it NRRD compatible. This can be a convenient way to make NRRD inputs for other tools e.g. CMTK and also allows the same data block to pointed to by different nhdr files with different spatial calibration.

**Usage**

```r
write.nrrd(x, file, enc = c("gzip", "raw", "text"), dtype = c("float", "byte", "short", "ushort", "int", "double"), header = attr(x, "header"), endian = .Platform$endian, datafile = NULL)
write.nrrd.header(header, file)
write.nrrd.header.for.file(infile, outfile = NULL)
```

**Arguments**

- `x` Data to write as an array, vector or im3d object.
- `file` Character string naming an output file (a detached nrrd header when file has extension 'nhdr').
**write.nrrd**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>enc</td>
<td>One of three supported nrrd encodings (&quot;gzip&quot;, &quot;raw&quot;, &quot;text&quot;)</td>
</tr>
<tr>
<td>dtype</td>
<td>The data type to write. One of &quot;float&quot;, &quot;byte&quot;, &quot;short&quot;, &quot;ushort&quot;, &quot;int&quot;, &quot;double&quot;</td>
</tr>
<tr>
<td>header</td>
<td>List containing fields of nrrd header - see Header section.</td>
</tr>
<tr>
<td>endian</td>
<td>One of &quot;big&quot; or &quot;little&quot;. Defaults to <code>.Platform$endian</code>.</td>
</tr>
<tr>
<td>datafile</td>
<td>Optional name of separate file into which data should be written (see details).</td>
</tr>
<tr>
<td>infile, outfile</td>
<td>Path to input and output file for write.nrrd.header.for.file. If outfile is NULL (the default) then it will be set to &lt;infilestem.nhdr&gt;.</td>
</tr>
</tbody>
</table>

**Detached NRRDs**

NRRD files can be written in detached format (see [http://teem.sourceforge.net/nrrd/format.html#detached](http://teem.sourceforge.net/nrrd/format.html#detached)) in which a text nhdr file is used to described the contents of a separate (usually binary) data file. This means that the nhdr file can be inspected and edited with a text editor, while the datablock can be in a completely raw format that can be opened even by programs that do not understand the NRRD format. Furthermore detached NRRD header files can be written to accompany non-NRRD image data so that it can be opened by nrrd readers.

If file has extension .nhdr or datafile is non-NULL, then write.nrrd will write a separate datafile. If datafile is set, then it is interpreted as specifying a path relative to the nhdr file. If datafile is not specified then default filenames will be chosen according to the encoding following the conventions of the teem library:

- raw '\<nhdrstem>.raw'
- gzip '\<nhdrstem>.raw.gz'
- text '\<nhdrstem>.ascii'

**Data file paths**

When a detached NRRD is written, the datafile can be specified either as relative or absolute path. Relative paths are strongly recommended - the best place is right next to the datafile. Relative paths are always specified with respect to the location of the nhdr file.

The datafile argument is not processed by write.nrrd so it is up to the caller to decide whether a relative or absolute path will be used.

For write.nrrd.header.for.file if outfile is not specified then the nhdr file will be placed next to the original image stack and the datafile field will therefore just be basename(infile). If outfile is specified explicitly, then datafile will be set to the full path in the infile argument. Therefore if you wish to specify outfile, you must set the current working directory (using setwd) to the location in which outfile will be written to ensure that the path to the datafile is correct. A future TODO would add the ability to convert an absolute datafile path to a relative one (by finding the common path between datafile and nhdr folders).

**Header**

For write.nrrd, arguments enc, dtype, and endian along with the dimensions of the input (x) will override the corresponding NRRD header fields from any supplied header argument. See [http://teem.sourceforge.net/nrrd/format.html](http://teem.sourceforge.net/nrrd/format.html) for details of the NRRD fields.
write.vtk

Write object to VTK file

Description

Write object to VTK file

Usage

write.vtk(x, file, ...)

## S3 method for class 'neuron'
write.vtk(
  x,
  file,
  datatype = c("float", "double"),
  title = file,
  WriteAllSubTrees = TRUE,
  ...
)

Arguments

- **x**: Object to write
- **file**: Path to output file
- **...**: Additional arguments to methods
- **datatype**: The VTK data type (one of float or double)
- **title**: Title of the .vtk file (defaults to file)
- **WriteAllSubTrees**: Whether to write all subtrees in the neuron or just the main tree.

Examples

## Not run:
n=Cell07PNs[[1]]
write.vtk(n, paste0(n$NeuronName, ".vtk"))
write.neuron(n, paste0(n$NeuronName, ".vtk"))

## End(Not run)
xform

Transform the 3D location of objects such as neurons

Description

xform is designed to operate on a variety of data types, especially objects encapsulating neurons. xform depends on two specialised downstream functions xformpoints and xformimage. These are user visible any contain some useful documentation, but should only be required for expert use; in almost all circumstances, you should use only xform.

xform.character is designed to work with files on disk. Presently it is restricted to images, although other datatypes may be supported in future.

Usage

xform(x, reg, ...)  
## Default S3 method:  
xform(x, reg, na.action = c("warn", "none", "drop", "error"), ...)  
## S3 method for class 'character'  
xform(x, reg, ...)  
## S3 method for class 'list'  
xform(x, reg, FallBackToAffine = TRUE, na.action = "error", ...)  
## S3 method for class 'shape3d'  
xform(x, reg, FallBackToAffine = TRUE, na.action = "error", ...)  
## S3 method for class 'neuron'  
xform(x, reg, FallBackToAffine = TRUE, na.action = "error", ...)  
## S3 method for class 'data.frame'  
xform(x, reg, subset = NULL, ...)  
## S3 method for class 'dotprops'  
xform(x, reg, FallBackToAffine = TRUE, ...)  
## S3 method for class 'neuronlist'  
xform(  
  x,  
  reg,  
  subset = NULL,  
  ...,  
  OmitFailures = NA,  
  VectoriseRegistrations = FALSE,  
  TransformDFCoords = TRUE
)
Arguments

- **x**: an object to transform
- **reg**: A registration defined by a matrix, a function, a cmtkreg object, or a character vector specifying a path to one or more registrations on disk (see Registrations section).
- **...**: additional arguments passed to methods and eventually to `xformpoints`
- **na.action**: How to handle NAs. NB drop may not work for some classes.
- **FallbackToAffine**: Whether to use an affine transform when a cmtk warping transformation fails.
- **subset**: For `xform.neuronlist` indices (character/logical/integer) that specify a subset of the members of `x` to be transformed.
- **OmitFailures**: Whether to omit neurons for which `FUN` gives an error. The default value (NA) will result in nlapply stopping with an error message the moment there is an error. For other values, see details.
- **VectoriseRegistrations**: When FALSE, the default, each element of `reg` will be applied sequentially to each element of `x`. When TRUE, it is assumed that there is one element of `reg` for each element of `x`.
- **TransformDFCoords**: If the metadata data.frame attached to `x` includes columns that look like x,y,z coordinates, transform those as well.

Details

Methods are provided for some specialised S3 classes. Further methods can of course be constructed for user-defined S3 classes. However this will probably not be necessary if the `xyzmatrix` and `xyzmatrix<-` generics are suitably overloaded and the S3 object inherits from list.

Note that given the behaviour of the `xyzmatrix` functions, the `xform.data.frame` method will transform the x,y,z or X,Y,Z columns of a data.frame if the data.frame has more than 3 columns, erroring out if no such unique columns exist.

**TODO** get this to work for matrices with more than 3 columns by working on `xyzmatrix` definition.

For the `xform.dotprops` method, dotprops tangent vectors will be recalculated from scratch after the points have been transformed (even though the tangent vectors could in theory be transformed more or less correctly). When there are multiple transformations, `xform` will take care to carry out all transformations before recalculating the vectors.

With `xform.neuronlist`, if you want to apply a different registration to each object in the neuronlist `x`, then you should use `VectoriseRegistrations=TRUE`.

When `x`’s attached data.frame contains columns called x,y,z or X,Y,Z then these are assumed to be coordinates and also transformed when `TransformDFCoords=TRUE` (the default). This provides a mechanism for transforming the soma positions of neuronlist objects containing dotprops objects (which do not otherwise store the soma position). Note that if transformation fails, a warning will be issued and the points will be replaced with NA values.
Registrations

When \texttt{reg} is a character vector, \texttt{xform}'s specialised downstream functions will check to see if it defines a path to one (or more) registrations on disk. These can be of two classes

- CMTK registrations
- \texttt{reglist} objects saved in R's RDS format (see \texttt{readRDS}) which can contain any sequence of registrations supported by \texttt{nat}.

If the path does indeed point to a CMTK registration, this method will hand off to \texttt{xformpoints.cmtkreg} or \texttt{xformimages.cmtkreg}. In this case, the character vector may optionally have an attribute, 'swap', a logical vector of the same length indicating whether the transformation direction should be swapped. At the moment only CMTK registration files are supported.

If \texttt{reg} is a character vector of length \( \geq 1 \) defining a sequence of registration files on disk they should proceed from sample to reference.

Where \texttt{reg} is a function, it should have a signature like \texttt{myfun(x,\ldots)} where the \ldots must be provided in order to swallow any arguments passed from higher level functions that are not relevant to this particular transformation function.

See Also

\texttt{xformpoints}

Examples

```r
## Not run:
k1=kcs20[[1]]
k1.default=xform(k1,function(x,\ldots)x)
stopifnot(isTRUE(all.equal(k1,k1.default)))
k1.5=xform(k1,function(x,\ldots)x, k=5)
stopifnot(isTRUE(all.equal(k1.5,k1.default)))
k1.20=xform(k1,function(x,\ldots)x, k=20)
stopifnot(!isTRUE(all.equal(k1,k1.20)))

# apply two registrations converting sample->IS2->JFRC2
reg_seq=c("IS2_sample.list", "JFRC2_IS2.list")
xform(k1, reg_seq)
# apply two registrations, swapping the direction of the second one
# i.e. sample -> IS2 -> FCWB
reg_seq=structure(c("IS2_sample.list", "IS2_FCWB.list"), swap=c(FALSE, TRUE))
xform(k1, reg_seq)

## End(Not run)
```

```r
## Not run:
# apply reg1 to Cell07PNs[[1]], reg2 to Cell07PNs[[2]] etc
regs=c(reg1, reg2, reg3)
x=xform(Cell07PNs[1:3], reg=regs, VectoriseRegistrations=TRUE)

## End(Not run)
```
xformimage

Transform image files using a registration or affine matrix

Description

You should almost always call \texttt{xform} rather calling than \texttt{xformimage} directly.

Usage

\begin{verbatim}
xformimage(reg, image, ...)

## S3 method for class 'character'
xformimage(reg, image, ...)

## S3 method for class 'cmtkreg'
xformimage(
    reg,
    image,
    transformtype = c("warp", "affine"),
    direction = NULL,
    ...
)

## S3 method for class 'reglist'
xformimage(reg, image, ...)

## Default S3 method:
xformimage(reg, image, ...)
\end{verbatim}

Arguments

\begin{itemize}
\item \texttt{reg} \hspace{1cm} A registration defined by a matrix or a \texttt{cmtkreg} object, or a character vector specifying a path to a CMTK registration on disk (see details). If \texttt{reg} is a character vector of length >1 defining a sequence of registration files on disk they should proceed from sample to reference.
\item \texttt{image} \hspace{1cm} Nx3 matrix of image
\item \texttt{...} \hspace{1cm} Additional arguments passed to methods (and then eventually to \texttt{cmtk.reformatx})
\item \texttt{transformtype} \hspace{1cm} Which transformation to use when the CMTK file contains both warp (default) and affine (TODO)
\item \texttt{direction} \hspace{1cm} Whether to transform image from sample space to reference space (called \texttt{forward} by CMTK) or from reference to sample space (called \texttt{inverse} by CMTK). Default (when \texttt{NULL} is forward).
\end{itemize}
Details

When passed a character vector, xformimage will check to see if it defines a path containing CMTK registration erroring out if this is not the case. If the path does indeed point to a CMTK registration, this method will hand off to xformimage.cmtkreg. A future TODO would be to provide a mechanism for extending this behaviour for other registration formats. If a list of transformations is passed in, these transformations are passed to the cmtk reformatx tool in the order received. Note that there is presently no support for

- using the inverse of a registration
- specifying a mask
- passing additional arguments to reformatx

Note that the direction of CMTK registrations can be the source of much confusion. This is because CMTK defines the forward direction as the transform required to reformat an image in sample (floating) space to an image in template space. Since this operation involves filling a regular grid in template space by looking up the corresponding positions in sample space, the transformation that is required is (somewhat counterintuitively) the one that maps template to sample. However in neuroanatomical work, one often has points in sample space that one would like to transform into template space. Here one needs CMTK's inverse transformation.

A second source of confusion is that when there are multiple transformations, CMTK's reformatx tool (wrapped by cmtk.reformatx) expects them to be listed:

ref_intermediate.list intermediate_sample.list

where ref_intermediate.list is the CMTK registration obtained with ref as target/reference and intermediate as sample/floating image.

For consistency, all xform.* methods expect multiple registrations to be listed from sample to reference and this order is then swapped when they are passed on to cmtk.reformatx.

whereas CMTK's streamxform tool (wrapped by xformpoints) expects them in the opposite order.

Value

Character vector with path to xformed image.

See Also

cmtk.reformatx, xformpoints, xform

Description

You should almost always call xform rather calling than xformpoints directly.
Usage

xformpoints(reg, points, ...)

## S3 method for class 'character'
xformpoints(reg, points, ...)

## S3 method for class 'cmtkreg'
xformpoints(
    reg,
    points,
    transformtype = c("warp", "affine"),
    direction = NULL,
    FallBackToAffine = FALSE,
    ...
)

## S3 method for class 'reglist'
xformpoints(reg, points, ...)

## Default S3 method:
xformpoints(reg, points, ...)

Arguments

reg A registration defined by a matrix, a function, a cmtkreg object, a reglist object containing a sequence of arbitrary registrations, or a character vector specifying path(s) to registrations on disk (see details).

points Nx3 matrix of points

... Additional arguments passed to methods

transformtype Which transformation to use when the CMTK file contains both warp (default) and affine
direction Whether to transform points from sample space to reference space (called inverse by CMTK) or from reference to sample space (called forward by CMTK). Default (when NULL is inverse).

FallBackToAffine Whether to use the affine transformation for points that fail to transform under a warping transformation.

Details

If a list of transformations is passed in, these transformations are performed in sequence order, such that xformpoints(c(a,b,c), x) == xformpoints(c, xformpoints(b, xformpoints(a, x))))

Note that the direction of CMTK registrations can be the source of much confusion. This is because CMTK defines the forward direction as the transform required to reformat an image in sample (floating) space to an image in template space. Since this operation involves filling a regular grid in template space by looking up the corresponding positions in sample space, the transformation that is required is (somewhat counterintuitively) the one that maps template to sample. However in
neuroanatomical work, one often has points in sample space that one would like to transform into template space. Here one needs the inverse transformation.

---

### xyzmatrix

**Get and assign coordinates for classes containing 3D vertex data**

**Description**

`xyzmatrix` gets coordinates from objects containing 3D vertex data

`xyzmatrix<-` assigns xyz elements of neuron or dotprops object and can also handle matrix like objects with columns named X, Y, Z or x, y, z.

**Usage**

```r
xyzmatrix(x, ...)

## Default S3 method:
xyzmatrix(x, y = NULL, z = NULL, ...)

## S3 method for class 'neuron'
xyzmatrix(x, ...)

## S3 method for class 'neuronlist'
xyzmatrix(x, ...)

## S3 method for class 'dotprops'
xyzmatrix(x, ...)

## S3 method for class 'hxsurf'
xyzmatrix(x, ...)

## S3 method for class 'igraph'
xyzmatrix(x, ...)

## S3 method for class 'mesh3d'
xyzmatrix(x, ...)

xyzmatrix(x) <- value

## S3 replacement method for class 'neuron'
xyzmatrix(x) <- value

## S3 replacement method for class 'dotprops'
xyzmatrix(x) <- value

## S3 replacement method for class 'hxsurf'
xyzmatrix(x) <- value
```
xyzmatrix

## S3 replacement method for class 'igraph'
xyzmatrix(x) <- value

## S3 replacement method for class 'shape3d'
xyzmatrix(x) <- value

## S3 replacement method for class 'neuronlist'
xyzmatrix(x) <- value

### Arguments

**x**
- object containing 3D coordinates

**...**
- additional arguments passed to methods

**y, z**
- separate y and z coordinates

**value**
- Nx3 matrix specifying new xyz coords

### Details

Note that `xyzmatrix` can extract or set 3D coordinates in a `matrix` or `data.frame` that **either** has exactly 3 columns **or** has 3 columns named X,Y,Z or x,y,z.

### Value

For `xyzmatrix`: Nx3 matrix containing 3D coordinates

For `xyzmatrix<-`: Original object with modified coords

### See Also

`xyzmatrix`

### Examples

```r
# see all available methods for different classes
methods('xyzmatrix')

# ... and for the assignment method
methods('xyzmatrix<-')

n=Cell07PNs[[1]]
xyzmatrix(n)<-xyzmatrix(n)
stopifnot(isTRUE(
  all.equal(xyzmatrix(n),xyzmatrix(Cell07PNs[[1]])))
))
```
[.neuronlistfh  

*Extract from neuronlistfh object or its attached data.frame*

---

**Description**

[.neuronlistfh extracts either a sublist from a neuronlistfh (converting it to a regular in memory list in the process) or its attached data.frame.

**Usage**

```r
## S3 method for class 'neuronlistfh'
x[i, j, drop]
```

**Arguments**

- `x`: A neuronlistfh object
- `i, j`: elements to extract or replace. Numeric, logical or character or, for the `[]` get method, empty. See details and the help for `[.data.frame`.
- `drop`: logical. If `TRUE` the result is coerced to the lowest possible dimension. The default is to drop if only one column is left, but **not** to drop if only one row is left.

**Details**

Note that if `i` is a numeric or logical indexing vector, it will be converted internally to a vector of names by using the (sorted) names of the objects in `x` (i.e. `names(x)[i]`)

**Value**

A new in-memory neuronlist or when using two subscripts, a `data.frame` - see examples.

**See Also**

- `neuronlistfh`, `.[.neuronlist`, `.[.data.frame`, `<-.data.frame`
- Other neuronlistfh: `neuronlistfh()`, `read.neuronlistfh()`, `remotesync()`, `write.neuronlistfh()`

**Examples**

```r
# make a test neuronlistfh backed by a temporary folder on disk
tf=tempfile('kcs20fh')
kcs20fh<-as.neuronlistfh(kcs20, dbdir=tf)

# get first neurons as an in memory neuronlist
class(kcs20fh[1:3])

# extract attached data.frame
str(kcs20fh[,])
# or part of the data.frame
```
str(kcs20fh[1:2,1:3])

# data.frame assignment (this one changes nothing)
kcs20fh[1:2, 'gene_name'] <- kcs20fh[1:2, 'gene_name']

# clean up
unlink(tf, recursive=TRUE)
Index

* amira
  amiratype, 14
  is.amiramesh, 56
  read.amiramesh, 112
  read.hxsurf, 114
  write.hxsurf, 156

* cmtk-commandline
  cmtk.dof2mat, 28
  cmtk.mat2dof, 30

* cmtk-geometry
  affmat2cmtkparams, 10
  cmtk.dof2mat, 28
  cmtk.mat2dof, 30
  cmtkparams2affmat, 36

* cmtk-io
  cmtk.extract_affine, 29
  read.cmtk, 113
  read.cmtkreg, 114
  write.cmtk, 155
  write.cmtkreg, 156

* hxsurf
  as.hxsurf, 16
  as.mesh3d, 18
  materials, 63
  plot3d.hxsurf, 98
  read.hxsurf, 114
  subset.hxsurf, 144
  write.hxsurf, 156

* im3d
  as.im3d, 17
  boundingbox, 21
  im3d, 47
  im3d-coords, 49
  im3d-io, 50
  imexpand.grid, 53
  imslice, 54
  is.im3d, 58
  mask, 62
  origin, 89
  projection, 106
  threshold, 150
  unmask, 152
  voxdims, 153

* nat-data
  Cell07PNs, 24
  kcs20, 61
  MBL.surf, 64

* neuronlistfh
  [.neuronlistfh, 172
  neuronlistfh, 75
  read.neuronlistfh, 122
  remotesync, 128
  write.neuronlistfh, 158

* neuronlist
  *.neuronlist, 9
  is.neuronlist, 59
  neuronlist, 71
  neuronlist-dataframe-methods, 72
  neuronlistfh, 75
  nlapply, 80
  read.neurons, 123
  write.neurons, 159

* neuron
  neuron, 68
  ngraph, 78
  plot.neuron, 91
  potential_synapses, 105
  prune, 108
  resample, 129
  rootpoints, 130
  spine, 139
  subset.neuron, 145

* package
  nat-package, 5
  *.dotprops, 7
  *.neuron, 8, 132
  *.neuronlist, 6, 9, 59, 72, 74, 77, 82, 124, 160
subset, 142
subset.data.frame, 148
subset.dotprops, 5, 109, 142, 146, 148
subset.hxsurf, 16, 19, 64, 98, 115, 142, 144, 157
subset.neuron, 5, 70, 80, 92, 106, 108, 111, 130, 140, 142, 145, 148
subset.neuronlist, 6, 44, 46, 142, 147
summary(summary.neuronlist), 149
summary.neuronlist, 149
system2, 27, 28
tail, 74
tail(neuronlist-dataframe-methods), 72
terrain.colors, 52
threshold, 18, 23, 48, 49, 51, 53, 55, 58, 63, 89, 107, 150, 152, 154
tmesh3d, 19
topo.colors, 52
union, 151, 151
unmask, 18, 23, 48, 49, 51, 53, 55, 58, 63, 89, 107, 151, 152, 154
vcgClean, 104
vector, 150
view3d, 89
voxdims, 6, 18, 23, 48, 49, 51, 53, 55, 58, 63, 89, 107, 151, 152, 153

with, 74
with(neuronlist-dataframe-methods), 72
with.neuronlist, 24, 61
write.amiramesh, 154
write.cmtk, 29, 113, 114, 155, 156
write.cmtkreg, 29, 38, 113, 114, 155, 156
write.hxsurf, 14, 16, 19, 57, 64, 98, 113, 115, 144, 156
write.im3d, 6
write.im3d(im3d-io), 50
write.landmarks(read.landmarks), 116
write.neuron, 43, 120, 157, 158, 160
write.neuronlistfh, 77, 123, 129, 158, 172
write.neurons, 6, 10, 59, 72, 74, 77, 82, 124, 159
write.nrrd, 51, 126, 154, 155, 161
write.vtk, 163
xform, 6, 7, 65, 66, 128, 138, 164, 167, 168
xformimage, 164, 167
xformpoints, 164–166, 168, 168
xformpoints.cmtkreg, 127
xmlParse, 118, 120, 121
xyzmatrix, 22, 111, 170, 171
xyzmatrix<= (xyzmatrix), 170
xyzpos, 6, 55
xyzpos(im3d-coords), 49