

# Package ‘semTools’

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**Title** Useful Tools for Structural Equation Modeling

**Description** Provides useful tools for structural equation modeling.

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**Imports** methods

**Suggests** MASS, foreign, parallel, boot, Amelia, mice, GPArotation, mnormt, blavaan, emmeans, testthat

**License** GPL (>= 2)

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**LazyLoad** yes

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## Description

Automatically add auxiliary variables to a lavaan model when using full information maximum likelihood (FIML) to handle missing data

## Usage

```
auxiliary(model, data, aux, fun, ...)
lavaan.auxiliary(model, data, aux, ...)
cfa.auxiliary(model, data, aux, ...)
sem.auxiliary(model, data, aux, ...)
growth.auxiliary(model, data, aux, ...)
```

## Arguments

model	The analysis model can be specified with 1 of 2 objects: <ol style="list-style-type: none"> <li>lavaan <a href="#">model.syntax</a> specifying a hypothesized model <i>without</i> mention of auxiliary variables in aux</li> <li>a parameter table, as returned by <a href="#">parTable</a>, specifying the target model <i>without</i> auxiliary variables. This option requires these columns (and silently ignores all others): c("lhs", "op", "rhs", "user", "group", "free", "label", "plabel", "start")</li> </ol>
data	data.frame that includes auxiliary variables as well as any observed variables in the model
aux	character. Names of auxiliary variables to add to model
fun	character. Name of a specific lavaan function used to fit model to data (i.e., "lavaan", "cfa", "sem", or "growth"). Only required for auxiliary.
...	additional arguments to pass to <a href="#">lavaan</a> .

## Details

These functions are wrappers around the corresponding lavaan functions. You can use them the same way you use [lavaan](#), but you *must* pass your full data.frame to the data argument. Because the saturated-correlates approaches (Enders, 2008) treats exogenous variables as random, `fixed.x` must be set to FALSE. Because FIML requires continuous data (although nonnormality corrections can still be requested), no variables in the model nor auxiliary variables specified in aux can be declared as ordered.

**Value**

a fitted `lavaan` object. Additional information is stored as a list in the `@external` slot:

- `baseline.model`. a fitted `lavaan` object. Results of fitting an appropriate independence model for the calculation of incremental fit indices (e.g., CFI, TLI) in which the auxiliary variables remain saturated, so only the target variables are constrained to be orthogonal. See Examples for how to send this baseline model to `fitMeasures`.
- `aux`. The character vector of auxiliary variable names.
- `baseline.syntax`. A character vector generated within the auxiliary function, specifying the `baseline.model` syntax.

**Author(s)**

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

Enders, C. K. (2008). A note on the use of missing auxiliary variables in full information maximum likelihood-based structural equation models. *Structural Equation Modeling*, 15(3), 434–448. doi:10.1080/10705510802154307

**Examples**

```
dat1 <- lavaan::HolzingerSwineford1939
set.seed(12345)
dat1$z <- rnorm(nrow(dat1))
dat1$x5 <- ifelse(dat1$z < quantile(dat1$z, .3), NA, dat1$x5)
dat1$x9 <- ifelse(dat1$z > quantile(dat1$z, .8), NA, dat1$x9)

targetModel <- "
  visual  =~ x1 + x2 + x3
  textual =~ x4 + x5 + x6
  speed   =~ x7 + x8 + x9
"

## works just like cfa(), but with an extra "aux" argument
fitaux1 <- cfa.auxiliary(targetModel, data = dat1, aux = "z",
  missing = "fiml", estimator = "mlr")

## with multiple auxiliary variables and multiple groups
fitaux2 <- cfa.auxiliary(targetModel, data = dat1, aux = c("z", "ageyr", "grade"),
  group = "school", group.equal = "loadings")

## calculate correct incremental fit indices (e.g., CFI, TLI)
fitMeasures(fitaux2, fit.measures = c("cfi", "tli"))
## NOTE: lavaan will use the internally stored baseline model, which
##       is the independence model plus saturated auxiliary parameters
lavInspect(fitaux2@external$baseline.model, "free")
```

---

 BootMiss-class

 Class For the Results of Bollen-Stine Bootstrap with Incomplete Data
 

---

## Description

This class contains the results of Bollen-Stine bootstrap with missing data.

## Usage

```
## S4 method for signature 'BootMiss'
show(object)

## S4 method for signature 'BootMiss'
summary(object)

## S4 method for signature 'BootMiss'
hist(x, ..., alpha = 0.05, nd = 2,
     printLegend = TRUE, legendArgs = list(x = "topleft"))
```

## Arguments

object, x	object of class BootMiss
...	Additional arguments to pass to <a href="#">hist</a>
alpha	alpha level used to draw confidence limits
nd	number of digits to display
printLegend	logical. If TRUE (default), a legend will be printed with the histogram
legendArgs	list of arguments passed to the <a href="#">legend</a> function. The default argument is a list placing the legend at the top-left of the figure.

## Value

The `hist` method returns a list of length == 2, containing the arguments for the call to `hist` and the arguments to the call for `legend`, respectively.

## Slots

time	A list containing 2 difftime objects (transform and fit), indicating the time elapsed for data transformation and for fitting the model to bootstrap data sets, respectively.
transData	Transformed data
bootDist	The vector of $\chi^2$ values from bootstrap data sets fitted by the target model
origChi	The $\chi^2$ value from the original data set
df	The degree of freedom of the model
bootP	The $p$ value comparing the original $\chi^2$ with the bootstrap distribution

## Objects from the Class

Objects can be created via the `bsBootMiss` function.

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## See Also

[bsBootMiss](#)

## Examples

```
# See the example from the bsBootMiss function
```

---

bsBootMiss

*Bollen-Stine Bootstrap with the Existence of Missing Data*

---

## Description

Implement the Bollen and Stine's (1992) Bootstrap when missing observations exist. The implemented method is proposed by Savalei and Yuan (2009). This can be used in two ways. The first and easiest option is to fit the model to incomplete data in lavaan using the FIML estimator, then pass that lavaan object to `bsBootMiss`.

## Usage

```
bsBootMiss(x, transformation = 2, nBoot = 500, model, rawData, Sigma, Mu,
  group, ChiSquared, EMcov, writeTransData = FALSE, transDataOnly = FALSE,
  writeBootData = FALSE, bootSamplesOnly = FALSE, writeArgs, seed = NULL,
  suppressWarn = TRUE, showProgress = TRUE, ...)
```

## Arguments

<code>x</code>	A target lavaan object used in the Bollen-Stine bootstrap
<code>transformation</code>	The transformation methods in Savalei and Yuan (2009). There are three methods in the article, but only the first two are currently implemented here. Use <code>transformation = 1</code> when there are few missing data patterns, each of which has a large size, such as in a planned-missing-data design. Use <code>transformation = 2</code> when there are more missing data patterns. The currently unavailable transformation = 3 would be used when several missing data patterns have $n = 1$ .
<code>nBoot</code>	The number of bootstrap samples.
<code>model</code>	Optional. The target model if <code>x</code> is not provided.
<code>rawData</code>	Optional. The target raw data set if <code>x</code> is not provided.

Sigma	Optional. The model-implied covariance matrix if $x$ is not provided.
Mu	Optional. The model-implied mean vector if $x$ is not provided.
group	Optional character string specifying the name of the grouping variable in <code>rawData</code> if $x$ is not provided.
ChiSquared	Optional. The model's $\chi^2$ test statistic if $x$ is not provided.
EMcov	Optional, if $x$ is not provided. The EM (or Two-Stage ML) estimated covariance matrix used to speed up Transformation 2 algorithm.
writeTransData	Logical. If TRUE, the transformed data set is written to a text file, <code>transDataOnly</code> is set to TRUE, and the transformed data is returned invisibly.
transDataOnly	Logical. If TRUE, the result will provide the transformed data only.
writeBootData	Logical. If TRUE, the stacked bootstrap data sets are written to a text file, <code>bootSamplesOnly</code> is set to TRUE, and the list of bootstrap data sets are returned invisibly.
bootSamplesOnly	Logical. If TRUE, the result will provide bootstrap data sets only.
writeArgs	Optional list. If <code>writeBootData = TRUE</code> or <code>writeBootData = TRUE</code> , user can pass arguments to the <code>write.table</code> function as a list. Some default values are provided: <code>file = "bootstrappedSamples.dat"</code> , <code>row.names = FALSE</code> , and <code>na = "-999"</code> , but the user can override all of these by providing other values for those arguments in the <code>writeArgs</code> list.
seed	The seed number used in randomly drawing bootstrap samples.
suppressWarn	Logical. If TRUE, warnings from lavaan function will be suppressed when fitting the model to each bootstrap sample.
showProgress	Logical. Indicating whether to display a progress bar while fitting models to bootstrap samples.
...	The additional arguments in the <code>lavaan</code> function. See also <code>lavOptions</code>

## Details

The second is designed for users of other software packages (e.g., LISREL, EQS, Amos, or Mplus). Users can import their data,  $\chi^2$  value, and model-implied moments from another package, and they have the option of saving (or writing to a file) either the transformed data or bootstrapped samples of that data, which can be analyzed in other programs. In order to analyze the bootstrapped samples and return a  $p$  value, users of other programs must still specify their model using lavaan syntax.

## Value

As a default, this function returns a `BootMiss` object containing the results of the bootstrap samples. Use `show`, `summary`, or `hist` to examine the results. Optionally, the transformed data set is returned if `transDataOnly = TRUE`. Optionally, the bootstrap data sets are returned if `bootSamplesOnly = TRUE`.

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

Syntax for transformations borrowed from <http://www2.psych.ubc.ca/~vsavalei/>



## References

Bollen, K. A., & Stine, R. A. (1992). Bootstrapping goodness-of-fit measures in structural equation models. *Sociological Methods & Research*, *21*(2), 205–229. doi:10.1177/0049124192021002004

Savalei, V., & Yuan, K.-H. (2009). On the model-based bootstrap with missing data: Obtaining a p-value for a test of exact fit. *Multivariate Behavioral Research*, *44*(6), 741–763. doi:10.1080/00273170903333590

## See Also

[BootMiss](#)

## Examples

```
## Not run:
dat1 <- HolzingerSwineford1939
dat1$x5 <- ifelse(dat1$x1 <= quantile(dat1$x1, .3), NA, dat1$x5)
dat1$x9 <- ifelse(is.na(dat1$x5), NA, dat1$x9)

targetModel <- "
visual =~ x1 + x2 + x3
textual =~ x4 + x5 + x6
speed =~ x7 + x8 + x9
"

targetFit <- sem(targetModel, dat1, meanstructure = TRUE, std.lv = TRUE,
                missing = "fiml", group = "school")
summary(targetFit, fit = TRUE, standardized = TRUE)

# The number of bootstrap samples should be much higher.
temp <- bsBootMiss(targetFit, transformation = 1, nBoot = 10, seed = 31415)

temp
summary(temp)
hist(temp)
hist(temp, printLegend = FALSE) # suppress the legend
## user can specify alpha level (default: alpha = 0.05), and the number of
## digits to display (default: nd = 2). Pass other arguments to hist(...),
## or a list of arguments to legend() via "legendArgs"
hist(temp, alpha = .01, nd = 3, xlab = "something else", breaks = 25,
     legendArgs = list("bottomleft", box.lty = 2))

## End(Not run)
```

## Description

This is a utility function used to calculate the "D2" statistic for pooling test statistics across multiple imputations. This function is called by several functions used for `lavaan.mi` objects, such as `lavTestLRT.mi`, `lavTestWald.mi`, and `lavTestScore.mi`. But this function can be used for any general scenario because it only requires a vector of  $\chi^2$  statistics (one from each imputation) and the degrees of freedom for the test statistic. See Li, Meng, Raghunathan, & Rubin (1991) and Enders (2010, chapter 8) for details about how it is calculated.

## Usage

```
calculate.D2(w, DF = 0L, asymptotic = FALSE)
```

## Arguments

<code>w</code>	numeric vector of Wald $\chi^2$ statistics. Can also be Wald $z$ statistics, which will be internally squared to make $\chi^2$ statistics with one <i>df</i> (must set <code>DF = 0L</code> ).
<code>DF</code>	degrees of freedom ( <i>df</i> ) of the $\chi^2$ statistics. If <code>DF = 0L</code> (default), <code>w</code> is assumed to contain $z$ statistics, which will be internally squared.
<code>asymptotic</code>	logical. If <code>FALSE</code> (default), the pooled test will be returned as an $F$ -distributed statistic with numerator ( <code>df1</code> ) and denominator ( <code>df2</code> ) degrees of freedom. If <code>TRUE</code> , the pooled $F$ statistic will be multiplied by its <code>df1</code> on the assumption that its <code>df2</code> is sufficiently large enough that the statistic will be asymptotically $\chi^2$ distributed with <code>df1</code> .

## Value

A numeric vector containing the test statistic, *df*, its *p* value, and 2 missing-data diagnostics: the relative increase in variance (RIV, or average for multiparameter tests: ARIV) and the fraction missing information (FMI = ARIV / (1 + ARIV)).

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## References

Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.

Li, K.-H., Meng, X.-L., Raghunathan, T. E., & Rubin, D. B. (1991). Significance levels from repeated *p*-values with multiply-imputed data. *Statistica Sinica*, 1(1), 65–92. Retrieved from <https://www.jstor.org/stable/24303994>

## See Also

[lavTestLRT.mi](#), [lavTestWald.mi](#), [lavTestScore.mi](#)

**Examples**

```
## generate a vector of chi-squared values, just for example
DF <- 3 # degrees of freedom
M <- 20 # number of imputations
CHI <- rchisq(M, DF)

## pool the "results"
calculate.D2(CHI, DF) # by default, an F statistic is returned
calculate.D2(CHI, DF, asymptotic = TRUE) # asymptotically chi-squared

## generate standard-normal values, for an example of Wald z tests
Z <- rnorm(M)
calculate.D2(Z) # default DF = 0 will square Z to make chisq(DF = 1)
## F test is equivalent to a t test with the denominator DF
```

---

chisqSmallN

*Small-N correction for  $\chi^2$  test statistic*


---

**Description**

Calculate small- $N$  corrections for  $\chi^2$  model-fit test statistic to adjust for small sample size (relative to model size).

**Usage**

```
chisqSmallN(fit0, fit1 = NULL, smallN.method = if (is.null(fit1))
  c("swain", "yuan.2015") else "yuan.2005", ..., omitimps = c("no.conv",
  "no.se"))
```

**Arguments**

fit0, fit1	<a href="#">lavaan</a> object(s) provided after running the <code>cfa</code> , <code>sem</code> , <code>growth</code> , or <code>lavaan</code> functions. <a href="#">lavaan.mi</a> object(s) also accepted.
smallN.method	character indicating the small- $N$ correction method to use. Multiple may be chosen (all of which assume normality), as described in Shi et al. (2018): <code>c("swain", "yuan.2015", "yuan.2005", "bartlett")</code> . Users may also simply select "all".
...	Additional arguments to the <a href="#">lavTestLRT</a> or <a href="#">lavTestLRT.mi</a> functions. Ignored when <code>is.null(fit1)</code> .
omitimps	character vector specifying criteria for omitting imputations from pooled results. Ignored unless <code>fit0</code> (and optionally <code>fit1</code> ) is a <a href="#">lavaan.mi</a> object. See <a href="#">lavTestLRT.mi</a> for a description of options and defaults.

## Details

Four finite-sample adjustments to the chi-squared statistic are currently available, all of which are described in Shi et al. (2018). These all assume normally distributed data, and may not work well with severely nonnormal data. Deng et al. (2018, section 4) review proposed small- $N$  adjustments that do not assume normality, which rarely show promise, so they are not implemented here. This function currently will apply small- $N$  adjustments to scaled test statistics with a warning that they do not perform well (Deng et al., 2018).

## Value

A list of numeric vectors: one for the originally requested statistic(s), along with one per requested smallN.method. All include the the (un)adjusted test statistic, its  $df$ , and the  $p$  value for the test under the null hypothesis that the model fits perfectly (or that the 2 models have equivalent fit). The adjusted chi-squared statistic(s) also include(s) the scaling factor for the small- $N$  adjustment.

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## References

- Deng, L., Yang, M., & Marcoulides, K. M. (2018). Structural equation modeling with many variables: A systematic review of issues and developments. *Frontiers in Psychology*, 9, 580. doi:10.3389/fpsyg.2018.00580
- Shi, D., Lee, T., & Terry, R. A. (2018). Revisiting the model size effect in structural equation modeling. *Structural Equation Modeling*, 25(1), 21–40. doi:10.1080/10705511.2017.1369088

## Examples

```
HS.model <- '
  visual =~ x1 + b1*x2 + x3
  textual =~ x4 + b2*x5 + x6
  speed  =~ x7 + b3*x8 + x9
'

fit1 <- cfa(HS.model, data = HolzingerSwineford1939[1:50,])
## test a single model (implicitly compared to a saturated model)
chisqSmallN(fit1)

## fit a more constrained model
fit0 <- cfa(HS.model, data = HolzingerSwineford1939[1:50,],
            orthogonal = TRUE)
## compare 2 models
chisqSmallN(fit1, fit0)
```

---

clipboard	<i>Copy or save the result of lavaan or FitDiff objects into a clipboard or a file</i>
-----------	--

---

### Description

Copy or save the result of lavaan or [FitDiff](#) object into a clipboard or a file. From the clipboard, users may paste the result into the Microsoft Excel or spreadsheet application to create a table of the output.

### Usage

```
clipboard(object, what = "summary", ...)

saveFile(object, file, what = "summary", tableFormat = FALSE,
         fit.measures = "default", writeArgs = list(), ...)
```

### Arguments

object	The lavaan or <a href="#">FitDiff</a> object
what	The attributes of the lavaan object to be copied in the clipboard. "summary" is to copy the screen provided from the summary function. "mifit" is to copy the result from the <a href="#">miPowerFit</a> function. Other attributes listed in the inspect method in the <a href="#">lavaan-class</a> could also be used, such as "coef", "se", "fit", "samp", and so on. For the The <a href="#">FitDiff</a> object, this argument is not active yet.
...	Additional argument listed in the <a href="#">miPowerFit</a> function (for lavaan object only).
file	A file name used for saving the result
tableFormat	If TRUE, save the result in the table format using tabs for separation. Otherwise, save the result as the output screen printed in the R console.
fit.measures	character vector specifying names of fit measures returned by <a href="#">fitMeasures</a> to be copied/saved. Only relevant if object is class <a href="#">FitDiff</a> .
writeArgs	list of additional arguments to be passed to <a href="#">write.table</a>

### Value

The resulting output will be saved into a clipboard or a file. If using the clipboard function, users may paste it in the other applications.

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**Examples**

```
## Not run:
library(lavaan)
HW.model <- ' visual =~ x1 + c1*x2 + x3
             textual =~ x4 + c1*x5 + x6
             speed  =~ x7 + x8 + x9 '

fit <- cfa(HW.model, data=HolzingerSwineford1939, group="school", meanstructure=TRUE)

# Copy the summary of the lavaan object
clipboard(fit)

# Copy the modification indices and the model fit from the miPowerFit function
clipboard(fit, "mifit")

# Copy the parameter estimates
clipboard(fit, "coef")

# Copy the standard errors
clipboard(fit, "se")

# Copy the sample statistics
clipboard(fit, "samp")

# Copy the fit measures
clipboard(fit, "fit")

# Save the summary of the lavaan object
saveFile(fit, "out.txt")

# Save the modification indices and the model fit from the miPowerFit function
saveFile(fit, "out.txt", "mifit")

# Save the parameter estimates
saveFile(fit, "out.txt", "coef")

# Save the standard errors
saveFile(fit, "out.txt", "se")

# Save the sample statistics
saveFile(fit, "out.txt", "samp")

# Save the fit measures
saveFile(fit, "out.txt", "fit")

## End(Not run)
```

**Description**

This function builds upon the [quark](#) function to provide a final dataset comprised of the original dataset provided to [quark](#) and enough principal components to be able to account for a certain level of variance in the data.

**Usage**

```
combinequark(quark, percent)
```

**Arguments**

quark	Provide the <a href="#">quark</a> object that was returned. It should be a list of objects. Make sure to include it in its entirety.
percent	Provide a percentage of variance that you would like to have explained. That many components (columns) will be extracted and kept with the output dataset. Enter this variable as a number WITHOUT a percentage sign.

**Value**

The output of this function is the original dataset used in [quark](#) combined with enough principal component scores to be able to account for the amount of variance that was requested.

**Author(s)**

Steven R. Chesnut (University of Southern Mississippi <[Steven.Chesnut@usm.edu](mailto:Steven.Chesnut@usm.edu)>)

**See Also**

[quark](#)

**Examples**

```
set.seed(123321)
dat <- HolzingerSwineford1939[,7:15]
misspat <- matrix(runif(nrow(dat) * 9) < 0.3, nrow(dat))
dat[misspat] <- NA
dat <- cbind(HolzingerSwineford1939[,1:3], dat)

quark.list <- quark(data = dat, id = c(1, 2))

final.data <- combinequark(quark = quark.list, percent = 80)
```

---

`compareFit`*Build an object summarizing fit indices across multiple models*

---

### Description

This function will create the template to compare fit indices across multiple fitted lavaan objects. The results can be exported to a clipboard or a file later.

### Usage

```
compareFit(..., nested = TRUE, argsLRT = list(), indices = TRUE,  
  baseline.model = NULL)
```

### Arguments

<code>...</code>	fitted lavaan models or list(s) of lavaan objects. <a href="#">lavaan.mi</a> objects are also accepted, but all models must belong to the same class.
<code>nested</code>	logical indicating whether the models in <code>...</code> are nested. See <a href="#">net</a> for an empirical test of nesting.
<code>argsLRT</code>	list of arguments to pass to <a href="#">lavTestLRT</a> , as well as to <a href="#">lavTestLRT.mi</a> and <a href="#">fitMeasures</a> when comparing <a href="#">lavaan.mi</a> models.
<code>indices</code>	logical indicating whether to return fit indices from the <a href="#">fitMeasures</a> function.
<code>baseline.model</code>	optional fitted <a href="#">lavaan</a> model passed to <a href="#">fitMeasures</a> to calculate incremental fit indices.

### Value

A [FitDiff](#) object that saves model fit comparisons across multiple models. If the models are not nested, only fit indices for each model are returned. If the models are nested, the differences in fit indices are additionally returned, as well as test statistics comparing each sequential pair of models (ordered by their degrees of freedom).

### Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)  
Sunthud Pornprasertmanit (<psunthud@gmail.com>)

### See Also

[FitDiff](#), [clipboard](#)



**Examples**

```

HS.model <- ' visual =~ x1 + x2 + x3
            textual =~ x4 + x5 + x6
            speed =~ x7 + x8 + x9 '

fit1 <- cfa(HS.model, data = HolzingerSwineford1939)

## non-nested model
m2 <- ' f1 =~ x1 + x2 + x3 + x4
      f2 =~ x5 + x6 + x7 + x8 + x9 '
fit2 <- cfa(m2, data = HolzingerSwineford1939)
compareFit(fit1, fit2, nested = FALSE)

## nested model comparisons:
out <- measurementInvariance(model = HS.model, data = HolzingerSwineford1939,
                             group = "school", quiet = TRUE)

compareFit(out)

## Not run:
## also applies to lavaan.mi objects (fit model to multiple imputations)
set.seed(12345)
HSMiss <- HolzingerSwineford1939[, paste("x", 1:9, sep = "")]
HSMiss$x5 <- ifelse(HSMiss$x1 <= quantile(HSMiss$x1, .3), NA, HSMiss$x5)
HSMiss$x9 <- ifelse(is.na(HSMiss$x5), NA, HSMiss$x9)
HSMiss$school <- HolzingerSwineford1939$school
HS.amelia <- amelia(HSMiss, m = 20, noms = "school")
imps <- HS.amelia$imputations

## request robust test statistics
mgfit2 <- cfa.mi(HS.model, data = imps, group = "school", estimator = "mlm")
mgfit1 <- cfa.mi(HS.model, data = imps, group = "school", estimator = "mlm",
               group.equal = "loadings")
mgfit0 <- cfa.mi(HS.model, data = imps, group = "school", estimator = "mlm",
               group.equal = c("loadings", "intercepts"))

## request the strictly-positive robust test statistics
compareFit(scalar = mgfit0, metric = mgfit1, config = mgfit2,
          argsLRT = list(asymptotic = TRUE,
                        method = "satorra.bentler.2010"))

## End(Not run)

```

**Description**

A simulated data set with 2 independent factors and 1 dependent factor where each factor has three indicators

**Usage**

```
dat2way
```

**Format**

A data.frame with 500 observations of 9 variables.

- x1** The first indicator of the first independent factor
- x2** The second indicator of the first independent factor
- x3** The third indicator of the first independent factor
- x4** The first indicator of the second independent factor
- x5** The second indicator of the second independent factor
- x6** The third indicator of the second independent factor
- x7** The first indicator of the dependent factor
- x8** The second indicator of the dependent factor
- x9** The third indicator of the dependent factor

**Source**

Data were generated by the [mvrnorm](#) function in the MASS package.

**Examples**

```
head(dat2way)
```

---

dat3way

*Simulated Dataset to Demonstrate Three-way Latent Interaction*

---

**Description**

A simulated data set with 3 independent factors and 1 dependent factor where each factor has three indicators

**Usage**

```
dat3way
```

**Format**

A data . frame with 500 observations of 12 variables.

- x1** The first indicator of the first independent factor
- x2** The second indicator of the first independent factor
- x3** The third indicator of the first independent factor
- x4** The first indicator of the second independent factor
- x5** The second indicator of the second independent factor
- x6** The third indicator of the second independent factor
- x7** The first indicator of the third independent factor
- x8** The second indicator of the third independent factor
- x9** The third indicator of the third independent factor
- x10** The first indicator of the dependent factor
- x11** The second indicator of the dependent factor
- x12** The third indicator of the dependent factor

**Source**

Data were generated by the [mvrnorm](#) function in the MASS package.

**Examples**

```
head(dat3way)
```

---

datCat	<i>Simulated Data set to Demonstrate Categorical Measurement Invariance</i>
--------	---

---

**Description**

A simulated data set with 2 factors with 4 indicators each separated into two groups

**Usage**

```
datCat
```

**Format**

A data . frame with 200 observations of 9 variables.

- g** Sex of respondents
- u1** Indicator 1
- u2** Indicator 2
- u3** Indicator 3

- u4** Indicator 4
- u5** Indicator 5
- u6** Indicator 6
- u7** Indicator 7
- u8** Indicator 8

### Source

Data were generated using the lavaan package.

### Examples

```
head(datCat)
```

---

discriminantValidity *Calculate discriminant validity statistics*

---

### Description

Calculate discriminant validity statistics based on a fitted lavaan object

### Usage

```
discriminantValidity(object, cutoff = 0.9, merge = FALSE, level = 0.95)
```

### Arguments

- |        |   |
|--------|---|
| object | The <a href="#">lavaan</a> model object returned by the <a href="#">cfa</a> function.                   |
| cutoff | A cutoff to be used in the constrained models in likelihood ratio tests.                                |
| merge  | Whether the constrained models should be constructed by merging two factors as one. Implies cutoff = 1. |
| level  | The confidence level required.  |

### Details

Evaluated on the measurement scale level, discriminant validity is commonly evaluated by checking if each pair of latent correlations is sufficiently below one (in absolute value) that the latent variables can be thought of representing two distinct constructs.

discriminantValidity function calculates two sets of statistics that are commonly used in discriminant validity evaluation. The first set are factor correlation estimates and their confidence intervals. The second set is a series of nested model tests, where the baseline model is compared against a set of constrained models that are constructed by constraining each factor correlation to the specified cutoff one at a time.

The function assume that the object is set of confirmatory factor analysis results where the latent variables are scaled by fixing their variances to 1s. If the model is not a CFA model, the function will

calculate the statistics for the correlations among exogenous latent variables, but for the *residual* variances with endogenous variables. If the latent variables are scaled in some other way (e.g. fixing the first loadings), the function issues a warning and re-estimates the model by fixing latent variances to 1 (and estimating all loadings) so that factor covariances are already estimated as correlations.

The likelihood ratio tests are done by comparing the original baseline model against more constrained alternatives. By default, these alternatives are constructed by fixing each correlation at a time to a cutoff value. The typical purpose of this test is to demonstrate that the estimated factor correlation is well below the cutoff and a significant *chi*<sup>2</sup> statistic thus indicates support for discriminant validity. In some cases, the original correlation estimate may already be greater than the cutoff, making it redundant to fit a "restricted" model. When this happens, the likelihood ratio test will be replaced by comparing the baseline model against itself. For correlations that are estimated to be negative, a negation of the cutoff is used in the constrained model.

Another alternative is to do a nested model comparison against a model where two factors are merged as one by setting the `merge` argument to `TRUE`. In this comparison, the constrained model is constructed by removing one of the correlated factors from the model and assigning its indicators to the factor that remains in the model.

### Value

A data frame of latent variable correlation estimates, their confidence intervals, and a likelihood ratio tests against constrained models. with the following attributes:

**baseline** The baseline model after possible rescaling.

**constrained** A list of the fitted constrained models used in the likelihood ratio test.

### Examples

```
library(lavaan)

HS.model <- ' visual =~ x1 + x2 + x3
            textual =~ x4 + x5 + x6
            speed  =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data = HolzingerSwineford1939)
discriminantValidity(fit)
discriminantValidity(fit, merge = TRUE)
```

### Description

This class contains the results of rotated exploratory factor analysis

**Usage**

```
## S4 method for signature 'EFA'
show(object)

## S4 method for signature 'EFA'
summary(object, suppress = 0.1, sort = TRUE)
```

**Arguments**

object	object of class EFA
suppress	any standardized loadings less than the specified value will not be printed to the screen
sort	logical. If TRUE (default), factor loadings will be sorted by their size in the console output

**Slots**

loading Rotated standardized factor loading matrix  
 rotate Rotation matrix  
 gradRotate gradient of the objective function at the rotated loadings  
 convergence Convergence status  
 phi: Factor correlation matrix. Will be an identity matrix if orthogonal rotation is used.  
 se Standard errors of the rotated standardized factor loading matrix  
 method Method of rotation  
 call The command used to generate this object

**Objects from the Class**

Objects can be created via the [orthRotate](#) or [oblqRotate](#) function.

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**See Also**

[efaUnrotate](#); [orthRotate](#); [oblqRotate](#)

**Examples**

```
unrotated <- efaUnrotate(HolzingerSwineford1939, nf = 3,
                        varList = paste0("x", 1:9), estimator = "mlr")
summary(unrotated, std = TRUE)
lavInspect(unrotated, "std")

# Rotated by Quartimin
```

```
rotated <- obliqueRotate(unrotated, method = "quartimin")
summary(rotated)
```

---

 efa.ekc

*Empirical Kaiser criterion*


---

## Description

Identify the number of factors to extract based on the Empirical Kaiser Criterion (EKC). The analysis can be run on a `data.frame` or data matrix (`data`), or on a correlation or covariance matrix (`sample.cov`) and the sample size (`sample.nobs`). A `data.frame` is returned with two columns: the eigenvalues from your data or covariance matrix and the reference eigenvalues. The number of factors suggested by the Empirical Kaiser Criterion (i.e. the sample eigenvalues greater than the reference eigenvalues), and the number of factors suggested by the original Kaiser Criterion (i.e. sample eigenvalues > 1) is printed above the output.

## Usage

```
efa.ekc(data = NULL, sample.cov = NULL, sample.nobs = NULL,
        missing = "default", ordered = NULL, plot = TRUE)
```

## Arguments

<code>data</code>	A <code>data.frame</code> or data matrix containing columns of variables to be factor-analyzed.
<code>sample.cov</code>	A covariance or correlation matrix can be used, instead of <code>data</code> , to estimate the eigenvalues.
<code>sample.nobs</code>	Number of observations (i.e. sample size) if <code>is.null(data)</code> and <code>sample.cov</code> is used.
<code>missing</code>	If "listwise", cases with missing values are removed listwise from the data frame. If "direct" or "ml" or "fiml" and the estimator is maximum likelihood, an EM algorithm is used to estimate the unrestricted covariance matrix (and mean vector). If "pairwise", pairwise deletion is used. If "default", the value is set depending on the estimator and the mimic option (see details in <a href="#">lavCor</a> ).
<code>ordered</code>	Character vector. Only used if object is a <code>data.frame</code> . Treat these variables as ordered (ordinal) variables. Importantly, all other variables will be treated as numeric (unless <code>is.ordered == TRUE</code> in <code>data</code> ). (see also <a href="#">lavCor</a> )
<code>plot</code>	logical. Whether to print a scree plot comparing the sample eigenvalues with the reference eigenvalues.

## Value

A `data.frame` showing the sample and reference eigenvalues.

The number of factors suggested by the Empirical Kaiser Criterion (i.e. the sample eigenvalues greater than the reference eigenvalues) is returned as an attribute (see Examples).

The number of factors suggested by the original Kaiser Criterion (i.e. sample eigenvalues > 1) is also printed as a header to the `data.frame`

**Author(s)**

Ylenio Longo (University of Nottingham; <yleniolongo@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

Braeken, J., & van Assen, M. A. L. M. (in press). An empirical Kaiser criterion. *Psychological Methods*, 22(3), 450–466. doi:10.1037/met0000074

**Examples**

```
## Simulate data with 3 factors
model <- '
  f1 =~ .3*x1 + .5*x2 + .4*x3
  f2 =~ .3*x4 + .5*x5 + .4*x6
  f3 =~ .3*x7 + .5*x8 + .4*x9
'
dat <- simulateData(model, seed = 123)
## save summary statistics
myCovMat <- cov(dat)
myCorMat <- cor(dat)
N <- nrow(dat)

## Run the EKC function
(out <- efa.ekc(dat))

## To extract the recommended number of factors using the EKC:
attr(out, "nfactors")

## If you do not have raw data, you can use summary statistics
(x1 <- efa.ekc(sample.cov = myCovMat, sample.nobs = N, plot = FALSE))
(x2 <- efa.ekc(sample.cov = myCorMat, sample.nobs = N, plot = FALSE))
```

---

 efaUnrotate

---

*Analyze Unrotated Exploratory Factor Analysis Model*


---

**Description**

This function will analyze unrotated exploratory factor analysis model. The unrotated solution can be rotated by the [orthRotate](#) and [oblqRotate](#) functions.

**Usage**

```
efaUnrotate(data = NULL, nf, varList = NULL, start = TRUE, aux = NULL, ...)
```



**Arguments**

<code>data</code>	A target data frame
<code>nf</code>	The desired number of factors
<code>varList</code>	Target observed variables. If not specified, all variables in <code>data</code> will be used (or <code>sample.cov</code> if <code>is.null(data)</code> ); see <code>cfa</code> for argument descriptions).
<code>start</code>	Use starting values in the analysis from the <code>factanal</code> function. If <code>FALSE</code> , the starting values from the <code>lavaan</code> package will be used. <code>TRUE</code> is ignored with a warning if the <code>aux</code> argument is used.
<code>aux</code>	The list of auxiliary variables. These variables will be included in the model by the saturated-correlates approach to account for missing information.
<code>...</code>	Other arguments in the <code>cfa</code> function in the <code>lavaan</code> package, such as <code>ordered</code> , <code>se</code> , <code>estimator</code> , or <code>sample.cov</code> and <code>sample.nobs</code> .

**Details**

This function will generate a `lavaan` script for unrotated exploratory factor analysis model such that (1) all factor loadings are estimated, (2) factor variances are fixed to 1, (3) factor covariances are fixed to 0, and (4) the dot products of any pairs of columns in the factor loading matrix are fixed to zero (Johnson & Wichern, 2002). The reason for creating this function to supplement the `factanal` function is that users can enjoy some advanced features from the `lavaan` package, such as scaled  $\chi^2$ , diagonally weighted least squares estimation for ordinal indicators, or full-information maximum likelihood (FIML) to handle incomplete data.

**Value**

A `lavaan` output of unrotated exploratory factor analysis solution.

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**Examples**

```
unrotated <- efaUnrotate(HolzingerSwineford1939, nf = 3,
                        varList = paste0("x", 1:9), estimator = "mlr")
summary(unrotated, std = TRUE)
inspect(unrotated, "std")

dat <- data.frame(HolzingerSwineford1939,
                 z = rnorm(nrow(HolzingerSwineford1939), 0, 1))
unrotated2 <- efaUnrotate(dat, nf = 2, varList = paste0("x", 1:9), aux = "z")
```

---

exLong	<i>Simulated Data set to Demonstrate Longitudinal Measurement Invariance</i>
--------	--

---

**Description**

A simulated data set with 1 factors with 3 indicators in three timepoints

**Usage**

exLong

**Format**

A data.frame with 200 observations of 10 variables.

**sex** Sex of respondents

**y1t1** Indicator 1 in Time 1

**y2t1** Indicator 2 in Time 1

**y3t1** Indicator 3 in Time 1

**y1t2** Indicator 1 in Time 2

**y2t2** Indicator 2 in Time 2

**y3t2** Indicator 3 in Time 2

**y1t3** Indicator 1 in Time 3

**y2t3** Indicator 2 in Time 3

**y3t3** Indicator 3 in Time 3

**Source**

Data were generated using the simsem package.

**Examples**

```
head(exLong)
```

---

`findRMSEApower`*Find the statistical power based on population RMSEA*

---

**Description**

Find the proportion of the samples from the sampling distribution of RMSEA in the alternative hypothesis rejected by the cutoff derived from the sampling distribution of RMSEA in the null hypothesis. This function can be applied for both test of close fit and test of not-close fit (MacCallum, Browne, & Suguwara, 1996)

**Usage**

```
findRMSEApower(rmseao, rmseaA, df, n, alpha = 0.05, group = 1)
```

**Arguments**

<code>rmseao</code>	Null RMSEA
<code>rmseaA</code>	Alternative RMSEA
<code>df</code>	Model degrees of freedom
<code>n</code>	Sample size of a dataset
<code>alpha</code>	Alpha level used in power calculations
<code>group</code>	The number of group that is used to calculate RMSEA.

**Details**

This function find the proportion of sampling distribution derived from the alternative RMSEA that is in the critical region derived from the sampling distribution of the null RMSEA. If `rmseaA` is greater than `rmseao`, the test of close fit is used and the critical region is in the right hand side of the null sampling distribution. On the other hand, if `rmseaA` is less than `rmseao`, the test of not-close fit is used and the critical region is in the left hand side of the null sampling distribution (MacCallum, Browne, & Suguwara, 1996).

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**References**

MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, *1*(2), 130–149. doi:10.1037/1082-989X.1.2.130

**See Also**

- [plotRMSEApower](#) to plot the statistical power based on population RMSEA given the sample size
- [plotRMSEAdist](#) to visualize the RMSEA distributions
- [findRMSEAsamplesize](#) to find the minimum sample size for a given statistical power based on population RMSEA

**Examples**

```
findRMSEApower(rmseA = .05, rmseB = .08, df = 20, n = 200)
```

---

```
findRMSEApowernested    Find power given a sample size in nested model comparison
```

---

**Description**

Find the sample size that the power in rejection the samples from the alternative pair of RMSEA is just over the specified power.

**Usage**

```
findRMSEApowernested(rmseA = NULL, rmseB = NULL, rmse1A,  
                      rmse1B = NULL, dfA, dfB, n, alpha = 0.05, group = 1)
```

**Arguments**

rmseA	The $H_0$ baseline RMSEA
rmseB	The $H_0$ alternative RMSEA (trivial misfit)
rmse1A	The $H_1$ baseline RMSEA
rmse1B	The $H_1$ alternative RMSEA (target misfit to be rejected)
dfA	degree of freedom of the more-restricted model
dfB	degree of freedom of the less-restricted model
n	Sample size
alpha	The alpha level
group	The number of group in calculating RMSEA

**Author(s)**

Bell Clinton  
 Pavel Panko (Texas Tech University; <pavel.panko@ttu.edu>)  
 Sunthud Pornprasertmanit (<psunthud@gmail.com>)

## References

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods, 11*(1), 19–35. doi:10.1037/1082-989X.11.1.19

## See Also

- [plotRMSEApowernested](#) to plot the statistical power for nested model comparison based on population RMSEA given the sample size
- [findRMSEAsamplesizenested](#) to find the minimum sample size for a given statistical power in nested model comparison based on population RMSEA

## Examples

```
findRMSEApowernested(rmseas0A = 0.06, rmseas0B = 0.05, rmsea1A = 0.08,
                    rmsea1B = 0.05, dfA = 22, dfB = 20, n = 200,
                    alpha = 0.05, group = 1)
```

---

findRMSEAsamplesize    *Find the minimum sample size for a given statistical power based on population RMSEA*

---

## Description

Find the minimum sample size for a specified statistical power based on population RMSEA. This function can be applied for both test of close fit and test of not-close fit (MacCallum, Browne, & Suguwara, 1996)

## Usage

```
findRMSEAsamplesize(rmseas0, rmseaA, df, power = 0.8, alpha = 0.05, group = 1)
```

## Arguments

rmseas0	Null RMSEA
rmseaA	Alternative RMSEA
df	Model degrees of freedom
power	Desired statistical power to reject misspecified model (test of close fit) or retain good model (test of not-close fit)
alpha	Alpha level used in power calculations
group	The number of group that is used to calculate RMSEA.

**Details**

This function find the minimum sample size for a specified power based on an iterative routine. The sample size keep increasing until the calculated power from [findRMSEApower](#) function is just over the specified power. If group is greater than 1, the resulting sample size is the sample size per group.

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**References**

MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, *1*(2), 130–149. doi:10.1037/1082-989X.1.2.130

**See Also**

- [plotRMSEApower](#) to plot the statistical power based on population RMSEA given the sample size
- [plotRMSEAdist](#) to visualize the RMSEA distributions
- [findRMSEApower](#) to find the statistical power based on population RMSEA given a sample size

**Examples**

```
findRMSEAsamplesize(rmse0 = .05, rmseaA = .08, df = 20, power = 0.80)
```

---

```
findRMSEAsamplesizenested
```

*Find sample size given a power in nested model comparison*

---

**Description**

Find the sample size that the power in rejection the samples from the alternative pair of RMSEA is just over the specified power.

**Usage**

```
findRMSEAsamplesizenested(rmse0A = NULL, rmsea0B = NULL, rmsea1A,  
  rmsea1B = NULL, dfA, dfB, power = 0.8, alpha = 0.05, group = 1)
```

**Arguments**

rmsea0A	The $H_0$ baseline RMSEA
rmsea0B	The $H_0$ alternative RMSEA (trivial misfit)
rmsea1A	The $H_1$ baseline RMSEA
rmsea1B	The $H_1$ alternative RMSEA (target misfit to be rejected)
dfA	degree of freedom of the more-restricted model.
dfB	degree of freedom of the less-restricted model.
power	The desired statistical power.
alpha	The alpha level.
group	The number of group in calculating RMSEA.

**Author(s)**

Bell Clinton

Pavel Panko (Texas Tech University; <pavel.panko@ttu.edu>)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**References**

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods, 11*(1), 19-35. doi:10.1037/1082-989X.11.1.19

**See Also**

- [plotRMSEApowernested](#) to plot the statistical power for nested model comparison based on population RMSEA given the sample size
- [findRMSEApowernested](#) to find the power for a given sample size in nested model comparison based on population RMSEA

**Examples**

```
findRMSEAsamplesizenested(rmsea0A = 0, rmsea0B = 0, rmsea1A = 0.06,  
                           rmsea1B = 0.05, dfA = 22, dfB = 20, power = 0.80,  
                           alpha = .05, group = 1)
```

FitDiff-class

*Class For Representing A Template of Model Fit Comparisons***Description**

This class contains model fit measures and model fit comparisons among multiple models

**Usage**

```
## S4 method for signature 'FitDiff'
show(object)

## S4 method for signature 'FitDiff'
summary(object, fit.measures = "default", nd = 3)
```

**Arguments**

object	object of class FitDiff
fit.measures	character vector naming fit indices the user can request from <a href="#">fitMeasures</a> . If "default", the fit measures will be c("chisq", "df", "pvalue", "cfi", "tli", "rmsea", "srmr", "aic"). If "all", all available fit measures will be returned.
nd	number of digits printed

**Slots**

name	character. The name of each model
model.class	character. One class to which each model belongs
nested	data.frame. Model fit comparisons between adjacently nested models that are ordered by their degrees of freedom ( <i>df</i> )
fit	data.frame. Fit measures of all models specified in the name slot, ordered by their <i>df</i>

**Objects from the Class**

Objects can be created via the [compareFit](#) function.

**Author(s)**

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**See Also**

[compareFit](#); [clipboard](#)



## Examples

```

HS.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed =~ x7 + x8 + x9 '

out <- measurementInvariance(model = HS.model, data = HolzingerSwineford1939,
                             group = "school", quiet = TRUE)
modelDiff <- compareFit(out)
summary(modelDiff)
summary(modelDiff, fit.measures = "all")
summary(modelDiff, fit.measures = c("aic", "bic"))

## Not run:
## Save results to a file
saveFile(modelDiff, file = "modelDiff.txt")

## Copy to a clipboard
clipboard(modelDiff)

## End(Not run)

```

---

fmi

*Fraction of Missing Information.*


---

## Description

This function estimates the Fraction of Missing Information (FMI) for summary statistics of each variable, using either an incomplete data set or a list of imputed data sets.

## Usage

```
fmi(data, method = "saturated", group = NULL, ords = NULL,
     varnames = NULL, exclude = NULL, fewImps = FALSE)
```

## Arguments

data	Either a single data.frame with incomplete observations, or a list of imputed data sets.
method	character. If "saturated" or "sat" (default), the model used to estimate FMI is a freely estimated covariance matrix and mean vector for numeric variables, and/or polychoric correlations and thresholds for ordered categorical variables, for each group (if applicable). If "null", only means and variances are estimated for numeric variables, and/or thresholds for ordered categorical variables (i.e., covariances and/or polychoric correlations are constrained to zero). See Details for more information.

group	character. The optional name of a grouping variable, to request FMI in each group.
ords	character. Optional vector of names of ordered-categorical variables, which are not already stored as class ordered in data.
varnames	character. Optional vector of variable names, to calculate FMI for a subset of variables in data. By default, all numeric and ordered variables will be included, unless data is a single incomplete data.frame, in which case only numeric variables can be used with FIML estimation. Other variable types will be removed.
exclude	character. Optional vector of variable names to exclude from the analysis.
fewImps	logical. If TRUE, use the estimate of FMI that applies a correction to the estimated between-imputation variance. Recommended when there are few imputations; makes little difference when there are many imputations. Ignored when data is not a list of imputed data sets.

### Details

The function estimates a saturated model with `lavaan` for a single incomplete data set using FIML, or with `lavaan.mi` for a list of imputed data sets. If `method = "saturated"`, FMI will be estimated for all summary statistics, which could take a lot of time with big data sets. If `method = "null"`, FMI will only be estimated for univariate statistics (e.g., means, variances, thresholds). The saturated model gives more reliable estimates, so it could also help to request a subset of variables from a large data set.

### Value

`fmi` returns a list with at least 2 of the following:

Covariances	A list of symmetric matrices: (1) the estimated/pooled covariance matrix, or a list of group-specific matrices (if applicable) and (2) a matrix of FMI, or a list of group-specific matrices (if applicable). Only available if <code>method = "saturated"</code> .
Variances	The estimated/pooled variance for each numeric variable. Only available if <code>method = "null"</code> (otherwise, it is on the diagonal of Covariances).
Means	The estimated/pooled mean for each numeric variable.
Thresholds	The estimated/pooled threshold(s) for each ordered-categorical variable.
message	A message indicating caution when the null model is used.

### Author(s)

Mauricio Garnier Villarreal (University of Kansas; <mauricio.garniervillarreal@marquette.edu>  
 Terrence Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### References

Rubin, D. B. (1987). *Multiple imputation for nonresponse in surveys*. New York, NY: Wiley.

Savalei, V. & Rhemtulla, M. (2012). On obtaining estimates of the fraction of missing information from full information maximum likelihood. *Structural Equation Modeling*, 19(3), 477–494. doi:10.1080/10705511.2012.687669

Wagner, J. (2010). The fraction of missing information as a tool for monitoring the quality of survey data. *Public Opinion Quarterly*, 74(2), 223–243. doi:10.1093/poq/nfq007

## Examples

```
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]
set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## calculate FMI (using FIML, provide partially observed data set)
(out1 <- fmi(HSMiss, exclude = "school"))
(out2 <- fmi(HSMiss, exclude = "school", method = "null"))
(out3 <- fmi(HSMiss, varnames = c("x5", "x6", "x7", "x8", "x9")))
(out4 <- fmi(HSMiss, group = "school"))

## Not run:
## ordered-categorical data
data(datCat)
lapply(datCat, class)
## impose missing values
set.seed(123)
for (i in 1:8) datCat[sample(1:nrow(datCat), size = .1*nrow(datCat)), i] <- NA
## impute data m = 3 times
library(Amelia)
set.seed(456)
impout <- amelia(datCat, m = 3, noms = "g", ords = paste0("u", 1:8), p2s = FALSE)
imps <- impout$imputations
## calculate FMI, using list of imputed data sets
fmi(imps, group = "g")

## End(Not run)
```

## Description

This function assesses discriminant validity through the heterotrait-monotrait ratio (HTMT) of the correlations (Henseler, Ringle & Sarstedt, 2015). Specifically, it assesses the geometric-mean correlation among indicators across constructs (i.e. heterotrait-heteromethod correlations) relative

to the geometric-mean correlation among indicators within the same construct (i.e. monotrait-heteromethod correlations). The resulting HTMT values are interpreted as estimates of inter-construct correlations. Absolute values of the correlations are recommended to calculate the HTMT matrix. Correlations are estimated using the `lavCor` function in the `lavaan` package.

### Usage

```
htmt(model, data = NULL, sample.cov = NULL, missing = "listwise",
      ordered = NULL, absolute = TRUE)
```

### Arguments

<code>model</code>	lavaan <a href="#">model.syntax</a> of a confirmatory factor analysis model where at least two factors are required for indicators measuring the same construct.
<code>data</code>	A <code>data.frame</code> or data matrix
<code>sample.cov</code>	A covariance or correlation matrix can be used, instead of <code>data</code> , to estimate the HTMT values.
<code>missing</code>	If "listwise", cases with missing values are removed listwise from the data frame. If "direct" or "ml" or "fiml" and the estimator is maximum likelihood, an EM algorithm is used to estimate the unrestricted covariance matrix (and mean vector). If "pairwise", pairwise deletion is used. If "default", the value is set depending on the estimator and the <code>mimic</code> option (see details in <a href="#">lavCor</a> ).
<code>ordered</code>	Character vector. Only used if object is a <code>data.frame</code> . Treat these variables as ordered (ordinal) variables. Importantly, all other variables will be treated as numeric (unless <code>is.ordered</code> in <code>data</code> ). (see also <a href="#">lavCor</a> )
<code>absolute</code>	logical. Whether HTMT values should be estimated based on absolute correlations (recommended and default is TRUE)

### Value

A matrix showing HTMT values (i.e., discriminant validity) between each pair of factors

### Author(s)

Ylenio Longo (University of Nottingham; <yleniolongo@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### References

- Henseler, J., Ringle, C. M., & Sarstedt, M. (2015). A new criterion for assessing discriminant validity in variance-based structural equation modeling. *Journal of the Academy of Marketing Science*, 43(1), 115–135. doi:10.1007/s11747-014-0403-8
- Voorhees, C. M., Brady, M. K., Calantone, R., & Ramirez, E. (2016). Discriminant validity testing in marketing: an analysis, causes for concern, and proposed remedies. *Journal of the Academy of Marketing Science*, 44(1), 119–134. doi:10.1007/s11747-015-0455-4

**Examples**

```

HS.model <- ' visual  =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed   =~ x7 + x8 + x9 '

dat <- HolzingerSwineford1939[, paste0("x", 1:9)]
htmt(HS.model, dat)

```

---

imposeStart

*Specify starting values from a lavaan output*


---

**Description**

This function will save the parameter estimates of a lavaan output and impose those parameter estimates as starting values for another analysis model. The free parameters with the same names or the same labels across two models will be imposed the new starting values. This function may help to increase the chance of convergence in a complex model (e.g., multitrait-multimethod model or complex longitudinal invariance model).

**Usage**

```
imposeStart(out, expr, silent = TRUE)
```

**Arguments**

out	The lavaan output that users wish to use the parameter estimates as starting values for an analysis model
expr	The original code that users use to run a lavaan model
silent	Logical to print the parameter table with new starting values

**Value**

A fitted lavaan model

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**Examples**

```

## The following example show that the longitudinal weak invariance model
## using effect coding was not convergent with three time points but convergent
## with two time points. Thus, the parameter estimates from the model with
## two time points are used as starting values of the three time points.
## The model with new starting values is convergent properly.

```

```

weak2time <- '
# Loadings
f1t1 =~ LOAD1*y1t1 + LOAD2*y2t1 + LOAD3*y3t1
      f1t2 =~ LOAD1*y1t2 + LOAD2*y2t2 + LOAD3*y3t2

# Factor Variances
f1t1 =~ f1t1
f1t2 =~ f1t2

# Factor Covariances
f1t1 =~ f1t2

# Error Variances
y1t1 =~ y1t1
y2t1 =~ y2t1
y3t1 =~ y3t1
y1t2 =~ y1t2
y2t2 =~ y2t2
y3t2 =~ y3t2

# Error Covariances
y1t1 =~ y1t2
y2t1 =~ y2t2
y3t1 =~ y3t2

# Factor Means
f1t1 ~ NA*1
f1t2 ~ NA*1

# Measurement Intercepts
y1t1 ~ INT1*1
y2t1 ~ INT2*1
y3t1 ~ INT3*1
y1t2 ~ INT4*1
y2t2 ~ INT5*1
y3t2 ~ INT6*1

# Constraints for Effect-coding Identification
LOAD1 == 3 - LOAD2 - LOAD3
INT1 == 0 - INT2 - INT3
INT4 == 0 - INT5 - INT6
'
model2time <- lavaan(weak2time, data = exLong)

weak3time <- '
# Loadings
f1t1 =~ LOAD1*y1t1 + LOAD2*y2t1 + LOAD3*y3t1
      f1t2 =~ LOAD1*y1t2 + LOAD2*y2t2 + LOAD3*y3t2
      f1t3 =~ LOAD1*y1t3 + LOAD2*y2t3 + LOAD3*y3t3

# Factor Variances
f1t1 =~ f1t1

```

```
f1t2 ~~ f1t2
f1t3 ~~ f1t3

# Factor Covariances
f1t1 ~~ f1t2 + f1t3
f1t2 ~~ f1t3

# Error Variances
y1t1 ~~ y1t1
y2t1 ~~ y2t1
y3t1 ~~ y3t1
y1t2 ~~ y1t2
y2t2 ~~ y2t2
y3t2 ~~ y3t2
y1t3 ~~ y1t3
y2t3 ~~ y2t3
y3t3 ~~ y3t3

# Error Covariances
y1t1 ~~ y1t2
y2t1 ~~ y2t2
y3t1 ~~ y3t2
y1t1 ~~ y1t3
y2t1 ~~ y2t3
y3t1 ~~ y3t3
y1t2 ~~ y1t3
y2t2 ~~ y2t3
y3t2 ~~ y3t3

# Factor Means
f1t1 ~ NA*1
f1t2 ~ NA*1
f1t3 ~ NA*1

# Measurement Intercepts
y1t1 ~ INT1*1
y2t1 ~ INT2*1
y3t1 ~ INT3*1
y1t2 ~ INT4*1
y2t2 ~ INT5*1
y3t2 ~ INT6*1
y1t3 ~ INT7*1
y2t3 ~ INT8*1
y3t3 ~ INT9*1

# Constraints for Effect-coding Identification
LOAD1 == 3 - LOAD2 - LOAD3
INT1 == 0 - INT2 - INT3
INT4 == 0 - INT5 - INT6
INT7 == 0 - INT8 - INT9
'

### The following command does not provide convergent result
# model3time <- lavaan(weak3time, data = exLong)
```

```
### Use starting values from the model with two time points
model3time <- imposeStart(model2time, lavaan(weak3time, data = exLong))
summary(model3time)
```

---

indProd                      *Make products of indicators using no centering, mean centering, double-mean centering, or residual centering*

---

### Description

The indProd function will make products of indicators using no centering, mean centering, double-mean centering, or residual centering. The orthogonalize function is the shortcut of the indProd function to make the residual-centered indicators products.

### Usage

```
indProd(data, var1, var2, var3 = NULL, match = TRUE, meanC = TRUE,
        residualC = FALSE, doubleMC = TRUE, namesProd = NULL)
```

```
orthogonalize(data, var1, var2, var3 = NULL, match = TRUE, namesProd = NULL)
```

### Arguments

data	The desired data to be transformed.
var1	Names or indices of the variables loaded on the first factor
var2	Names or indices of the variables loaded on the second factor
var3	Names or indices of the variables loaded on the third factor (for three-way interaction)
match	Specify TRUE to use match-paired approach (Marsh, Wen, & Hau, 2004). If FALSE, the resulting products are all possible products.
meanC	Specify TRUE for mean centering the main effect indicator before making the products
residualC	Specify TRUE for residual centering the products by the main effect indicators (Little, Bovaird, & Widaman, 2006).
doubleMC	Specify TRUE for centering the resulting products (Lin et. al., 2010)
namesProd	The names of resulting products

### Value

The original data attached with the products.

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>) Alexander Schoemann (East Carolina University; <schoemanna@ecu.edu>)



## References

- Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9(3), 275–300. doi:10.1037/1082-989X.9.3.275
- Lin, G. C., Wen, Z., Marsh, H. W., & Lin, H. S. (2010). Structural equation models of latent interactions: Clarification of orthogonalizing and double-mean-centering strategies. *Structural Equation Modeling*, 17(3), 374–391. doi:10.1080/10705511.2010.488999
- Little, T. D., Bovaird, J. A., & Widaman, K. F. (2006). On the merits of orthogonalizing powered and product terms: Implications for modeling interactions among latent variables. *Structural Equation Modeling*, 13(4), 497–519. doi:10.1207/s15328007sem1304\_1

## See Also

- [probe2WayMC](#) For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- [probe3WayMC](#) For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering.
- [probe2WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [probe3WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [plotProbe](#) Plot the simple intercepts and slopes of the latent interaction.

## Examples

```
## Mean centering / two-way interaction / match-paired
dat <- indProd(attitude[ , -1], var1 = 1:3, var2 = 4:6)

## Residual centering / two-way interaction / match-paired
dat2 <- indProd(attitude[ , -1], var1 = 1:3, var2 = 4:6, match = FALSE,
               meanC = FALSE, residualC = TRUE, doubleMC = FALSE)

## Double-mean centering / two-way interaction / match-paired
dat3 <- indProd(attitude[ , -1], var1 = 1:3, var2 = 4:6, match = FALSE,
               meanC = TRUE, residualC = FALSE, doubleMC = TRUE)

## Mean centering / three-way interaction / match-paired
dat4 <- indProd(attitude[ , -1], var1 = 1:2, var2 = 3:4, var3 = 5:6)

## Residual centering / three-way interaction / match-paired
dat5 <- orthogonalize(attitude[ , -1], var1 = 1:2, var2 = 3:4, var3 = 5:6,
                    match = FALSE)

## Double-mean centering / three-way interaction / match-paired
dat6 <- indProd(attitude[ , -1], var1 = 1:2, var2 = 3:4, var3 = 5:6,
               match = FALSE, meanC = TRUE, residualC = TRUE,
               doubleMC = TRUE)
```

```

## To add product-indicators to multiple-imputed data sets
## Not run:
HSMiss <- HolzingerSwineford1939[ , c(paste0("x", 1:9), "ageyr", "agemo")]
set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)
library(Amelia)
set.seed(12345)
HS.amelia <- amelia(HSMiss, m = 3, p2s = FALSE)
imps <- HS.amelia$imputations # extract a list of imputations
## apply indProd() to the list of data.frames
imps2 <- lapply(imps, indProd,
                var1 = c("x1", "x2", "x3"), var2 = c("x4", "x5", "x6"))
## verify:
lapply(imps2, head)

## End(Not run)

```

---

kd

*Generate data via the Kaiser-Dickman (1962) algorithm.*


---

## Description

Given a covariance matrix and sample size, generate raw data that correspond to the covariance matrix. Data can be generated to match the covariance matrix exactly, or to be a sample from the population covariance matrix.

## Usage

```
kd(covmat, n, type = c("exact", "sample"))
```

## Arguments

covmat	a symmetric, positive definite covariance matrix
n	the sample size for the data that will be generated
type	type of data generation. exact generates data that exactly correspond to covmat. sample treats covmat as a population covariance matrix, generating a sample of size n.

## Details

By default, R's `cov()` function divides by  $n-1$ . The data generated by this algorithm result in a covariance matrix that matches `covmat`, but you must divide by  $n$  instead of  $n-1$ .

**Value**

kd returns a data matrix of dimension n by nrow(covmat).

**Author(s)**

Ed Merkle (University of Missouri; <merklee@missouri.edu>)

**References**

Kaiser, H. F. and Dickman, K. (1962). Sample and population score matrices and sample correlation matrices from an arbitrary population correlation matrix. *Psychometrika*, 27(2), 179–182. doi:10.1007/BF02289635

**Examples**

```
#### First Example

## Get data
dat <- HolzingerSwineford1939[ , 7:15]
hs.n <- nrow(dat)

## Covariance matrix divided by n
hscov <- ((hs.n-1)/hs.n) * cov(dat)

## Generate new, raw data corresponding to hscov
newdat <- kd(hscov, hs.n)

## Difference between new covariance matrix and hscov is minimal
newcov <- (hs.n-1)/hs.n * cov(newdat)
summary(as.numeric(hscov - newcov))

## Generate sample data, treating hscov as population matrix
newdat2 <- kd(hscov, hs.n, type = "sample")

#### Another example

## Define a covariance matrix
covmat <- matrix(0, 3, 3)
diag(covmat) <- 1.5
covmat[2:3,1] <- c(1.3, 1.7)
covmat[3,2] <- 2.1
covmat <- covmat + t(covmat)

## Generate data of size 300 that have this covariance matrix
rawdat <- kd(covmat, 300)

## Covariances are exact if we compute sample covariance matrix by
## dividing by n (vs by n - 1)
summary(as.numeric((299/300)*cov(rawdat) - covmat))

## Generate data of size 300 where covmat is the population covariance matrix
```

```
rawdatt2 <- kd(covmat, 300)
```

---

kurtosis	<i>Finding excessive kurtosis</i>
----------	-----------------------------------

---

### Description

Finding excessive kurtosis ( $g_2$ ) of an object

### Usage

```
kurtosis(object, population = FALSE)
```

### Arguments

object	A vector used to find a excessive kurtosis
population	TRUE to compute the parameter formula. FALSE to compute the sample statistic formula.

### Details

The excessive kurtosis computed is  $g_2$ . The parameter excessive kurtosis  $\gamma_2$  formula is

$$\gamma_2 = \frac{\mu_4}{\mu_2^2} - 3,$$

where  $\mu_i$  denotes the  $i$  order central moment.

The excessive kurtosis formula for sample statistic  $g_2$  is

$$g_2 = \frac{k_4}{k_2^2},$$

where  $k_i$  are the  $i$  order  $k$ -statistic.

The standard error of the excessive kurtosis is

$$Var(\hat{g}_2) = \frac{24}{N}$$

where  $N$  is the sample size.

### Value

A value of an excessive kurtosis with a test statistic if the population is specified as FALSE

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

## References

Weisstein, Eric W. (n.d.). *Kurtosis*. Retrived from *MathWorld*—A Wolfram Web Resource: <http://mathworld.wolfram.com/Kurtosis.html>

## See Also

- [skew](#) Find the univariate skewness of a variable
- [mardiaSkew](#) Find the Mardia's multivariate skewness of a set of variables
- [mardiaKurtosis](#) Find the Mardia's multivariate kurtosis of a set of variables

## Examples

```
kurtosis(1:5)
```

---

lavaan.mi-class	<i>Class for a lavaan Model Fitted to Multiple Imputations</i>
-----------------	--

---

## Description

This class extends the [lavaanList](#) class, created by fitting a lavaan model to a list of data sets. In this case, the list of data sets are multiple imputations of missing data.

## Usage

```
## S4 method for signature 'lavaan.mi'
show(object)

## S4 method for signature 'lavaan.mi'
summary(object, se = TRUE, ci = FALSE,
  level = 0.95, standardized = FALSE, rsquare = FALSE, fmi = FALSE,
  scale.W = !asymptotic, omitimps = c("no.conv", "no.se"),
  asymptotic = FALSE, header = TRUE, output = "text",
  fit.measures = FALSE)

## S4 method for signature 'lavaan.mi'
nobs(object, total = TRUE)

## S4 method for signature 'lavaan.mi'
coef(object, type = "free", labels = TRUE, omitimps = c("no.conv", "no.se"))

## S4 method for signature 'lavaan.mi'
vcov(object, type = c("pooled", "between", "within",
  "ariv"), scale.W = TRUE, omitimps = c("no.conv", "no.se"))
```

```

## S4 method for signature 'lavaan.mi'
anova(object, ...)

## S4 method for signature 'lavaan.mi'
fitMeasures(object, fit.measures = "all",
  baseline.model = NULL, output = "vector", omitimps = c("no.conv",
  "no.se"), ...)

## S4 method for signature 'lavaan.mi'
fitmeasures(object, fit.measures = "all",
  baseline.model = NULL, output = "vector", omitimps = c("no.conv",
  "no.se"), ...)

## S4 method for signature 'lavaan.mi'
fitted(object, omitimps = c("no.conv", "no.se"))

## S4 method for signature 'lavaan.mi'
fitted.values(object, omitimps = c("no.conv", "no.se"))

## S4 method for signature 'lavaan.mi'
residuals(object, type = c("raw", "cor"), omitimps = c("no.conv", "no.se"))

## S4 method for signature 'lavaan.mi'
resid(object, type = c("raw", "cor"), omitimps = c("no.conv", "no.se"))

```

## Arguments

object	An object of class <code>lavaan.mi</code>
se, ci, level, standardized, rsquare, header, output	See <a href="#">parameterEstimates</a> . output can also be passed to <a href="#">fitMeasures</a> .
fmi	logical indicating whether to include the Fraction Missing Information (FMI) for parameter estimates in the summary output (see <b>Value</b> section).
scale.w	logical. If TRUE (default), the <code>vcov</code> method will calculate the pooled covariance matrix by scaling the within-imputation component by the ARIV (see Enders, 2010, p. 235, for definition and formula). Otherwise, the pooled matrix is calculated as the weighted sum of the within-imputation and between-imputation components (see Enders, 2010, ch. 8, for details). This in turn affects how the <code>summary</code> method calculates its pooled standard errors, as well as the Wald test ( <a href="#">lavTestWald.mi</a> ).
omitimps	character vector specifying criteria for omitting imputations from pooled results. Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD

solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation. Specific imputation numbers can also be included in this argument, in case users want to apply their own custom omission criteria (or simulations can use different numbers of imputations without redundantly refitting the model).

asymptotic	logical. If FALSE (typically a default, but see <b>Value</b> section for details using various methods), pooled tests (of fit or pooled estimates) will be $F$ or $t$ statistics with associated degrees of freedom ( $df$ ). If TRUE, the (denominator) $df$ are assumed to be sufficiently large for a $t$ statistic to follow a normal distribution, so it is printed as a $z$ statistic; likewise, $F$ times its numerator $df$ is printed, assumed to follow a $\chi^2$ distribution.
fit.measures, baseline.model	See <a href="#">fitMeasures</a> . <code>summary(object, fit.measures = TRUE)</code> will print (but not return) a table of fit measures to the console.
total	logical (default: TRUE) indicating whether the <code>nobs</code> method should return the total sample size or (if FALSE) a vector of group sample sizes.
type	The meaning of this argument varies depending on which method it is used for. Find detailed descriptions in the <b>Value</b> section under <code>coef</code> , <code>vcov</code> , and <code>residuals</code> .
labels	logical indicating whether the <code>coef</code> output should include parameter labels. Default is TRUE.
...	Additional arguments passed to <a href="#">lavTestLRT.mi</a> , or subsequently to <a href="#">lavTestLRT</a> .

### Value

<code>coef</code>	<code>signature(object = "lavaan.mi", type = "free", labels = TRUE, omit.imps = c("no.conv", "no.se"))</code> : See <a href="#">lavaan</a> . Returns the pooled point estimates (i.e., averaged across imputed data sets; see Rubin, 1987).
<code>vcov</code>	<code>signature(object = "lavaan.mi", scale.W = TRUE, omit.imps = c("no.conv", "no.se"), type = c("pooled", "between", "within", "ariv"))</code> : By default, returns the pooled covariance matrix of parameter estimates ( <code>type = "pooled"</code> ), the within-imputations covariance matrix ( <code>type = "within"</code> ), the between-imputations covariance matrix ( <code>type = "between"</code> ), or the average relative increase in variance ( <code>type = "ariv"</code> ) due to missing data.
<code>fitted.values</code>	<code>signature(object = "lavaan.mi", omit.imps = c("no.conv", "no.se"))</code> : See <a href="#">lavaan</a> . Returns model-implied moments, evaluated at the pooled point estimates.
<code>fitted</code>	alias for <code>fitted.values</code>
<code>residuals</code>	<code>signature(object = "lavaan.mi", type = c("raw", "cor"), omit.imps = c("no.conv", "no.se"))</code> : See <a href="#">lavaan</a> . By default ( <code>type = "raw"</code> ), returns the difference between the model-implied moments from <code>fitted.values</code> and the pooled observed moments (i.e., averaged across imputed data sets). Standardized residuals are also available, using Bollen's ( <code>type = "cor"</code> or <code>"cor.bollen"</code> ) or Bentler's ( <code>type = "cor.bentler"</code> ) formulas.
<code>resid</code>	alias for <code>residuals</code>

nobs	signature(object = "lavaan.mi", total = TRUE): either the total (default) sample size or a vector of group sample sizes (total = FALSE).
anova	signature(object = "lavaan.mi", ...): Returns a test of model fit for a single model (object) or test(s) of the difference(s) in fit between nested models passed via .... See <a href="#">lavTestLRT.mi</a> and <a href="#">compareFit</a> for details.
fitMeasures	signature(object = "lavaan.mi", fit.measures = "all", baseline.model = NULL, output = "vector", omitimps = c("no.conv", "no.se"), ...): See lavaan's <a href="#">fitMeasures</a> for details. Pass additional arguments to <a href="#">lavTestLRT.mi</a> via ....
fitmeasures	alias for fitMeasures.
show	signature(object = "lavaan.mi"): returns a message about convergence rates and estimation problems (if applicable) across imputed data sets.
summary	signature(object = "lavaan.mi", se = TRUE, ci = FALSE, level = .95, standardized = FALSE, rsquare = FALSE, fmi = FALSE, scale.W = !asymptotic, omitimps = c("no.conv", "no.se"), asymptotic = FALSE, header = TRUE, output = "text", fit.measures = FALSE): see <a href="#">parameterEstimates</a> for details. By default, summary returns pooled point and <i>SE</i> estimates, along with <i>t</i> test statistics and their associated <i>df</i> and <i>p</i> values. If ci = TRUE, confidence intervals are returned with the specified confidence level (default 95% CI). If asymptotic = TRUE, <i>z</i> instead of <i>t</i> tests are returned. standardized solution(s) can also be requested by name ("std.lv" or "std.all") or both are returned with TRUE. <i>R</i> -squared for endogenous variables can be requested, as well as the Fraction Missing Information (FMI) for parameter estimates. By default, the output will appear like lavaan's summary output, but if output == "data.frame", the returned data.frame will resemble the parameterEstimates output. The scale.W argument is passed to vcov (see description above). Setting fit.measures=TRUE will additionally print fit measures to the console, but they will not be returned.

## Slots

coefList	list of estimated coefficients in matrix format (one per imputation) as output by <a href="#">lavInspect</a> (fit, "est")
phiList	list of model-implied latent-variable covariance matrices (one per imputation) as output by <a href="#">lavInspect</a> (fit, "cov.lv")
miList	list of modification indices output by <a href="#">modindices</a>
seed	integer seed set before running imputations
lavListCall	call to <a href="#">lavaanList</a> used to fit the model to the list of imputed data sets in @DataList, stored as a list of arguments
imputeCall	call to imputation function (if used), stored as a list of arguments
convergence	list of logical vectors indicating whether, for each imputed data set, (1) the model converged on a solution, (2) <i>SEs</i> could be calculated, (3) the (residual) covariance matrix of latent variables ( $\Psi$ ) is non-positive-definite, and (4) the residual covariance matrix of observed variables ( $\Theta$ ) is non-positive-definite.
lavaanList_slots	All remaining slots are from <a href="#">lavaanList</a> , but <a href="#">runMI</a> only populates a subset of the list slots, two of them with custom information:
DataList	The list of imputed data sets



SampleStatsList List of output from `lavInspect(fit, "sampstat")` applied to each fitted model

ParTableList See [lavaanList](#)

vcovList See [lavaanList](#)

testList See [lavaanList](#)

h1List See [lavaanList](#). An additional element is added to the list: \$PT is the "saturated" model's parameter table, returned by `lav_partable_unrestricted`.

baselineList See [lavaanList](#)

### Objects from the Class

See the `runMI` function for details. Wrapper functions include [lavaan.mi](#), [cfa.mi](#), [sem.mi](#), and [growth.mi](#).

### Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### References

- Asparouhov, T., & Muthen, B. (2010). *Chi-square statistics with multiple imputation*. Technical Report. Retrieved from [www.statmodel.com](http://www.statmodel.com)
- Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.
- Li, K.-H., Meng, X.-L., Raghunathan, T. E., & Rubin, D. B. (1991). Significance levels from repeated *p*-values with multiply-imputed data. *Statistica Sinica*, *1*(1), 65–92. Retrieved from <https://www.jstor.org/stable/24303994>
- Meng, X.-L., & Rubin, D. B. (1992). Performing likelihood ratio tests with multiply-imputed data sets. *Biometrika*, *79*(1), 103–111. doi:10.2307/2337151
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### Examples

```
## See ?runMI help page
```

---

lavaan2emmeans

emmeans *Support Functions for lavaan Models*

---

### Description

Provide emmeans support for lavaan objects

### Usage

```
recover_data.lavaan(object, lavaan.DV, ...)
```

```
emm_basis.lavaan(object, trms, xlev, grid, lavaan.DV, ...)
```

## Arguments

object	An object of class <code>lavaan</code> . See <b>Details</b> .
lavaan.DV	character string naming the variable(s) for which expected marginal means / trends should be produced. A vector of names indicates a multivariate outcome, treated by default as repeated measures.
...	Further arguments passed to <code>emmeans::recover_data.lm</code> or <code>emmeans::emm_basis.lm</code>
trms, xlev, grid	See <code>emmeans::emm_basis</code>

## Details

**Supported DVs:** `lavaan.DV` must be an *endogenous variable*, by appearing on the left-hand side of either a regression operator ("`~`") or an intercept operator ("`~1`"), or both.

`lavaan.DV` can also be a vector of endogenous variable, in which case they will be treated by `emmeans` as a multivariate outcome (often, this indicates repeated measures) represented by an additional factor named `rep.meas` by default. The `rep.meas=` argument can be used to overwrite this default name.

**Unsupported Models:** This functionality does not support the following models:

- Multi-level models are not supported.
- Models not fit to a `data.frame` (i.e., models fit to a covariance matrix).

**Dealing with Fixed Parameters:** Fixed parameters (set with `lavaan`'s modifiers) are treated as-is: their values are set by the users, and they have a *SE* of 0 (as such, they do not co-vary with any other parameter).

**Dealing with Multigroup Models:** If a multigroup model is supplied, a factor is added to the reference grid, the name matching the `group` argument supplied when fitting the model. *Note that you must set `nesting = NULL`.*

**Dealing with Missing Data:** Limited testing suggests that these functions do work when the model was fit to incomplete data.

**Dealing with Factors:** By default `emmeans` recognizes binary variables (0,1) as a "factor" with two levels (and not a continuous variable). With some clever contrast definitions it should be possible to get the desired `emmeans` / contrasts. See example below.

## Author(s)

Mattan S. Ben-Shachar (Ben-Gurion University of the Negev; <matanshm@post.bgu.ac.il>)

## Examples

```
## Not run:

library(lavaan)
library(emmeans)
```

```

#### Moderation Analysis ####

mean_sd <- function(x) mean(x) + c(-sd(x), 0, sd(x))

model <- '
# regressions
Sepal.Length ~ b1 * Sepal.Width + b2 * Petal.Length + b3 * Sepal.Width:Petal.Length

# define mean parameter label for centered math for use in simple slopes
Sepal.Width ~ Sepal.Width.mean * 1

# define variance parameter label for centered math for use in simple slopes
Sepal.Width ~~ Sepal.Width.var * Sepal.Width

# simple slopes for condition effect
SD.below := b2 + b3 * (Sepal.Width.mean - sqrt(Sepal.Width.var))
mean      := b2 + b3 * (Sepal.Width.mean)
SD.above  := b2 + b3 * (Sepal.Width.mean + sqrt(Sepal.Width.var))
'

semFit <- sem(model = model,
              data = iris)

## Compare simple slopes
# From `emtrends`
test(
  emtrends(semFit, ~ Sepal.Width, "Petal.Length",
            lavaan.DV = "Sepal.Length",
            cov.red = mean_sd)
)

# From lavaan
parameterEstimates(semFit, output = "pretty")[13:15, ]
# Identical slopes.
# SEs differ due to lavaan estimating uncertainty of the mean / SD
# of Sepal.Width, whereas emmeans uses the mean+-SD as is (fixed).

#### Latent DV ####

model <- '
LAT1 =~ Sepal.Length + Sepal.Width

LAT1 ~ b1 * Petal.Width + 1 * Petal.Length

Petal.Length ~ Petal.Length.mean * 1

V1 := 1 * Petal.Length.mean + 1 * b1
V2 := 1 * Petal.Length.mean + 2 * b1
'

semFit <- sem(model = model,

```

```

      data = iris, std.lv = TRUE)

## Compare emmeans
# From emmeans
test(
  emmeans(semFit, ~ Petal.Width,
           lavaan.DV = "LAT1",
           at = list(Petal.Width = 1:2))
)

# From lavaan
parameterEstimates(semFit, output = "pretty")[15:16, ]
# Identical means.
# SEs differ due to lavaan estimating uncertainty of the mean
# of Petal.Length, whereas emmeans uses the mean as is.

#### Multi-Variate DV ####

model <- '
ind60 =~ x1 + x2 + x3

# metric invariance
dem60 =~ y1 + a*y2 + b*y3 + c*y4
dem65 =~ y5 + a*y6 + b*y7 + c*y8

# scalar invariance
y1 + y5 ~ d*1
y2 + y6 ~ e*1
y3 + y7 ~ f*1
y4 + y8 ~ g*1

# regressions (slopes differ: interaction with time)
dem60 ~ b1*ind60
dem65 ~ b2*ind60 + NA*1 + Mean.Diff*1

# residual correlations
y1 ~~ y5
y2 ~~ y4 + y6
y3 ~~ y7
y4 ~~ y8
y6 ~~ y8

# conditional mean differences (besides mean(ind60) == 0)
low := (-1*b2 + Mean.Diff) - (-1*b1) # 1 SD below M
high := (b2 + Mean.Diff) - b1      # 1 SD above M

semFit <- sem(model, data = PoliticalDemocracy)

## Compare contrasts
# From emmeans
emmeans(semFit, pairwise ~ rep.meas|ind60,

```

```

lavaan.DV = c("dem60", "dem65"),
at = list(ind60 = c(-1, 1))[[2]]

# From lavaan
parameterEstimates(semFit, output = "pretty")[49:50, ]

#### Multi Group ####

model <- 'x1 ~ c(int1, int2)*1 + c(b1, b2)*ageyr
diff_11 := (int2 + b2*11) - (int1 + b1*11)
diff_13 := (int2 + b2*13) - (int1 + b1*13)
diff_15 := (int2 + b2*15) - (int1 + b1*15)
,
semFit <- sem(model, group = "school", data = HolzingerSwineford1939)

## Compare contrasts
# From emmeans (note `nesting = NULL`)
emmeans(semFit, pairwise ~ school | ageyr, lavaan.DV = "x1",
         at = list(ageyr = c(11, 13, 15)), nesting = NULL)[[2]]

# From lavaan
parameterEstimates(semFit, output = "pretty")

#### Dealing with factors ####

warpbreaks <- cbind(warpbreaks,
                   model.matrix(~ wool + tension, data = warpbreaks))

model <- "
# Split for convenience
breaks ~ 1
breaks ~ woolB
breaks ~ tensionM + tensionH
breaks ~ woolB:tensionM + woolB:tensionH
"

semFit <- sem(model, warpbreaks)

## Compare contrasts
# From lm -> emmeans
lmFit <- lm(breaks ~ wool * tension, data = warpbreaks)
lmEM <- emmeans(lmFit, ~ tension + wool)
contrast(lmEM, method = data.frame(L_all = c(-1, .05, 0.5),
                                   M_H   = c(0, 1, -1)), by = "wool")

# From lavaan -> emmeans
lavEM <- emmeans(semFit, ~ tensionM + tensionH + woolB,
                 lavaan.DV = "breaks")
contrast(lavEM,
         method = list(
           "L_all|A" = c(c(-1, .05, 0.5, 0), rep(0, 4)),

```

```

      "M_H |A" = c(c(0, 1, -1, 0),      rep(0, 4)),
      "L_all|A" = c(rep(0, 4),        c(-1, .05, 0.5, 0)),
      "M_H |A" = c(rep(0, 4),        c(0, 1, -1, 0))
    ))

## End(Not run)

```

---

lavTestLRT.mi

*Likelihood Ratio Test for Multiple Imputations*


---

## Description

Likelihood ratio test (LRT) for lavaan models fitted to multiple imputed data sets. Statistics for comparing nested models can be calculated by pooling the likelihood ratios across imputed data sets, as described by Meng & Rubin (1992), or by pooling the LRT statistics from each imputation, as described by Li, Meng, Raghunathan, & Rubin (1991).

## Usage

```

lavTestLRT.mi(object, h1 = NULL, test = c("D3", "D2"),
  omitimps = c("no.conv", "no.se"), asymptotic = FALSE,
  pool.robust = FALSE, ...)

```

## Arguments

object, h1	An object of class <code>lavaan.mi</code> . object should be nested within (more constrained than) h1.
test	character indicating which pooling method to use. "D3", "mr", or "meng.rubin" (default) requests the method described by Meng & Rubin (1992). "D2", "LMRR", or "Li.et.al" requests the complete-data LRT statistic should be calculated using each imputed data set, which will then be pooled across imputations, as described in Li, Meng, Raghunathan, & Rubin (1991). Find additional details in Enders (2010, chapter 8).
omitimps	character vector specifying criteria for omitting imputations from pooled results. Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. Specific imputation numbers can also be included in this argument, in case users want to apply their own custom omission criteria (or simulations can use different numbers of imputations without redundantly refitting the model).
asymptotic	logical. If FALSE (default), the pooled test will be returned as an $F$ -distributed statistic with numerator (df1) and denominator (df2) degrees of freedom. If TRUE, the pooled $F$ statistic will be multiplied by its df1 on the assumption that its df2 is sufficiently large enough that the statistic will be asymptotically $\chi^2$ distributed with df1.

pool.robust      logical. Ignored unless test = "D2" and a robust test was requested. If pool.robust = TRUE, the robust test statistic is pooled, whereas pool.robust = FALSE will pool the naive test statistic (or difference statistic) and apply the average scale/shift parameter to it (unavailable for mean- and variance-adjusted difference statistics, so pool.robust will be set TRUE).

...                Additional arguments passed to [lavTestLRT](#), only if test = "D2" and pool.robust = TRUE

## Details

The Meng & Rubin (1992) method, also referred to as the "D3" statistic, is only applicable when using a likelihood-based estimator. Otherwise (e.g., DWLS for categorical outcomes), users are notified that test was set to "D2".

test = "Mplus" implies "D3" and asymptotic = TRUE (see Asparouhov & Muthen, 2010).

Note that unlike [lavTestLRT](#), [lavTestLRT](#) can only be used to compare a single pair of models, not a longer list of models. To compare several nested models fitted to multiple imputations, see examples on the [compareFit](#) help page.

## Value

A vector containing the LRT statistic (either an F or  $\chi^2$  statistic, depending on the asymptotic argument), its degrees of freedom (numerator and denominator, if asymptotic = FALSE), its *p* value, and 2 missing-data diagnostics: the relative increase in variance (RIV, or average for multiparameter tests: ARIV) and the fraction missing information (FMI = ARIV / (1 + ARIV)). Robust statistics will also include the average (across imputations) scaling factor and (if relevant) shift parameter(s), unless pool.robust = TRUE.

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## References

- Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.
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- Meng, X.-L., & Rubin, D. B. (1992). Performing likelihood ratio tests with multiply-imputed data sets. *Biometrika*, *79*(1), 103–111. doi:10.2307/2337151
- Rubin, D. B. (1987). *Multiple imputation for nonresponse in surveys*. New York, NY: Wiley.

## See Also

[lavTestLRT](#), [compareFit](#)

## Examples

```

## Not run:
## impose missing data for example
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]

set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## impute missing data
library(Amelia)
set.seed(12345)
HS.amelia <- amelia(HSMiss, m = 20, noms = "school", p2s = FALSE)
imps <- HS.amelia$imputations

## specify CFA model from lavaan's ?cfa help page
HS.model <- '
  visual  =~ x1 + b1*x2 + x3
  textual =~ x4 + b2*x5 + x6
  speed   =~ x7 + b3*x8 + x9
'

fit1 <- cfa.mi(HS.model, data = imps, estimator = "mlm")
fit0 <- cfa.mi(HS.model, data = imps, estimator = "mlm", orthogonal = TRUE)

## By default, use D3.
## Must request a chi-squared statistic to be robustified.
lavTestLRT.mi(fit1, h1 = fit0, asymptotic = TRUE)

## Using D2, you can either robustify the pooled naive statistic ...
lavTestLRT.mi(fit1, h1 = fit0, asymptotic = TRUE, test = "D2")
## ... or pool the robust chi-squared statistic
lavTestLRT.mi(fit1, h1 = fit0, asymptotic = TRUE, test = "D2",
              pool.robust = TRUE)

## End(Not run)

```

---

lavTestScore.mi

*Score Test for Multiple Imputations*

---

## Description

Score test (or "Lagrange multiplier" test) for lavaan models fitted to multiple imputed data sets. Statistics for releasing one or more fixed or constrained parameters in model can be calculated by pooling the gradient and information matrices pooled across imputed data sets in a method proposed by Mansolf, Jorgensen, & Enders (in press)—analogous to the "D1" Wald test proposed by Li, Meng, Raghunathan, & Rubin's (1991)—or by pooling the complete-data score-test statistics across imputed data sets (i.e., "D2"; Li et al., 1991).



**Usage**

```
lavTestScore.mi(object, add = NULL, release = NULL, test = c("D2", "D1"),
  scale.W = !asymptotic, omit.imps = c("no.conv", "no.se"),
  asymptotic = is.null(add), univariate = TRUE, cumulative = FALSE,
  epc = FALSE, standardized = epc, cov.std = epc, verbose = FALSE,
  warn = TRUE, information = "expected")
```

**Arguments**

object	An object of class <code>lavaan.mi</code> .
add	Either a character string (typically between single quotes) or a parameter table containing additional (currently fixed-to-zero) parameters for which the score test must be computed.
release	Vector of integers. The indices of the <i>equality</i> constraints that should be released. The indices correspond to the order of the equality constraints as they appear in the parameter table.
test	character indicating which pooling method to use. "D1" requests Mansolf, Jorgensen, & Enders' (in press) proposed Wald-like test for pooling the gradient and information, which are then used to calculate score-test statistics in the usual manner. "D2" (default because it is less computationally intensive) requests to pool the complete-data score-test statistics from each imputed data set, then pool them across imputations, described by Li et al. (1991) and Enders (2010).
scale.W	logical. If FALSE, the pooled information matrix is calculated as the weighted sum of the within-imputation and between-imputation components. Otherwise, the pooled information is calculated by scaling the within-imputation component by the average relative increase in variance (ARIV; Enders, 2010, p. 235), which is <i>only</i> consistent when requesting the <i>F</i> test (i.e., <code>asymptotic = FALSE</code> ). Ignored (irrelevant) if <code>test = "D2"</code> .
omit.imps	character vector specifying criteria for omitting imputations from pooled results. Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. Specific imputation numbers can also be included in this argument, in case users want to apply their own custom omission criteria (or simulations can use different numbers of imputations without redundantly refitting the model).
asymptotic	logical. If FALSE (default when using <code>add</code> to test adding fixed parameters to the model), the pooled test will be returned as an <i>F</i> -distributed variable with numerator ( <code>df1</code> ) and denominator ( <code>df2</code> ) degrees of freedom. If TRUE, the pooled <i>F</i> statistic will be multiplied by its <code>df1</code> on the assumption that its <code>df2</code> is sufficiently large enough that the statistic will be asymptotically $\chi^2$ distributed with <code>df1</code> . When using the <code>release</code> argument, <code>asymptotic</code> will be set to TRUE because (A)RIV can only be calculated for added parameters.
univariate	logical. If TRUE, compute the univariate score statistics, one for each constraint.

cumulative	logical. If TRUE, order the univariate score statistics from large to small, and compute a series of multivariate score statistics, each time including an additional constraint in the test.
epc	logical. If TRUE, and we are releasing existing constraints, compute the expected parameter changes for the existing (free) parameters (and any specified with add), if all constraints were released. For EPCs associated with a particular (1- <i>df</i> ) constraint, only specify one parameter in add or one constraint in release.
standardized	If TRUE, two extra columns (sepc.lv and sepc.all) in the \$epc table will contain standardized values for the EPCs. See <a href="#">lavTestScore</a> .
cov.std	logical. See <a href="#">standardizedSolution</a> .
verbose	logical. Not used for now.
warn	logical. If TRUE, print warnings if they occur.
information	character indicating the type of information matrix to use (check <a href="#">lavInspect</a> for available options). "expected" information is the default, which provides better control of Type I errors.

### Value

A list containing at least one data.frame:

- \$test: The total score test, with columns for the score test statistic ( $X^2$ ), its degrees of freedom (df), its  $p$  value under the  $\chi^2$  distribution (p.value), and if asymptotic=FALSE, the average relative increase in variance (ARIV) used to calculate the denominator  $df$  is also returned as a missing-data diagnostic, along with the fraction missing information (FMI = ARIV / (1 + ARIV)).
- \$uni: Optional (if univariate=TRUE). Each 1- $df$  score test, equivalent to modification indices. Also includes EPCs if epc=TRUE, and RIV and FMI if asymptotic=FALSE.
- \$cumulative: Optional (if cumulative=TRUE). Cumulative score tests, with ARIV and FMI if asymptotic=FALSE.
- \$sepc: Optional (if epc=TRUE). Parameter estimates, expected parameter changes, and expected parameter values if ALL the tested constraints were freed.

See [lavTestScore](#) for details.

### Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

Adapted from **lavaan** source code, written by Yves Rosseel (Ghent University; <Yves.Rosseel@UGent.be>)

test = "D1" method proposed by Maxwell Mansolf (University of California, Los Angeles; <mamansolf@gmail.com>)

### References

- Bentler, P. M., & Chou, C.-P. (1992). Some new covariance structure model improvement statistics. *Sociological Methods & Research*, 21(2), 259–282. doi:10.1177/0049124192021002006
- Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.

Li, K.-H., Meng, X.-L., Raghunathan, T. E., & Rubin, D. B. (1991). Significance levels from repeated  $p$ -values with multiply-imputed data. *Statistica Sinica*, 1(1), 65–92. Retrieved from <https://www.jstor.org/stable/24303994>

Mansolf, M., Jorgensen, T. D., & Enders, C. K. (in press). A multiple imputation score test for model modification in structural equation models. *Psychological Methods*. doi:10.1037/met0000243

## See Also

[lavTestScore](#)

## Examples

```
## Not run:
## impose missing data for example
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]

set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## impute missing data
library(Amelia)
set.seed(12345)
HS.amelia <- amelia(HSMiss, m = 20, noms = "school", p2s = FALSE)
imps <- HS.amelia$imputations

## specify CFA model from lavaan's ?cfa help page
HS.model <- '
  speed =~ c(L1, L1)*x7 + c(L1, L1)*x8 + c(L1, L1)*x9
'

out <- cfa.mi(HS.model, data = imps, group = "school", std.lv = TRUE)

## Mode 1: Score test for releasing equality constraints

## default test: Li et al.'s (1991) "D2" method
lavTestScore.mi(out, cumulative = TRUE)
## Li et al.'s (1991) "D1" method
lavTestScore.mi(out, test = "D1")

## Mode 2: Score test for adding currently fixed-to-zero parameters
lavTestScore.mi(out, add = 'x7 ~~ x8 + x9')

## End(Not run)
```

lavTestWald.mi

*Wald Test for Multiple Imputations***Description**

Wald test for testing a linear hypothesis about the parameters of lavaan models fitted to multiple imputed data sets. Statistics for constraining one or more free parameters in a model can be calculated from the pooled point estimates and asymptotic covariance matrix of model parameters using Rubin's (1987) rules, or by pooling the Wald test statistics across imputed data sets (Li, Meng, Raghunathan, & Rubin, 1991).

**Usage**

```
lavTestWald.mi(object, constraints = NULL, test = c("D1", "D2"),
  asymptotic = FALSE, scale.W = !asymptotic, omitimps = c("no.conv",
  "no.se"), verbose = FALSE, warn = TRUE)
```

**Arguments**

object	An object of class <code>lavaan.mi</code> .
constraints	A character string (typically between single quotes) containing one or more equality constraints. See examples for more details
test	character indicating which pooling method to use. "D1" or "Rubin" (default) indicates Rubin's (1987) rules will be applied to the point estimates and the asymptotic covariance matrix of model parameters, and those pooled values will be used to calculate the Wald test in the usual manner. "D2", "LMRR", or "Li.et.al" indicate that the complete-data Wald test statistic should be calculated using each imputed data set, which will then be pooled across imputations, as described in Li, Meng, Raghunathan, & Rubin (1991) and Enders (2010, chapter 8).
asymptotic	logical. If FALSE (default), the pooled test will be returned as an $F$ -distributed statistic with numerator (df1) and denominator (df2) degrees of freedom. If TRUE, the pooled $F$ statistic will be multiplied by its df1 on the assumption that its df2 is sufficiently large enough that the statistic will be asymptotically $\chi^2$ distributed with df1.
scale.W	logical. If FALSE, the pooled asymptotic covariance matrix of model parameters is calculated as the weighted sum of the within-imputation and between-imputation components. Otherwise, the pooled asymptotic covariance matrix of model parameters is calculated by scaling the within-imputation component by the average relative increase in variance (ARIV; see Enders, 2010, p. 235), which is <i>only</i> consistent when requesting the $F$ test (i.e., <code>asymptotic = FALSE</code> ). Ignored (irrelevant) if <code>test = "D2"</code> .
omitimps	character vector specifying criteria for omitting imputations from pooled results. Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option

("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. Specific imputation numbers can also be included in this argument, in case users want to apply their own custom omission criteria (or simulations can use different numbers of imputations without redundantly refitting the model).

verbose        logical. If TRUE, print the restriction matrix and the estimated restricted values.  
warn            logical. If TRUE, print warnings if they occur.

### Details

The constraints are specified using the "=" operator. Both the left-hand side and the right-hand side of the equality can contain a linear combination of model parameters, or a constant (like zero). The model parameters must be specified by their user-specified labels from the `link[lavaan]{model.syntax}`. Names of defined parameters (using the ":=" operator) can be included too.

### Value

A vector containing the Wald test statistic (either an F or  $\chi^2$  statistic, depending on the asymptotic argument), the degrees of freedom (numerator and denominator, if `asymptotic = FALSE`), and a *p* value. If `asymptotic = FALSE`, the relative increase in variance (RIV, or average for multiparameter tests: ARIV) used to calculate the denominator *df* is also returned as a missing-data diagnostic, along with the fraction missing information (FMI = ARIV / (1 + ARIV)).

### Author(s)

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Adapted from **lavaan** source code, written by Yves Rosseeel (Ghent University; <Yves.Rosseeel@UGent.be>)

### References

Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.

Li, K.-H., Meng, X.-L., Raghunathan, T. E., & Rubin, D. B. (1991). Significance levels from repeated *p*-values with multiply-imputed data. *Statistica Sinica*, 1(1), 65–92. Retrieved from <https://www.jstor.org/stable/24303994>

Rubin, D. B. (1987). *Multiple imputation for nonresponse in surveys*. New York, NY: Wiley.

### See Also

[lavTestWald](#)

### Examples

```
## Not run:
## impose missing data for example
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]
set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
```

```

age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## impute missing data
library(Amelia)
set.seed(12345)
HS.amelia <- amelia(HSMiss, m = 20, noms = "school", p2s = FALSE)
imps <- HS.amelia$imputations

## specify CFA model from lavaan's ?cfa help page
HS.model <- '
  visual =~ x1 + b1*x2 + x3
  textual =~ x4 + b2*x5 + x6
  speed   =~ x7 + b3*x8 + x9
'

fit <- cfa.mi(HS.model, data = imps)

## Testing whether a single parameter equals zero yields the 'chi-square'
## version of the Wald z statistic from the summary() output, or the
## 'F' version of the t statistic from the summary() output, depending
## whether asymptotic = TRUE or FALSE
lavTestWald.mi(fit, constraints = "b1 == 0")      # default D1 statistic
lavTestWald.mi(fit, constraints = "b1 == 0", test = "D2") # D2 statistic

## The real advantage is simultaneously testing several equality
## constraints, or testing more complex constraints:
con <- '
  2*b1 == b3
  b2 - b3 == 0
'

lavTestWald.mi(fit, constraints = con) # default F statistic
lavTestWald.mi(fit, constraints = con, asymptotic = TRUE) # chi-squared

## End(Not run)

```

---

loadingFromAlpha

*Find standardized factor loading from coefficient alpha*


---

## Description

Find standardized factor loading from coefficient alpha assuming that all items have equal loadings.

## Usage

```
loadingFromAlpha(alpha, ni)
```

**Arguments**

alpha	A desired coefficient alpha value.
ni	A desired number of items.

**Value**

result	The standardized factor loadings that make desired coefficient alpha with specified number of items.
--------	--

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**Examples**

```
loadingFromAlpha(0.8, 4)
```

---

mardiaKurtosis	<i>Finding Mardia's multivariate kurtosis</i>
----------------	---

---

**Description**

Finding Mardia's multivariate kurtosis of multiple variables

**Usage**

```
mardiaKurtosis(dat, use = "everything")
```

**Arguments**

dat	The target matrix or data frame with multiple variables
use	Missing data handling method from the <code>cov</code> function.

**Details**

The Mardia's multivariate kurtosis formula (Mardia, 1970) is

$$b_{2,d} = \frac{1}{n} \sum_{i=1}^n \left[ (\mathbf{X}_i - \bar{\mathbf{X}})' \mathbf{S}^{-1} (\mathbf{X}_i - \bar{\mathbf{X}}) \right]^2,$$

where  $d$  is the number of variables,  $X$  is the target dataset with multiple variables,  $n$  is the sample size,  $\mathbf{S}$  is the sample covariance matrix of the target dataset, and  $\bar{\mathbf{X}}$  is the mean vectors of the target dataset binded in  $n$  rows. When the population multivariate kurtosis is normal, the  $b_{2,d}$  is asymptotically distributed as normal distribution with the mean of  $d(d+2)$  and variance of  $8d(d+2)/n$ .

**Value**

A value of a Mardia's multivariate kurtosis with a test statistic

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**References**

Mardia, K. V. (1970). Measures of multivariate skewness and kurtosis with applications. *Biometrika*, 57(3), 519-530. doi:10.2307/2334770

**See Also**

- [skew](#) Find the univariate skewness of a variable
- [kurtosis](#) Find the univariate excessive kurtosis of a variable
- [mardiaSkew](#) Find the Mardia's multivariate skewness of a set of variables

**Examples**

```
library(lavaan)
mardiaKurtosis(HolzingerSwineford1939[ , paste0("x", 1:9)])
```

---

mardiaSkew

*Finding Mardia's multivariate skewness*

---

**Description**

Finding Mardia's multivariate skewness of multiple variables

**Usage**

```
mardiaSkew(dat, use = "everything")
```

**Arguments**

**dat** The target matrix or data frame with multiple variables

**use** Missing data handling method from the [cov](#) function.



**Details**

The Mardia's multivariate skewness formula (Mardia, 1970) is

$$b_{1,d} = \frac{1}{n^2} \sum_{i=1}^n \sum_{j=1}^n \left[ (\mathbf{X}_i - \bar{\mathbf{X}})' \mathbf{S}^{-1} (\mathbf{X}_j - \bar{\mathbf{X}}) \right]^3,$$

where  $d$  is the number of variables,  $X$  is the target dataset with multiple variables,  $n$  is the sample size,  $S$  is the sample covariance matrix of the target dataset, and  $\bar{X}$  is the mean vectors of the target dataset binded in  $n$  rows. When the population multivariate skewness is normal, the  $\frac{n}{6}b_{1,d}$  is asymptotically distributed as  $\chi^2$  distribution with  $d(d+1)(d+2)/6$  degrees of freedom.

**Value**

A value of a Mardia's multivariate skewness with a test statistic

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**References**

Mardia, K. V. (1970). Measures of multivariate skewness and kurtosis with applications. *Biometrika*, 57(3), 519-530. doi:10.2307/2334770

**See Also**

- [skew](#) Find the univariate skewness of a variable
- [kurtosis](#) Find the univariate excessive kurtosis of a variable
- [mardiaKurtosis](#) Find the Mardia's multivariate kurtosis of a set of variables

**Examples**

```
library(lavaan)
mardiaSkew(HolzingerSwineford1939[, paste0("x", 1:9)])
```

---

maximalRelia

*Calculate maximal reliability*


---

**Description**

Calculate maximal reliability of a scale

**Usage**

```
maximalRelia(object, omit.imps = c("no.conv", "no.se"))
```

## Arguments

object	A <a href="#">lavaan</a> or <a href="#">lavaan.mi</a> object, expected to contain only exogenous common factors (i.e., a CFA model).
omit.imps	character vector specifying criteria for omitting imputations from pooled results. Can include any of c("no.conv", "no.se", "no.npd"), the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

## Details

Given that a composite score ( $W$ ) is a weighted sum of item scores:

$$W = \mathbf{w}'\mathbf{x},$$

where  $\mathbf{x}$  is a  $k \times 1$  vector of the scores of each item,  $\mathbf{w}$  is a  $k \times 1$  weight vector of each item, and  $k$  represents the number of items. Then, maximal reliability is obtained by finding  $\mathbf{w}$  such that reliability attains its maximum (Li, 1997; Raykov, 2012). Note that the reliability can be obtained by

$$\rho = \frac{\mathbf{w}'\mathbf{S}_T\mathbf{w}}{\mathbf{w}'\mathbf{S}_X\mathbf{w}}$$

where  $\mathbf{S}_T$  is the covariance matrix explained by true scores and  $\mathbf{S}_X$  is the observed covariance matrix. Numerical method is used to find  $\mathbf{w}$  in this function.

For continuous items,  $\mathbf{S}_T$  can be calculated by

$$\mathbf{S}_T = \Lambda\Psi\Lambda',$$

where  $\Lambda$  is the factor loading matrix and  $\Psi$  is the covariance matrix among factors.  $\mathbf{S}_X$  is directly obtained by covariance among items.

For categorical items, Green and Yang's (2009) method is used for calculating  $\mathbf{S}_T$  and  $\mathbf{S}_X$ . The element  $i$  and  $j$  of  $\mathbf{S}_T$  can be calculated by

$$[\mathbf{S}_T]_{ij} = \sum_{c_i=1}^{C_i-1} \sum_{c_j=1}^{C_j-1} \Phi_2(\tau_{x_{c_i}}, \tau_{x_{c_j}}, [\Lambda\Psi\Lambda']_{ij}) - \sum_{c_i=1}^{C_i-1} \Phi_1(\tau_{x_{c_i}}) \sum_{c_j=1}^{C_j-1} \Phi_1(\tau_{x_{c_j}}),$$

where  $C_i$  and  $C_j$  represents the number of thresholds in Items  $i$  and  $j$ ,  $\tau_{x_{c_i}}$  represents the threshold  $c_i$  of Item  $i$ ,  $\tau_{x_{c_j}}$  represents the threshold  $c_j$  of Item  $j$ ,  $\Phi_1(\tau_{x_{c_i}})$  is the cumulative probability of

$\tau_{x_{c_i}}$  given a univariate standard normal cumulative distribution and  $\Phi_2(\tau_{x_{c_i}}, \tau_{x_{c_j}}, \rho)$  is the joint cumulative probability of  $\tau_{x_{c_i}}$  and  $\tau_{x_{c_j}}$  given a bivariate standard normal cumulative distribution with a correlation of  $\rho$

Each element of  $\mathbf{S}_X$  can be calculated by

$$[\mathbf{S}_T]_{ij} = \sum_{c_i=1}^{C_i-1} \sum_{c_j=1}^{C_j-1} \Phi_2(\tau_{V_{c_i}}, \tau_{V_{c_j}}, \rho_{ij}^*) - \sum_{c_i=1}^{C_i-1} \Phi_1(\tau_{V_{c_i}}) \sum_{c_j=1}^{C_j-1} \Phi_1(\tau_{V_{c_j}}),$$

where  $\rho_{ij}^*$  is a polychoric correlation between Items  $i$  and  $j$ .

### Value

Maximal reliability values of each group. The maximal-reliability weights are also provided. Users may extract the weights by the `attr` function (see example below).

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

### References

- Li, H. (1997). A unifying expression for the maximal reliability of a linear composite. *Psychometrika*, 62(2), 245–249. doi:10.1007/BF02295278
- Raykov, T. (2012). Scale construction and development using structural equation modeling. In R. H. Hoyle (Ed.), *Handbook of structural equation modeling* (pp. 472–494). New York, NY: Guilford.

### See Also

[reliability](#) for reliability of an unweighted composite score

### Examples

```
total <- 'f =~ x1 + x2 + x3 + x4 + x5 + x6 + x7 + x8 + x9 '
fit <- cfa(total, data = HolzingerSwineford1939)
maximalRelia(fit)

# Extract the weight
mr <- maximalRelia(fit)
attr(mr, "weight")
```

---

measEq.syntax                      *Syntax for measurement equivalence*

---

## Description

Automatically generates lavaan model syntax to specify a confirmatory factor analysis (CFA) model with equality constraints imposed on user-specified measurement (or structural) parameters. Optionally returns the fitted model (if data are provided) representing some chosen level of measurement equivalence/invariance across groups and/or repeated measures.

## Usage

```
measEq.syntax(configural.model, ..., ID.fac = "std.lv",
  ID.cat = "Wu.Estabrook.2016", ID.thr = c(1L, 2L), group = NULL,
  group.equal = "", group.partial = "", longFacNames = list(),
  longIndNames = list(), long.equal = "", long.partial = "",
  auto = "all", warn = TRUE, debug = FALSE, return.fit = FALSE)
```

## Arguments

`configural.model`

A model with no measurement-invariance constraints (i.e., representing only configural invariance), unless required for model identification. `configural.model` can be either:

- lavaan [model.syntax](#) or a parameter table (as returned by [parTable](#)) specifying the configural model. Using this option, the user can also provide either raw data or summary statistics via `sample.cov` and (optionally) `sample.mean`. See argument descriptions in [lavaan](#). In order to include thresholds in the generated syntax, either users must provide raw data, or the `configural.model` syntax must specify all thresholds (see first example). If raw data are not provided, the number of blocks (groups, levels, or combination) must be indicated using an arbitrary `sample.nobs` argument (e.g., 3 groups could be specified using `sample.nobs=rep(1,3)`).
- a fitted [lavaan](#) model (e.g., as returned by [cfa](#)) estimating the configural model

Note that the specified or fitted model must not contain any latent structural parameters (i.e., it must be a CFA model), unless they are higher-order constructs with latent indicators (i.e., a second-order CFA).

... Additional arguments (e.g., `data`, `ordered`, or parameterization) passed to the [lavaan](#) function. See also [lavOptions](#).

`ID.fac` character. The method for identifying common-factor variances and (if `meanstructure = TRUE`) means. Three methods are available, which go by different names in the literature:

- Standardize the common factor (mean = 0,  $SD = 1$ ) by specifying any of: `"std.lv"`, `"unit.variance"`, `"UV"`, `"fixed.factor"`, `"fixed-factor"`

- Choose a reference indicator by specifying any of: "auto.fix.first", "unit.loading", "UL", "marker", "ref", "ref.indicator", "reference.indicator", "reference-indicator", "marker.variable", "marker-variable"
- Apply effects-code constraints to loadings and intercepts by specifying any of: "FX", "EC", "effects", "effects.coding", "effects-coding", "effects.code", "effects-code"

See Kloessner & Klopp (2019) for details about all three methods.

ID.cat	<p>character. The method for identifying (residual) variances and intercepts of latent item-responses underlying any ordered indicators. Four methods are available:</p> <ul style="list-style-type: none"> <li>• To follow Wu &amp; Estabrook's (2016) guidelines (default), specify any of: "Wu.Estabrook.2016", "Wu.2016", "Wu.Estabrook", "Wu", "Wu2016".</li> <li>• To use the default settings of <i>Mplus</i> and lavaan, specify any of: "default", "Mplus", "Muthen". Details provided in Millsap &amp; Tein (2004).</li> <li>• To use the constraints recommended by Millsap &amp; Tein (2004; see also Liu et al., 2017, for the longitudinal case) specify any of: "millsap", "millsap.2004", "millsap.tein.2004"</li> <li>• To use the default settings of LISREL, specify "LISREL" or "Joreskog". Details provided in Millsap &amp; Tein (2004).</li> </ul> <p>See <b>Details</b> and <b>References</b> for more information.</p>
ID.thr	<p>integer. Only relevant when ID.cat = "Millsap.Tein.2004". Used to indicate which thresholds should be constrained for identification. The first integer indicates the threshold used for all indicators, the second integer indicates the additional threshold constrained for a reference indicator (ignored if binary).</p>
group	<p>optional character indicating the name of a grouping variable. See <a href="#">cfa</a>.</p>
group.equal	<p>optional character vector indicating type(s) of parameter to equate across groups. Ignored if is.null(group). See <a href="#">lavOptions</a>.</p>
group.partial	<p>optional character vector or a parameter table indicating exceptions to group.equal (see <a href="#">lavOptions</a>). Any variables not appearing in the configural.model will be ignored, and any parameter constraints needed for identification (e.g., two thresholds per indicator when ID.cat = "Millsap") will be removed.</p>
longFacNames	<p>optional named list of character vectors, each indicating multiple factors in the model that are actually the same construct measured repeatedly. See <b>Details</b> and <b>Examples</b>.</p>
longIndNames	<p>optional named list of character vectors, each indicating multiple indicators in the model that are actually the same indicator measured repeatedly. See <b>Details</b> and <b>Examples</b>.</p>
long.equal	<p>optional character vector indicating type(s) of parameter to equate across repeated measures. Ignored if no factors are indicated as repeatedly measured in longFacNames.</p>
long.partial	<p>optional character vector or a parameter table indicating exceptions to long.equal. Any longitudinal variable names not appearing in names(longFacNames) or names(longIndNames) will be ignored, and any parameter constraints needed for identification will be removed.</p>

auto	Used to automatically included autocorrelated measurement errors among repeatedly measured indicators in <code>longIndNames</code> . Specify a single integer to set the maximum order (e.g., <code>auto = 1L</code> indicates that an indicator's unique factors should only be correlated between adjacently measured occasions). <code>auto = TRUE</code> or "all" will specify residual covariances among all possible lags per repeatedly measured indicator in <code>longIndNames</code> .
warn, debug	logical. Passed to <code>lavaan</code> and <code>lavParseModelString</code> . See <code>lavOptions</code> .
return.fit	logical indicating whether the generated syntax should be fitted to the provided data (or summary statistics, if provided via <code>sample.cov</code> ). If <code>configural.model</code> is a fitted lavaan model, the generated syntax will be fitted using the update method (see <code>lavaan</code> ), and <code>...</code> will be passed to <code>lavaan</code> . If neither data nor a fitted lavaan model were provided, this must be FALSE. If TRUE, the generated <code>measEq.syntax</code> object will be included in the lavaan object's <code>@external</code> slot, accessible by <code>fit@external\$measEq.syntax</code> .

## Details

This function is a pedagogical and analytical tool to generate model syntax representing some level of measurement equivalence/invariance across any combination of multiple groups and/or repeated measures. Support is provided for confirmatory factor analysis (CFA) models with simple or complex structure (i.e., cross-loadings and correlated residuals are allowed). For any complexities that exceed the limits of automation, this function is intended to still be useful by providing a means to generate syntax that users can easily edit to accommodate their unique situations.

Limited support is provided for bifactor models and higher-order constructs. Because bifactor models have cross-loadings by definition, the option `ID.fac = "effects.code"` is unavailable. `ID.fac = "UV"` is recommended for bifactor models, but `ID.fac = "UL"` is available on the condition that each factor has a unique first indicator in the `configural.model`. In order to maintain generality, higher-order factors may include a mix of manifest and latent indicators, but they must therefore require `ID.fac = "UL"` to avoid complications with differentiating lower-order vs. higher-order (or mixed-level) factors. The keyword "loadings" in `group.equal` or `long.equal` constrains factor loadings of all manifest indicators (including loadings on higher-order factors that also have latent indicators), whereas the keyword "regressions" constrains factor loadings of latent indicators. Users can edit the model syntax manually to adjust constraints as necessary, or clever use of the `group.partial` or `long.partial` arguments could make it possible for users to still automated their model syntax. The keyword "intercepts" constrains the intercepts of all manifest indicators, and the keyword "means" constrains intercepts and means of all latent common factors, regardless of whether they are latent indicators of higher-order factors. To test equivalence of lower-order and higher-order intercepts/means in separate steps, the user can either manually edit their generated syntax or conscientiously exploit the `group.partial` or `long.partial` arguments as necessary.

**ID.fac:** If the `configural.model` fixes any (e.g., the first) factor loadings, the generated syntax object will retain those fixed values. This allows the user to retain additional constraints that might be necessary (e.g., if there are only 1 or 2 indicators). Some methods must be used in conjunction with other settings:

- `ID.cat = "Millsap"` requires `ID.fac = "UL"` and `parameterization = "theta"`.
- `ID.cat = "LISREL"` requires `parameterization = "theta"`.
- `ID.fac = "effects.code"` is unavailable when there are any cross-loadings.

ID.cat: Wu & Estabrook (2016) recommended constraining thresholds to equality first, and doing so should allow releasing any identification constraints no longer needed. For each ordered indicator, constraining one threshold to equality will allow the item's intercepts to be estimated in all but the first group or repeated measure. Constraining a second threshold (if applicable) will allow the item's (residual) variance to be estimated in all but the first group or repeated measure. For binary data, there is no independent test of threshold, intercept, or residual-variance equality. Equivalence of thresholds must also be assumed for three-category indicators. These guidelines provide the least restrictive assumptions and tests, and are therefore the default.

The default setting in *Mplus* is similar to Wu & Estabrook (2016), except that intercepts are always constrained to zero (so they are assumed to be invariant without testing them). Millsap & Tein (2004) recommended parameterization = "theta" and identified an item's residual variance in all but the first group (or occasion; Liu et al., 2017) by constraining its intercept to zero and one of its thresholds to equality. A second threshold for the reference indicator (so ID.fac = "UL") is used to identify the common-factor means in all but the first group/occasion. The LISREL software fixes the first threshold to zero and (if applicable) the second threshold to 1, and assumes any remaining thresholds to be equal across groups / repeated measures; thus, the intercepts are always identified, and residual variances (parameterization = "theta") are identified except for binary data, when they are all fixed to one.

**Repeated Measures:** If each repeatedly measured factor is measured by the same indicators (specified in the same order in the configural.model) on each occasion, without any cross-loadings, the user can let longIndNames be automatically generated. Generic names for the repeatedly measured indicators are created using the name of the repeatedly measured factors (i.e., names(longFacNames)) and the number of indicators. So the repeatedly measured first indicator ("ind") of a longitudinal construct called "factor" would be generated as ".\_factor\_ind.1".

The same types of parameter can be specified for long.equal as for group.equal (see [lavOptions](#) for a list), except for "residual.covariances" or "lv.covariances". Instead, users can constrain autocovariances using keywords "resid.autocov" or "lv.autocov". Note that group.equal = "lv.covariances" or group.equal = "residual.covariances" will constrain any autocovariances across groups, along with any other covariances the user specified in the configural.model. Note also that autocovariances cannot be specified as exceptions in long.partial, so anything more complex than the auto argument automatically provides should instead be manually specified in the configural.model.

When users set orthogonal=TRUE in the configural.model (e.g., in bifactor models of repeatedly measured constructs), autocovariances of each repeatedly measured factor will still be freely estimated in the generated syntax.

**Missing Data:** If users wish to utilize the [auxiliary](#) function to automatically include auxiliary variables in conjunction with missing = "FIML", they should first generate the hypothesized-model syntax, then submit that syntax as the model to auxiliary(). If users utilized [runMI](#) to fit their configural.model to multiply imputed data, that model can also be passed to the configural.model argument, and if return.fit = TRUE, the generated model will be fitted to the multiple imputations.

## Value

By default, an object of class [measEq.syntax](#). If return.fit = TRUE, a fitted [lavaan](#) model, with the measEq.syntax object stored in the @external slot, accessible by fit@external\$measEq.syntax.

**Author(s)**

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**References**

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- Wu, H., & Estabrook, R. (2016). Identification of confirmatory factor analysis models of different levels of invariance for ordered categorical outcomes. *Psychometrika*, 81(4), 1014–1045. doi:10.1007/s11336-016-9506-0

**See Also**

[compareFit](#)

**Examples**

```
mod.cat <- ' FU1 =~ u1 + u2 + u3 + u4
           FU2 =~ u5 + u6 + u7 + u8 '
## the 2 factors are actually the same factor (FU) measured twice
longFacNames <- list(FU = c("FU1", "FU2"))

## CONFIGURAL model: no constraints across groups or repeated measures
syntax.config <- measEq.syntax(configural.model = mod.cat,
                               # NOTE: data provides info about numbers of
                               #       groups and thresholds
                               data = datCat,
                               ordered = paste0("u", 1:8),
                               parameterization = "theta",
                               ID.fac = "std.lv", ID.cat = "Wu.Estabrook.2016",
                               group = "g", longFacNames = longFacNames)

## print lavaan syntax to the Console
cat(as.character(syntax.config))
## print a summary of model features
summary(syntax.config)

## THRESHOLD invariance:
## only necessary to specify thresholds if you have no data
mod.th <- '
  u1 | t1 + t2 + t3 + t4
  u2 | t1 + t2 + t3 + t4
  u3 | t1 + t2 + t3 + t4
  u4 | t1 + t2 + t3 + t4
  u5 | t1 + t2 + t3 + t4
```



```

    u6 | t1 + t2 + t3 + t4
    u7 | t1 + t2 + t3 + t4
    u8 | t1 + t2 + t3 + t4
  ,
syntax.thresh <- measEq.syntax(configural.model = c(mod.cat, mod.th),
  # NOTE: data not provided, so syntax must
  #       include thresholds, and number of
  #       groups == 2 is indicated by:
  sample.nobs = c(1, 1),
  parameterization = "theta",
  ID.fac = "std.lv", ID.cat = "Wu.Estabrook.2016",
  group = "g", group.equal = "thresholds",
  longFacNames = longFacNames,
  long.equal = "thresholds")
## notice that constraining 4 thresholds allows intercepts and residual
## variances to be freely estimated in all but the first group & occasion
cat(as.character(syntax.thresh))
## print a summary of model features
summary(syntax.thresh)

## Fit a model to the data either in a subsequent step (recommended):
mod.config <- as.character(syntax.config)
fit.config <- cfa(mod.config, data = datCat, group = "g",
  ordered = paste0("u", 1:8), parameterization = "theta")
## or in a single step (not generally recommended):
fit.thresh <- measEq.syntax(configural.model = mod.cat, data = datCat,
  ordered = paste0("u", 1:8),
  parameterization = "theta",
  ID.fac = "std.lv", ID.cat = "Wu.Estabrook.2016",
  group = "g", group.equal = "thresholds",
  longFacNames = longFacNames,
  long.equal = "thresholds", return.fit = TRUE)
## compare their fit to test threshold invariance
anova(fit.config, fit.thresh)

## -----
## RECOMMENDED PRACTICE: fit one invariance model at a time
## -----

## - A downside of setting return.fit=TRUE is that if the model has trouble
##   converging, you don't have the opportunity to investigate the syntax,
##   or even to know whether an error resulted from the syntax-generator or
##   from lavaan itself.
## - A downside of automatically fitting an entire set of invariance models
##   (like the old measurementInvariance() function did) is that you might
##   end up testing models that shouldn't even be fitted because less
##   restrictive models already fail (e.g., don't test full scalar
##   invariance if metric invariance fails! Establish partial metric
##   invariance first, then test equivalent of intercepts ONLY among the
##   indicators that have invariate loadings.)

```

```

## The recommended sequence is to (1) generate and save each syntax object,
## (2) print it to the screen to verify you are fitting the model you expect
## to (and potentially learn which identification constraints should be
## released when equality constraints are imposed), and (3) fit that model
## to the data, as you would if you had written the syntax yourself.

## Continuing from the examples above, after establishing invariance of
## thresholds, we proceed to test equivalence of loadings and intercepts
## (metric and scalar invariance, respectively)
## simultaneously across groups and repeated measures.

## Not run:

## metric invariance
syntax.metric <- measEq.syntax(configural.model = mod.cat, data = datCat,
                              ordered = paste0("u", 1:8),
                              parameterization = "theta",
                              ID.fac = "std.lv", ID.cat = "Wu.Estabrook.2016",
                              group = "g", longFacNames = longFacNames,
                              group.equal = c("thresholds", "loadings"),
                              long.equal = c("thresholds", "loadings"))
summary(syntax.metric) # summarize model features
mod.metric <- as.character(syntax.metric) # save as text
cat(mod.metric) # print/view lavaan syntax
## fit model to data
fit.metric <- cfa(mod.metric, data = datCat, group = "g",
                 ordered = paste0("u", 1:8), parameterization = "theta")
## test equivalence of loadings, given equivalence of thresholds
anova(fit.thresh, fit.metric)

## scalar invariance
syntax.scalar <- measEq.syntax(configural.model = mod.cat, data = datCat,
                              ordered = paste0("u", 1:8),
                              parameterization = "theta",
                              ID.fac = "std.lv", ID.cat = "Wu.Estabrook.2016",
                              group = "g", longFacNames = longFacNames,
                              group.equal = c("thresholds", "loadings",
                                              "intercepts"),
                              long.equal = c("thresholds", "loadings",
                                              "intercepts"))
summary(syntax.scalar) # summarize model features
mod.scalar <- as.character(syntax.scalar) # save as text
cat(mod.scalar) # print/view lavaan syntax
## fit model to data
fit.scalar <- cfa(mod.scalar, data = datCat, group = "g",
                 ordered = paste0("u", 1:8), parameterization = "theta")
## test equivalence of intercepts, given equal thresholds & loadings
anova(fit.metric, fit.scalar)

## For a single table with all results, you can pass the models to
## summarize to the compareFit() function
compareFit(fit.config, fit.thresh, fit.metric, fit.scalar)

```

```

## -----
## NOT RECOMMENDED: fit several invariance models at once
## -----
test.seq <- c("thresholds","loadings","intercepts","means","residuals")
meq.list <- list()
for (i in 0:length(test.seq)) {
  if (i == 0L) {
    meq.label <- "configural"
    group.equal <- ""
    long.equal <- ""
  } else {
    meq.label <- test.seq[i]
    group.equal <- test.seq[1:i]
    long.equal <- test.seq[1:i]
  }
  meq.list[[meq.label]] <- measEq.syntax(configural.model = mod.cat,
                                       data = datCat,
                                       ordered = paste0("u", 1:8),
                                       parameterization = "theta",
                                       ID.fac = "std.lv",
                                       ID.cat = "Wu.Estabrook.2016",
                                       group = "g",
                                       group.equal = group.equal,
                                       longFacNames = longFacNames,
                                       long.equal = long.equal,
                                       return.fit = TRUE)
}

compareFit(meq.list)

## -----
## Binary indicators
## -----

## borrow example data from Mplus user guide
myData <- read.table("http://www.statmodel.com/usersguide/chap5/ex5.16.dat")
names(myData) <- c("u1","u2","u3","u4","u5","u6","x1","x2","x3","g")
bin.mod <- '
  FU1 =~ u1 + u2 + u3
  FU2 =~ u4 + u5 + u6
'

## Must SIMULTANEOUSLY constrain thresholds, loadings, and intercepts
test.seq <- list(strong = c("thresholds","loadings","intercepts"),
                means = "means",
                strict = "residuals")
meq.list <- list()
for (i in 0:length(test.seq)) {
  if (i == 0L) {
    meq.label <- "configural"

```

```

    group.equal <- ""
    long.equal <- ""
  } else {
    meq.label <- names(test.seq)[i]
    group.equal <- unlist(test.seq[1:i])
    # long.equal <- unlist(test.seq[1:i])
  }
  meq.list[[meq.label]] <- measEq.syntax(configural.model = bin.mod,
                                       data = myData,
                                       ordered = paste0("u", 1:6),
                                       parameterization = "theta",
                                       ID.fac = "std.lv",
                                       ID.cat = "Wu.Estabrook.2016",
                                       group = "g",
                                       group.equal = group.equal,
                                       #longFacNames = longFacNames,
                                       #long.equal = long.equal,
                                       return.fit = TRUE)
}

compareFit(meq.list)

## -----
## Multilevel Invariance
## -----

## To test invariance across levels in a MLSEM, specify syntax as though
## you are fitting to 2 groups instead of 2 levels.

mlsem <- ' f1 =~ y1 + y2 + y3
          f2 =~ y4 + y5 + y6 '
## metric invariance
syntax.metric <- measEq.syntax(configural.model = mlsem, meanstructure = TRUE,
                               ID.fac = "std.lv", sample.nobs = c(1, 1),
                               group = "cluster", group.equal = "loadings")
## by definition, Level-1 means must be zero, so fix them
syntax.metric <- update(syntax.metric,
                       change.syntax = paste0("y", 1:6, " ~ c(0, NA)*1"))
## save as a character string
mod.metric <- as.character(syntax.metric, groups.as.blocks = TRUE)
## convert from multigroup to multilevel
mod.metric <- gsub(pattern = "group:", replacement = "level:",
                  x = mod.metric, fixed = TRUE)
## fit model to data
fit.metric <- lavaan(mod.metric, data = Demo.twolevel, cluster = "cluster")
summary(fit.metric)

## End(Not run)

```

## Description

This class of object stores information used to automatically generate lavaan model syntax to represent user-specified levels of measurement equivalence/invariance across groups and/or repeated measures. See [measEq.syntax](#) for details.

## Usage

```
## S4 method for signature 'measEq.syntax'
as.character(x, package = "lavaan",
  params = NULL, single = TRUE, groups.as.blocks = FALSE)

## S4 method for signature 'measEq.syntax'
show(object)

## S4 method for signature 'measEq.syntax'
summary(object, verbose = TRUE)

## S4 method for signature 'measEq.syntax'
update(object, ..., evaluate = TRUE, change.syntax = NULL)
```

## Arguments

x, object	an object of class <code>measEq.syntax</code>
package	character indicating the package for which the model syntax should be generated. Currently, only "lavaan" and "mplus" are supported.
params	character vector indicating which type(s) of parameter to print syntax for. Must match a type that can be passed to <code>group.equal</code> or <code>long.equal</code> , but "residual.covariances" and "lv.covariances" will be silently ignored. Instead, requesting "residuals" or "lv.variances" will return covariances along with variances. By default (NULL), all types are printed.
single	logical indicating whether to concatenate lavaan <a href="#">model.syntax</a> into a single character string. Setting FALSE will return a vector of strings, which may be convenient (or even necessary to prevent an error) in models with long variable names, many variables, or many groups.
groups.as.blocks	logical indicating whether to write lavaan <a href="#">model.syntax</a> using vectors of labels and values for multiple groups (the default: FALSE), or whether to write a separate "block" of syntax per group. The block structure could allow users to apply the generated multigroup syntax (after some editing) to test invariance across levels in a multilevel SEM (see final example on <a href="#">measEq.syntax</a> help page).
verbose	logical indicating whether to print a summary to the screen (default). If FALSE, only a pattern matrix is returned.
...	Additional arguments to the call, or arguments with changed values.
evaluate	If TRUE, evaluate the new call; otherwise, return the new call.

`change.syntax` lavaan [model.syntax](#) specifying labels or fixed/free values of parameters in object. These provide some flexibility to customize existing parameters without having to copy/paste the output of `as.character(object)` into an R script. For example, `group.partial` will free a parameter across all groups, but `update` allows users to free the parameter in just one group while maintaining equality constraints among other groups.

## Value

`summary` `signature(object = "measEq.syntax", verbose = TRUE)`: A character matrix indicating the pattern of numeric, ordered, or latent indicators loading on common factors. By default (`verbose = TRUE`), `summary` also prints descriptive details about the model, including the numbers of indicators and factors, and which parameters are constrained to equality.

`show` `signature(object = "measEq.syntax")`: Prints a message about how to use the object for model fitting. Invisibly returns the object.

`update` `signature(object = "measEq.syntax"), ..., evaluate = TRUE, change.syntax = NULL`: Creates a new object with updated arguments in `...`, or updated parameter labels or fixed/free specifications in object.

`as.character` `signature(x = "measEq.syntax", package = "lavaan")`: Converts the `measEq.syntax` object to model syntax that can be copy/pasted or written to a syntax file to be edited before analysis, or simply passed to [lavaan](#) to fit the model to data. Generated *Mplus* syntax could also be utilized using the **MplusAutomation** package.

## Slots

`package` character indicating the software package used to represent the model. Currently, only "lavaan" is available, which uses the LISREL representation (see [lavOptions](#)). In the future, "OpenMx" may become available, using RAM representation.

`model.type` character. Currently, only "cfa" is available. Future versions may allow for MIMIC / RFA models, where invariance can be tested across levels of exogenous variables explicitly included as predictors of indicators, controlling for their effects on (or correlation with) the common factors.

`call` The function call as returned by `match.call()`, with some arguments updated if necessary for logical consistency.

`meanstructure` logical indicating whether a mean structure is included in the model.

`numeric` character vector naming numeric manifest indicators.

`ordered` character vector naming ordered indicators.

`parameterization` character. See [lavOptions](#).

`specify` list of parameter matrices, similar in form to the output of `lavInspect(fit, "free")`. These matrices are logical, indicating whether each parameter should be specified in the model syntax.

`values` list of parameter matrices, similar in form to the output of `lavInspect(fit, "free")`. These matrices are numeric, indicating whether each parameter should be freely estimated (indicated by NA) or fixed to a particular value.

labels list of parameter matrices, similar in form to the output of `lavInspect(fit, "free")`. These matrices contain character labels used to constrain parameters to equality.

constraints character vector containing additional equality constraints used to identify the model when `ID.fac = "fx"`.

ngroups integer indicating the number of groups.

### Author(s)

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### Examples

```
## See ?measEq.syntax help page for examples using lavaan
```

---

miPowerFit

*Modification indices and their power approach for model fit evaluation*

---

### Description

The model fit evaluation approach using modification indices and expected parameter changes.

### Usage

```
miPowerFit(lavaanObj, stdLoad = 0.4, cor = 0.1, stdBeta = 0.1,
  intcept = 0.2, stdDelta = NULL, delta = NULL, cilevel = 0.9)
```

### Arguments

lavaanObj	The lavaan model object used to evaluate model fit
stdLoad	The amount of standardized factor loading that one would like to be detected (rejected). The default value is 0.4, which is suggested by Saris and colleagues (2009, p. 571).
cor	The amount of factor or error correlations that one would like to be detected (rejected). The default value is 0.1, which is suggested by Saris and colleagues (2009, p. 571).
stdBeta	The amount of standardized regression coefficients that one would like to be detected (rejected). The default value is 0.1, which is suggested by Saris and colleagues (2009, p. 571).
intcept	The amount of standardized intercept (similar to Cohen's <i>d</i> that one would like to be detected (rejected). The default value is 0.2, which is equivalent to a low effect size proposed by Cohen (1988, 1992).

stdDelta	The vector of the standardized parameters that one would like to be detected (rejected). If this argument is specified, the value here will overwrite the other arguments above. The order of the vector must be the same as the row order from modification indices from the lavaan object. If a single value is specified, the value will be applied to all parameters.
delta	The vector of the unstandardized parameters that one would like to be detected (rejected). If this argument is specified, the value here will overwrite the other arguments above. The order of the vector must be the same as the row order from modification indices from the lavaan object. If a single value is specified, the value will be applied to all parameters.
cilevel	The confidence level of the confidence interval of expected parameter changes. The confidence intervals are used in the equivalence testing.

### Details

In the lavaan object, one can inspect the modification indices and expected parameter changes. Those values can be used to evaluate model fit by two methods.

First, Saris, Satorra, and van der Veld (2009, pp. 570-573) used the power to detect modification indices and expected parameter changes to evaluate model fit. First, one should evaluate whether the modification index of each parameter is significant. Second, one should evaluate whether the power to detect a target expected parameter change is high enough. If the modification index is not significant and the power is high, there is no misspecification. If the modification index is significant and the power is low, the fixed parameter is misspecified. If the modification index is significant and the power is high, the expected parameter change is investigated. If the expected parameter change is large (greater than the target expected parameter change), the parameter is misspecified. If the expected parameter change is low (lower than the target expected parameter change), the parameter is not misspecified. If the modification index is not significant and the power is low, the decision is inconclusive.

Second, the confidence intervals of the expected parameter changes are formed. These confidence intervals are compared with the range of trivial misspecification, which could be  $(-\text{delta}, \text{delta})$  or  $(0, \text{delta})$  for nonnegative parameters. If the confidence intervals are outside of the range of trivial misspecification, the fixed parameters are severely misspecified. If the confidence intervals are inside the range of trivial misspecification, the fixed parameters are trivially misspecified. If confidence intervals are overlapped the range of trivial misspecification, the decision is inconclusive.

### Value

A data frame with these variables:

1. lhs: The left-hand side variable, with respect to the operator in in the lavaan `model.syntax`
2. op: The lavaan syntax operator: "`~~`" represents covariance, "`=~`" represents factor loading, "`~`" represents regression, and "`~1`" represents intercept.
3. rhs: The right-hand side variable
4. group: The level of the group variable for the parameter in question
5. mi: The modification index of the fixed parameter
6. epc: The expected parameter change if the parameter is freely estimated



7. target.epc: The target expected parameter change that represents the minimum size of misspecification that one would like to be detected by the test with a high power
8. std.epc: The standardized expected parameter change if the parameter is freely estimated
9. std.target.epc: The standardized target expected parameter change
10. significant.mi: Represents whether the modification index value is significant
11. high.power: Represents whether the power is enough to detect the target expected parameter change
12. decision.pow: The decision whether the parameter is misspecified or not based on Saris et al's method: "M" represents the parameter is misspecified, "NM" represents the parameter is not misspecified, "EPC:M" represents the parameter is misspecified decided by checking the expected parameter change value, "EPC:NM" represents the parameter is not misspecified decided by checking the expected parameter change value, and "I" represents the decision is inconclusive.
13. se.epc: The standard errors of the expected parameter changes.
14. lower.epc: The lower bound of the confidence interval of expected parameter changes.
15. upper.epc: The upper bound of the confidence interval of expected parameter changes.
16. lower.std.epc: The lower bound of the confidence interval of standardized expected parameter changes.
17. upper.std.epc: The upper bound of the confidence interval of standardized expected parameter changes.
18. decision.ci: The decision whether the parameter is misspecified or not based on the confidence interval method: "M" represents the parameter is misspecified, "NM" represents the parameter is not misspecified, and "I" represents the decision is inconclusive.

The row numbers matches with the results obtained from the `inspect(object, "mi")` function.

#### Author(s)

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#### References

- Cohen, J. (1988). *Statistical power analysis for the behavioral sciences* (2nd ed.). Hillsdale, NJ: Erlbaum.
- Cohen, J. (1992). A power primer. *Psychological Bulletin*, *112*(1), 155–159. doi:10.1037/0033-2909.112.1.155
- Saris, W. E., Satorra, A., & van der Veld, W. M. (2009). Testing structural equation models or detection of misspecifications? *Structural Equation Modeling*, *16*(4), 561–582. doi:10.1080/10705510903203433

#### See Also

[moreFitIndices](#) For the additional fit indices information

## Examples

```

library(lavaan)

HS.model <- ' visual  =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed   =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data = HolzingerSwineford1939,
           group = "sex", meanstructure = TRUE)
miPowerFit(fit)

model <- '
# latent variable definitions
ind60 =~ x1 + x2 + x3
dem60 =~ y1 + a*y2 + b*y3 + c*y4
dem65 =~ y5 + a*y6 + b*y7 + c*y8

# regressions
dem60 ~ ind60
dem65 ~ ind60 + dem60

# residual correlations
y1 ~~ y5
y2 ~~ y4 + y6
y3 ~~ y7
y4 ~~ y8
y6 ~~ y8
,

fit2 <- sem(model, data = PoliticalDemocracy, meanstructure = TRUE)
miPowerFit(fit2, stdLoad = 0.3, cor = 0.2, stdBeta = 0.2, intcept = 0.5)

```

---

modindices.mi

*Modification Indices for Multiple Imputations*

---

## Description

Modification indices (*1-df* Lagrange multiplier tests) from a latent variable model fitted to multiple imputed data sets. Statistics for releasing one or more fixed or constrained parameters in model can be calculated by pooling the gradient and information matrices across imputed data sets in a method proposed by Mansolf, Jorgensen, & Enders (in press)—analogous to the "D1" Wald test proposed by Li, Meng, Raghunathan, & Rubin (1991)—or by pooling the complete-data score-test statistics across imputed data sets (i.e., "D2"; Li et al., 1991).

## Usage

```

modindices.mi(object, test = c("D2", "D1"), omit.imps = c("no.conv",
  "no.se"), standardized = TRUE, cov.std = TRUE,

```

```
information = "expected", power = FALSE, delta = 0.1, alpha = 0.05,
high.power = 0.75, sort. = FALSE, minimum.value = 0,
maximum.number = nrow(LIST), na.remove = TRUE, op = NULL)
```

```
modificationIndices.mi(object, test = c("D2", "D1"),
omitimps = c("no.conv", "no.se"), standardized = TRUE, cov.std = TRUE,
information = "expected", power = FALSE, delta = 0.1, alpha = 0.05,
high.power = 0.75, sort. = FALSE, minimum.value = 0,
maximum.number = nrow(LIST), na.remove = TRUE, op = NULL)
```

## Arguments

object	An object of class <a href="#">lavaan.mi</a>
test	character indicating which pooling method to use. "D1" requests Mansolf, Jorgensen, & Enders' (in press) proposed Wald-like test for pooling the gradient and information, which are then used to calculate score-test statistics in the usual manner. "D2" (default because it is less computationally intensive) requests to pool the complete-data score-test statistics from each imputed data set, then pool them across imputations, described by Li et al. (1991) and Enders (2010).
omitimps	character vector specifying criteria for omitting imputations from pooled results. Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. Specific imputation numbers can also be included in this argument, in case users want to apply their own custom omission criteria (or simulations can use different numbers of imputations without redundantly refitting the model).
standardized	logical. If TRUE, two extra columns ( <code>\$sepc.lv</code> and <code>\$sepc.all</code> ) will contain standardized values for the EPCs. In the first column ( <code>\$sepc.lv</code> ), standardization is based on the variances of the (continuous) latent variables. In the second column ( <code>\$sepc.all</code> ), standardization is based on both the variances of both (continuous) observed and latent variables. (Residual) covariances are standardized using (residual) variances.
cov.std	logical. TRUE if <code>test == "D2"</code> . If TRUE (default), the (residual) observed covariances are scaled by the square-root of the diagonal elements of the $\Theta$ matrix, and the (residual) latent covariances are scaled by the square-root of the diagonal elements of the $\Psi$ matrix. If FALSE, the (residual) observed covariances are scaled by the square-root of the diagonal elements of the model-implied covariance matrix of observed variables ( $\Sigma$ ), and the (residual) latent covariances are scaled by the square-root of the diagonal elements of the model-implied covariance matrix of the latent variables.
information	character indicating the type of information matrix to use (check <a href="#">lavInspect</a> for available options). "expected" information is the default, which provides better control of Type I errors.
power	logical. If TRUE, the (post-hoc) power is computed for each modification index, using the values of delta and alpha.

delta	The value of the effect size, as used in the post-hoc power computation, currently using the unstandardized metric of the \$epc column.
alpha	The significance level used for deciding if the modification index is statistically significant or not.
high.power	If the computed power is higher than this cutoff value, the power is considered 'high'. If not, the power is considered 'low'. This affects the values in the \$decision column in the output.
sort.	logical. If TRUE, sort the output using the values of the modification index values. Higher values appear first.
minimum.value	numeric. Filter output and only show rows with a modification index value equal or higher than this minimum value.
maximum.number	integer. Filter output and only show the first maximum number rows. Most useful when combined with the sort. option.
na.remove	logical. If TRUE (default), filter output by removing all rows with NA values for the modification indices.
op	character string. Filter the output by selecting only those rows with operator op.

### Value

A data.frame containing modification indices and (S)EPCs.

### Note

When test = "D2", each (S)EPC will be pooled by taking its average across imputations. When test = "D1", EPCs will be calculated in the standard way using the pooled gradient and information, and SEPCs will be calculated by standardizing the EPCs using model-implied (residual) variances.

### Author(s)

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Adapted from **lavaan** source code, written by Yves Rosseel (Ghent University; <Yves.Rosseel@UGent.be>)

test = "D1" method proposed by Maxwell Mansolf (University of California, Los Angeles; <mamansolf@gmail.com>)

### References

Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.

Li, K.-H., Meng, X.-L., Raghunathan, T. E., & Rubin, D. B. (1991). Significance levels from repeated *p*-values with multiply-imputed data. *Statistica Sinica*, 1(1), 65–92. Retrieved from <https://www.jstor.org/stable/24303>

Mansolf, M., Jorgensen, T. D., & Enders, C. K. (in press). A multiple imputation score test for model modification in structural equation models. *Psychological Methods*. doi:10.1037/met0000243

## Examples

```
## Not run:
## impose missing data for example
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]

set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## impute missing data
library(Amelia)
set.seed(12345)
HS.amelia <- amelia(HSMiss, m = 20, noms = "school", p2s = FALSE)
imps <- HS.amelia$imputations

## specify CFA model from lavaan's ?cfa help page
HS.model <- '
  visual =~ x1 + x2 + x3
  textual =~ x4 + x5 + x6
  speed   =~ x7 + x8 + x9
'

out <- cfa.mi(HS.model, data = imps)

modindices.mi(out) # default: Li et al.'s (1991) "D2" method
modindices.mi(out, test = "D1") # Li et al.'s (1991) "D1" method

## End(Not run)
```

## Description

This function takes an expression for an indirect effect, the parameters and standard errors associated with the expression and returns a confidence interval based on a Monte Carlo test of mediation (MacKinnon, Lockwood, & Williams, 2004).

## Usage

```
monteCarloMed(expression, ..., ACM = NULL, object = NULL, rep = 20000,
              CI = 95, plot = FALSE, outputValues = FALSE)
```

**Arguments**

expression	A character scalar representing the computation of an indirect effect. Different parameters in the expression should have different alphanumeric values. Expressions can use either addition (+) or multiplication (*) operators.
...	Parameter estimates for all parameters named in expression. The order of parameters should follow from expression (the first parameter named in expression should be the first parameter listed in ...). Alternatively ... can be a vector of parameter estimates.
ACM	A matrix representing the asymptotic covariance matrix of the parameters described in expression. This matrix should be a symmetric matrix with dimensions equal to the number of parameters names in expression. Information on finding the ACOV is popular SEM software is described below.)
object	A lavaan model object fitted after running the running the cfa, sem, growth, or lavaan functions. The model must have parameters labelled with the same labels used in expression. When using this option do not specify values for ... or ACM
rep	The number of replications to compute. Many thousand are recommended.
CI	Width of the confidence interval computed.
plot	Should the function output a plot of simulated values of the indirect effect?
outputValues	Should the function output all simulated values of the indirect effect?

**Details**

This function implements the Monte Carlo test of mediation first described in MacKinnon, Lockwood, & Williams (2004) and extends it to complex cases where the indirect effect is more than a function of two parameters. The function takes an expression for the indirect effect, randomly simulated values of the indirect effect based on the values of the parameters (and the associated standard errors) comprising the indirect effect, and outputs a confidence interval of the indirect effect based on the simulated values. For further information on the Monte Carlo test of mediation see MacKinnon, Lockwood, & Williams (2004) and Preacher & Selig (2012).

The asymptotic covariance matrix can be easily found in many popular SEM software applications.

- LISREL: Including the EC option on the OU line will print the ACM to a separate file. The file contains the lower triangular elements of the ACM in free format and scientific notation
- Mplus Include the command TECH3; in the OUTPUT section. The ACM will be printed in the output.
- lavaan: Use the command vcov on the fitted lavaan object to print the ACM to the screen

**Value**

A list with two elements. The first element is the point estimate for the indirect effect. The second element is a matrix with values for the upper and lower limits of the confidence interval generated from the Monte Carlo test of mediation. If outputValues = TRUE, output will be a list with a list with the point estimate and values for the upper and lower limits of the confidence interval as the first element and a vector of simulated values of the indirect effect as the second element.

**Author(s)**

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Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

MacKinnon, D. P., Lockwood, C. M., & Williams, J. (2004). Confidence limits for the indirect effect: Distribution of the product and resampling methods. *Multivariate Behavioral Research*, 39(1) 99–128. doi:10.1207/s15327906mbr3901\_4

Preacher, K. J., & Selig, J. P. (2010, July). Monte Carlo method for assessing multilevel mediation: An interactive tool for creating confidence intervals for indirect effects in 1-1-1 multilevel models [Computer software]. Available from <http://quantpsy.org/>.

Preacher, K. J., & Selig, J. P. (2012). Advantages of Monte Carlo confidence intervals for indirect effects. *Communication Methods and Measures*, 6(2), 77–98. doi:10.1080/19312458.2012.679848

Selig, J. P., & Preacher, K. J. (2008, June). Monte Carlo method for assessing mediation: An interactive tool for creating confidence intervals for indirect effects [Computer software]. Available from <http://quantpsy.org/>.

**Examples**

```
## Simple two path mediation
## Write expression of indirect effect
med <- 'a*b'
## Parameter values from analyses
aparam <- 1
bparam <- 2
## Asymptotic covariance matrix from analyses
AC <- matrix(c(.01,.00002,
               .00002,.02), nrow=2, byrow=TRUE)
## Compute CI, include a plot
monteCarloMed(med, coef1 = aparam, coef2 = bparam, outputValues = FALSE,
              plot = TRUE, ACM = AC)

## Use a vector of parameter estimates as input
aparam <- c(1,2)
monteCarloMed(med, coef1 = aparam, outputValues = FALSE,
              plot = TRUE, ACM = AC)

## Complex mediation with two paths for the indirect effect
## Write expression of indirect effect
med <- 'a1*b1 + a1*b2'
## Parameter values and standard errors from analyses
aparam <- 1
b1param <- 2
b2param <- 1
```

```

## Asymptotic covariance matrix from analyses
AC <- matrix(c(1, .00002, .00003,
               .00002, 1, .00002,
               .00003, .00002, 1), nrow = 3, byrow = TRUE)
## Compute CI do not include a plot
monteCarloMed(med, coef1 = aparam, coef2 = b1param,
              coef3 = b2param, ACM = AC)

## WORKING WITH lavaan MODELS. From the mediation tutorial:
## http://lavaan.ugent.be/tutorial/mediation.html

set.seed(1234)
X <- rnorm(100)
M <- 0.5*X + rnorm(100)
Y <- 0.7*M + rnorm(100)
Data <- data.frame(X = X, Y = Y, M = M)
model <- ' # direct effect
Y ~ c*X
# mediator
M ~ a*X
Y ~ b*M
# indirect effect (a*b)
ab := a*b
# total effect
total := c + (a*b)
'
fit <- sem(model, data = Data)

med <- 'a*b'
## Automatically extract information from lavaan object
monteCarloMed(med, object = fit)

## or (unnecessary) manually extract the information first
myParams <- c("a", "b")
myCoefs <- coef(fit)[myParams]
myACM <- vcov(fit)[myParams, myParams]
monteCarloMed(med, myCoefs, ACM = myACM)

```

---

moreFitIndices

*Calculate more fit indices*


---

## Description

Calculate more fit indices that are not already provided in lavaan.

## Usage

```
moreFitIndices(object, fit.measures = "all", nPrior = 1)
```



**Arguments**

object	The lavaan model object provided after running the cfa, sem, growth, or lavaan functions.
fit.measures	Additional fit measures to be calculated. All additional fit measures are calculated by default
nPrior	The sample size on which prior is based. This argument is used to compute BIC*.

**Details**

Gamma Hat (gammaHat; West, Taylor, & Wu, 2012) is a global fit index which can be computed (assuming equal number of indicators across groups) by

$$gammaHat = \frac{p}{p + 2 \times \frac{\chi_k^2 - df_k}{N}},$$

where  $p$  is the number of variables in the model,  $\chi_k^2$  is the  $\chi^2$  test statistic value of the target model,  $df_k$  is the degree of freedom when fitting the target model, and  $N$  is the sample size (or sample size minus the number of groups if mimic is set to "EQS").

Adjusted Gamma Hat (adjGammaHat; West, Taylor, & Wu, 2012) is a global fit index which can be computed by

$$adjGammaHat = \left(1 - \frac{K \times p \times (p + 1)}{2 \times df_k}\right) \times (1 - gammaHat),$$

where  $K$  is the number of groups (please refer to Dudgeon, 2004 for the multiple-group adjustment for agfi\*).

Corrected Akaike Information Criterion (aic.smallN; Burnham & Anderson, 2003) is a corrected version of AIC for small sample size, often abbreviated AICc:

$$aic.smallN = AIC + \frac{2k(k + 1)}{N - k - 1},$$

where  $AIC$  is the original AIC:  $-2 \times LL + 2k$  (where  $k$  = the number of estimated parameters in the target model). Note that AICc is a small-sample correction derived for univariate regression models, so it is probably *not* appropriate for comparing SEMs.

Corrected Bayesian Information Criterion (bic.priorN; Kuha, 2004) is similar to BIC but explicitly specifying the sample size on which the prior is based ( $N_{prior}$ ).

$$bic.priorN = f + k \log(1 + N/N_{prior}),$$

Stochastic information criterion (SIC; Preacher, 2006) is similar to AIC or BIC. This index will account for model complexity in the model's function form, in addition to the number of free parameters. This index will be provided only when the  $\chi^2$  value is not scaled. The SIC can be computed by

$$sic = \frac{1}{2} \left( f - \log \det I(\hat{\theta}) \right),$$

where  $I(\hat{\theta})$  is the information matrix of the parameters.

Hannan-Quinn Information Criterion (hqc; Hannan & Quinn, 1979) is used for model selection similar to AIC or BIC.

$$hqc = f + 2k \log(\log N),$$

Note that if Satorra–Bentler or Yuan–Bentler’s method is used, the fit indices using the scaled  $\chi^2$  values are also provided.

See [nullRMSEA](#) for the further details of the computation of RMSEA of the null model.

### Value

1. gammaHat: Gamma Hat
2. adjGammaHat: Adjusted Gamma Hat
3. baseline.rmsea: RMSEA of the Baseline (Null) Model
4. aic.smallN: Corrected (for small sample size) Akaike Information Criterion
5. bic.priorN: Bayesian Information Criterion with specified prior sample size
6. sic: Stochastic Information Criterion
7. hqc: Hannan-Quinn Information Criterion
8. gammaHat.scaled: Gamma Hat using scaled  $\chi^2$
9. adjGammaHat.scaled: Adjusted Gamma Hat using scaled  $\chi^2$
10. baseline.rmsea.scaled: RMSEA of the Baseline (Null) Model using scaled  $\chi^2$

### Author(s)

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Aaron Boulton (University of North Carolina, Chapel Hill; <aboulton@email.unc.edu>)

Ruben Arslan (Humboldt-University of Berlin, <rubenarslan@gmail.com>)

Yves Rosseel (Ghent University; <Yves.Rosseel@UGent.be>)

### References

- Burnham, K., & Anderson, D. (2003). *Model selection and multimodel inference: A practical–theoretic approach*. New York, NY: Springer–Verlag.
- Dudgeon, P. (2004). A note on extending Steiger’s (1998) multiple sample RMSEA adjustment to other noncentrality parameter-based statistic. *Structural Equation Modeling*, *11*(3), 305–319. doi:10.1207/s15328007sem1103\_1

Kuha, J. (2004). AIC and BIC: Comparisons of assumptions and performance. *Sociological Methods Research*, 33(2), 188–229. doi:10.1177/0049124103262065

Preacher, K. J. (2006). Quantifying parsimony in structural equation modeling. *Multivariate Behavioral Research*, 43(3), 227–259. doi:10.1207/s15327906mbr4103\_1

West, S. G., Taylor, A. B., & Wu, W. (2012). Model fit and model selection in structural equation modeling. In R. H. Hoyle (Ed.), *Handbook of Structural Equation Modeling* (pp. 209–231). New York, NY: Guilford.

### See Also

- [miPowerFit](#) For the modification indices and their power approach for model fit evaluation
- [nullRMSEA](#) For RMSEA of the null model

### Examples

```
HS.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed  =~ x7 + x8 + x9 '
```

```
fit <- cfa(HS.model, data = HolzingerSwineford1939)
moreFitIndices(fit)
```

```
fit2 <- cfa(HS.model, data = HolzingerSwineford1939, estimator = "mlr")
moreFitIndices(fit2)
```

---

mvrnonnorm

*Generate Non-normal Data using Vale and Maurelli (1983) method*


---

### Description

Generate Non-normal Data using Vale and Maurelli (1983) method. The function is designed to be as similar as the popular `mvrnorm` function in the MASS package. The codes are copied from `mvrnorm` function in the MASS package for argument checking and `lavaan` package for data generation using Vale and Maurelli (1983) method.

### Usage

```
mvrnonnorm(n, mu, Sigma, skewness = NULL, kurtosis = NULL, empirical = FALSE)
```

### Arguments

n	Sample size
mu	A mean vector. If elements are named, those will be used as variable names in the returned data matrix.

Sigma	A positive-definite symmetric matrix specifying the covariance matrix of the variables. If rows or columns are named (and mu is unnamed), those will be used as variable names in the returned data matrix.
skewness	A vector of skewness of the variables
kurtosis	A vector of excessive kurtosis of the variables
empirical	If TRUE, mu and Sigma specify the empirical rather than population mean and covariance matrix

**Value**

A data matrix

**Author(s)**

The original function is the [simulateData](#) function written by Yves Rosseel in the lavaan package. The function is adjusted for a convenient usage by Sunthud Pornprasertmanit (<psunthud@gmail.com>). Terrence D. Jorgensen added the feature to retain variable names from mu or Sigma.

**References**

Vale, C. D. & Maurelli, V. A. (1983). Simulating multivariate nonnormal distributions. *Psychometrika*, 48(3), 465–471. doi:10.1007/BF02293687

**Examples**

```
set.seed(123)
mvrnonnorm(20, c(1, 2), matrix(c(10, 2, 2, 5), 2, 2),
skewness = c(5, 2), kurtosis = c(3, 3))
## again, with variable names specified in mu
set.seed(123)
mvrnonnorm(20, c(a = 1, b = 2), matrix(c(10, 2, 2, 5), 2, 2),
skewness = c(5, 2), kurtosis = c(3, 3))
```

---

net

*Nesting and Equivalence Testing*

---

**Description**

This test examines whether pairs of SEMs are nested or equivalent.

**Usage**

```
net(..., crit = 1e-04)
```

## Arguments

...	The lavaan objects used for test of nesting and equivalence
crit	The upper-bound criterion for testing the equivalence of models. Models are considered nested (or equivalent) if the difference between their $\chi^2$ fit statistics is less than this criterion.

## Details

The concept of nesting/equivalence should be the same regardless of estimation method. However, the particular method of testing nesting/equivalence (as described in Bentler & Satorra, 2010) employed by the `net` function analyzes summary statistics (model-implied means and covariance matrices, not raw data). In the case of robust methods like MLR, the raw data is only utilized for the robust adjustment to SE and chi-sq, and the `net` function only checks the unadjusted chi-sq for the purposes of testing nesting/equivalence. This method also applies to models for categorical data, following the procedure described by Asparouhov & Muthen (2019).

## Value

The `Net` object representing the outputs for nesting and equivalent testing, including a logical matrix of test results and a vector of degrees of freedom for each model.

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## References

Bentler, P. M., & Satorra, A. (2010). Testing model nesting and equivalence. *Psychological Methods*, 15(2), 111–123. doi:10.1037/a0019625

Asparouhov, T., & Muthen, B. (2019). Nesting and equivalence testing for structural equation models. *Structural Equation Modeling*, 26(2), 302–309. doi:10.1080/10705511.2018.1513795

## Examples

```
## Not run:
m1 <- ' visual =~ x1 + x2 + x3
      textual =~ x4 + x5 + x6
      speed  =~ x7 + x8 + x9 '

m2 <- ' f1 =~ x1 + x2 + x3 + x4
      f2 =~ x5 + x6 + x7 + x8 + x9 '

m3 <- ' visual =~ x1 + x2 + x3
      textual =~ eq*x4 + eq*x5 + eq*x6
      speed  =~ x7 + x8 + x9 '

fit1 <- cfa(m1, data = HolzingerSwineford1939)
fit1a <- cfa(m1, data = HolzingerSwineford1939, std.lv = TRUE) # Equivalent to fit1
```

```

fit2 <- cfa(m2, data = HolzingerSwineford1939) # Not equivalent to or nested in fit1
fit3 <- cfa(m3, data = HolzingerSwineford1939) # Nested in fit1 and fit1a

tests <- net(fit1, fit1a, fit2, fit3)
tests
summary(tests)

## End(Not run)

```

---

Net-class

*Class For the Result of Nesting and Equivalence Testing*

---

### Description

This class contains the results of nesting and equivalence testing among multiple models

### Usage

```

## S4 method for signature 'Net'
show(object)

## S4 method for signature 'Net'
summary(object)

```

### Arguments

object            An object of class Net.

### Value

show              signature(object = "Net"): prints the logical matrix of test results. NA indicates a model did not converge.

summary           signature(object = "Net"): prints a narrative description of results. The original object is invisibly returned.

### Slots

test   Logical matrix indicating nesting/equivalence among models

df    The degrees of freedom of tested models

### Objects from the Class

Objects can be created via the `net` function.

### Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**See Also**[net](#)**Examples**

```
# See the example in the net function.
```

---

nullRMSEA	<i>Calculate the RMSEA of the null model</i>
-----------	--

---

**Description**

Calculate the RMSEA of the null (baseline) model

**Usage**

```
nullRMSEA(object, scaled = FALSE, silent = FALSE)
```

**Arguments**

object	The lavaan model object provided after running the <code>cfa</code> , <code>sem</code> , <code>growth</code> , or <code>lavaan</code> functions.
scaled	If TRUE, the scaled (or robust, if available) RMSEA is returned. Ignored if a robust test statistic was not requested.
silent	If TRUE, do not print anything on the screen.

**Details**

RMSEA of the null model is calculated similar to the formula provided in the lavaan package. The standard formula of RMSEA is

$$RMSEA = \sqrt{\frac{\chi^2}{N \times df} - \frac{1}{N}} \times \sqrt{G}$$

where  $\chi^2$  is the chi-square test statistic value of the target model,  $N$  is the total sample size,  $df$  is the degree of freedom of the hypothesized model,  $G$  is the number of groups. Kenny proposed in his website that

"A reasonable rule of thumb is to examine the RMSEA for the null model and make sure that is no smaller than 0.158. An RMSEA for the model of 0.05 and a TLI of .90, implies that the RMSEA of the null model is 0.158. If the RMSEA for the null model is less than 0.158, an incremental measure of fit may not be that informative."

See also <http://davidakenny.net/cm/fit.htm>

**Value**

A value of RMSEA of the null model (a numeric vector) returned invisibly.

**Author(s)**

Ruben Arslan (Humboldt-University of Berlin, <rubenarslan@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

Kenny, D. A., Kaniskan, B., & McCoach, D. B. (2015). The performance of RMSEA in models with small degrees of freedom. *Sociological Methods Research*, 44(3), 486–507. doi:10.1177/0049124114543236

**See Also**

- [miPowerFit](#) For the modification indices and their power approach for model fit evaluation
- [moreFitIndices](#) For other fit indices

**Examples**

```
HS.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed  =~ x7 + x8 + x9 '
```

```
fit <- cfa(HS.model, data = HolzingerSwineford1939)
nullRMSEA(fit)
```

---

orthRotate

*Implement orthogonal or oblique rotation*

---

**Description**

These functions will implement orthogonal or oblique rotation on standardized factor loadings from a lavaan output.

**Usage**

```
orthRotate(object, method = "varimax", ...)
```

```
oblqRotate(object, method = "quartimin", ...)
```

```
funRotate(object, fun, ...)
```



**Arguments**

object	A lavaan output
method	The method of rotations, such as "varimax", "quartimax", "geomin", "oblimin", or any gradient projection algorithms listed in the <a href="#">GPA</a> function in the <code>GPARotation</code> package.
...	Additional arguments for the <a href="#">GPForth</a> function (for <code>orthRotate</code> ), the <a href="#">GPFoblq</a> function (for <code>oblqRotate</code> ), or the function that users provide in the <code>fun</code> argument.
fun	The name of the function that users wish to rotate the standardized solution. The functions must take the first argument as the standardized loading matrix and return the <code>GPARotation</code> object. Check this page for available functions: <a href="#">rotations</a> .

**Details**

These functions will rotate the unrotated standardized factor loadings by orthogonal rotation using the [GPForth](#) function or oblique rotation using the [GPFoblq](#) function the `GPARotation` package. The resulting rotation matrix will be used to calculate standard errors of the rotated standardized factor loading by delta method by numerically computing the Jacobian matrix by the [lav\\_func\\_jacobian\\_simple](#) function.

**Value**

An `linkS4class{EFA}` object that saves the rotated EFA solution

**Author(s)**

Sunthud Pornprasertmanit (<[psunthud@gmail.com](mailto:psunthud@gmail.com)>)

**Examples**

```
## Not run:

unrotated <- efaUnrotate(HolzingerSwineford1939, nf = 3,
                        varList = paste0("x", 1:9), estimator = "mlr")

# Orthogonal varimax
out.varimax <- orthRotate(unrotated, method = "varimax")
summary(out.varimax, sort = FALSE, suppress = 0.3)

# Orthogonal Quartimin
orthRotate(unrotated, method = "quartimin")

# Oblique Quartimin
oblqRotate(unrotated, method = "quartimin")

# Geomin
oblqRotate(unrotated, method = "geomin")
```

```

# Target rotation
library(GPARotation)
target <- matrix(0, 9, 3)
target[1:3, 1] <- NA
target[4:6, 2] <- NA
target[7:9, 3] <- NA
colnames(target) <- c("factor1", "factor2", "factor3")
## This function works with GPARotation version 2012.3-1
funRotate(unrotated, fun = "targetQ", Target = target)

## End(Not run)

```

---

parcelAllocation

*Random Allocation of Items to Parcels in a Structural Equation Model*


---

## Description

This function generates a given number of randomly generated item-to-parcel allocations, fits a model to each allocation, and provides averaged results over all allocations.

## Usage

```

parcelAllocation(model, data, parcel.names, item.syntax, nAlloc = 100,
  fun = "sem", alpha = 0.05, fit.measures = c("chisq", "df", "cfi",
  "tli", "rmsea", "srmr"), ..., show.progress = FALSE, iseed = 12345,
  do.fit = TRUE, return.fit = FALSE, warn = FALSE)

```

## Arguments

model	<a href="#">lavaan</a> model syntax specifying the model fit to (at least some) parceled data. Note that there can be a mixture of items and parcels (even within the same factor), in case certain items should never be parceled. Can be a character string or parameter table. Also see <a href="#">lavaanify</a> for more details.
data	A data.frame containing all observed variables appearing in the model, as well as those in the <code>item.syntax</code> used to create parcels. If the data have missing values, multiple imputation before parcelling is recommended: submit a stacked data set (with a variable for the imputation number, so they can be separated later) and set <code>do.fit = FALSE</code> to return the list of data.frames (one per allocation), each of which is a stacked, imputed data set with parcels.
parcel.names	character vector containing names of all parcels appearing as indicators in model.
item.syntax	<a href="#">lavaan</a> model syntax specifying the model that would be fit to all of the un-parceled items, including items that should be randomly allocated to parcels appearing in model.
nAlloc	The number of random items-to-parcels allocations to generate.

fun	character string indicating the name of the <code>lavaan</code> function used to fit model to data. Can only take the values "lavaan", "sem", "cfa", or "growth".
alpha	Alpha level used as criterion for significance.
fit.measures	character vector containing names of fit measures to request from each fitted <code>lavaan</code> model. See the output of <code>fitMeasures</code> for a list of available measures.
...	Additional arguments to be passed to <code>lavaanList</code> . See also <code>lavOptions</code>
show.progress	If TRUE, show a <code>txtProgressBar</code> indicating how fast the model-fitting iterates over allocations.
iseed	(Optional) Random seed used for parceling items. When the same random seed is specified and the program is re-run, the same allocations will be generated. Using the same <code>iseed</code> argument will ensure the any model is fit to the same parcel allocations. <i>Note:</i> When using <b>parallel</b> options, you must first type <code>RNGkind("L'Ecuyer-CMRG")</code> into the R Console, so that the seed will be controlled across cores.
do.fit	If TRUE (default), the model is fitted to each parceled data set, and the summary of results is returned (see the Value section below). If FALSE, the items are randomly parceled, but the model is not fit; instead, the <code>list</code> of <code>data.frames</code> is returned (so assign it to an object).
return.fit	If TRUE, a <code>lavaanList</code> object is returned with the list of results across allocations
warn	Whether to print warnings when fitting model to each allocation

## Details

This function implements the random item-to-parcel allocation procedure described in Sterba (2011) and Sterba and MacCallum (2010). The function takes a single data set with item-level data, randomly assigns items to parcels, fits a structural equation model to the parceled data (using `lavaanList`), and repeats this process for a user-specified number of random allocations. Results from all fitted models are summarized in the output. For further details on the benefits of randomly allocating items to parcels, see Sterba (2011) and Sterba and MccCallum (2010).

## Value

Estimates	A <code>data.frame</code> containing results related to parameter estimates with columns corresponding to their names; average and standard deviation across allocations; minimum, maximum, and range across allocations; and the proportion of allocations in which each parameter estimate was significant.
SE	A <code>data.frame</code> containing results similar to <code>Estimates</code> , but related to the standard errors of parameter estimates.
Fit	A <code>data.frame</code> containing results related to model fit, with columns corresponding to fit index names; their average and standard deviation across allocations; the minimum, maximum, and range across allocations; and (if the test statistic or RMSEA is included in <code>fit.measures</code> ) the proportion of allocations in which each test of (exact or close) fit was significant.
Model	A <code>lavaanList</code> object containing results of the model fitted to each parcel allocation. Only returned if <code>return.fit = TRUE</code> .

**Author(s)**

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

Sterba, S. K. (2011). Implications of parcel-allocation variability for comparing fit of item-solutions and parcel-solutions. *Structural Equation Modeling*, *18*(4), 554–577. doi:10.1080/10705511.2011.607073

Sterba, S. K. & MacCallum, R. C. (2010). Variability in parameter estimates and model fit across random allocations of items to parcels. *Multivariate Behavioral Research*, *45*(2), 322–358. doi:10.1080/00273171003680302

Sterba, S. K., & Rights, J. D. (2016). Accounting for parcel-allocation variability in practice: Combining sources of uncertainty and choosing the number of allocations. *Multivariate Behavioral Research*, *51*(2–3), 296–313. doi:10.1080/00273171.2016.1144502

Sterba, S. K., & Rights, J. D. (2017). Effects of parceling on model selection: Parcel-allocation variability in model ranking. *Psychological Methods*, *22*(1), 47–68. doi:10.1037/met0000067

**See Also**

[PAVranking](#) for comparing 2 models, [poolMAlloc](#) for choosing the number of allocations

**Examples**

```
## Fit 2-factor CFA to simulated data. Each factor has 9 indicators.

## Specify the item-level model (if NO parcels were created)
item.syntax <- c(paste0("f1 =~ f1item", 1:9),
                paste0("f2 =~ f2item", 1:9))
cat(item.syntax, sep = "\n")
## Below, we reduce the size of this same model by
## applying different parceling schemes

## 3-indicator parcels
mod.parcels <- '
f1 =~ par1 + par2 + par3
f2 =~ par4 + par5 + par6
'

## names of parcels
(parcel.names <- paste0("par", 1:6))

## Not run:
## override default random-number generator to use parallel options
RNGkind("L'Ecuyer-CMRG")

parcelAllocation(mod.parcels, data = simParcel, nAlloc = 100,
                parcel.names = parcel.names, item.syntax = item.syntax,
                std.lv = TRUE, # any addition lavaan arguments
                parallel = "snow") # parallel options
```

```

## POOL RESULTS by treating parcel allocations as multiple imputations
## Details provided in Sterba & Rights (2016); see ?poolMAlloc.

## save list of data sets instead of fitting model yet
dataList <- parcelAllocation(mod.parcels, data = simParcel, nAlloc = 100,
                             parcel.names = parcel.names,
                             item.syntax = item.syntax,
                             do.fit = FALSE)
## now fit the model to each data set
fit.parcels <- cfa.mi(mod.parcels, data = dataList, std.lv = TRUE)
summary(fit.parcels) # uses Rubin's rules
anova(fit.parcels)  # pooled test statistic
class?lavaan.mi     # find more methods for pooling results

## End(Not run)

## multigroup example
simParcel$group <- 0:1 # arbitrary groups for example
mod.mg <- '
f1 =~ par1 + c(L2, L2)*par2 + par3
f2 =~ par4 + par5 + par6
'

## names of parcels
(parcel.names <- paste0("par", 1:6))

parcelAllocation(mod.mg, data = simParcel, parcel.names, item.syntax,
                 std.lv = TRUE, group = "group", group.equal = "loadings",
                 nAlloc = 20, show.progress = TRUE)

## parcels for first factor, items for second factor
mod.items <- '
f1 =~ par1 + par2 + par3
f2 =~ f2item2 + f2item7 + f2item8
'

## names of parcels
(parcel.names <- paste0("par", 1:3))

parcelAllocation(mod.items, data = simParcel, parcel.names, item.syntax,
                 nAlloc = 20, std.lv = TRUE)

## mixture of 1- and 3-indicator parcels for second factor
mod.mix <- '
f1 =~ par1 + par2 + par3
f2 =~ f2item2 + f2item7 + f2item8 + par4 + par5 + par6
'

## names of parcels
(parcel.names <- paste0("par", 1:6))

```

```
parcelAllocation(mod.mix, data = simParcel, parcel.names, item.syntax,
  nAlloc = 20, std.lv = TRUE)
```

---

partialInvariance      *Partial Measurement Invariance Testing Across Groups*

---

### Description

This test will provide partial invariance testing by (a) freeing a parameter one-by-one from nested model and compare with the original nested model or (b) fixing (or constraining) a parameter one-by-one from the parent model and compare with the original parent model. This function only works with congeneric models. The `partialInvariance` is used for continuous variable. The `partialInvarianceCat` is used for categorical variables.

### Usage

```
partialInvariance(fit, type, free = NULL, fix = NULL, refgroup = 1,
  poolvar = TRUE, p.adjust = "none", fbound = 2, return.fit = FALSE,
  method = "satorra.bentler.2001")
```

```
partialInvarianceCat(fit, type, free = NULL, fix = NULL, refgroup = 1,
  poolvar = TRUE, p.adjust = "none", return.fit = FALSE,
  method = "satorra.bentler.2001")
```

### Arguments

<code>fit</code>	A list of models for invariance testing. Each model should be assigned by appropriate names (see details). The result from <code>measurementInvariance</code> or <code>measurementInvarianceCat</code> could be used in this argument directly.
<code>type</code>	The types of invariance testing: "metric", "scalar", "strict", or "means"
<code>free</code>	A vector of variable names that are free across groups in advance. If partial mean invariance is tested, this argument represents a vector of factor names that are free across groups.
<code>fix</code>	A vector of variable names that are constrained to be equal across groups in advance. If partial mean invariance is tested, this argument represents a vector of factor names that are fixed across groups.
<code>refgroup</code>	The reference group used to make the effect size comparison with the other groups.
<code>poolvar</code>	If TRUE, the variances are pooled across group for standardization. Otherwise, the variances of the reference group are used for standardization.
<code>p.adjust</code>	The method used to adjust p values. See <code>p.adjust</code> for the options for adjusting p values. The default is to not use any corrections.
<code>fbound</code>	The z-scores of factor that is used to calculate the effect size of the loading difference proposed by Millsap and Olivera-Aguilar (2012).

return.fit	Return the submodels fitted by this function
method	The method used to calculate likelihood ratio test. See <a href="#">lavTestLRT</a> for available options

## Details

There are four types of partial invariance testing:

- Partial weak invariance. The model named 'fit.configural' from the list of models is compared with the model named 'fit.loadings'. Each loading will be freed or fixed from the metric and configural invariance models respectively. The modified models are compared with the original model. Note that the objects in the list of models must have the names of "fit.configural" and "fit.loadings". Users may use "metric", "weak", "loading", or "loadings" in the type argument. Note that, for testing invariance on marker variables, other variables will be assigned as marker variables automatically.
- Partial strong invariance. The model named 'fit.loadings' from the list of models is compared with the model named either 'fit.intercepts' or 'fit.thresholds'. Each intercept will be freed or fixed from the scalar and metric invariance models respectively. The modified models are compared with the original model. Note that the objects in the list of models must have the names of "fit.loadings" and either "fit.intercepts" or "fit.thresholds". Users may use "scalar", "strong", "intercept", "intercepts", "threshold", or "thresholds" in the type argument. Note that, for testing invariance on marker variables, other variables will be assigned as marker variables automatically. Note that if all variables are dichotomous, scalar invariance testing is not available.
- Partial strict invariance. The model named either 'fit.intercepts' or 'fit.thresholds' (or 'fit.loadings') from the list of models is compared with the model named 'fit.residuals'. Each residual variance will be freed or fixed from the strict and scalar (or metric) invariance models respectively. The modified models are compared with the original model. Note that the objects in the list of models must have the names of "fit.residuals" and either "fit.intercepts", "fit.thresholds", or "fit.loadings". Users may use "strict", "residual", "residuals", "error", or "errors" in the type argument.
- Partial mean invariance. The model named either 'fit.intercepts' or 'fit.thresholds' (or 'fit.residuals' or 'fit.loadings') from the list of models is compared with the model named 'fit.means'. Each factor mean will be freed or fixed from the means and scalar (or strict or metric) invariance models respectively. The modified models are compared with the original model. Note that the objects in the list of models must have the names of "fit.means" and either "fit.residuals", "fit.intercepts", "fit.thresholds", or "fit.loadings". Users may use "means" or "mean" in the type argument.

Two types of comparisons are used in this function:

1. free: The nested model is used as a template. Then, one parameter indicating the differences between two models is free. The new model is compared with the nested model. This process is repeated for all differences between two models. The likelihood-ratio test and the difference in CFI are provided.
2. fix: The parent model is used as a template. Then, one parameter indicating the differences between two models is fixed or constrained to be equal to other parameters. The new model is then compared with the parent model. This process is repeated for all differences between two models. The likelihood-ratio test and the difference in CFI are provided.

3. `wald`: This method is similar to the `fix` method. However, instead of building a new model and compare them with likelihood-ratio test, multivariate wald test is used to compare equality between parameter estimates. See `lavTestWald` for further details. Note that if any rows of the contrast cannot be summed to 0, the Wald test is not provided, such as comparing two means where one of the means is fixed as 0. This test statistic is not as accurate as likelihood-ratio test provided in `fix`. I provide it here in case that likelihood-ratio test fails to converge.

Note that this function does not adjust for the inflated Type I error rate from multiple tests. The degree of freedom of all tests would be the number of groups minus 1.

The details of standardized estimates and the effect size used for each parameters are provided in the vignettes by running `vignette("partialInvariance")`.

## Value

A list of results are provided. The list will consists of at least two elements:

1. `estimates`: The results of parameter estimates including pooled estimates (`poollest`), the estimates for each group, standardized estimates for each group (`std`), the difference in standardized values, and the effect size statistic ( $g$  for factor loading difference and  $h$  for error variance difference). See the details of this effect size statistic by running `vignette("partialInvariance")`. In the `partialInvariance` function, the additional effect statistics proposed by Millsap and Olivera-Aguilar (2012) are provided. For factor loading, the additional outputs are the observed mean difference (`diff_mean`), the mean difference if factor scores are low (`low_fscore`), and the mean difference if factor scores are high (`high_fscore`). The low factor score is calculated by (a) finding the factor scores that its  $z$  score equals `-bound` (the default is `-2`) from all groups and (b) picking the minimum value among the factor scores. The high factor score is calculated by (a) finding the factor scores that its  $z$  score equals `bound` (default = `2`) from all groups and (b) picking the maximum value among the factor scores. For measurement intercepts, the additional outputs are the observed means difference (`diff_mean`) and the proportion of the differences in the intercepts over the observed means differences (`propdiff`). For error variances, the additional outputs are the proportion of the difference in error variances over the difference in observed variances (`propdiff`).
2. `results`: Statistical tests as well as the change in CFI are provided.  $\chi^2$  and  $p$  value are provided for all methods.
3. `models`: The submodels used in the `free` and `fix` methods, as well as the nested and parent models. The nested and parent models will be changed from the original models if `free` or `fit` arguments are specified.

## Author(s)

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## References

Millsap, R. E., & Olivera-Aguilar, M. (2012). Investigating measurement invariance using confirmatory factor analysis. In R. H. Hoyle (Ed.), *Handbook of structural equation modeling* (pp. 380–392). New York, NY: Guilford.



**See Also**

[measurementInvariance](#) for measurement invariance for continuous variables; [measurementInvarianceCat](#) for measurement invariance for categorical variables; [lavTestWald](#) for multivariate Wald test

**Examples**

```
## Conduct weak invariance testing manually by using fixed-factor
## method of scale identification

library(lavaan)

conf <- "
f1 =~ NA*x1 + x2 + x3
f2 =~ NA*x4 + x5 + x6
f1 ~~ c(1, 1)*f1
f2 ~~ c(1, 1)*f2
"

weak <- "
f1 =~ NA*x1 + x2 + x3
f2 =~ NA*x4 + x5 + x6
f1 ~~ c(1, NA)*f1
f2 ~~ c(1, NA)*f2
"

configural <- cfa(conf, data = HolzingerSwineford1939, std.lv = TRUE, group="school")
weak <- cfa(weak, data = HolzingerSwineford1939, group="school", group.equal="loadings")
models <- list(fit.configural = configural, fit.loadings = weak)
partialInvariance(models, "metric")

## Not run:
partialInvariance(models, "metric", free = "x5") # "x5" is free across groups in advance
partialInvariance(models, "metric", fix = "x4") # "x4" is fixed across groups in advance

## Use the result from the measurementInvariance function
HW.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed =~ x7 + x8 + x9 '

models2 <- measurementInvariance(model = HW.model, data=HolzingerSwineford1939,
                                group="school")
partialInvariance(models2, "scalar")

## Conduct weak invariance testing manually by using fixed-factor
## method of scale identification for dichotomous variables

f <- rnorm(1000, 0, 1)
u1 <- 0.9*f + rnorm(1000, 1, sqrt(0.19))
u2 <- 0.8*f + rnorm(1000, 1, sqrt(0.36))
u3 <- 0.6*f + rnorm(1000, 1, sqrt(0.64))
u4 <- 0.7*f + rnorm(1000, 1, sqrt(0.51))
```

```

u1 <- as.numeric(cut(u1, breaks = c(-Inf, 0, Inf)))
u2 <- as.numeric(cut(u2, breaks = c(-Inf, 0.5, Inf)))
u3 <- as.numeric(cut(u3, breaks = c(-Inf, 0, Inf)))
u4 <- as.numeric(cut(u4, breaks = c(-Inf, -0.5, Inf)))
g <- rep(c(1, 2), 500)
dat2 <- data.frame(u1, u2, u3, u4, g)

configural2 <- "
f1 =~ NA*u1 + u2 + u3 + u4
u1 | c(t11, t11)*t1
u2 | c(t21, t21)*t1
u3 | c(t31, t31)*t1
u4 | c(t41, t41)*t1
f1 ~~ c(1, 1)*f1
f1 ~ c(0, NA)*1
u1 ~~ c(1, 1)*u1
u2 ~~ c(1, NA)*u2
u3 ~~ c(1, NA)*u3
u4 ~~ c(1, NA)*u4
"

outConfigural2 <- cfa(configural2, data = dat2, group = "g",
  parameterization = "theta", estimator = "wlsmv",
  ordered = c("u1", "u2", "u3", "u4"))

weak2 <- "
f1 =~ NA*u1 + c(f11, f11)*u1 + c(f21, f21)*u2 + c(f31, f31)*u3 + c(f41, f41)*u4
u1 | c(t11, t11)*t1
u2 | c(t21, t21)*t1
u3 | c(t31, t31)*t1
u4 | c(t41, t41)*t1
f1 ~~ c(1, NA)*f1
f1 ~ c(0, NA)*1
u1 ~~ c(1, 1)*u1
u2 ~~ c(1, NA)*u2
u3 ~~ c(1, NA)*u3
u4 ~~ c(1, NA)*u4
"

outWeak2 <- cfa(weak2, data = dat2, group = "g", parameterization = "theta",
  estimator = "wlsmv", ordered = c("u1", "u2", "u3", "u4"))
modelsCat <- list(fit.configural = outConfigural2, fit.loadings = outWeak2)

partialInvarianceCat(modelsCat, type = "metric")

partialInvarianceCat(modelsCat, type = "metric", free = "u2")
partialInvarianceCat(modelsCat, type = "metric", fix = "u3")

## Use the result from the measurementInvarianceCat function

model <- ' f1 =~ u1 + u2 + u3 + u4
          f2 =~ u5 + u6 + u7 + u8'
```

```

modelsCat2 <- measurementInvarianceCat(model = model, data = datCat, group = "g",
                                       parameterization = "theta",
                                       estimator = "wlsmv", strict = TRUE)

partialInvarianceCat(modelsCat2, type = "scalar")

## End(Not run)

```

### Description

This function quantifies and assesses the consequences of parcel-allocation variability for model ranking of structural equation models (SEMs) that differ in their structural specification but share the same parcel-level measurement specification (see Sterba & Rights, 2016). This function calls [parcelAllocation](#)—which can be used with only one SEM in isolation—to fit two (assumed) nested models to each of a specified number of random item-to-parcel allocations. Output includes summary information about the distribution of model selection results (including plots) and the distribution of results for each model individually, across allocations within-sample. Note that this function can be used when selecting among more than two competing structural models as well (see instructions below involving the `seed` argument).

### Usage

```

PAVranking(model0, model1, data, parcel.names, item.syntax, nAlloc = 100,
           fun = "sem", alpha = 0.05, bic.crit = 10, fit.measures = c("chisq",
                           "df", "cfi", "tli", "rmsea", "srmr", "logl", "aic", "bic", "bic2"), ...,
           show.progress = FALSE, iseed = 12345, warn = FALSE)

```

### Arguments

- |                             |   |
|-----------------------------|---|
| <code>model0, model1</code> | <code>lavaan</code> model syntax specifying nested models ( <code>model0</code> within <code>model1</code> ) to be fitted to the same parceled data. Note that there can be a mixture of items and parcels (even within the same factor), in case certain items should never be parceled. Can be a character string or parameter table. Also see <a href="#">lavaanify</a> for more details.  |
| <code>data</code>           | A <code>data.frame</code> containing all observed variables appearing in the model, as well as those in the <code>item.syntax</code> used to create parcels. If the data have missing values, multiple imputation before parcelling is recommended: submit a stacked data set (with a variable for the imputation number, so they can be separated later) and set <code>do.fit = FALSE</code> to return the list of <code>data.frames</code> (one per allocation), each of which is a stacked, imputed data set with parcels. |
| <code>parcel.names</code>   | character vector containing names of all parcels appearing as indicators in model.  |

<code>item.syntax</code>	<code>lavaan</code> model syntax specifying the model that would be fit to all of the un-parceled items, including items that should be randomly allocated to parcels appearing in <code>model</code> .
<code>nAlloc</code>	The number of random items-to-parcels allocations to generate.
<code>fun</code>	character string indicating the name of the <code>lavaan</code> function used to fit <code>model</code> to data. Can only take the values "lavaan", "sem", "cfa", or "growth".
<code>alpha</code>	Alpha level used as criterion for significance.
<code>bic.crit</code>	Criterion for assessing evidence in favor of one model over another. See Rafferty (1995) for guidelines (default is "very strong evidence" in favor of the model with lower BIC).
<code>fit.measures</code>	character vector containing names of fit measures to request from each fitted <code>lavaan</code> model. See the output of <code>fitMeasures</code> for a list of available measures.
<code>...</code>	Additional arguments to be passed to <code>lavaanList</code> . See also <code>lavOptions</code>
<code>show.progress</code>	If TRUE, show a <code>txtProgressBar</code> indicating how fast each model-fitting iterates over allocations.
<code>iseed</code>	(Optional) Random seed used for parceling items. When the same random seed is specified and the program is re-run, the same allocations will be generated. The seed argument can be used to assess parcel-allocation variability in model ranking when considering more than two models. For each pair of models under comparison, the program should be rerun using the same random seed. Doing so ensures that multiple model comparisons will employ the same set of parcel datasets. <i>Note:</i> When using <b>parallel</b> options, you must first type <code>RNGkind("L'Ecuyer-CMRG")</code> into the R Console, so that the seed will be controlled across cores.
<code>warn</code>	Whether to print warnings when fitting models to each allocation

## Details

This is based on a SAS macro `ParcelAlloc` (Sterba & MacCallum, 2010). The `PAVranking` function produces results discussed in Sterba and Rights (2016) relevant to the assessment of parcel-allocation variability in model selection and model ranking. Specifically, the `PAVranking` function first calls `parcelAllocation` to generate a given number (`nAlloc`) of item-to-parcel allocations, fitting both specified models to each allocation, and providing summaries of PAV for each model. Additionally, `PAVranking` provides the following new summaries:

- PAV in model selection index values and model ranking between Models `model0` and `model1`.
- The proportion of allocations that converged and the proportion of proper solutions (results are summarized for allocations with both converged and proper allocations only).

For further details on the benefits of the random allocation of items to parcels, see Sterba (2011) and Sterba and MacCallum (2010).

To test whether nested models have equivalent fit, results can be pooled across allocations using the same methods available for pooling results across multiple imputations of missing data (see **Examples**).

*Note:* This function requires the `lavaan` package. Missing data must be coded as NA. If the function returns "Error in plot.new() : figure margins too large", the user may need to increase size of the plot window (e.g., in RStudio) and rerun the function.

**Value**

`model0.results` Results returned by `parcelAllocation` for `model0` (see the **Value** section).

`model1.results` Results returned by `parcelAllocation` for `model1` (see the **Value** section).

`model0.v.model1`

A list of model-comparison results, including the following:

- `LRT_Summary`: The average likelihood ratio test across allocations, as well as the *SD*, minimum, maximum, range, and the proportion of allocations for which the test was significant.
- `Fit_Index_Differences`: Differences in fit indices, organized by what proportion favored each model and among those, what the average difference was.
- `Favored_by_BIC`: The proportion of allocations in which each model met the criterion (`bic.crit`) for a substantial difference in fit.
- `Convergence_Summary`: The proportion of allocations in which each model (and both models) converged on a solution.

Histograms are also printed to the current plot-output device.

**Author(s)**

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**References**

Raftery, A. E. (1995). Bayesian model selection in social research. *Sociological Methodology*, 25, 111–163. doi:10.2307/271063

Sterba, S. K. (2011). Implications of parcel-allocation variability for comparing fit of item-solutions and parcel-solutions. *Structural Equation Modeling: A Multidisciplinary Journal*, 18(4), 554–577. doi:10.1080/10705511.2011.607073

Sterba, S. K., & MacCallum, R. C. (2010). Variability in parameter estimates and model fit across repeated allocations of items to parcels. *Multivariate Behavioral Research*, 45(2), 322–358. doi:10.1080/00273171003680302

Sterba, S. K., & Rights, J. D. (2016). Accounting for parcel-allocation variability in practice: Combining sources of uncertainty and choosing the number of allocations. *Multivariate Behavioral Research*, 51(2–3), 296–313. doi:10.1080/00273171.2016.1144502

Sterba, S. K., & Rights, J. D. (2017). Effects of parceling on model selection: Parcel-allocation variability in model ranking. *Psychological Methods*, 22(1), 47–68. doi:10.1037/met0000067

**See Also**

`parcelAllocation` for fitting a single model, `poolMAlloc` for choosing the number of allocations

**Examples**

```
## Specify the item-level model (if NO parcels were created)
## This must apply to BOTH competing models
```

```

item.syntax <- c(paste0("f1 =~ f1item", 1:9),
                paste0("f2 =~ f2item", 1:9))
cat(item.syntax, sep = "\n")
## Below, we reduce the size of this same model by
## applying different parceling schemes

## Specify a 2-factor CFA with correlated factors, using 3-indicator parcels
mod1 <- '
f1 =~ par1 + par2 + par3
f2 =~ par4 + par5 + par6
'

## Specify a more restricted model with orthogonal factors
mod0 <- '
f1 =~ par1 + par2 + par3
f2 =~ par4 + par5 + par6
f1 ~~ 0*f2
'

## names of parcels (must apply to BOTH models)
(parcel.names <- paste0("par", 1:6))

## Not run:
## override default random-number generator to use parallel options
RNGkind("L'Ecuyer-CMRG")

PAVranking(model0 = mod0, model1 = mod1, data = simParcel, nAlloc = 100,
           parcel.names = parcel.names, item.syntax = item.syntax,
           std.lv = TRUE,          # any addition lavaan arguments
           parallel = "snow")    # parallel options

## POOL RESULTS by treating parcel allocations as multiple imputations.
## Details provided in Sterba & Rights (2016); see ?poolMAlloc.

## save list of data sets instead of fitting model yet
datalist <- parcelAllocation(mod.parcels, data = simParcel, nAlloc = 100,
                           parcel.names = parcel.names,
                           item.syntax = item.syntax,
                           do.fit = FALSE)

## now fit each model to each data set
fit0 <- cfa.mi(mod0, data = datalist, std.lv = TRUE)
fit1 <- cfa.mi(mod1, data = datalist, std.lv = TRUE)
anova(fit0, fit1) # pooled test statistic comparing models
class?lavaan.mi  # find more methods for pooling results

## End(Not run)

```

---

permutMeasEq                      *Permutation Randomization Tests of Measurement Equivalence and Differential Item Functioning (DIF)*

---

### Description

The function `permutMeasEq` provides tests of hypotheses involving measurement equivalence, in one of two frameworks: multigroup CFA or MIMIC models.

### Usage

```
permutMeasEq(nPermute, modelType = c("mgcfa", "mimic"), con, uncon = NULL,
  null = NULL, param = NULL, freeParam = NULL, covariates = NULL,
  AFIs = NULL, moreAFIs = NULL, maxSparse = 10, maxNonconv = 10,
  showProgress = TRUE, warn = -1, datafun, extra,
  parallelType = c("none", "multicore", "snow"), ncpus = NULL, cl = NULL,
  iseed = 12345)
```

### Arguments

<code>nPermute</code>	An integer indicating the number of random permutations used to form empirical distributions under the null hypothesis.
<code>modelType</code>	A character string indicating type of model employed: multiple-group CFA ("mgcfa") or MIMIC ("mimic").
<code>con</code>	The constrained lavaan object, in which the parameters specified in <code>param</code> are constrained to equality across all groups when <code>modelType = "mgcfa"</code> , or which regression paths are fixed to zero when <code>modelType = "mimic"</code> . In the case of testing <i>configural</i> invariance when <code>modelType = "mgcfa"</code> , <code>con</code> is the configural model (implicitly, the unconstrained model is the saturated model, so use the defaults <code>uncon = NULL</code> and <code>param = NULL</code> ). When <code>modelType = "mimic"</code> , <code>con</code> is the MIMIC model in which the covariate predicts the latent construct(s) but no indicators (unless they have already been identified as DIF items).
<code>uncon</code>	Optional. The unconstrained lavaan object, in which the parameters specified in <code>param</code> are freely estimated in all groups. When <code>modelType = "mgcfa"</code> , only in the case of testing <i>configural</i> invariance should <code>uncon = NULL</code> . When <code>modelType = "mimic"</code> , any non-NULL <code>uncon</code> is silently set to <code>NULL</code> .
<code>null</code>	Optional. A lavaan object, in which an alternative null model is fit (besides the default independence model specified by lavaan) for the calculation of incremental fit indices. See Widamin & Thompson (2003) for details. If <code>NULL</code> , lavaan's default independence model is used.
<code>param</code>	An optional character vector or list of character vectors indicating which parameters the user would test for DIF following a rejection of the omnibus null hypothesis tested using (more)AFIs. Note that <code>param</code> does not guarantee certain parameters <i>are</i> constrained in <code>con</code> ; that is for the user to specify when fitting the model. If users have any "anchor items" that they would never intend to free across groups (or levels of a covariate), these should be excluded from <code>param</code> ; exceptions to a type of parameter can be specified in <code>freeParam</code> . When

	<p>modelType = "mgcfa", param indicates which parameters of interest are constrained across groups in con and are unconstrained in uncon. Parameter names must match those returned by names(coef(con)), but omitting any group-specific suffixes (e.g., "f1~1" rather than "f1~1.g2") or user-specified labels (that is, the parameter names must follow the rules of lavaan's <code>model.syntax</code>). Alternatively (or additionally), to test all constraints of a certain type (or multiple types) of parameter in con, param may take any combination of the following values: "loadings", "intercepts", "thresholds", "residuals", "residual.covariances", "means", "lv.variances", and/or "lv.covariances". When modelType = "mimic", param must be a vector of individual parameters or a list of character strings to be passed one-at-a-time to <code>lavTestScore(object = con, add = param[i])</code>, indicating which (sets of) regression paths fixed to zero in con that the user would consider freeing (i.e., exclude anchor items). If modelType = "mimic" and param is a list of character strings, the multivariate test statistic will be saved for each list element instead of 1-df modification indices for each individual parameter, and names(param) will name the rows of the MI.obs slot (see <code>permuteMeasEq</code>). Set param = NULL (default) to avoid collecting modification indices for any follow-up tests.</p>
freeParam	<p>An optional character vector, silently ignored when modelType = "mimic". If param includes a type of parameter (e.g., "loadings"), freeParam indicates exceptions (i.e., anchor items) that the user would <i>not</i> intend to free across groups and should therefore be ignored when calculating <i>p</i> values adjusted for the number of follow-up tests. Parameter types that are already unconstrained across groups in the fitted con model (i.e., a <i>partial</i> invariance model) will automatically be ignored, so they do not need to be specified in freeParam. Parameter names must match those returned by names(coef(con)), but omitting any group-specific suffixes (e.g., "f1~1" rather than "f1~1.g2") or user-specified labels (that is, the parameter names must follow the rules of lavaan <code>model.syntax</code>).</p>
covariates	<p>An optional character vector, only applicable when modelType = "mimic". The observed data are partitioned into columns indicated by covariates, and the rows are permuted simultaneously for the entire set before being merged with the remaining data. Thus, the covariance structure is preserved among the covariates, which is necessary when (e.g.) multiple dummy codes are used to represent a discrete covariate or when covariates interact. If covariates = NULL when modelType = "mimic", the value of covariates is inferred by searching param for predictors (i.e., variables appearing after the "~" operator).</p>
AFIs	<p>A character vector indicating which alternative fit indices (or chi-squared itself) are to be used to test the multiparameter omnibus null hypothesis that the constraints specified in con hold in the population. Any fit measures returned by <code>fitMeasures</code> may be specified (including constants like "df", which would be nonsensical). If both AFIs and moreAFIs are NULL, only "chisq" will be returned.</p>
moreAFIs	<p>Optional. A character vector indicating which (if any) alternative fit indices returned by <code>moreFitIndices</code> are to be used to test the multiparameter omnibus null hypothesis that the constraints specified in con hold in the population.</p>
maxSparse	<p>Only applicable when modelType = "mgcfa" and at least one indicator is ordered. An integer indicating the maximum number of consecutive times that randomly</p>



permuted group assignment can yield a sample in which at least one category (of an ordered indicator) is unobserved in at least one group, such that the same set of parameters cannot be estimated in each group. If such a sample occurs, group assignment is randomly permuted again, repeatedly until a sample is obtained with all categories observed in all groups. If `maxSparse` is exceeded, NA will be returned for that iteration of the permutation distribution.

<code>maxNonconv</code>	An integer indicating the maximum number of consecutive times that a random permutation can yield a sample for which the model does not converge on a solution. If such a sample occurs, permutation is attempted repeatedly until a sample is obtained for which the model does converge. If <code>maxNonconv</code> is exceeded, NA will be returned for that iteration of the permutation distribution, and a warning will be printed when using <code>show</code> or <code>summary</code> .
<code>showProgress</code>	Logical. Indicating whether to display a progress bar while permuting. Silently set to FALSE when using parallel options.
<code>warn</code>	Sets the handling of warning messages when fitting model(s) to permuted data sets. See <a href="#">options</a> .
<code>datafun</code>	An optional function that can be applied to the data (extracted from <code>con</code> ) after each permutation, but before fitting the model(s) to each permutation. The <code>datafun</code> function must have an argument named <code>data</code> that accepts a <code>data.frame</code> , and it must return a <code>data.frame</code> containing the same column names. The column order may differ, the values of those columns may differ (so be careful!), and any additional columns will be ignored when fitting the model, but an error will result if any column names required by the model syntax do not appear in the transformed data set. Although available for any <code>modelType</code> , <code>datafun</code> may be useful when using the MIMIC method to test for nonuniform DIF (metric/weak invariance) by using product indicators for a latent factor representing the interaction between a factor and one of the covariates, in which case the product indicators would need to be recalculated after each permutation of the covariates. To access other R objects used within <code>permuteMeasEq</code> , the arguments to <code>datafun</code> may also contain any subset of the following: <code>"con"</code> , <code>"uncon"</code> , <code>"null"</code> , <code>"param"</code> , <code>"freeParam"</code> , <code>"covariates"</code> , <code>"AFIs"</code> , <code>"moreAFIs"</code> , <code>"maxSparse"</code> , <code>"maxNonconv"</code> , and/or <code>"iseed"</code> . The values for those arguments will be the same as the values supplied to <code>permuteMeasEq</code> .
<code>extra</code>	An optional function that can be applied to any (or all) of the fitted lavaan objects ( <code>con</code> , <code>uncon</code> , and/or <code>null</code> ). This function will also be applied after fitting the model(s) to each permuted data set. To access the R objects used within <code>permuteMeasEq</code> , the arguments to <code>extra</code> must be any subset of the following: <code>"con"</code> , <code>"uncon"</code> , <code>"null"</code> , <code>"param"</code> , <code>"freeParam"</code> , <code>"covariates"</code> , <code>"AFIs"</code> , <code>"moreAFIs"</code> , <code>"maxSparse"</code> , <code>"maxNonconv"</code> , and/or <code>"iseed"</code> . The values for those arguments will be the same as the values supplied to <code>permuteMeasEq</code> . The <code>extra</code> function must return a named numeric vector or a named list of scalars (i.e., a list of numeric vectors of length == 1). Any unnamed elements (e.g., <code>"</code> or <code>NULL</code> ) of the returned object will result in an error.
<code>parallelType</code>	The type of parallel operation to be used (if any). The default is <code>"none"</code> . Forking is not possible on Windows, so if <code>"multicore"</code> is requested on a Windows machine, the request will be changed to <code>"snow"</code> with a message.

ncpus	Integer: number of processes to be used in parallel operation. If NULL (the default) and parallelType c("multicore", "snow"), the default is one less than the maximum number of processors detected by <code>detectCores</code> . This default is also silently set if the user specifies more than the number of processors detected.
cl	An optional <b>parallel</b> or <b>snow</b> cluster for use when parallelType = "snow". If NULL, a "PSOCK" cluster on the local machine is created for the duration of the permuteMeasEq call. If a valid <code>makeCluster</code> object is supplied, parallelType is silently set to "snow", and ncpus is silently set to length(cl).
iseed	Integer: Only used to set the states of the RNG when using parallel options, in which case <code>RNGkind</code> is set to "L'Ecuyer-CMRG" with a message. See <code>clusterSetRNGStream</code> and Section 6 of <code>vignette("parallel", "parallel")</code> for more details. If user supplies an invalid value, iseed is silently set to the default (12345). To set the state of the RNG when not using parallel options, call <code>set.seed</code> before calling permuteMeasEq.

## Details

The function permuteMeasEq provides tests of hypotheses involving measurement equivalence, in one of two frameworks:

- 1 For multiple-group CFA models, provide a pair of nested lavaan objects, the less constrained of which (uncon) freely estimates a set of measurement parameters (e.g., factor loadings, intercepts, or thresholds; specified in param) in all groups, and the more constrained of which (con) constrains those measurement parameters to equality across groups. Group assignment is repeatedly permuted and the models are fit to each permutation, in order to produce an empirical distribution under the null hypothesis of no group differences, both for (a) changes in user-specified fit measures (see AFIs and moreAFIs) and for (b) the maximum modification index among the user-specified equality constraints. Configural invariance can also be tested by providing that fitted lavaan object to con and leaving uncon = NULL, in which case param must be NULL as well.
- 2 In MIMIC models, one or a set of continuous and/or discrete covariates can be permuted, and a constrained model is fit to each permutation in order to provide a distribution of any fit measures (namely, the maximum modification index among fixed parameters in param) under the null hypothesis of measurement equivalence across levels of those covariates.

In either framework, modification indices for equality constraints or fixed parameters specified in param are calculated from the constrained model (con) using the function `lavTestScore`.

For multiple-group CFA models, the multiparameter omnibus null hypothesis of measurement equivalence/invariance is that there are no group differences in any measurement parameters (of a particular type). This can be tested using the anova method on nested lavaan objects, as seen in the output of `measurementInvariance`, or by inspecting the change in alternative fit indices (AFIs) such as the CFI. The permutation randomization method employed by permuteMeasEq generates an empirical distribution of any AFIs under the null hypothesis, so the user is not restricted to using fixed cutoffs proposed by Cheung & Rensvold (2002), Chen (2007), or Meade, Johnson, & Braddy (2008).

If the multiparameter omnibus null hypothesis is rejected, partial invariance can still be established by freeing invalid equality constraints, as long as equality constraints are valid for at least two indicators per factor. Modification indices can be calculated from the constrained model (con), but

multiple testing leads to inflation of Type I error rates. The permutation randomization method employed by permuteMeasEq creates a distribution of the maximum modification index if the null hypothesis is true, which allows the user to control the familywise Type I error rate in a manner similar to Tukey's  $q$  (studentized range) distribution for the Honestly Significant Difference (HSD) post hoc test.

For MIMIC models, DIF can be tested by comparing modification indices of regression paths to the permutation distribution of the maximum modification index, which controls the familywise Type I error rate. The MIMIC approach could also be applied with multiple-group models, but the grouping variable would not be permuted; rather, the covariates would be permuted separately within each group to preserve between-group differences. So whether parameters are constrained or unconstrained across groups, the MIMIC approach is only for testing null hypotheses about the effects of covariates on indicators, controlling for common factors.

In either framework, lavaan's group.label argument is used to preserve the order of groups seen in con when permuting the data.

### Value

The permuteMeasEq object representing the results of testing measurement equivalence (the multiparameter omnibus test) and DIF (modification indices), as well as diagnostics and any extra output.

### Author(s)

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### References

#### Papers about permutation tests of measurement equivalence:

Jorgensen, T. D., Kite, B. A., Chen, P.-Y., & Short, S. D. (in press). Permutation randomization methods for testing measurement equivalence and detecting differential item functioning in multiple-group confirmatory factor analysis. *Psychological Methods*, 23(4), 708–728. doi:10.1037/met0000152

Kite, B. A., Jorgensen, T. D., & Chen, P.-Y. (in press). Random permutation testing applied to measurement invariance testing with ordered-categorical indicators. *Structural Equation Modeling* 25(4), 573–587. doi:10.1080/10705511.2017.1421467

Jorgensen, T. D. (2017). Applying permutation tests and multivariate modification indices to configurationally invariant models that need respecification. *Frontiers in Psychology*, 8(1455). doi:10.3389/fpsyg.2017.01455

#### Additional reading:

Chen, F. F. (2007). Sensitivity of goodness of fit indexes to lack of measurement invariance. *Structural Equation Modeling*, 14(3), 464–504. doi:10.1080/10705510701301834

Cheung, G. W., & Rensvold, R. B. (2002). Evaluating goodness-of-fit indexes for testing measurement invariance. *Structural Equation Modeling*, 9(2), 233–255. doi:10.1207/S15328007SEM0902\_5

Meade, A. W., Johnson, E. C., & Braddy, P. W. (2008). Power and sensitivity of alternative fit indices in tests of measurement invariance. *Journal of Applied Psychology*, 93(3), 568–592. doi:10.1037/0021-9010.93.3.568

Widamin, K. F., & Thompson, J. S. (2003). On specifying the null model for incremental fit indices in structural equation modeling. *Psychological Methods*, 8(1), 16–37. doi:10.1037/1082-989X.8.1.16

**See Also**

[TukeyHSD](#), [lavTestScore](#), [measurementInvariance](#), [measurementInvarianceCat](#)

**Examples**

```
## Not run:

#####
## Multiple-Group CFA ##
#####

## create 3-group data in lavaan example(cfa) data
HS <- lavaan::HolzingerSwineford1939
HS$ageGroup <- ifelse(HS$ageyr < 13, "preteen",
                     ifelse(HS$ageyr > 13, "teen", "thirteen"))

## specify and fit an appropriate null model for incremental fit indices
mod.null <- c(paste0("x", 1:9, " ~ c(T", 1:9, ", T", 1:9, ")*1"),
             paste0("x", 1:9, " ~~ c(L", 1:9, ", L", 1:9, ")*x", 1:9))
fit.null <- cfa(mod.null, data = HS, group = "ageGroup")

## fit target model with varying levels of measurement equivalence
mod.config <- '
visual =~ x1 + x2 + x3
textual =~ x4 + x5 + x6
speed   =~ x7 + x8 + x9
'

miout <- measurementInvariance(model = mod.config, data = HS, std.lv = TRUE,
                              group = "ageGroup")

(fit.config <- miout[["fit.configural"]])
(fit.metric <- miout[["fit.loadings"]])
(fit.scalar <- miout[["fit.intercepts"]])

##### Permutation Method

## fit indices of interest for multiparameter omnibus test
myAFIs <- c("chisq", "cfi", "rmsea", "mfi", "aic")
moreAFIs <- c("gammaHat", "adjGammaHat")

## Use only 20 permutations for a demo. In practice,
## use > 1000 to reduce sampling variability of estimated p values

## test configural invariance
set.seed(12345)
out.config <- permuteMeasEq(nPermute = 20, con = fit.config)
out.config

## test metric equivalence
set.seed(12345) # same permutations
```

```

out.metric <- permutMeasEq(nPermute = 20, uncon = fit.config, con = fit.metric,
                          param = "loadings", AFIs = myAFIs,
                          moreAFIs = moreAFIs, null = fit.null)
summary(out.metric, nd = 4)

## test scalar equivalence
set.seed(12345) # same permutations
out.scalar <- permutMeasEq(nPermute = 20, uncon = fit.metric, con = fit.scalar,
                          param = "intercepts", AFIs = myAFIs,
                          moreAFIs = moreAFIs, null = fit.null)
summary(out.scalar)

## Not much to see without significant DIF.
## Try using an absurdly high alpha level for illustration.
outsum <- summary(out.scalar, alpha = .50)

## notice that the returned object is the table of DIF tests
outsum

## visualize permutation distribution
hist(out.config, AFI = "chisq")
hist(out.metric, AFI = "chisq", nd = 2, alpha = .01,
      legendArgs = list(x = "topright"))
hist(out.scalar, AFI = "cfi", printLegend = FALSE)

##### Extra Output

## function to calculate expected change of Group-2 and -3 latent means if
## each intercept constraint were released
extra <- function(con) {
  output <- list()
  output["x1.vis2"] <- lavTestScore(con, release = 19:20, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[70]
  output["x1.vis3"] <- lavTestScore(con, release = 19:20, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[106]
  output["x2.vis2"] <- lavTestScore(con, release = 21:22, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[70]
  output["x2.vis3"] <- lavTestScore(con, release = 21:22, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[106]
  output["x3.vis2"] <- lavTestScore(con, release = 23:24, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[70]
  output["x3.vis3"] <- lavTestScore(con, release = 23:24, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[106]
  output["x4.txt2"] <- lavTestScore(con, release = 25:26, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[71]
  output["x4.txt3"] <- lavTestScore(con, release = 25:26, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[107]
  output["x5.txt2"] <- lavTestScore(con, release = 27:28, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[71]
  output["x5.txt3"] <- lavTestScore(con, release = 27:28, univariate = FALSE,
                                   epc = TRUE, warn = FALSE)$epc$epc[107]
  output["x6.txt2"] <- lavTestScore(con, release = 29:30, univariate = FALSE,

```

```

                                epc = TRUE, warn = FALSE)$epc$epc[71]
output["x6.txt3"] <- lavTestScore(con, release = 29:30, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[107]
output["x7.spd2"] <- lavTestScore(con, release = 31:32, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[72]
output["x7.spd3"] <- lavTestScore(con, release = 31:32, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[108]
output["x8.spd2"] <- lavTestScore(con, release = 33:34, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[72]
output["x8.spd3"] <- lavTestScore(con, release = 33:34, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[108]
output["x9.spd2"] <- lavTestScore(con, release = 35:36, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[72]
output["x9.spd3"] <- lavTestScore(con, release = 35:36, univariate = FALSE,
                                epc = TRUE, warn = FALSE)$epc$epc[108]

  output
}

## observed EPC
extra(fit.scalar)

## permutation results, including extra output
set.seed(12345) # same permutations
out.scalar <- permuteMeasEq(nPermute = 20, uncon = fit.metric, con = fit.scalar,
                           param = "intercepts", AFIs = myAFIs,
                           moreAFIs = moreAFIs, null = fit.null, extra = extra)

## summarize extra output
summary(out.scalar, extra = TRUE)

#####
## MIMIC ##
#####

## Specify Restricted Factor Analysis (RFA) model, equivalent to MIMIC, but
## the factor covaries with the covariate instead of being regressed on it.
## The covariate defines a single-indicator construct, and the
## double-mean-centered products of the indicators define a latent
## interaction between the factor and the covariate.
mod.mimic <- '
visual =~ x1 + x2 + x3
age =~ ageyr
age.by.vis =~ x1.ageyr + x2.ageyr + x3.ageyr

x1 ~~ x1.ageyr
x2 ~~ x2.ageyr
x3 ~~ x3.ageyr
'

HS.orth <- indProd(var1 = paste0("x", 1:3), var2 = "ageyr", match = FALSE,
                  data = HS[ , c("ageyr", paste0("x", 1:3))] )
fit.mimic <- cfa(mod.mimic, data = HS.orth, meanstructure = TRUE)
summary(fit.mimic, stand = TRUE)

```

```

## Whereas MIMIC models specify direct effects of the covariate on an indicator,
## DIF can be tested in RFA models by specifying free loadings of an indicator
## on the covariate's construct (uniform DIF, scalar invariance) and the
## interaction construct (nonuniform DIF, metric invariance).
param <- as.list(paste0("age + age.by.vis =~ x", 1:3))
names(param) <- paste0("x", 1:3)
# param <- as.list(paste0("x", 1:3, " ~ age + age.by.vis")) # equivalent

## test both parameters simultaneously for each indicator
do.call(rbind, lapply(param, function(x) lavTestScore(fit.mimic, add = x)$test))
## or test each parameter individually
lavTestScore(fit.mimic, add = as.character(param))

##### Permutation Method

## function to recalculate interaction terms after permuting the covariate
datafun <- function(data) {
  d <- data[, c(paste0("x", 1:3), "ageyr")]
  indProd(var1 = paste0("x", 1:3), var2 = "ageyr", match = FALSE, data = d)
}

set.seed(12345)
perm.mimic <- permuteMeasEq(nPermute = 20, modelType = "mimic",
                           con = fit.mimic, param = param,
                           covariates = "ageyr", datafun = datafun)

summary(perm.mimic)

## End(Not run)

```

---

permuteMeasEq-class     *Class for the Results of Permutation Randomization Tests of Measurement Equivalence and DIF*

---

## Description

This class contains the results of tests of Measurement Equivalence and Differential Item Functioning (DIF).

## Usage

```

## S4 method for signature 'permuteMeasEq'
show(object)

## S4 method for signature 'permuteMeasEq'
summary(object, alpha = 0.05, nd = 3, extra = FALSE)

```

```
## S4 method for signature 'permuteMeasEq'
hist(x, ..., AFI, alpha = 0.05, nd = 3,
     printLegend = TRUE, legendArgs = list(x = "topleft"))
```

### Arguments

object, x	object of class <code>permuteMeasEq</code>
alpha	alpha level used to draw confidence limits in <code>hist</code> and flag significant statistics in summary output
nd	number of digits to display
extra	logical indicating whether the summary output should return permutation-based $p$ values for each statistic returned by the <code>extra</code> function. If <code>FALSE</code> (default), summary will return permutation-based $p$ values for each modification index.
...	Additional arguments to pass to <code>hist</code>
AFI	character indicating the fit measure whose permutation distribution should be plotted
printLegend	logical. If <code>TRUE</code> (default), a legend will be printed with the histogram
legendArgs	list of arguments passed to the <code>legend</code> function. The default argument is a list placing the legend at the top-left of the figure.

### Value

- The `show` method prints a summary of the multiparameter omnibus test results, using the user-specified AFIs. The parametric  $(\Delta)\chi^2$  test is also displayed.
- The `summary` method prints the same information from the `show` method, but when `extra = FALSE` (the default) it also provides a table summarizing any requested follow-up tests of DIF using modification indices in slot `MI.obs`. The user can also specify an alpha level for flagging modification indices as significant, as well as `nd` (the number of digits displayed). For each modification index, the  $p$  value is displayed using a central  $\chi^2$  distribution with the  $df$  shown in that column. Additionally, a  $p$  value is displayed using the permutation distribution of the maximum index, which controls the familywise Type I error rate in a manner similar to Tukey's studentized range test. If any indices are flagged as significant using the `tukey.p.value`, then a message is displayed for each flagged index. The invisibly returned `data.frame` is the displayed table of modification indices, unless `permuteMeasEq` was called with `param = NULL`, in which case the invisibly returned object is `object`. If `extra = TRUE`, the permutation-based  $p$  values for each statistic returned by the `extra` function are displayed and returned in a `data.frame` instead of the modification indices requested in the `param` argument.
- The `hist` method returns a list of length == 2, containing the arguments for the call to `hist` and the arguments to the call for `legend`, respectively. This list may facilitate creating a customized histogram of `AFI.dist`, `MI.dist`, or `extra.dist`

### Slots

`PT` A `data.frame` returned by a call to `parTable` on the constrained model

`modelType` A character indicating the specified `modelType` in the call to `permuteMeasEq`



- ANOVA** A numeric vector indicating the results of the observed  $(\Delta)\chi^2$  test, based on the central  $\chi^2$  distribution
- AFI.obs** A vector of observed (changes in) user-selected fit measures
- AFI.dist** The permutation distribution(s) of user-selected fit measures. A data.frame with `n.Permutations` rows and one column for each `AFI.obs`.
- AFI.pval** A vector of  $p$  values (one for each element in slot `AFI.obs`) calculated using slot `AFI.dist`, indicating the probability of observing a change at least as extreme as `AFI.obs` if the null hypothesis were true
- MI.obs** A data.frame of observed Lagrange Multipliers (modification indices) associated with the equality constraints or fixed parameters specified in the `param` argument. This is a subset of the output returned by a call to `lavTestScore` on the constrained model.
- MI.dist** The permutation distribution of the maximum modification index (among those seen in slot `MI.obs`\$X2) at each permutation of group assignment or of covariates
- extra.obs** If `permutMeasEq` was called with an `extra` function, the output when applied to the original data is concatenated into this vector
- extra.dist** A data.frame, each column of which contains the permutation distribution of the corresponding statistic in slot `extra.obs`
- n.Permutations** An integer indicating the number of permutations requested by the user
- n.Converged** An integer indicating the number of permutation iterations which yielded a converged solution
- n.nonConverged** An integer vector of length `n.Permutations` indicating how many times group assignment was randomly permuted (at each iteration) before converging on a solution
- n.Sparse** Only relevant with ordered indicators when `modelType == "mgcfa"`. An integer vector of length `n.Permutations` indicating how many times group assignment was randomly permuted (at each iteration) before obtaining a sample with all categories observed in all groups.
- oldSeed** An integer vector storing the value of `.Random.seed` before running `permutMeasEq`. Only relevant when using a parallel/multicore option and the original `RNGkind() != "L'Ecuyer-CMRG"`. This enables users to restore their previous `.Random.seed` state, if desired, by running: `.Random.seed[-1] <-permutedResults@oldSeed[-1]`

### Objects from the Class

Objects can be created via the `permutMeasEq` function.

### Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### See Also

`permutMeasEq`

## Examples

```
# See the example from the permuteMeasEq function
```

---

plausibleValues	<i>Plausible-Values Imputation of Factor Scores Estimated from a lavaan Model</i>
-----------------	---

---

## Description

Draw plausible values of factor scores estimated from a fitted [lavaan](#) model, then treat them as multiple imputations of missing data using [runMI](#).

## Usage

```
plausibleValues(object, nDraws = 20L, seed = 12345,
  omitimps = c("no.conv", "no.se"), ...)
```

## Arguments

object	A fitted model of class <a href="#">lavaan</a> , <a href="#">blavaan</a> , or <a href="#">lavaan.mi</a>
nDraws	integer specifying the number of draws, analogous to the number of imputed data sets. If object is of class <a href="#">lavaan.mi</a> , this will be the number of draws taken <i>per imputation</i> . Ignored if object is of class <a href="#">blavaan</a> , in which case the number of draws is the number of MCMC samples from the posterior.
seed	integer passed to <a href="#">set.seed()</a> . Ignored if object is of class <a href="#">blavaan</a> ,
omitimps	character vector specifying criteria for omitting imputations when object is of class <a href="#">lavaan.mi</a> . Can include any of <code>c("no.conv", "no.se", "no.npd")</code> .
...	Optional arguments to pass to <a href="#">lavPredict</a> . <code>assemble</code> will be ignored because multiple groups are always assembled into a single <code>data.frame</code> per draw. <code>type</code> will be ignored because it is set internally to <code>type="lv"</code> .

## Details

Because latent variables are unobserved, they can be considered as missing data, which can be imputed using Monte Carlo methods. This may be of interest to researchers with sample sizes too small to fit their complex structural models. Fitting a factor model as a first step, [lavPredict](#) provides factor-score estimates, which can be treated as observed values in a path analysis (Step 2). However, the resulting standard errors and test statistics could not be trusted because the Step-2 analysis would not take into account the uncertainty about the estimated factor scores. Using the asymptotic sampling covariance matrix of the factor scores provided by [lavPredict](#), [plausibleValues](#) draws a set of `nDraws` imputations from the sampling distribution of each factor score, returning a list of data sets that can be treated like multiple imputations of incomplete data. If the data were already imputed to handle missing data, [plausibleValues](#) also accepts an object of class [lavaan.mi](#), and will draw `nDraws` plausible values from each imputation. Step 2 would then take into account

uncertainty about both missing values and factor scores. Bayesian methods can also be used to generate factor scores, as available with the **blavaan** package, in which case plausible values are simply saved parameters from the posterior distribution. See Asparouhov and Muthen (2010) for further technical details and references.

Each returned `data.frame` includes a `case.idx` column that indicates the corresponding rows in the data set to which the model was originally fitted (unless the user requests only Level-2 variables). This can be used to merge the plausible values with the original observed data, but users should note that including any new variables in a Step-2 model might not accurately account for their relationship(s) with factor scores because they were not accounted for in the Step-1 model from which factor scores were estimated.

If object is a multilevel lavaan model, users can request plausible values for latent variables at particular levels of analysis by setting the `lavPredict` argument `level=1` or `level=2`. If the `level` argument is not passed via `...`, then both levels are returned in a single merged data set per draw. For multilevel models, each returned `data.frame` also includes a column indicating to which cluster each row belongs (unless the user requests only Level-2 variables).

### Value

A list of length `nDraws`, each of which is a `data.frame` containing plausible values, which can be treated as a list of imputed data sets to be passed to `runMI` (see **Examples**). If object is of class `lavaan.mi`, the list will be of length `nDraws*m`, where `m` is the number of imputations.

### Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### References

Asparouhov, T. & Muthen, B. O. (2010). *Plausible values for latent variables using Mplus*. Technical Report. Retrieved from [www.statmodel.com/download/Plausible.pdf](http://www.statmodel.com/download/Plausible.pdf)

### See Also

`runMI`, `lavaan.mi`

### Examples

```
## example from ?cfa and ?lavPredict help pages
HS.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed  =~ x7 + x8 + x9 '

fit1 <- cfa(HS.model, data = HolzingerSwineford1939)
fs1 <- plausibleValues(fit1, nDraws = 3,
                      ## lavPredict() can add only the modeled data
                      append.data = TRUE)

lapply(fs1, head)

## To merge factor scores to original data.frame (not just modeled data)
```

```

fs1 <- plausibleValues(fit1, nDraws = 3)
idx <- lavInspect(fit1, "case.idx") # row index for each case
if (is.list(idx)) idx <- do.call(c, idx) # for multigroup models
data(HolzingerSwineford1939) # copy data to workspace
HolzingerSwineford1939$case.idx <- idx # add row index as variable
## loop over draws to merge original data with factor scores
for (i in seq_along(fs1)) {
  fs1[[i]] <- merge(fs1[[i]], HolzingerSwineford1939, by = "case.idx")
}
lapply(fs1, head)

## multiple-group analysis, in 2 steps
step1 <- cfa(HS.model, data = HolzingerSwineford1939, group = "school",
  group.equal = c("loadings","intercepts"))
PV.list <- plausibleValues(step1)

## subsequent path analysis
path.model <- ' visual ~ c(t1, t2)*textual + c(s1, s2)*speed '
## Not run:
step2 <- sem.mi(path.model, data = PV.list, group = "school")
## test equivalence of both slopes across groups
lavTestWald.mi(step2, constraints = 't1 == t2 ; s1 == s2')

## End(Not run)

## multilevel example from ?Demo.twolevel help page
model <- '
  level: 1
    fw =~ y1 + y2 + y3
    fw ~ x1 + x2 + x3
  level: 2
    fb =~ y1 + y2 + y3
    fb ~ w1 + w2
'
msem <- sem(model, data = Demo.twolevel, cluster = "cluster")
m1PVs <- plausibleValues(msem, nDraws = 3) # both levels by default
lapply(m1PVs, head, n = 10)
## only Level 1
m1PV1 <- plausibleValues(msem, nDraws = 3, level = 1)
lapply(m1PV1, head)
## only Level 2
m1PV2 <- plausibleValues(msem, nDraws = 3, level = 2)
lapply(m1PV2, head)

```

**Description**

This function will plot the line graphs representing the simple effect of the independent variable given the values of the moderator. For multigroup models, it will only generate a plot for 1 group, as specified in the function used to obtain the first argument.

**Usage**

```
plotProbe(object, xlim, xlab = "Independent Variable",
          ylab = "Dependent Variable", legend = TRUE, legendArgs = list(), ...)
```

**Arguments**

object	The result of probing latent interaction obtained from <a href="#">probe2WayMC</a> , <a href="#">probe2WayRC</a> , <a href="#">probe3WayMC</a> , or <a href="#">probe3WayRC</a> function.
xlim	The vector of two numbers: the minimum and maximum values of the independent variable
xlab	The label of the x-axis
ylab	The label of the y-axis
legend	logical. If TRUE (default), a legend is printed.
legendArgs	list of arguments passed to <a href="#">legend</a> function if legend=TRUE.
...	Any addition argument for the <a href="#">plot</a> function

**Value**

None. This function will plot the simple main effect only.

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**See Also**

- [indProd](#) For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- [probe2WayMC](#) For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe3WayMC](#) For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe2WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [probe3WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.

**Examples**

```

library(lavaan)

dat2wayMC <- indProd(dat2way, 1:3, 4:6)

model1 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f12 =~ x1.x4 + x2.x5 + x3.x6
f3 =~ x7 + x8 + x9
f3 ~ f1 + f2 + f12
f12 ~~ 0*f1
f12 ~~ 0*f2
x1 ~ 0*x1
x4 ~ 0*x1
x1.x4 ~ 0*x1
x7 ~ 0*x1
f1 ~ NA*x1
f2 ~ NA*x1
f12 ~ NA*x1
f3 ~ NA*x1
"

fitMC2way <- sem(model1, data = dat2wayMC, std.lv = FALSE,
                 meanstructure = TRUE)
result2wayMC <- probe2WayMC(fitMC2way, c("f1", "f2", "f12"),
                            "f3", "f2", c(-1, 0, 1))
plotProbe(result2wayMC, xlim = c(-2, 2))

dat3wayMC <- indProd(dat3way, 1:3, 4:6, 7:9)

model3 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f3 =~ x7 + x8 + x9
f12 =~ x1.x4 + x2.x5 + x3.x6
f13 =~ x1.x7 + x2.x8 + x3.x9
f23 =~ x4.x7 + x5.x8 + x6.x9
f123 =~ x1.x4.x7 + x2.x5.x8 + x3.x6.x9
f4 =~ x10 + x11 + x12
f4 ~ f1 + f2 + f3 + f12 + f13 + f23 + f123
f1 ~~ 0*f12
f1 ~~ 0*f13
f1 ~~ 0*f123
f2 ~~ 0*f12
f2 ~~ 0*f23
f2 ~~ 0*f123
f3 ~~ 0*f13
f3 ~~ 0*f23
f3 ~~ 0*f123

```

```

f12 ~~ 0*f123
f13 ~~ 0*f123
f23 ~~ 0*f123
x1 ~ 0*1
x4 ~ 0*1
x7 ~ 0*1
x10 ~ 0*1
x1.x4 ~ 0*1
x1.x7 ~ 0*1
x4.x7 ~ 0*1
x1.x4.x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f3 ~ NA*1
f12 ~ NA*1
f13 ~ NA*1
f23 ~ NA*1
f123 ~ NA*1
f4 ~ NA*1
"

fitMC3way <- sem(model3, data = dat3wayMC, std.lv = FALSE,
                 meanstructure = TRUE)
result3wayMC <- probe3WayMC(fitMC3way,
                             c("f1", "f2", "f3", "f12", "f13", "f23", "f123"),
                             "f4", c("f1", "f2"), c(-1, 0, 1), c(-1, 0, 1))
plotProbe(result3wayMC, xlim = c(-2, 2))

```

---

plotRMSEAdist

*Plot the sampling distributions of RMSEA*


---

### Description

Plots the sampling distributions of RMSEA based on the noncentral chi-square distributions

### Usage

```
plotRMSEAdist(rmseas, n, df, ptile = NULL, caption = NULL,
             rmseaScale = TRUE, group = 1)
```

### Arguments

rmseas	The vector of RMSEA values to be plotted
n	Sample size of a dataset
df	Model degrees of freedom
ptile	The percentile rank of the distribution of the first RMSEA that users wish to plot a vertical line in the resulting graph

caption	The name vector of each element of rmsea
rmseaScale	If TRUE, the RMSEA scale is used in the x-axis. If FALSE, the chi-square scale is used in the x-axis.
group	The number of group that is used to calculate RMSEA.

### Details

This function creates overlapping plots of the sampling distribution of RMSEA based on noncentral  $\chi^2$  distribution (MacCallum, Browne, & Suguwara, 1996). First, the noncentrality parameter ( $\lambda$ ) is calculated from RMSEA (Steiger, 1998; Dudgeon, 2004) by

$$\lambda = (N - 1)d\varepsilon^2/K,$$

where  $N$  is sample size,  $d$  is the model degree of freedom,  $K$  is the number of group, and  $\varepsilon$  is the population RMSEA. Next, the noncentral  $\chi^2$  distribution with a specified  $df$  and noncentrality parameter is plotted. Thus, the x-axis represents the sample  $\chi^2$  value. The sample  $\chi^2$  value can be transformed to the sample RMSEA scale ( $\hat{\varepsilon}$ ) by

$$\hat{\varepsilon} = \sqrt{K} \sqrt{\frac{\chi^2 - d}{(N - 1)d}},$$

where  $\chi^2$  is the  $\chi^2$  value obtained from the noncentral  $\chi^2$  distribution.

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

### References

- Dudgeon, P. (2004). A note on extending Steiger's (1998) multiple sample RMSEA adjustment to other noncentrality parameter-based statistic. *Structural Equation Modeling*, 11(3), 305–319. doi:10.1207/s15328007sem1103\_1
- MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, 1(2), 130–149. doi:10.1037/1082-989X.1.2.130
- Steiger, J. H. (1998). A note on multiple sample extensions of the RMSEA fit index. *Structural Equation Modeling*, 5(4), 411–419. doi:10.1080/10705519809540115

### See Also

- [plotRMSEApower](#) to plot the statistical power based on population RMSEA given the sample size
- [findRMSEApower](#) to find the statistical power based on population RMSEA given a sample size
- [findRMSEAsamplesize](#) to find the minimum sample size for a given statistical power based on population RMSEA



**Examples**

```
plotRMSEAdist(c(.05, .08), n = 200, df = 20, ptile = .95, rmseaScale = TRUE)
plotRMSEAdist(c(.05, .01), n = 200, df = 20, ptile = .05, rmseaScale = FALSE)
```

---

plotRMSEApower      *Plot power curves for RMSEA*

---

**Description**

Plots power of RMSEA over a range of sample sizes

**Usage**

```
plotRMSEApower(rmseas0, rmseaA, df, nlow, nhigh, steps = 1, alpha = 0.05,
  group = 1, ...)
```

**Arguments**

rmseas0	Null RMSEA
rmseaA	Alternative RMSEA
df	Model degrees of freedom
nlow	Lower sample size
nhigh	Upper sample size
steps	Increase in sample size for each iteration. Smaller values of steps will lead to more precise plots. However, smaller step sizes means a longer run time.
alpha	Alpha level used in power calculations
group	The number of group that is used to calculate RMSEA.
...	The additional arguments for the plot function.

**Details**

This function creates plot of power for RMSEA against a range of sample sizes. The plot places sample size on the horizontal axis and power on the vertical axis. The user should indicate the lower and upper values for sample size and the sample size between each estimate ("step size") We strongly urge the user to read the sources below (see References) before proceeding. A web version of this function is available at: <http://quantpsy.org/rmsea/rmseaplot.htm>.

**Value**

Plot of power for RMSEA against a range of sample sizes

**Author(s)**

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Kristopher J. Preacher (Vanderbilt University; <kris.preacher@vanderbilt.edu>)

Donna L. Coffman (Pennsylvania State University; <dlc30@psu.edu.>)

**References**

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods*, *11*(1), 19–35. doi:10.1037/1082-989X.11.1.19

MacCallum, R. C., Browne, M. W., & Sugawara, H. M. (1996). Power analysis and determination of sample size for covariance structure modeling. *Psychological Methods*, *1*(2), 130–149. doi:10.1037/1082-989X.1.2.130

MacCallum, R. C., Lee, T., & Browne, M. W. (2010). The issue of isopower in power analysis for tests of structural equation models. *Structural Equation Modeling*, *17*(1), 23–41. doi:10.1080/10705510903438906

Preacher, K. J., Cai, L., & MacCallum, R. C. (2007). Alternatives to traditional model comparison strategies for covariance structure models. In T. D. Little, J. A. Bovaird, & N. A. Card (Eds.), *Modeling contextual effects in longitudinal studies* (pp. 33–62). Mahwah, NJ: Lawrence Erlbaum Associates.

Steiger, J. H. (1998). A note on multiple sample extensions of the RMSEA fit index. *Structural Equation Modeling*, *5*(4), 411–419. doi:10.1080/10705519809540115

Steiger, J. H., & Lind, J. C. (1980, June). *Statistically based tests for the number of factors*. Paper presented at the annual meeting of the Psychometric Society, Iowa City, IA.

**See Also**

- [plotRMSEAdist](#) to visualize the RMSEA distributions
- [findRMSEApower](#) to find the statistical power based on population RMSEA given a sample size
- [findRMSEAsamplesize](#) to find the minimum sample size for a given statistical power based on population RMSEA

**Examples**

```
plotRMSEApower(rmseA0 = .025, rmseA = .075, df = 23,  
               nlow = 100, nhigh = 500, steps = 10)
```

---

plotRMSEApowernested *Plot power of nested model RMSEA*

---

### Description

Plot power of nested model RMSEA over a range of possible sample sizes.

### Usage

```
plotRMSEApowernested(rmse0A = NULL, rmse0B = NULL, rmse1A,
  rmse1B = NULL, dfA, dfB, nlow, nhigh, steps = 1, alpha = 0.05,
  group = 1, ...)
```

### Arguments

rmse0A	The $H_0$ baseline RMSEA
rmse0B	The $H_0$ alternative RMSEA (trivial misfit)
rmse1A	The $H_1$ baseline RMSEA
rmse1B	The $H_1$ alternative RMSEA (target misfit to be rejected)
dfA	degree of freedom of the more-restricted model
dfB	degree of freedom of the less-restricted model
nlow	Lower bound of sample size
nhigh	Upper bound of sample size
steps	Step size
alpha	The alpha level
group	The number of group in calculating RMSEA
...	The additional arguments for the plot function.

### Author(s)

Bell Clinton

Pavel Panko (Texas Tech University; <pavel.panko@ttu.edu>)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

### References

MacCallum, R. C., Browne, M. W., & Cai, L. (2006). Testing differences between nested covariance structure models: Power analysis and null hypotheses. *Psychological Methods, 11*(1), 19-35. doi:10.1037/1082-989X.11.1.19

**See Also**

- [findRMSEApowernested](#) to find the power for a given sample size in nested model comparison based on population RMSEA
- [findRMSEAsamplesizenested](#) to find the minimum sample size for a given statistical power in nested model comparison based on population RMSEA

**Examples**

```
plotRMSEApowernested(rmse0A = 0, rmse0B = 0, rmse1A = 0.06,
                     rmse1B = 0.05, dfA = 22, dfB = 20, nlow = 50,
                     nhigh = 500, steps = 1, alpha = .05, group = 1)
```

---

poolMAlloc

*Pooled estimates and standard errors across M parcel-allocations:  
Combining sampling variability and parcel-allocation variability.*

---

**Description**

This function employs an iterative algorithm to pick the number of random item-to-parcel allocations needed to meet user-defined stability criteria for a fitted structural equation model (SEM) (see **Details** below for more information). Pooled point and standard-error estimates from this SEM can be outputted at this final selected number of allocations (however, it is more efficient to save the allocations and treat them as multiple imputations using [runMI](#); see **See Also** for links with examples). Additionally, new indices (see Sterba & Rights, 2016) are outputted for assessing the relative contributions of parcel-allocation variability vs. sampling variability in each estimate. At each iteration, this function generates a given number of random item-to-parcel allocations, fits a SEM to each allocation, pools estimates across allocations from that iteration, and then assesses whether stopping criteria are met. If stopping criteria are not met, the algorithm increments the number of allocations used (generating all new allocations).

**Usage**

```
poolMAlloc(nPerPar, facPlc, nAllocStart, nAllocAdd = 0,
           parceloutput = NULL, syntax, dataset, stopProp, stopValue,
           selectParam = NULL, indices = "default", double = FALSE,
           checkConv = FALSE, names = "default", leaveout = 0,
           useTotalAlloc = FALSE, ...)
```

**Arguments**

**nPerPar** A list in which each element is a vector, corresponding to each factor, indicating sizes of parcels. If variables are left out of parceling, they should not be accounted for here (i.e., there should not be parcels of size "1").

facPlc	A list of vectors, each corresponding to a factor, specifying the item indicators of that factor (whether included in parceling or not). Either variable names or column numbers. Variables not listed will not be modeled or included in output datasets.
nAllocStart	The number of random allocations of items to parcels to generate in the first iteration of the algorithm.
nAllocAdd	The number of allocations to add with each iteration of the algorithm. Note that if only one iteration is desired, nAllocAdd can be set to 0 and results will be output for nAllocStart allocations only.
parceloutput	Optional character. Path (folder/directory) where $M$ (the final selected number of allocations) parceled data sets will be outputted from the iteration where the algorithm met stopping criteria. Note for Windows users: file path must be specified using forward slashes (/), not backslashes (\). See <a href="#">path.expand</a> for details. If NULL (default), nothing is saved to disk.
syntax	lavaan syntax that defines the model.
dataset	Item-level dataset
stopProp	Value used in defining stopping criteria of the algorithm ( $\delta_a$ in Sterba & Rights, 2016). This is the minimum proportion of change (in any pooled parameter or pooled standard error estimate listed in selectParam) that is allowable from one iteration of the algorithm to the next. That is, change in pooled estimates and pooled standard errors from one iteration to the next must all be less than (stopProp) x (value from former iteration). Note that stopValue can override this criterion (see below). Also note that values less than .01 are unlikely to lead to more substantively meaningful precision. Also note that if only stopValue is a desired criterion, stopProp can be set to 0.
stopValue	Value used in defining stopping criteria of the algorithm ( $\delta_b$ in Sterba & Rights, 2016). stopValue is a minimum allowable amount of absolute change (in any pooled parameter or pooled standard error estimate listed in selectParam) from one iteration of the algorithm to the next. For a given pooled estimate or pooled standard error, stopValue is only invoked as a stopping criteria when the minimum change required by stopProp is less than stopValue. Note that values less than .01 are unlikely to lead to more substantively meaningful precision. Also note that if only stopProp is a desired criterion, stopValue can be set to 0.
selectParam	(Optional) A list of the pooled parameters to be used in defining stopping criteria (i.e., stopProp and stopValue). These parameters should appear in the order they are listed in the lavaan syntax. By default, all pooled parameters are used. Note that selectParam should only contain freely-estimated parameters. In one example from Sterba & Rights (2016) selectParam included all free parameters except item intercepts and in another example selectParam included only structural parameters.
indices	Optional character vector indicating the names of available <a href="#">fitMeasures</a> to be included in the output. The first and second elements should be a chi-squared test statistic and its associated degrees of freedom, both of which will be added if missing. If "default", the indices will be c("chisq", "df", "cfi", "tli", "rmsea", "srmr"). If a robust test statistic is requested (see <a href="#">lavOptions</a> ), c("chisq", "df") will

	be replaced by <code>c("chisq.scaled", "df.scaled")</code> . For the output to include both the naive and robust test statistics, indices should include both, but put the scaled test statistics first, as in <code>indices = c("chisq.scaled", "df.scaled", "chisq", "df")</code>
double	(Optional) If set to TRUE, requires stopping criteria ( <code>stopProp</code> and <code>stopValue</code> ) to be met for all parameters (in <code>selectParam</code> ) for two consecutive iterations of the algorithm. By default, this is set to FALSE, meaning stopping criteria need only be met at one iteration of the algorithm.
checkConv	(Optional) If set to TRUE, function will output pooled estimates and standard errors from 10 iterations post-convergence.
names	(Optional) A character vector containing the names of parceled variables.
leaveout	(Optional) A vector of variables to be left out of randomized parceling. Either variable names or column numbers are allowed.
useTotalAlloc	(Optional) If set to TRUE, function will output a separate set of results that uses all allocations created by the algorithm, rather than $M$ allocations (see "Allocations needed for stability" below). This distinction is further discussed in Sterba and Rights (2016).
...	Additional arguments to be passed to <code>lavaan</code> . See also <code>lavOptions</code>

## Details

For further details on the benefits of the random allocation of items to parcels, see Sterba (2011) and Sterba & MacCallum (2010).

This function implements an algorithm for choosing the number of allocations ( $M$ ; described in Sterba & Rights, 2016), pools point and standard-error estimates across these  $M$  allocations, and produces indices for assessing the relative contributions of parcel-allocation variability vs. sampling variability in each estimate.

To obtain pooled test statistics for model fit or model comparison, the `list` or `parcel` allocations can be passed to `runMI` (find **Examples** on the help pages for `parcelAllocation` and `PAVranking`).

This function randomly generates a given number (`nAllocStart`) of item-to-parcel allocations, fits a SEM to each allocation, and then increments the number of allocations used (by `nAllocAdd`) until the pooled point and standard-error estimates fulfill stopping criteria (`stopProp` and `stopValue`, defined above). A summary of results from the model that was fit to the  $M$  allocations are returned.

Additionally, this function outputs the proportion of allocations with solutions that converged (using a maximum likelihood estimator) as well as the proportion of allocations with solutions that were converged and proper. The converged and proper solutions among the final  $M$  allocations are used in computing pooled results.

Additionally, after each iteration of the algorithm, information useful in monitoring the algorithm is outputted. The number of allocations used at that iteration, the proportion of pooled parameter estimates meeting stopping criteria at the previous iteration, the proportion of pooled standard errors meeting stopping criteria at the previous iteration, and the runtime of that iteration are outputted. When stopping criteria are satisfied, the full set of results are outputted.

## Value

**Estimates** A table containing pooled results across  $M$  allocations at the iteration where stopping criteria were met. Columns correspond to individual parameter name,

pooled estimate, pooled standard error,  $p$ -value for a  $z$ -test of the parameter,  $z$ -based 95% confidence interval,  $p$ -value for a  $t$ -test of the parameter (using degrees of freedom described in Sterba & Rights, 2016), and  $t$ -based 95% confidence interval for the parameter.

**Fit** A table containing results related to model fit from the  $M$  allocations at the iteration where stopping criteria were met. Columns correspond to fit index names, the average of each index across allocations, the standard deviation of each fit index across allocations, the maximum of each fit index across allocations, the minimum of each fit index across allocations, the range of each fit index across allocations, and the percent of the  $M$  allocations where the chi-square test of absolute fit was significant.

**Proportion of converged and proper allocations**

A table containing the proportion of the final  $M$  allocations that converged (using a maximum likelihood estimator) and the proportion of allocations that converged to proper solutions. Note that pooled estimates, pooled standard errors, and other results are computed using only the converged, proper allocations.

**Allocations needed for stability (M)**

The number of allocations ( $M$ ) at which the algorithm's stopping criteria (defined above) were met.

**Indices used to quantify uncertainty in estimates due to sample vs. allocation variability**

A table containing individual parameter names, an estimate of the proportion of total variance of a pooled parameter estimate that is attributable to parcel-allocation variability (PPAV), and an estimate of the ratio of the between-allocation variance of a pooled parameter estimate to the within-allocation variance (RPAV). See Sterba & Rights (2016) for more detail.

**Total runtime (minutes)**

The total runtime of the function, in minutes. Note that the total runtime will be greater when the specified model encounters convergence problems for some allocations, as is the case with the `simParcel` dataset used below.

## Author(s)

Jason D. Rights (Vanderbilt University; <jason.d.rights@vanderbilt.edu>)

The author would also like to credit Corbin Quick and Alexander Schoemann for providing the original `parcelAllocation` function on which this function is based.

## References

- Sterba, S. K. (2011). Implications of parcel-allocation variability for comparing fit of item-solutions and parcel-solutions. *Structural Equation Modeling*, 18(4), 554–577. doi:10.1080/10705511.2011.607073
- Sterba, S. K., & MacCallum, R. C. (2010). Variability in parameter estimates and model fit across random allocations of items to parcels. *Multivariate Behavioral Research*, 45(2), 322–358. doi:10.1080/00273171003680302
- Sterba, S. K., & Rights, J. D. (2016). Accounting for parcel-allocation variability in practice: Combining sources of uncertainty and choosing the number of allocations. *Multivariate Behavioral Research*, 51(2–3), 296–313. doi:10.1080/00273171.2016.1144502

Sterba, S. K., & Rights, J. D. (2017). Effects of parceling on model selection: Parcel-allocation variability in model ranking. *Psychological Methods*, 22(1), 47–68. doi:10.1037/met0000067

### See Also

[runMI](#) for treating allocations as multiple imputations to pool results across allocations. See **Examples** on help pages for:

- [parcelAllocation](#) for fitting a single model
- [PAVranking](#) for comparing 2 models

### Examples

```
## Not run:
## lavaan syntax: A 2 Correlated
## factor CFA model to be fit to parceled data

parmodel <- '
  f1 =~ NA*p1f1 + p2f1 + p3f1
  f2 =~ NA*p1f2 + p2f2 + p3f2
  p1f1 ~ 1
  p2f1 ~ 1
  p3f1 ~ 1
  p1f2 ~ 1
  p2f2 ~ 1
  p3f2 ~ 1
  p1f1 ~~ p1f1
  p2f1 ~~ p2f1
  p3f1 ~~ p3f1
  p1f2 ~~ p1f2
  p2f2 ~~ p2f2
  p3f2 ~~ p3f2
  f1 ~~ 1*f1
  f2 ~~ 1*f2
  f1 ~~ f2
'

## specify items for each factor
f1name <- colnames(simParcel)[1:9]
f2name <- colnames(simParcel)[10:18]

## run function
poolMAlloc(nPerPar = list(c(3,3,3), c(3,3,3)),
  facPlc = list(f1name, f2name), nAllocStart = 10, nAllocAdd = 10,
  syntax = parmodel, dataset = simParcel, stopProp = .03,
  stopValue = .03, selectParam = c(1:6, 13:18, 21),
  names = list("p1f1", "p2f1", "p3f1", "p1f2", "p2f2", "p3f2"),
  double = FALSE, useTotalAlloc = FALSE)

## End(Not run)
```



```
## See examples on ?parcelAllocation and ?PAVranking for how to obtain
## pooled test statistics and other pooled lavaan output.
## Details provided in Sterba & Rights (2016).
```

---

probe2WayMC	<i>Probing two-way interaction on the no-centered or mean-centered latent interaction</i>
-------------	---

---

## Description

Probing interaction for simple intercept and simple slope for the no-centered or mean-centered latent two-way interaction

## Usage

```
probe2WayMC(fit, nameX, nameY, modVar, valProbe, group,
  omitimps = c("no.conv", "no.se"))
```

## Arguments

fit	A fitted <a href="#">lavaan</a> or <a href="#">lavaan.mi</a> object with a latent 2-way interaction.
nameX	character vector of all 3 factor names used as the predictors. The lower-order factors must be listed first, and the final name must be the latent interaction factor.
nameY	The name of factor that is used as the dependent variable.
modVar	The name of factor that is used as a moderator. The effect of the other independent factor will be probed at each value of the moderator variable listed in valProbe.
valProbe	The values of the moderator that will be used to probe the effect of the focal predictor.
group	In multigroup models, the label of the group for which the results will be returned. Must correspond to one of <code>lavInspect(fit, "group.label")</code> , or an integer corresponding to which of those group labels.
omitimps	character vector specifying criteria for omitting imputations from pooled results. Ignored unless fit is of class <code>lavaan.mi</code> . Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

## Details

Before using this function, researchers need to make the products of the indicators between the first-order factors using mean centering (Marsh, Wen, & Hau, 2004). Note that the double-mean centering may not be appropriate for probing interaction if researchers are interested in simple intercepts. The mean or double-mean centering can be done by the `indProd` function. The indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

Let that the latent interaction model regressing the dependent variable ( $Y$ ) on the independent variable ( $X$ ) and the moderator ( $Z$ ) be

$$Y = b_0 + b_1X + b_2Z + b_3XZ + r,$$

where  $b_0$  is the estimated intercept or the expected value of  $Y$  when both  $X$  and  $Z$  are 0,  $b_1$  is the effect of  $X$  when  $Z$  is 0,  $b_2$  is the effect of  $Z$  when  $X$  is 0,  $b_3$  is the interaction effect between  $X$  and  $Z$ , and  $r$  is the residual term.

For probing two-way interaction, the simple intercept of the independent variable at each value of the moderator (Aiken & West, 1991; Cohen, Cohen, West, & Aiken, 2003; Preacher, Curran, & Bauer, 2006) can be obtained by

$$b_{0|X=0,Z} = b_0 + b_2Z.$$

The simple slope of the independent variable at each value of the moderator can be obtained by

$$b_{X|Z} = b_1 + b_3Z.$$

The variance of the simple intercept formula is

$$Var(b_{0|X=0,Z}) = Var(b_0) + 2ZCov(b_0, b_2) + Z^2Var(b_2)$$

where  $Var$  denotes the variance of a parameter estimate and  $Cov$  denotes the covariance of two parameter estimates.

The variance of the simple slope formula is

$$Var(b_{X|Z}) = Var(b_1) + 2ZCov(b_1, b_3) + Z^2Var(b_3)$$

Wald  $z$  statistic is used for test statistic (even for objects of class `lavaan.mi`).

## Value

A list with two elements:

1. `SimpleIntercept`: The intercepts given each value of the moderator. This element will be `NULL` unless the factor intercept is estimated (e.g., not fixed at 0).
2. `SimpleSlope`: The slopes given each value of the moderator.

In each element, the first column represents the values of the moderators specified in the `valProbe` argument. The second column is the simple intercept or simple slope. The third column is the *SE* of the simple intercept or simple slope. The fourth column is the Wald ( $z$ ) statistic. The fifth column is the  $p$  value testing whether the simple intercepts or slopes are different from 0.

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

Aiken, L. S., & West, S. G. (1991). *Multiple regression: Testing and interpreting interactions*. Newbury Park, CA: Sage.

Cohen, J., Cohen, P., West, S. G., & Aiken, L. S. (2003). *Applied multiple regression/correlation analysis for the behavioral sciences* (3rd ed.). New York, NY: Routledge.

Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9(3), 275–300. doi:10.1037/1082-989X.9.3.275

Preacher, K. J., Curran, P. J., & Bauer, D. J. (2006). Computational tools for probing interactions in multiple linear regression, multilevel modeling, and latent curve analysis. *Journal of Educational and Behavioral Statistics*, 31(4), 437–448. doi:10.3102/10769986031004437

**See Also**

- [indProd](#) For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- [probe3WayMC](#) For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe2WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [probe3WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [plotProbe](#) Plot the simple intercepts and slopes of the latent interaction.

**Examples**

```
library(lavaan)

dat2wayMC <- indProd(dat2way, 1:3, 4:6)

model1 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f12 =~ x1.x4 + x2.x5 + x3.x6
f3 =~ x7 + x8 + x9
f3 ~ f1 + f2 + f12
f12 ~~0*f1
f12 ~~ 0*f2
x1 ~ 0*1
x4 ~ 0*1
x1.x4 ~ 0*1
```

```

x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f12 ~ NA*1
f3 ~ NA*1
"

fitMC2way <- sem(model1, data = dat2wayMC, std.lv = FALSE,
                 meanstructure = TRUE)
summary(fitMC2way)

probe2WayMC(fitMC2way, nameX = c("f1", "f2", "f12"), nameY = "f3",
            modVar = "f2", valProbe = c(-1, 0, 1))

```

---

probe2WayRC	<i>Probing two-way interaction on the residual-centered latent interaction</i>
-------------	--

---

### Description

Probing interaction for simple intercept and simple slope for the residual-centered latent two-way interaction (Geldhof et al., 2013)

### Usage

```

probe2WayRC(fit, nameX, nameY, modVar, valProbe, group,
            omitimps = c("no.conv", "no.se"))

```

### Arguments

fit	A fitted <a href="#">lavaan</a> or <a href="#">lavaan.mi</a> object with a latent 2-way interaction.
nameX	character vector of all 3 factor names used as the predictors. The lower-order factors must be listed first, and the final name must be the latent interaction factor.
nameY	The name of factor that is used as the dependent variable.
modVar	The name of factor that is used as a moderator. The effect of the other independent factor will be probed at each value of the moderator variable listed in <code>valProbe</code> .
valProbe	The values of the moderator that will be used to probe the effect of the focal predictor.
group	In multigroup models, the label of the group for which the results will be returned. Must correspond to one of <code>lavInspect(fit, "group.label")</code> , or an integer corresponding to which of those group labels.

`omit.imps` character vector specifying criteria for omitting imputations from pooled results. Ignored unless `fit` is of class `lavaan.mi`. Can include any of `c("no.conv", "no.se", "no.npd")`, the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option (`"no.npd"`) would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

### Details

Before using this function, researchers need to make the products of the indicators between the first-order factors and residualize the products by the original indicators (Lance, 1988; Little, Bovaird, & Widaman, 2006). The process can be automated by the `indProd` function. Note that the indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms. To use this function the model must be fit with a mean structure. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

The probing process on residual-centered latent interaction is based on transforming the residual-centered result into the no-centered result. See Geldhof et al. (2013) for further details. Note that this approach based on a strong assumption that the first-order latent variables are normally distributed. The probing process is applied after the no-centered result (parameter estimates and their covariance matrix among parameter estimates) has been computed. See the `probe2WayMC` for further details.

### Value

A list with two elements:

1. `SimpleIntercept`: The intercepts given each value of the moderator. This element will be `NULL` unless the factor intercept is estimated (e.g., not fixed at 0).
2. `SimpleSlope`: The slopes given each value of the moderator.

In each element, the first column represents the values of the moderators specified in the `valProbe` argument. The second column is the simple intercept or simple slope. The third column is the standard error of the simple intercept or simple slope. The fourth column is the Wald ( $z$ ) statistic. The fifth column is the  $p$  value testing whether the simple intercepts or slopes are different from 0.

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## References

- Lance, C. E. (1988). Residual centering, exploratory and confirmatory moderator analysis, and decomposition of effects in path models containing interactions. *Applied Psychological Measurement*, *12*(2), 163–175. doi:10.1177/014662168801200205
- Little, T. D., Bovaird, J. A., & Widaman, K. F. (2006). On the merits of orthogonalizing powered and product terms: Implications for modeling interactions. *Structural Equation Modeling*, *13*(4), 497–519. doi:10.1207/s15328007sem1304\_1
- Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, *9*(3), 275–300. doi:10.1037/1082-989X.9.3.275
- Geldhof, G. J., Pornprasertmanit, S., Schoemann, A. M., & Little, T. D. (2013). Orthogonalizing through residual centering: Extended applications and caveats *Educational and Psychological Measurement*, *73*(1), 27–46. doi:10.1177/0013164412445473

## See Also

- [indProd](#) For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- [probe2WayMC](#) For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe3WayMC](#) For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe3WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [plotProbe](#) Plot the simple intercepts and slopes of the latent interaction.

## Examples

```
library(lavaan)

dat2wayRC <- orthogonalize(dat2way, 1:3, 4:6)

model1 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f12 =~ x1.x4 + x2.x5 + x3.x6
f3 =~ x7 + x8 + x9
f3 ~ f1 + f2 + f12
f12 ~~0*f1
f12 ~~ 0*f2
x1 ~ 0*1
x4 ~ 0*1
x1.x4 ~ 0*1
x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f12 ~ NA*1
```

```

f3 ~ NA*1
"

fitRC2way <- sem(model1, data = dat2wayRC, std.lv = FALSE,
                 meanstructure = TRUE)
summary(fitRC2way)

probe2WayRC(fitRC2way, nameX = c("f1", "f2", "f12"), nameY = "f3",
            modVar = "f2", valProbe = c(-1, 0, 1))

```

---

probe3WayMC	<i>Probing three-way interaction on the no-centered or mean-centered latent interaction</i>
-------------	---

---

### Description

Probing interaction for simple intercept and simple slope for the no-centered or mean-centered latent two-way interaction

### Usage

```

probe3WayMC(fit, nameX, nameY, modVar, valProbe1, valProbe2, group,
            omitimps = c("no.conv", "no.se"))

```

### Arguments

fit	A fitted <code>lavaan</code> or <code>lavaan.mi</code> object with a latent 2-way interaction.
nameX	character vector of all 7 factor names used as the predictors. The 3 lower-order factors must be listed first, followed by the 3 second-order factors (specifically, the 4th element must be the interaction between the factors listed first and second, the 5th element must be the interaction between the factors listed first and third, and the 6th element must be the interaction between the factors listed second and third). The final name will be the factor representing the 3-way interaction.
nameY	The name of factor that is used as the dependent variable.
modVar	The name of two factors that are used as the moderators. The effect of the independent factor on each combination of the moderator variable values will be probed.
valProbe1	The values of the first moderator that will be used to probe the effect of the independent factor.
valProbe2	The values of the second moderator that will be used to probe the effect of the independent factor.
group	In multigroup models, the label of the group for which the results will be returned. Must correspond to one of <code>lavInspect(fit, "group.label")</code> .

`omitimps` character vector specifying criteria for omitting imputations from pooled results. Ignored unless `fit` is of class `lavaan.mi`. Can include any of `c("no.conv", "no.se", "no.npd")`, the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option (`"no.npd"`) would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

## Details

Before using this function, researchers need to make the products of the indicators between the first-order factors using mean centering (Marsh, Wen, & Hau, 2004). Note that the double-mean centering may not be appropriate for probing interaction if researchers are interested in simple intercepts. The mean or double-mean centering can be done by the `indProd` function. The indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

Let that the latent interaction model regressing the dependent variable ( $Y$ ) on the independent variable ( $X$ ) and two moderators ( $Z$  and  $W$ ) be

$$Y = b_0 + b_1X + b_2Z + b_3W + b_4XZ + b_5XW + b_6ZW + b_7XZW + r,$$

where  $b_0$  is the estimated intercept or the expected value of  $Y$  when  $X$ ,  $Z$ , and  $W$  are 0,  $b_1$  is the effect of  $X$  when  $Z$  and  $W$  are 0,  $b_2$  is the effect of  $Z$  when  $X$  and  $W$  is 0,  $b_3$  is the effect of  $W$  when  $X$  and  $Z$  are 0,  $b_4$  is the interaction effect between  $X$  and  $Z$  when  $W$  is 0,  $b_5$  is the interaction effect between  $X$  and  $W$  when  $Z$  is 0,  $b_6$  is the interaction effect between  $Z$  and  $W$  when  $X$  is 0,  $b_7$  is the three-way interaction effect between  $X$ ,  $Z$ , and  $W$ , and  $r$  is the residual term.

For probing three-way interaction, the simple intercept of the independent variable at the specific values of the moderators (Aiken & West, 1991) can be obtained by

$$b_{0|X=0,Z,W} = b_0 + b_2Z + b_3W + b_6ZW.$$

The simple slope of the independent variable at the specific values of the moderators can be obtained by

$$b_{X|Z,W} = b_1 + b_3Z + b_4W + b_7ZW.$$

The variance of the simple intercept formula is

$$Var(b_{0|X=0,Z,W}) = Var(b_0) + Z^2Var(b_2) + W^2Var(b_3) + Z^2W^2Var(b_6) + 2ZCov(b_0, b_2) + 2WCov(b_0, b_3) + 2ZWCov(b_0, b_6)$$

where  $Var$  denotes the variance of a parameter estimate and  $Cov$  denotes the covariance of two parameter estimates.

The variance of the simple slope formula is

$$Var(b_{X|Z,W}) = Var(b_1) + Z^2Var(b_4) + W^2Var(b_5) + Z^2W^2Var(b_7) + 2ZCov(b_1, b_4) + 2WCov(b_1, b_5) + 2ZWCov(b_1, b_7)$$

Wald  $z$  statistic is used for test statistic (even for objects of class `lavaan.mi`).



**Value**

A list with two elements:

1. `SimpleIntercept`: The intercepts given each combination of moderator values. This element will be shown only if the factor intercept is estimated (e.g., not fixed at 0).
2. `SimpleSlope`: The slopes given each combination of moderator values.

In each element, the first column represents values of the first moderator specified in the `valProbe1` argument. The second column represents values of the second moderator specified in the `valProbe2` argument. The third column is the simple intercept or simple slope. The fourth column is the standard error of the simple intercept or simple slope. The fifth column is the Wald ( $z$ ) statistic. The sixth column is the  $p$  value testing whether the simple intercepts or slopes are different from 0.

**Author(s)**

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Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

Aiken, L. S., & West, S. G. (1991). *Multiple regression: Testing and interpreting interactions*. Newbury Park, CA: Sage.

Marsh, H. W., Wen, Z., & Hau, K. T. (2004). Structural equation models of latent interactions: Evaluation of alternative estimation strategies and indicator construction. *Psychological Methods*, 9(3), 275–300. doi:10.1037/1082-989X.9.3.275

**See Also**

- [indProd](#) For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- [probe2WayMC](#) For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe2WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [probe3WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [plotProbe](#) Plot the simple intercepts and slopes of the latent interaction.

**Examples**

```
library(lavaan)

dat3wayMC <- indProd(dat3way, 1:3, 4:6, 7:9)

model3 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
```

```

f3 =~ x7 + x8 + x9
f12 =~ x1.x4 + x2.x5 + x3.x6
f13 =~ x1.x7 + x2.x8 + x3.x9
f23 =~ x4.x7 + x5.x8 + x6.x9
f123 =~ x1.x4.x7 + x2.x5.x8 + x3.x6.x9
f4 =~ x10 + x11 + x12
f4 ~ f1 + f2 + f3 + f12 + f13 + f23 + f123
f1 ~~ 0*f12
f1 ~~ 0*f13
f1 ~~ 0*f123
f2 ~~ 0*f12
f2 ~~ 0*f23
f2 ~~ 0*f123
f3 ~~ 0*f13
f3 ~~ 0*f23
f3 ~~ 0*f123
f12 ~~ 0*f123
f13 ~~ 0*f123
f23 ~~ 0*f123
x1 ~ 0*1
x4 ~ 0*1
x7 ~ 0*1
x10 ~ 0*1
x1.x4 ~ 0*1
x1.x7 ~ 0*1
x4.x7 ~ 0*1
x1.x4.x7 ~ 0*1
f1 ~ NA*1
f2 ~ NA*1
f3 ~ NA*1
f12 ~ NA*1
f13 ~ NA*1
f23 ~ NA*1
f123 ~ NA*1
f4 ~ NA*1
"

fitMC3way <- sem(model3, data = dat3wayMC, std.lv = FALSE,
                 meanstructure = TRUE)
summary(fitMC3way)

probe3WayMC(fitMC3way, nameX = c("f1" ,"f2" ,"f3",
                                "f12","f13","f23", # the order matters!
                                "f123"),           # 3-way interaction
            nameY = "f4", modVar = c("f1", "f2"),
            valProbe1 = c(-1, 0, 1), valProbe2 = c(-1, 0, 1))

```

**Description**

Probing interaction for simple intercept and simple slope for the residual-centered latent three-way interaction (Geldhof et al., 2013)

**Usage**

```
probe3WayRC(fit, nameX, nameY, modVar, valProbe1, valProbe2, group,
  omitimps = c("no.conv", "no.se"))
```

**Arguments**

<code>fit</code>	A fitted <code>lavaan</code> or <code>lavaan.mi</code> object with a latent 2-way interaction.
<code>nameX</code>	character vector of all 7 factor names used as the predictors. The 3 lower-order factors must be listed first, followed by the 3 second-order factors (specifically, the 4th element must be the interaction between the factors listed first and second, the 5th element must be the interaction between the factors listed first and third, and the 6th element must be the interaction between the factors listed second and third). The final name will be the factor representing the 3-way interaction.
<code>nameY</code>	The name of factor that is used as the dependent variable.
<code>modVar</code>	The name of two factors that are used as the moderators. The effect of the independent factor on each combination of the moderator variable values will be probed.
<code>valProbe1</code>	The values of the first moderator that will be used to probe the effect of the independent factor.
<code>valProbe2</code>	The values of the second moderator that will be used to probe the effect of the independent factor.
<code>group</code>	In multigroup models, the label of the group for which the results will be returned. Must correspond to one of <code>lavInspect(fit, "group.label")</code> .
<code>omitimps</code>	character vector specifying criteria for omitting imputations from pooled results. Ignored unless <code>fit</code> is of class <code>lavaan.mi</code> . Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

**Details**

Before using this function, researchers need to make the products of the indicators between the first-order factors and residualize the products by the original indicators (Lance, 1988; Little, Bovaird,

& Widaman, 2006). The process can be automated by the `indProd` function. Note that the indicator products can be made for all possible combination or matched-pair approach (Marsh et al., 2004). Next, the hypothesized model with the regression with latent interaction will be used to fit all original indicators and the product terms (Geldhof et al., 2013). To use this function the model must be fit with a mean structure. See the example for how to fit the product term below. Once the lavaan result is obtained, this function will be used to probe the interaction.

The probing process on residual-centered latent interaction is based on transforming the residual-centered result into the no-centered result. See Geldhof et al. (2013) for further details. Note that this approach based on a strong assumption that the first-order latent variables are normally distributed. The probing process is applied after the no-centered result (parameter estimates and their covariance matrix among parameter estimates) has been computed. See the `probe3WayMC` for further details.

### Value

A list with two elements:

1. `SimpleIntercept`: The intercepts given each value of the moderator. This element will be shown only if the factor intercept is estimated (e.g., not fixed as 0).
2. `SimpleSlope`: The slopes given each value of the moderator.

In each element, the first column represents values of the first moderator specified in the `valProbe1` argument. The second column represents values of the second moderator specified in the `valProbe2` argument. The third column is the simple intercept or simple slope. The fourth column is the *SE* of the simple intercept or simple slope. The fifth column is the Wald (*z*) statistic. The sixth column is the *p* value testing whether the simple intercepts or slopes are different from 0.

### Author(s)

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Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### References

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- Pornprasertmanit, S., Schoemann, A. M., Geldhof, G. J., & Little, T. D. (submitted). *Probing latent interaction estimated with a residual centering approach.*

**See Also**

- [indProd](#) For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.
- [probe2WayMC](#) For probing the two-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe3WayMC](#) For probing the three-way latent interaction when the results are obtained from mean-centering, or double-mean centering
- [probe2WayRC](#) For probing the two-way latent interaction when the results are obtained from residual-centering approach.
- [plotProbe](#) Plot the simple intercepts and slopes of the latent interaction.

**Examples**

```

library(lavaan)

dat3wayRC <- orthogonalize(dat3way, 1:3, 4:6, 7:9)

model3 <- "
f1 =~ x1 + x2 + x3
f2 =~ x4 + x5 + x6
f3 =~ x7 + x8 + x9
f12 =~ x1.x4 + x2.x5 + x3.x6
f13 =~ x1.x7 + x2.x8 + x3.x9
f23 =~ x4.x7 + x5.x8 + x6.x9
f123 =~ x1.x4.x7 + x2.x5.x8 + x3.x6.x9
f4 =~ x10 + x11 + x12
f4 ~ f1 + f2 + f3 + f12 + f13 + f23 + f123
f1 ~~ 0*f12
f1 ~~ 0*f13
f1 ~~ 0*f123
f2 ~~ 0*f12
f2 ~~ 0*f23
f2 ~~ 0*f123
f3 ~~ 0*f13
f3 ~~ 0*f23
f3 ~~ 0*f123
f12 ~~ 0*f123
f13 ~~ 0*f123
f23 ~~ 0*f123
x1 ~ 0*x1
x4 ~ 0*x1
x7 ~ 0*x1
x10 ~ 0*x1
x1.x4 ~ 0*x1
x1.x7 ~ 0*x1
x4.x7 ~ 0*x1
x1.x4.x7 ~ 0*x1
f1 ~ NA*x1
f2 ~ NA*x1

```

```

f3 ~ NA*1
f12 ~ NA*1
f13 ~ NA*1
f23 ~ NA*1
f123 ~ NA*1
f4 ~ NA*1
"

fitRC3way <- sem(model3, data = dat3wayRC, std.lv = FALSE,
                meanstructure = TRUE)
summary(fitRC3way)

probe3WayMC(fitRC3way, nameX = c("f1" , "f2" , "f3",
                                "f12", "f13", "f23", # the order matters!
                                "f123"),           # 3-way interaction
            nameY = "f4", modVar = c("f1", "f2"),
            valProbe1 = c(-1, 0, 1), valProbe2 = c(-1, 0, 1))

```

---

quark

*Quark*


---

### Description

The quark function provides researchers with the ability to calculate and include component scores calculated by taking into account the variance in the original dataset and all of the interaction and polynomial effects of the data in the dataset.

### Usage

```
quark(data, id, order = 1, silent = FALSE, ...)
```

### Arguments

data	The data frame is a required component for quark. In order for quark to process a data frame, it must not contain any factors or text-based variables. All variables must be in numeric format. Identifiers and dates can be left in the data; however, they will need to be identified under the id argument.
id	Identifiers and dates within the dataset will need to be acknowledged as quark cannot process these. By acknowledging the identifiers and dates as a vector of column numbers or variable names, quark will remove them from the data temporarily to complete its main processes. Among many potential issues of not acknowledging identifiers and dates are issues involved with imputation, product and polynomial effects, and principal component analysis.
order	Order is an optional argument provided by quark that can be used when the imputation procedures in mice fail. Under some circumstances, mice cannot calculate missing values due to issues with extreme missingness. Should an error present itself stating a failure due to not having any columns selected, set

	the argument <code>order = 2</code> in order to reorder the imputation method procedure. Otherwise, use the default <code>order = 1</code> .
<code>silent</code>	If <code>FALSE</code> , the details of the quark process are printed.
<code>...</code>	additional arguments to pass to <code>mice</code> .

### Details

The `quark` function calculates these component scores by first filling in the data via means of multiple imputation methods and then expanding the dataset by aggregating the non-overlapping interaction effects between variables by calculating the mean of the interactions and polynomial effects. The multiple imputation methods include one of iterative sampling and group mean substitution and multiple imputation using a polytomous regression algorithm (`mice`). During the expansion process, the dataset is expanded to three times its normal size (in width). The first third of the dataset contains all of the original data post imputation, the second third contains the means of the polynomial effects (squares and cubes), and the final third contains the means of the non-overlapping interaction effects. A full principal component analysis is conducted and the individual components are retained. The subsequent `combinequark` function provides researchers the control in determining how many components to extract and retain. The function returns the dataset as submitted (with missing values) and the component scores as requested for a more accurate multiple imputation in subsequent steps.

### Value

The output value from using the `quark` function is a list. It will return a list with 7 components.

ID Columns	Is a vector of the identifier columns entered when running <code>quark</code> .
ID Variables	Is a subset of the dataset that contains the identifiers as acknowledged when running <code>quark</code> .
Used Data	Is a matrix / dataframe of the data provided by user as the basis for <code>quark</code> to process.
Imputed Data	Is a matrix / dataframe of the data after the multiple method imputation process.
Big Matrix	Is the expanded product and polynomial matrix.
Principal Components	Is the entire dataframe of principal components for the dataset. This dataset will have the same number of rows of the big matrix, but will have 1 less column (as is the case with principal component analyses).
Percent Variance Explained	Is a vector of the percent variance explained with each column of principal components.

### Author(s)

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Terrence D. Jorgensen (University of Amsterdam)

The PCA code is copied and modified from the `FactoMineR` package.

## References

Howard, W. J., Rhemtulla, M., & Little, T. D. (2015). Using Principal Components as Auxiliary Variables in Missing Data Estimation. *Multivariate Behavioral Research*, *50*(3), 285–299. doi:10.1080/00273171.2014.999267

## See Also

[combinequark](#)

## Examples

```
set.seed(123321)

dat <- HolzingerSwineford1939[,7:15]
misspat <- matrix(runif(nrow(dat) * 9) < 0.3, nrow(dat))
dat[misspat] <- NA
dat <- cbind(HolzingerSwineford1939[,1:3], dat)
## Not run:
quark.list <- quark(data = dat, id = c(1, 2))

final.data <- combinequark(quark = quark.list, percent = 80)

## Example to rerun quark after imputation failure:
quark.list <- quark(data = dat, id = c(1, 2), order = 2)

## End(Not run)
```

---

reliability

*Calculate reliability values of factors*

---

## Description

Calculate reliability values of factors by coefficient omega

## Usage

```
reliability(object, return.total = FALSE, dropSingle = TRUE,
  omitimps = c("no.conv", "no.se"))
```

## Arguments

object	A <a href="#">lavaan</a> or <a href="#">lavaan.mi</a> object, expected to contain only exogenous common factors (i.e., a CFA model).
return.total	logical indicating whether to return a final column containing the reliability of a composite of all items. Ignored in 1-factor models, and should only be set TRUE if all factors represent scale dimensions that could nonetheless be collapsed to a single scale composite (scale sum or scale mean).



dropSingle	logical indicating whether to exclude factors defined by a single indicator from the returned results. If TRUE (default), single indicators will still be included in the total column when return.total = TRUE.
omit.imps	character vector specifying criteria for omitting imputations from pooled results. Can include any of c("no.conv", "no.se", "no.npd"), the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

### Details

The coefficient alpha (Cronbach, 1951) can be calculated by

$$\alpha = \frac{k}{k-1} \left[ 1 - \frac{\sum_{i=1}^k \sigma_{ii}}{\sum_{i=1}^k \sigma_{ii} + 2 \sum_{i<j} \sigma_{ij}} \right],$$

where  $k$  is the number of items in a factor,  $\sigma_{ii}$  is the item  $i$  observed variances,  $\sigma_{ij}$  is the observed covariance of items  $i$  and  $j$ .

The coefficient omega (Bollen, 1980; see also Raykov, 2001) can be calculated by

$$\omega_1 = \frac{\left( \sum_{i=1}^k \lambda_i \right)^2 \text{Var}(\psi)}{\left( \sum_{i=1}^k \lambda_i \right)^2 \text{Var}(\psi) + \sum_{i=1}^k \theta_{ii} + 2 \sum_{i<j} \theta_{ij}},$$

where  $\lambda_i$  is the factor loading of item  $i$ ,  $\psi$  is the factor variance,  $\theta_{ii}$  is the variance of measurement errors of item  $i$ , and  $\theta_{ij}$  is the covariance of measurement errors from item  $i$  and  $j$ .

The second coefficient omega (Bentler, 1972, 2009) can be calculated by

$$\omega_2 = \frac{\left( \sum_{i=1}^k \lambda_i \right)^2 \text{Var}(\psi)}{\mathbf{1}' \hat{\Sigma} \mathbf{1}},$$

where  $\hat{\Sigma}$  is the model-implied covariance matrix, and  $\mathbf{1}$  is the  $k$ -dimensional vector of 1. The first and the second coefficients omega will have the same value when the model has simple structure, but different values when there are (for example) cross-loadings or method factors. The first coefficient omega can be viewed as the reliability controlling for the other factors (like  $\eta_p^2$  partial in ANOVA). The second coefficient omega can be viewed as the unconditional reliability (like  $\eta^2$  in ANOVA).

The third coefficient omega (McDonald, 1999), which is sometimes referred to hierarchical omega, can be calculated by

$$\omega_3 = \frac{\left(\sum_{i=1}^k \lambda_i\right)^2 \text{Var}(\psi)}{\mathbf{1}'\Sigma\mathbf{1}},$$

where  $\Sigma$  is the observed covariance matrix. If the model fits the data well, the third coefficient omega will be similar to the  $\omega_2$ . Note that if there is a directional effect in the model, all coefficients omega will use the total factor variances, which is calculated by `lavInspect(object, "cov.lv")`.

In conclusion,  $\omega_1$ ,  $\omega_2$ , and  $\omega_3$  are different in the denominator. The denominator of the first formula assumes that a model is congeneric factor model where measurement errors are not correlated. The second formula accounts for correlated measurement errors. However, these two formulas assume that the model-implied covariance matrix explains item relationships perfectly. The residuals are subject to sampling error. The third formula use observed covariance matrix instead of model-implied covariance matrix to calculate the observed total variance. This formula is the most conservative method in calculating coefficient omega.

The average variance extracted (AVE) can be calculated by

$$AVE = \frac{\mathbf{1}'\text{diag}(\Lambda\Psi\Lambda')\mathbf{1}}{\mathbf{1}'\text{diag}(\hat{\Sigma})\mathbf{1}},$$

Note that this formula is modified from Fornell & Larcker (1981) in the case that factor variances are not 1. The proposed formula from Fornell & Larcker (1981) assumes that the factor variances are 1. Note that AVE will not be provided for factors consisting of items with dual loadings. AVE is the property of items but not the property of factors.

Regarding categorical indicators, coefficient alpha and AVE are calculated based on polychoric correlations. The coefficient alpha from this function may be not the same as the standard alpha calculation for categorical items. Researchers may check the `alpha` function in the `psych` package for the standard coefficient alpha calculation.

Item thresholds are not accounted for. Coefficient omega for categorical items, however, is calculated by accounting for both item covariances and item thresholds using Green and Yang's (2009, formula 21) approach. Three types of coefficient omega indicate different methods to calculate item total variances. The original formula from Green and Yang is equivalent to  $\omega_3$  in this function. Green and Yang did not propose a method for calculating reliability with a mixture of categorical and continuous indicators, and we are currently unaware of an appropriate method. Therefore, when `reliability` detects both categorical and continuous indicators in the model, an error is returned. If the categorical indicators load on a different factor(s) than continuous indicators, then reliability can be calculated separately for those scales by fitting separate models and submitting each to the `reliability` function.

### Value

Reliability values (coefficient alpha, coefficients omega, average variance extracted) of each factor in each group. If there are multiple factors, a total column can optionally be included.

### Author(s)

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## References

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## See Also

[reliabilityL2](#) for reliability value of a desired second-order factor, [maximalRelia](#) for the maximal reliability of weighted composite

## Examples

```
library(lavaan)

HS.model <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed  =~ x7 + x8 + x9 '

fit <- cfa(HS.model, data = HolzingerSwineford1939)
reliability(fit)
reliability(fit, return.total = TRUE)
```

---

reliabilityL2                      *Calculate the reliability values of a second-order factor*

---

### Description

Calculate the reliability values (coefficient omega) of a second-order factor

### Usage

```
reliabilityL2(object, secondFactor, omit.imps = c("no.conv", "no.se"))
```

### Arguments

object	A <a href="#">lavaan</a> or <a href="#">lavaan.mi</a> object, expected to contain a least one exogenous higher-order common factor.
secondFactor	The name of a single second-order factor in the model fitted in object. The function must be called multiple times to estimate reliability for each higher-order factor.
omit.imps	character vector specifying criteria for omitting imputations from pooled results. Can include any of <code>c("no.conv", "no.se", "no.npd")</code> , the first 2 of which are the default setting, which excludes any imputations that did not converge or for which standard errors could not be computed. The last option ("no.npd") would exclude any imputations which yielded a nonpositive definite covariance matrix for observed or latent variables, which would include any "improper solutions" such as Heywood cases. NPD solutions are not excluded by default because they are likely to occur due to sampling error, especially in small samples. However, gross model misspecification could also cause NPD solutions, users can compare pooled results with and without this setting as a sensitivity analysis to see whether some imputations warrant further investigation.

### Details

The first formula of the coefficient omega (in the [reliability](#)) will be mainly used in the calculation. The model-implied covariance matrix of a second-order factor model can be separated into three sources: the second-order common-factor variance, the residual variance of the first-order common factors (i.e., not accounted for by the second-order factor), and the measurement error of observed indicators:

$$\hat{\Sigma} = \Lambda \mathbf{B} \Phi_2 \mathbf{B}' \Lambda' + \Lambda \Psi_u \Lambda' + \Theta,$$

where  $\hat{\Sigma}$  is the model-implied covariance matrix,  $\Lambda$  contains first-order factor loadings,  $\mathbf{B}$  contains second-order factor loadings,  $\Phi_2$  is the covariance matrix of the second-order factor(s),  $\Psi_u$  is the covariance matrix of residuals from first-order factors, and  $\Theta$  is the covariance matrix of the measurement errors from observed indicators. Thus, we can calculate the proportion of variance of a composite score calculated from the observed indicators (e.g., a total score or scale mean) that is attributable to the second-order factor, i.e. coefficient omega at Level 1:

$$\omega_{L1} = \frac{\mathbf{1}'\Lambda\mathbf{B}\Phi_2\mathbf{B}'\Lambda'\mathbf{1}}{\mathbf{1}'\Lambda\mathbf{B}\Phi_2\mathbf{B}'\Lambda'\mathbf{1} + \mathbf{1}'\Lambda\Psi_u\Lambda'\mathbf{1} + \mathbf{1}'\Theta\mathbf{1}},$$

where  $\mathbf{1}$  is the  $k$ -dimensional vector of 1 and  $k$  is the number of observed variables.

The model-implied covariance matrix among first-order factors ( $\Phi_1$ ) can be calculated as:

$$\Phi_1 = \mathbf{B}\Phi_2\mathbf{B}' + \Psi_u,$$

Thus, the proportion of variance among first-order common factors that is attributable to the second-order factor (i.e., coefficient omega at Level 2) can be calculated as:

$$\omega_{L2} = \frac{\mathbf{1}'_F\mathbf{B}\Phi_2\mathbf{B}'\mathbf{1}_F}{\mathbf{1}'_F\mathbf{B}\Phi_2\mathbf{B}'\mathbf{1}_F + \mathbf{1}'_F\Psi_u\mathbf{1}_F},$$

where  $\mathbf{1}_F$  is the  $F$ -dimensional vector of 1 and  $F$  is the number of first-order factors. This Level-2 omega can be interpreted as an estimate of the reliability of a hypothetical composite calculated from error-free observable variables representing the first-order common factors. This might only be meaningful as a thought experiment.

An additional thought experiment is possible: If the observed indicators contained only the second-order common-factor variance and unsystematic measurement error, then there would be no first-order common factors because their unique variances would be excluded from the observed measures. An estimate of this hypothetical composite reliability can be calculated as the partial coefficient omega at Level 1, or the proportion of observed variance explained by the second-order factor after partialling out the uniqueness from the first-order factors:

$$\omega_{L1} = \frac{\mathbf{1}'\Lambda\mathbf{B}\Phi_2\mathbf{B}'\Lambda'\mathbf{1}}{\mathbf{1}'\Lambda\mathbf{B}\Phi_2\mathbf{B}'\Lambda'\mathbf{1} + \mathbf{1}'\Theta\mathbf{1}},$$

Note that if the second-order factor has a direct factor loading on some observed variables, the observed variables will be counted as first-order factors, which might not be desirable.

### Value

Reliability values at Levels 1 and 2 of the second-order factor, as well as the partial reliability value at Level 1

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

### See Also

[reliability](#) for the reliability of the first-order factors.

## Examples

```
library(lavaan)

HS.model3 <- ' visual =~ x1 + x2 + x3
             textual =~ x4 + x5 + x6
             speed  =~ x7 + x8 + x9
             higher =~ visual + textual + speed'

fit6 <- cfa(HS.model3, data = HolzingerSwineford1939)
reliability(fit6) # Should provide a warning for the endogenous variables
reliabilityL2(fit6, "higher")
```

---

residualCovariate	<i>Residual-center all target indicators by covariates</i>
-------------------	--

---

## Description

This function will regress target variables on the covariate and replace the target variables by the residual of the regression analysis. This procedure is useful to control the covariate from the analysis model (Geldhof, Pornprasertmanit, Schoemann, & Little, 2013).

## Usage

```
residualCovariate(data, targetVar, covVar)
```

## Arguments

data	The desired data to be transformed.
targetVar	Variable names or the position of indicators that users wish to be residual centered (as dependent variables)
covVar	Covariate names or the position of the covariates using for residual centering (as independent variables) onto target variables

## Value

The data that the target variables replaced by the residuals

## Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

## References

Geldhof, G. J., Pornprasertmanit, S., Schoemann, A. M., & Little, T. D. (2013). Orthogonalizing through residual centering: Extended applications and caveats. *Educational and Psychological Measurement, 73*(1), 27–46. doi:10.1177/0013164412445473

**See Also**

[indProd](#) For creating the indicator products with no centering, mean centering, double-mean centering, or residual centering.

**Examples**

```
dat <- residualCovariate(attitude, 2:7, 1)
```

---

runMI

*Fit a lavaan Model to Multiple Imputed Data Sets*


---

**Description**

This function fits a lavaan model to a list of imputed data sets, and can also implement multiple imputation for a single data.frame with missing observations, using either the Amelia package or the mice package.

**Usage**

```
runMI(model, data, fun = "lavaan", ..., m, miArgs = list(),
      miPackage = "Amelia", seed = 12345)
```

```
lavaan.mi(model, data, ..., m, miArgs = list(), miPackage = "Amelia",
          seed = 12345)
```

```
cfa.mi(model, data, ..., m, miArgs = list(), miPackage = "Amelia",
       seed = 12345)
```

```
sem.mi(model, data, ..., m, miArgs = list(), miPackage = "Amelia",
       seed = 12345)
```

```
growth.mi(model, data, ..., m, miArgs = list(), miPackage = "Amelia",
          seed = 12345)
```

**Arguments**

model	The analysis model can be specified using lavaan <a href="#">model.syntax</a> or a parameter table (as returned by <a href="#">parTable</a> ).
data	A data.frame with missing observations, or a list of imputed data sets (if data are imputed already). If runMI has already been called, then imputed data sets are stored in the @DataList slot, so data can also be a lavaan.mi object from which the same imputed data will be used for additional analyses.
fun	character. Name of a specific lavaan function used to fit model to data (i.e., "lavaan", "cfa", "sem", or "growth"). Only required for runMI.

...	additional arguments to pass to <code>lavaan</code> or <code>lavaanList</code> . See also <code>lavOptions</code> . Note that <code>lavaanList</code> provides parallel computing options, as well as a <code>FUN</code> argument so the user can extract custom output after the model is fitted to each imputed data set (see <b>Examples</b> ). TIP: If a custom <code>FUN</code> is used <i>and</i> <code>parallel = "snow"</code> is requested, the user-supplied function should explicitly call <code>library</code> or use <code>::</code> for any functions not part of the base distribution.
<code>m</code>	integer. Request the number of imputations. Ignored if data is already a list of imputed data sets or a <code>lavaan.mi</code> object.
<code>miArgs</code>	Addition arguments for the multiple-imputation function ( <code>miPackage</code> ). The arguments should be put in a list (see example below). Ignored if data is already a list of imputed data sets or a <code>lavaan.mi</code> object.
<code>miPackage</code>	Package to be used for imputation. Currently these functions only support "Amelia" or "mice" for imputation. Ignored if data is already a list of imputed data sets or a <code>lavaan.mi</code> object.
<code>seed</code>	integer. Random number seed to be set before imputing the data. Ignored if data is already a list of imputed data sets or a <code>lavaan.mi</code> object.

**Value**

A `lavaan.mi` object

**Author(s)**

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

**References**

- Enders, C. K. (2010). *Applied missing data analysis*. New York, NY: Guilford.
- Rubin, D. B. (1987). *Multiple imputation for nonresponse in surveys*. New York, NY: Wiley.

**Examples**

```
## Not run:
## impose missing data for example
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]

set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## specify CFA model from lavaan's ?cfa help page
HS.model <- '
  visual =~ x1 + x2 + x3
  textual =~ x4 + x5 + x6
  speed   =~ x7 + x8 + x9
'
```

## impute data within runMI...



```

out1 <- cfa.mi(HS.model, data = HSMiss, m = 20, seed = 12345,
              miArgs = list(noms = "school"))

## ... or impute missing data first
library(Amelia)
set.seed(12345)
HS.amelia <- amelia(HSMiss, m = 20, noms = "school", p2s = FALSE)
imps <- HS.amelia$imputations
out2 <- cfa.mi(HS.model, data = imps)

## same results (using the same seed results in the same imputations)
cbind(impute.within = coef(out1), impute.first = coef(out2))

summary(out1, fit.measures = TRUE)
summary(out1, ci = FALSE, fmi = TRUE, output = "data.frame")
summary(out1, ci = FALSE, stand = TRUE, rsq = TRUE)

## model fit. D3 includes information criteria
anova(out1)
## equivalently:
lavTestLRT.mi(out1)
## request D2
anova(out1, test = "D2")
## request fit indices
fitMeasures(out1)

## fit multigroup model without invariance constraints
mgfit.config <- cfa.mi(HS.model, data = imps, estimator = "mlm",
                     group = "school")
## add invariance constraints, and use previous fit as "data"
mgfit.metric <- cfa.mi(HS.model, data = mgfit.config, estimator = "mlm",
                     group = "school", group.equal = "loadings")
mgfit.scalar <- cfa.mi(HS.model, data = mgfit.config, estimator = "mlm",
                     group = "school",
                     group.equal = c("loadings","intercepts"))

## compare fit of 2 models to test metric invariance
## (scaled likelihood ratio test)
lavTestLRT.mi(mgfit.metric, h1 = mgfit.config)
## To compare multiple models, you must use anova()
anova(mgfit.config, mgfit.metric, mgfit.scalar)
## or compareFit(), which also includes fit indices for comparison
## (optional: name the models)
compareFit(config = mgfit.config, metric = mgfit.metric,
           scalar = mgfit.scalar,
           argsLRT = list(test = "D2", method = "satorra.bentler.2010"))

## correlation residuals to investigate local misfit
resid(mgfit.scalar, type = "cor.bentler")
## modification indices for fixed parameters, to investigate local misfit
modindices.mi(mgfit.scalar)
## or lavTestScore.mi for modification indices about equality constraints

```

```

lavTestScore.mi(mgfit.scalar)

## Wald test of whether latent means are == (fix 3 means to zero in group 2)
eq.means <- ' .p70. == 0
             .p71. == 0
             .p72. == 0 '
lavTestWald.mi(mgfit.scalar, constraints = eq.means)

## ordered-categorical data
data(datCat)
lapply(datCat, class) # indicators already stored as ordinal
## impose missing values
set.seed(123)
for (i in 1:8) datCat[sample(1:nrow(datCat), size = .1*nrow(datCat)), i] <- NA

## impute ordinal missing data using mice package
library(mice)
set.seed(456)
miceImps <- mice(datCat)
## save imputations in a list of data.frames
implist <- list()
for (i in miceImps$m) implist[[i]] <- complete(miceImps, action = i)

## fit model, save zero-cell tables and obsolete "WRMR" fit indices
catout <- cfa.mi(' f =~ 1*u1 + 1*u2 + 1*u3 + 1*u4 ', data = implist,
                FUN = function(fit) {
                  list(wrmr = lavaan::fitMeasures(fit, "wrmr"),
                       zeroCells = lavaan::lavInspect(fit, "zero.cell.tables"))
                })
summary(catout)
lavTestLRT.mi(catout, test = "D2", pool.robust = TRUE)
fitMeasures(catout, fit.measures = c("rmsea", "srmr", "cfi"),
            test = "D2", pool.robust = TRUE)

## extract custom output
sapply(catout@funList, function(x) x$wrmr) # WRMR for each imputation
catout@funList[[1]]$zeroCells # zero-cell tables for first imputation
catout@funList[[2]]$zeroCells # zero-cell tables for second imputation ...

## End(Not run)

```

## Description

The **semTools** package provides many miscellaneous functions that are useful for statistical analysis involving SEM in R. Many functions extend the functionality of the **lavaan** package. Some sets of functions in **semTools** correspond to the same theme. We call such a collection of functions a *suite*. Our suites include:

- Model Fit Evaluation: [moreFitIndices](#), [nullRMSEA](#), [singleParamTest](#), [miPowerFit](#), and [chisqSmallN](#)
- Measurement Invariance: [measEq.syntax](#), [partialInvariance](#), [partialInvarianceCat](#), and [permuteMeasEq](#)
- Power Analysis: [SSpower](#), [findRMSEApower](#), [plotRMSEApower](#), [plotRMSEAdist](#), [findRMSEAsamplesize](#), [findRMSEApowernested](#), [plotRMSEApowernested](#), and [findRMSEAsamplesizenested](#)
- Missing Data Analysis: [auxiliary](#), [runMI](#), [twostage](#), [fmi](#), [bsBootMiss](#), [quark](#), and [combinequark](#)
- Latent Interactions: [indProd](#), [orthogonalize](#), [probe2WayMC](#), [probe3WayMC](#), [probe2WayRC](#), [probe3WayRC](#), and [plotProbe](#)
- Exploratory Factor Analysis (EFA): [efa.ekc](#), [efaUnrotate](#), [orthRotate](#), [oblqRotate](#), and [funRotate](#)
- Reliability Estimation: [reliability](#), [reliabilityL2](#), and [maximalRelia](#)
- Parceling: [parcelAllocation](#), [PAVranking](#), and [poolMAlloc](#)
- Non-Normality: [skew](#), [kurtosis](#), [mardiaSkew](#), [mardiaKurtosis](#), and [mvrnonnorm](#)

All users of R (or SEM) are invited to submit functions or ideas for functions by contacting the maintainer, Terrence Jorgensen (<TJorgensen314@gmail.com>). Contributors are encouraged to use Roxygen comments to document their contributed code, which is consistent with the rest of **semTools**. Read the vignette from the **roxygen2** package for details: `vignette("rd", package = "roxygen2")`

---

simParcel

*Simulated Data set to Demonstrate Random Allocations of Parcels*

---

## Description

A simulated data set with 2 factors with 9 indicators for each factor

## Usage

```
simParcel
```

## Format

A data.frame with 800 observations of 18 variables.

**f1item1** Item 1 loading on factor 1

**f1item2** Item 2 loading on factor 1

**f1item3** Item 3 loading on factor 1

**f1item4** Item 4 loading on factor 1  
**f1item5** Item 5 loading on factor 1  
**f1item6** Item 6 loading on factor 1  
**f1item7** Item 7 loading on factor 1  
**f1item8** Item 8 loading on factor 1  
**f1item9** Item 9 loading on factor 1  
**f2item1** Item 1 loading on factor 2  
**f2item2** Item 2 loading on factor 2  
**f2item3** Item 3 loading on factor 2  
**f2item4** Item 4 loading on factor 2  
**f2item5** Item 5 loading on factor 2  
**f2item6** Item 6 loading on factor 2  
**f2item7** Item 7 loading on factor 2  
**f2item8** Item 8 loading on factor 2  
**f2item9** Item 9 loading on factor 2

### Source

Data were generated using the simsem package.

### Examples

```
head(simParcel)
```

---

singleParamTest

*Single Parameter Test Divided from Nested Model Comparison*

---

### Description

In comparing two nested models,  $\Delta\chi^2$  test may indicate that two models are different. However, like other omnibus tests, researchers do not know which fixed parameters or constraints make these two models different. This function will help researchers identify the significant parameter.

### Usage

```
singleParamTest(model1, model2, return.fit = FALSE,  
  method = "satorra.bentler.2001")
```

**Arguments**

model1	Model 1.
model2	Model 2. Note that two models must be nested models. Further, the order of parameters in their parameter tables are the same. That is, nested models with different scale identifications may not be able to test by this function.
return.fit	Return the submodels fitted by this function
method	The method used to calculate likelihood ratio test. See <a href="#">lavTestLRT</a> for available options

**Details**

This function first identify the differences between these two models. The model with more free parameters is referred to as parent model and the model with less free parameters is referred to as nested model. Three tests are implemented here:

1. free: The nested model is used as a template. Then, one parameter indicating the differences between two models is free. The new model is compared with the nested model. This process is repeated for all differences between two models.
2. fix: The parent model is used as a template. Then, one parameter indicating the differences between two models is fixed or constrained to be equal to other parameters. The new model is then compared with the parent model. This process is repeated for all differences between two models.
3. mi: No longer available because the test of modification indices is not consistent. For example, if two parameters are equality constrained, the modification index from the first parameter is not equal to the second parameter.

Note that this function does not adjust for the inflated Type I error rate from multiple tests.

**Value**

If `return.fit = FALSE`, the result tables are provided.  $\chi^2$  and  $p$  value are provided for all methods. Note that the  $\chi^2$  is all based on 1 *df*. Expected parameter changes and their standardized forms are also provided.

If `return.fit = TRUE`, a list with two elements are provided. The first element is the tabular result. The second element is the submodels used in the free and fix methods.

**Author(s)**

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

**Examples**

```
library(lavaan)

# Nested model comparison by hand
HS.model1 <- ' visual =~ x1 + x2 + x3
              textual =~ x4 + x5 + x6'
```

```

HS.model2 <- ' visual =~ a*x1 + a*x2 + a*x3
             textual =~ b*x4 + b*x5 + b*x6'

m1 <- cfa(HS.model1, data = HolzingerSwineford1939, std.lv = TRUE,
          estimator = "MLR")
m2 <- cfa(HS.model2, data = HolzingerSwineford1939, std.lv = TRUE,
          estimator = "MLR")
anova(m1, m2)
singleParamTest(m1, m2)

## Nested model comparison from the measurementInvariance function
HW.model <- ' visual =~ x1 + x2 + x3
            textual =~ x4 + x5 + x6
            speed =~ x7 + x8 + x9 '

models <- measurementInvariance(model = HW.model, data = HolzingerSwineford1939,
                               group = "school")
singleParamTest(models[[1]], models[[2]])

## Note that the comparison between weak (Model 2) and scalar invariance
## (Model 3) cannot be done by this function # because the weak invariance
## model fixes factor means as 0 in Group 2 but the strong invariance model
## frees the factor means in Group 2. Users may try to compare
## strong (Model 3) and means invariance models by this function.

```

---

skew

*Finding skewness*


---

### Description

Finding skewness ( $g_1$ ) of an object

### Usage

```
skew(object, population = FALSE)
```

### Arguments

object	A vector used to find a skewness
population	TRUE to compute the parameter formula. FALSE to compute the sample statistic formula.

### Details

The skewness computed is  $g_1$ . The parameter skewness  $\gamma_2$  formula is

$$\gamma_2 = \frac{\mu_3}{\mu_2^{3/2}},$$

where  $\mu_i$  denotes the  $i$  order central moment.

The excessive kurtosis formula for sample statistic  $g_2$  is

$$g_2 = \frac{k_3}{k_2^2},$$

where  $k_i$  are the  $i$  order  $k$ -statistic.

The standard error of the skewness is

$$Var(\hat{g}_2) = \frac{6}{N}$$

where  $N$  is the sample size.

### Value

A value of a skewness with a test statistic if the population is specified as FALSE

### Author(s)

Sunthud Pornprasertmanit (<psunthud@gmail.com>)

### References

Weisstein, Eric W. (n.d.). *Skewness*. Retrived from *MathWorld*—A Wolfram Web Resource: <http://mathworld.wolfram.com/Skewness.html>

### See Also

- [kurtosis](#) Find the univariate excessive kurtosis of a variable
- [mardiaSkew](#) Find Mardia's multivariate skewness of a set of variables
- [mardiaKurtosis](#) Find the Mardia's multivariate kurtosis of a set of variables

### Examples

```
skew(1:5)
```

---

splitSample

*Randomly Split a Data Set into Halves*


---

### Description

This function randomly splits a data set into two halves, and saves the resulting data sets to the same folder as the original.

### Usage

```
splitSample(dataset, path = "default", div = 2, type = "default",
            name = "splitSample")
```

### Arguments

dataset	The original data set to be divided. Can be a file path to a *.csv or *.dat file (headers will automatically be detected) or an R object (matrix or dataframe). (Windows users: file path must be specified using FORWARD SLASHES (/) ONLY.)
path	File path to folder for output data sets. NOT REQUIRED if dataset is a filename. Specify ONLY if dataset is an R object, or desired output folder is not that of original data set. If path is specified as "object", output data sets will be returned as a list, and not saved to hard drive.
div	Number of output data sets. NOT REQUIRED if default, 2 halves.
type	Output file format ("dat" or "csv"). NOT REQUIRED unless desired output formatting differs from that of input, or dataset is an R object and csv formatting is desired.
name	Output file name. NOT REQUIRED unless desired output name differs from that of input, or input dataset is an R object. (If input is an R object and name is not specified, name will be "splitSample".)

### Details

This function randomly orders the rows of a data set, divides the data set into two halves, and saves the halves to the same folder as the original data set, preserving the original formatting. Data set type (\*.csv or \*.dat) and formatting (headers) are automatically detected, and output data sets will preserve input type and formatting unless specified otherwise. Input can be in the form of a file path (\*.dat or \*.csv), or an R object (matrix or dataframe). If input is an R object and path is default, output data sets will be returned as a list object.

### Value

If path = "object", list of output data sets. Otherwise, output will saved to hard drive in the same format as input.



**Author(s)**

Corbin Quick (University of Michigan; <corbinq@umich.edu>)

**Examples**

```
#### Input is .dat file
#splitSample("C:/Users/Default/Desktop/MYDATA.dat")
#### Output saved to "C:/Users/Default/Desktop/" in .dat format
#### Names are "MYDATA_s1.dat" and "MYDATA_s2.dat"

#### Input is R object
## Split C02 dataset from the datasets package
library(datasets)
splitMyData <- splitSample(CO2, path = "object")
summary(splitMyData[[1]])
summary(splitMyData[[2]])
#### Output object splitMyData becomes list of output data sets

#### Input is .dat file in "C:/" folder
#splitSample("C:/testdata.dat", path = "C:/Users/Default/Desktop/", type = "csv")
#### Output saved to "C:/Users/Default/Desktop/" in *.csv format
#### Names are "testdata_s1.csv" and "testdata_s2.csv"

#### Input is R object
#splitSample(myData, path = "C:/Users/Default/Desktop/", name = "splitdata")
#### Output saved to "C:/Users/Default/Desktop/" in *.dat format
#### Names are "splitdata_s1.dat" and "splitdata_s2.dat"
```

---

SSpower

*Power for model parameters*


---

**Description**

Apply Satorra & Saris (1985) method for chi-squared power analysis.

**Usage**

```
SSpower(powerModel, n, nparam, popModel, mu, Sigma, fun = "cfa",
        alpha = 0.05, ...)
```

**Arguments**

powerModel	lavaan <a href="#">model.syntax</a> for the model to be analyzed. This syntax should constrain at least one nonzero parameter to 0 (or another number).
n	integer. Sample size used in power calculation, or a vector of sample sizes if analyzing a multigroup model. If <code>length(n) &lt; length(Sigma)</code> when Sigma is a list, n will be recycled.

nparam	integer. Number of invalid constraints in powerModel.
popModel	lavaan <a href="#">model.syntax</a> specifying the data-generating model. This syntax should specify values for all nonzero parameters in the model. If <code>length(n) &gt; 1</code> , the same population values will be used for each group. Different population values per group can only be specified by utilizing Sigma (and mu).
mu	numeric or list. For a single-group model, a vector of population means. For a multigroup model, a list of vectors (one per group). If mu and popModel are missing, mean structure will be excluded from the analysis.
Sigma	matrix or list. For a single-group model, a population covariance matrix. For a multigroup model, a list of matrices (one per group). If missing, popModel will be used to generate a model-implied Sigma.
fun	character. Name of lavaan function used to fit powerModel (i.e., "cfa", "sem", "growth", or "lavaan").
alpha	Type I error rate used to set a criterion for rejecting H0.
...	additional arguments to pass to <a href="#">lavaan</a> .

### Details

Specify all non-zero parameters in a population model, either by using lavaan syntax (popModel) or by submitting a population covariance matrix (Sigma) and optional mean vector (mu) implied by the population model. Then specify an analysis model that constrains at least one nonzero parameter to an incorrect value. Note the number in the nparam argument.

### Author(s)

Alexander M. Schoemann (East Carolina University; <schoemanna@ecu.edu>)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

### References

Satorra, A., & Saris, W. E. (1985). Power of the likelihood ratio test in covariance structure analysis. *Psychometrika*, 50, 83–90. doi:10.1007/BF02294150

### Examples

```
## Specify population values. Note every parameter has a fixed value.
modelP <- '
  f1 =~ .7*v1 + .7*v2 + .7*v3 + .7*v4
  f2 =~ .7*v5 + .7*v6 + .7*v7 + .7*v8
  f1 =~ .3*f2
  f1 =~ 1*f1
  f2 =~ 1*f2
  V1 =~ .51*v1
  V2 =~ .51*v2
  V3 =~ .51*v3
  V4 =~ .51*v4
  V5 =~ .51*v5
  V6 =~ .51*v6
```

```

V7 ~~ .51*V7
V8 ~~ .51*V8
,

## Specify analysis model. Note parameter of interest f1~~f2 is fixed to 0.
modelA <- '
  f1 =~ V1 + V2 + V3 + V4
  f2 =~ V5 + V6 + V7 + V8
  f1 ~~ 0*f2
,

## Calculate power
SSpower(powerModel = modelA, popModel = modelP, n = 150, nparam = 1,
        std.lv = TRUE)

## Get power for a range of sample sizes

Ns <- seq(100, 500, 40)
Power <- rep(NA, length(Ns))
for(i in 1:length(Ns)) {
  Power[i] <- SSpower(powerModel = modelA, popModel = modelP,
                    n = Ns[i], nparam = 1, std.lv = TRUE)
}
plot(x = Ns, y = Power, type = "l", xlab = "Sample Size")

## Specify second population to calculate power for multigroup model

popMoments1 <- fitted(cfa(modelP))
modelP2 <- '
  f1 =~ .7*V1 + .7*V2 + .7*V3 + .7*V4
  f2 =~ .7*V5 + .7*V6 + .7*V7 + .7*V8
  f1 ~~ .5*f2    ## higher correlation in Group 2
  f1 ~~ 1*f1
  f2 ~~ 1*f2
  V1 ~~ .51*V1
  V2 ~~ .51*V2
  V3 ~~ .51*V3
  V4 ~~ .51*V4
  V5 ~~ .51*V5
  V6 ~~ .51*V6
  V7 ~~ .51*V7
  V8 ~~ .51*V8
,

popMoments2 <- fitted(cfa(modelP2))
modelA2 <- '
  f1 =~ V1 + V2 + V3 + V4
  f2 =~ V5 + V6 + V7 + V8
  f1 ~~ c(0, 0)*f2
,

mu <- list(popMoments1$mean, popMoments2$mean) # ignored if NULL
Sigma <- list(popMoments1$cov, popMoments2$cov)
SSpower(powerModel = modelA2, mu = mu, Sigma = Sigma,
        n = c(60, 65), nparam = 2)

```

---

tukeySEM	<i>Tukey's WSD post-hoc test of means for unequal variance and sample size</i>
----------	--

---

### Description

This function computes Tukey's WSD post hoc test of means when variances and sample sizes are not equal across groups. It can be used as a post hoc test when comparing latent means in multiple group SEM.

### Usage

```
tukeySEM(m1, m2, var1, var2, n1, n2, ng)
```

### Arguments

m1	Mean of group 1.
m2	Mean of group 2.
var1	Variance of group 1.
var2	Variance of group 2.
n1	Sample size of group 1.
n2	Sample size of group 2.
ng	Total number of groups to be compared (i.e., the number of groups compared in the omnibus test).

### Details

After conducting an omnibus test of means across three or more groups, researchers often wish to know which sets of means differ at a particular Type I error rate. Tukey's WSD test holds the error rate stable across multiple comparisons of means. This function implements an adaptation of Tukey's WSD test from Maxwell & Delaney (2004), that allows variances and sample sizes to differ across groups.

### Value

A vector with three elements:

1.  $q$ : The  $q$  statistic
2.  $df$ : The degrees of freedom for the  $q$  statistic
3.  $p$ : A  $p$  value based on the  $q$  statistic,  $df$ , and the total number of groups to be compared

### Author(s)

Alexander M. Schoemann (East Carolina University; <schoemanna@ecu.edu>)

## References

Maxwell, S. E., & Delaney, H. D. (2004). *Designing experiments and analyzing data: A model comparison perspective* (2nd ed.). Mahwah, NJ: Lawrence Erlbaum Associates.

## Examples

```
## For a case where three groups have been compared:
## Group 1: mean = 3.91, var = 0.46, n = 246
## Group 2: mean = 3.96, var = 0.62, n = 465
## Group 3: mean = 2.94, var = 1.07, n = 64

## compare group 1 and group 2
tukeySEM(3.91, 3.96, 0.46, 0.62, 246, 425, 3)

## compare group 1 and group 3
tukeySEM(3.91, 2.94, 0.46, 1.07, 246, 64, 3)

## compare group 2 and group 3
tukeySEM(3.96, 2.94, 0.62, 1.07, 465, 64, 3)
```

---

twostage

*Fit a lavaan model using 2-Stage Maximum Likelihood (TSML) estimation for missing data.*

---

## Description

This function automates 2-Stage Maximum Likelihood (TSML) estimation, optionally with auxiliary variables. Step 1 involves fitting a saturated model to the partially observed data set (to variables in the hypothesized model as well as auxiliary variables related to missingness). Step 2 involves fitting the hypothesized model to the model-implied means and covariance matrix (also called the "EM" means and covariance matrix) as if they were complete data. Step 3 involves correcting the Step-2 standard errors (*SEs*) and chi-squared statistic to account for additional uncertainty due to missing data (using information from Step 1; see References section for sources with formulas).

## Usage

```
twostage(..., aux, fun, baseline.model = NULL)

lavaan.2stage(..., aux = NULL, baseline.model = NULL)

cfa.2stage(..., aux = NULL, baseline.model = NULL)

sem.2stage(..., aux = NULL, baseline.model = NULL)

growth.2stage(..., aux = NULL, baseline.model = NULL)
```

## Arguments

...	Arguments passed to the <code>lavaan</code> function specified in the <code>fun</code> argument. See also <code>lavOptions</code> . At a minimum, the user must supply the first two named arguments to <code>lavaan</code> (i.e., <code>model</code> and <code>data</code> ).
<code>aux</code>	An optional character vector naming auxiliary variable(s) in data
<code>fun</code>	The character string naming the lavaan function used to fit the Step-2 hypothesized model (" <code>cfa</code> ", " <code>sem</code> ", " <code>growth</code> ", or " <code>lavaan</code> ").
<code>baseline.model</code>	An optional character string, specifying the lavaan <code>model.syntax</code> for a user-specified baseline model. Interested users can use the fitted baseline model to calculate incremental fit indices (e.g., CFI and TLI) using the corrected chi-squared values (see the <code>anova</code> method in <code>twostage</code> ). If <code>NULL</code> , the default "independence model" (i.e., freely estimated means and variances, but all covariances constrained to zero) will be specified internally.

## Details

All variables (including auxiliary variables) are treated as endogenous variables in the Step-1 saturated model (`fixed.x = FALSE`), so data are assumed continuous, although not necessarily multivariate normal (dummy-coded auxiliary variables may be included in Step 1, but categorical endogenous variables in the Step-2 hypothesized model are not allowed). To avoid assuming multivariate normality, request `se = "robust.huber.white"`. CAUTION: In addition to setting `fixed.x = FALSE` and `conditional.x = FALSE` in `lavaan`, this function will automatically set `meanstructure = TRUE`, `estimator = "ML"`, `missing = "fiml"`, and `test = "standard"`. `lavaan`'s `se` option can only be set to "`standard`" to assume multivariate normality or to "`robust.huber.white`" to relax that assumption.

## Value

The `twostage` object contains 3 fitted lavaan models (saturated, target/hypothesized, and baseline) as well as the names of auxiliary variables. None of the individual models provide the correct model results (except the point estimates in the target model are unbiased). Use the methods in `twostage` to extract corrected *SEs* and test statistics.

## Author(s)

Terrence D. Jorgensen (University of Amsterdam; <TJorgensen314@gmail.com>)

## References

- Savalei, V., & Bentler, P. M. (2009). A two-stage approach to missing data: Theory and application to auxiliary variables. *Structural Equation Modeling, 16*(3), 477–497. doi:10.1080/10705510903008238
- Savalei, V., & Falk, C. F. (2014). Robust two-stage approach outperforms robust full information maximum likelihood with incomplete nonnormal data. *Structural Equation Modeling, 21*(2), 280–302. doi:10.1080/10705511.2014.882692

## See Also

[twostage](#)

**Examples**

```

## impose missing data for example
HSMiss <- HolzingerSwineford1939[ , c(paste("x", 1:9, sep = ""),
                                     "ageyr", "agemo", "school")]

set.seed(12345)
HSMiss$x5 <- ifelse(HSMiss$x5 <= quantile(HSMiss$x5, .3), NA, HSMiss$x5)
age <- HSMiss$ageyr + HSMiss$agemo/12
HSMiss$x9 <- ifelse(age <= quantile(age, .3), NA, HSMiss$x9)

## specify CFA model from lavaan's ?cfa help page
HS.model <- '
  visual  =~ x1 + x2 + x3
  textual =~ x4 + x5 + x6
  speed   =~ x7 + x8 + x9
'

## use ageyr and agemo as auxiliary variables
out <- cfa.2stage(model = HS.model, data = HSMiss, aux = c("ageyr", "agemo"))

## two versions of a corrected chi-squared test results are shown
out
## see Savalei & Bentler (2009) and Savalei & Falk (2014) for details

## the summary additionally provides the parameter estimates with corrected
## standard errors, test statistics, and confidence intervals, along with
## any other options that can be passed to parameterEstimates()
summary(out, standardized = TRUE)

## use parameter labels to fit a more constrained model
modc <- '
  visual  =~ x1 + x2 + x3
  textual =~ x4 + x5 + x6
  speed   =~ x7 + a*x8 + a*x9
'
outc <- cfa.2stage(model = modc, data = HSMiss, aux = c("ageyr", "agemo"))

## use the anova() method to test this constraint
anova(out, outc)
## like for a single model, two corrected statistics are provided

```

## Description

This class contains the results of 2-Stage Maximum Likelihood (TSML) estimation for missing data. The `summary`, `anova`, `vcov` methods return corrected *SEs* and test statistics. Other methods are simply wrappers around the corresponding `lavaan` methods.

## Usage

```
## S4 method for signature 'twostage'
show(object)

## S4 method for signature 'twostage'
summary(object, ...)

## S4 method for signature 'twostage'
anova(object, h1 = NULL, baseline = FALSE)

## S4 method for signature 'twostage'
nobs(object, type = c("ntotal", "ngroups",
  "n.per.group", "norig", "patterns", "coverage"))

## S4 method for signature 'twostage'
coef(object, type = c("free", "user"))

## S4 method for signature 'twostage'
vcov(object, baseline = FALSE)

## S4 method for signature 'twostage'
fitted.values(object, model = c("target", "saturated",
  "baseline"), type = "moments", labels = TRUE)

## S4 method for signature 'twostage'
fitted(object, model = c("target", "saturated",
  "baseline"), type = "moments", labels = TRUE)

## S4 method for signature 'twostage'
residuals(object, type = c("raw", "cor", "normalized", "standardized"))

## S4 method for signature 'twostage'
resid(object, type = c("raw", "cor", "normalized", "standardized"))
```

## Arguments

<code>object</code>	An object of class <code>twostage</code> .
<code>...</code>	arguments passed to <code>parameterEstimates</code> .
<code>h1</code>	An object of class <code>twostage</code> in which <code>object</code> is nested, so that their difference in fit can be tested using <code>anova</code> (see <b>Value</b> section for details).
<code>baseline</code>	logical indicating whether to return results for the baseline model, rather than the default target (hypothesized) model.



type	The meaning of this argument varies depending on which method it is used for. Find detailed descriptions in the <b>Value</b> section under <code>coef</code> , <code>nobs</code> , and <code>residuals</code> .
model	character naming the slot for which to return the model-implied sample moments (see <code>fitted.values</code> description.)
labels	logical indicating whether the model-implied sample moments should have (row/column) labels.

**Value**

show	<code>signature(object = "twostage")</code> : The show function is used to display the results of the anova method, as well as the header of the (uncorrected) target model results.
summary	<code>signature(object = "twostage", ...)</code> : The summary function prints the same information from the show method, but also provides (and returns) the output of <code>parameterEstimates(object@target, ...)</code> with corrected <i>SEs</i> , test statistics, and confidence intervals. Additional arguments can be passed to <code>parameterEstimates</code> , including <code>fmi = TRUE</code> to provide an estimate of the fraction of missing information.
anova	<code>signature(object = "twostage", h1 = NULL, baseline = FALSE)</code> : The anova function returns the residual-based $\chi^2$ test statistic result, as well as the scaled $\chi^2$ test statistic result, for the model in the target slot, or for the model in the baseline slot if <code>baseline = TRUE</code> . The user can also provide a single additional twostage object to the <code>h1</code> argument, in which case anova returns residual-based and scaled $(\Delta)\chi^2$ test results, under the assumption that the models are nested. The models will be automatically sorted according to their degrees of freedom.
nobs	<code>signature(object = "twostage", type = c("ntotal", "ngroups", "n.per.group", "norig", "patterns"))</code> : The nobs function will return the total sample size used in the analysis by default. Also available are the number of groups or the sample size per group, the original sample size (if any rows were deleted because all variables were missing), the missing data patterns, and the matrix of coverage (diagonal is the proportion of sample observed on each variable, and off-diagonal is the proportion observed for both of each pair of variables).
coef	<code>signature(object = "twostage", type = c("free", "user"))</code> : This is simply a wrapper around the corresponding <code>lavaan</code> method, providing point estimates from the target slot.
vcov	<code>signature(object = "twostage", baseline = FALSE)</code> : Returns the asymptotic covariance matrix of the estimated parameters (corrected for additional uncertainty due to missing data) for the model in the target slot, or for the model in the baseline slot if <code>baseline = TRUE</code> .
fitted.values, fitted	<code>signature(object = "twostage", model = c("target", "saturated", "baseline"))</code> : This is simply a wrapper around the corresponding <code>lavaan</code> method, providing model-implied sample moments from the slot specified in the model argument.
residuals, resid	<code>signature(object = "twostage", type = c("raw", "cor", "normalized", "standardized"))</code> : This is simply a wrapper around the corresponding <code>lavaan</code> method, providing residuals of the specified type from the target slot.

**Slots**

saturated A fitted [lavaan](#) object containing the saturated model results

target A fitted [lavaan](#) object containing the target/hypothesized model results

baseline A fitted [lavaan](#) object containing the baseline/null model results

auxNames A character string (potentially of length == 0) of any auxiliary variable names, if used

**Objects from the Class**

Objects can be created via the [twostage](#) function.

**Author(s)**

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**See Also**

[twostage](#)

**Examples**

```
# See the example from the twostage function
```

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